



<u>Supplementary Figure 1</u> Comparing Shh with GBS-GFP at different developmental times reveals a non-linear relationship.

A To provide more accurate staging of neural tube development we exploited the linear relationship between somite number and neural tube size after E9.5 of development (along the ventral-dorsal axis). The mean is indicated by the red cross, the standard deviation by the error bar and the max and min by the blue circles. Each somite represents approximately 2 hours of developmental time, hence somite stage and embryonic day staging are related as labeled on the x-axis.

B A section from an embryo at the 30 somite stage immunostained for Shh (C) and Nkx2.2 (C'). The ventral limit of Nkx2.2 expression at this stage corresponds to the floor plate boundary as shown in Supplementary Figure 1B.

C An independent data set was derived for the GBS-GFP. The data were collected from 23 differently staged embryos and comprises 65 sections. The plot was derived in the same way as in Figure 2 (in this case there is no interpolation of the curves and error bars). The figure reveals the same temporal adaptation over time for each different position of the neural tube (data has been reproduced from ref. 1).

Α	dna	-tr	ptc	deg 👂	Ó	$\frac{d([pto])}{dt}$	=	$= + \left(\frac{te_{\mathcal{D}}te_{\mathcal{D}}(\mathbf{x}, \mathcal{O}\mathbb{E}_{\mathcal{D}}) + (\mathcal{K}, \mathcal{O}\mathbb{E}) + (\mathcal{K}, \mathcal{O}\mathbb{E}) + (K$
	ptc	_tl	Ptc_i	nactiv	/e deq ∳ Ø	<u>d(PteInsetive))</u> ds	=	= + (iL.Pto: [ptc]) - (dog.Pto - [Ptc hastre]) - (acLPto - [Ptc hastre])
	Ptc_inactive	act	Ptc +	Shh -	→ Ptc:Shh	<u>d ([Pto])</u> de	=	= + (acLPto : [PtcInactiod]) - (dcp.Pto : [Ptc]) - (k.SthPto : [Ptc] : [Sthi])
	dna	tr →	x	deg	φ	$\frac{d([a])}{d\delta}$	=	$= + \left(\frac{tr_{\mathcal{S}} \cdot (K \cdot \mathcal{O} l.x \ [\operatorname{Gals}] - c \cdot \mathcal{O} l.A + 1)^2 + (K \cdot \mathcal{O} l.x \ [\operatorname{Gals}] - c \cdot \mathcal{O} l.A + 1)^2 + (K \cdot \mathcal{O} l.x \ [\operatorname{Gals}] + (K \cdot \mathcal{O} l.x \ [$
	х	_tl →	х	deg	ø	4([X]) 48	=	$= + (d, X \cdot [q]) \\ - (dog X \cdot [X])$
	dna	_tr →	gliFL	deg	ø	<u>d(gaFL])</u> तः	=	$= + \left(\frac{i\tau_{c}gk^{2}L.K.Fo(gk; Poil (i + K.X.gk; P) \cdot c_{c}X^{2}}{(i + K.X.gk; Pi + K.Z.gk; Pi - c_{c}X^{2})}\right)$ $- \left(\frac{i\sigma_{c}ck^{2}L}{(i + L.X.gk; Pi + K.Z.gk; Pi - c_{c}X^{2})}\right)$
	gliFL	_tl	GliFL	conv conv deq	GliA GliR ø	$\frac{d([GEFL])}{d \delta}$	=	$= + (4.0 \text{ MFL}) \frac{(4.0 \text{ MFL})}{\text{ Km}_{-} \text{ For } \text{ point}} + \alpha \text{ const. } GhFL + \alpha $
	GliFL	conv	GliA	deg	ø	d([GhA])	=	$= + \left(\frac{\cos q_c J(M_1 + \log L_1 + M_m, P c_c)}{Km_m + T c_c + \log A}\right) \\ - \left(\frac{d_c J(M_1 + \log L_1 + \log A)}{Km_m + M_m + $
	GliFL	conv	GliR	deg	ø	d([GhR]) d b	=	$= + (cons.GkR \cdot [GhFL]) \\ - (dog.GkR \cdot [GhFR])$
	dna	-tr	gfp	deg	ø	<u>d([s\$9])</u> d0	=	= + (<u>KFoldp</u> [mai] (K.Oildp [mai] < <u>Oild+1</u>) ² + (<u>K.Oildp</u> [mai] ² + (<u>K.Oildp</u> [mai]) ² + (<u>K.Oildp</u> [mai

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	11	Discussion to a	Uniform Prior Range (Log10 scale, null = zero value)				
Parameter	Units	Description	Fuli Modei	Gil Degradation	Gil Regulation	Ptc Regulation	
deg_GliFL	h ⁻¹	degradation rate of ful-length Gli protein	-2:1.5	-2:1.5	-2:1.5	-2:1.5	
deg_GliR	h ⁻¹	degradation rate of repressor Gli protein	-2:1.5	-2:1.5	=degGliFL	=degGliFL	
deg_GliA	h'1	degradation rate of active Gli protein	-2:1.5	-2:1.5	=degGliFL	=degGliFL	
deg_Ptc	h''	degradation rate of Ptch1 protein	-2:1.5	-2:1.5	-2:1.5	-2:1.5	
deg_X	h ⁻¹	degradation rate of transcription factor 'X' protein	-2:1.5	null	-2:1.5	null	
deg_gfp	h ⁻¹	degradation rate of gfp RNA	-2:1.5	-2:1.5	-2:1.5	-2:1.5	
deg_gliFL	h'1	degradation rate of ful-length gli RNA	-2:1.5	-2:1.5	-2:1.5	-2:1.5	
deg_ptc	h'1	degradation rate of ptch1 RNA	-2:1.5	-2:1.5	-2:1.5	-2:1.5	
deg_x	h ⁻¹	degradation rate of transcription factor 'x' RNA	-2:1.5	null	-2:1.5	null	
k_ShhPtc	h'1	forward reaction rate of Shh Ptch1 binding	-2:2	-2:2	-2:2	-2:2	
K_Gli_gfp	uM1	binding affinity of Gli to gfp enhancer	-2:2	-2:2	-2:2	-2:2	
K_Gli_ptc	uM ¹	binding affinity of Gli to ptch1 enhancer	-2:2	null	null	-2:2	
K_Gli_x	цM ¹	binding affinity of Glitox enhancer	-2:2	-2:2	-2:2	-2:2	
K_X_gli	uM ¹	binding affinity of X to gli enhancer	-2:2	null	-2:2	null	
K_Pol_gfp	uM ¹	binding affinity of Polymerase to gfp promoter	-2:2	-2:2	-2:2	-2:2	
K_Pol_gli	цM ¹	binding affinity of Polymerase to gli promoter	-2:2	-2:2	-2:2	-2:2	
K_Pol_ptc	uМ1	binding affinity of Polymerase to ptch1 promoter	-2:2	-2:2	-2:2	-2:2	
K_Pol_x	uM1	binding affinity of Polymerase to x promoter	-2:2	null	-2:2	null	
Km_Ptc	µМ	Michaelis constant for Ptch1 inhibtion of Gli conversion	-2:2	-2:2	-2:2	-2:2	
Pol	щM	background concentration of Polymerase	0	0	0	0	
tl_GliFL	h ⁻¹	translation rate of Gli (into full length form)	-2:2	-2:2	-2:2	-2:2	
tl_Ptc	h'1	translation rate of Ptch1 (into inactive form)	-2:2	-2:2	-2:2	-2:2	
tl_X	h'1	translation rate of X	-2:2	null	-2:2	null	
tr_gfp	шMh ⁻¹	maximal transcription rate of gfp	-2:2	-2:2	-2:2	-2:2	
tr_gliFL	μMh-1	maximal transcription rate of gli	-2:2	-2:2	-2:2	-2 : 2	
tr_ptc	μMh-1	maximal transcription rate of ptch1	-2:2	-2:2	-2:2	-2:2	
tr_x	μMh-1	maximal transcription rate of x	-2:2	null	-2:2	null	
act_Ptc	h"*	activation rate of Ptch1 (or transport rate to membrane)	-2:2	-2:2	-2:2	-2:2	
conv_GliA	h'1	conversion rate of GliFL to GliA	-2:2	-2:2	-2:2	-2:2	
conv_GliR	h ⁻¹	conversion rate of GIIFL to GIIR	-2:2	-2:2	-2:2	-2 : 2	
c_GliA		transcriptional power of GliA	1	1	1	1	
(_X		transcriptional power of X	null	null	null	null	



<u>Supplementary Figure 2</u> Bayesian model analysis and posterior parameter distributions.

A The full set of reactions used in the model. The reaction schema for each protein or RNA are illustrated (left) alongside a set of ordinary differential equations (ODE)s (right) which detail the dynamics and regulatory functions associated with each reaction. Degradation rates are denoted *deg_*, transcription rates *tr_*, translation rates *tl_*, protein conversion rates *conv_*, protein activation rates *act_*, forward reaction rates *k_*, and binding affinities *K_*. Shh binds to Ptch1 (abbreviated to Ptc) by mass action binding. Ptch1 inhibits GliFL conversion to GliA via a first order Hill function. Thermodynamic regulation functions are used to determine gene expression in all cases – binding of all the transcription factors is assumed to occur at two independent sites. (Note that GliR is assumed to always act as a strong repressor with *c_GliR*=0 hence these terms are not shown in the regulatory function).

B The full set of parameters used in the model and their associated units. The prior ranges used in the Bayesian analysis are shown for each of the model mechanisms. Log uniform ranges were used in all cases - hence a value of 0 indicates 1, where a zero value was used in the model this is indicated by the *null* value in the table. When the Gli degradation mechanism was disabled the degradation rates of GliA and GliR were set to equal the degradation rate of GliFL.

C-AF The marginal posterior distribution is shown for each of the derived parameters in the different models, coloured according to the key in W. The box plots show the median (central bar), 25-75 percentiles (box) and ~1-99 percentiles (whiskers) of the marginal distribution for each of the parameters indicated. Note that in the cases where no boxplot is shown this indicates that the parameter did not feature in that particular mechanism (i.e. was fixed at zero). The parameters are described in full in Supplementary Figure 2B where the prior ranges and units are specified.



Supplementary Figure 3 Gli2 transcriptional regulation in the neural tube

This figure is reproduced from ref. 2. G-J In situ staining of Gli2 transcripts reveals that Gli2 transcriptional down regulation between E9.5 and E10.5 is restricted to ventral regions of the neural tube.

REFERENCES

- 1. Balaskas, N. *et al.* Gene regulatory logic for reading the Sonic Hedgehog signaling gradient in the vertebrate neural tube. *Cell* **148**, 273–284 (2012).
- 2. Ribes, V. *et al.* Distinct Sonic Hedgehog signaling dynamics specify floor plate and ventral neuronal progenitors in the vertebrate neural tube. *Genes Dev.* **24**, 1186–1200 (2010).