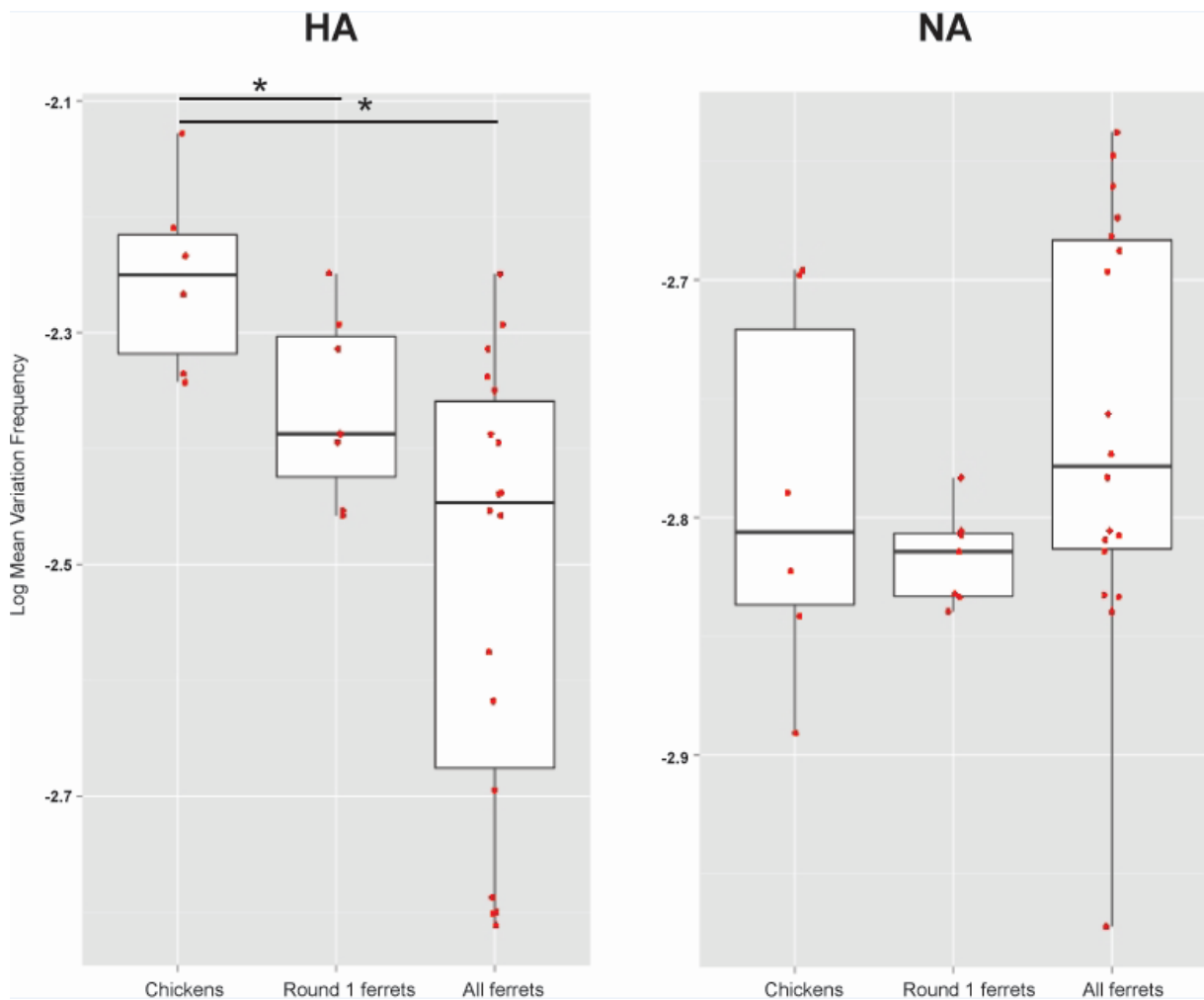
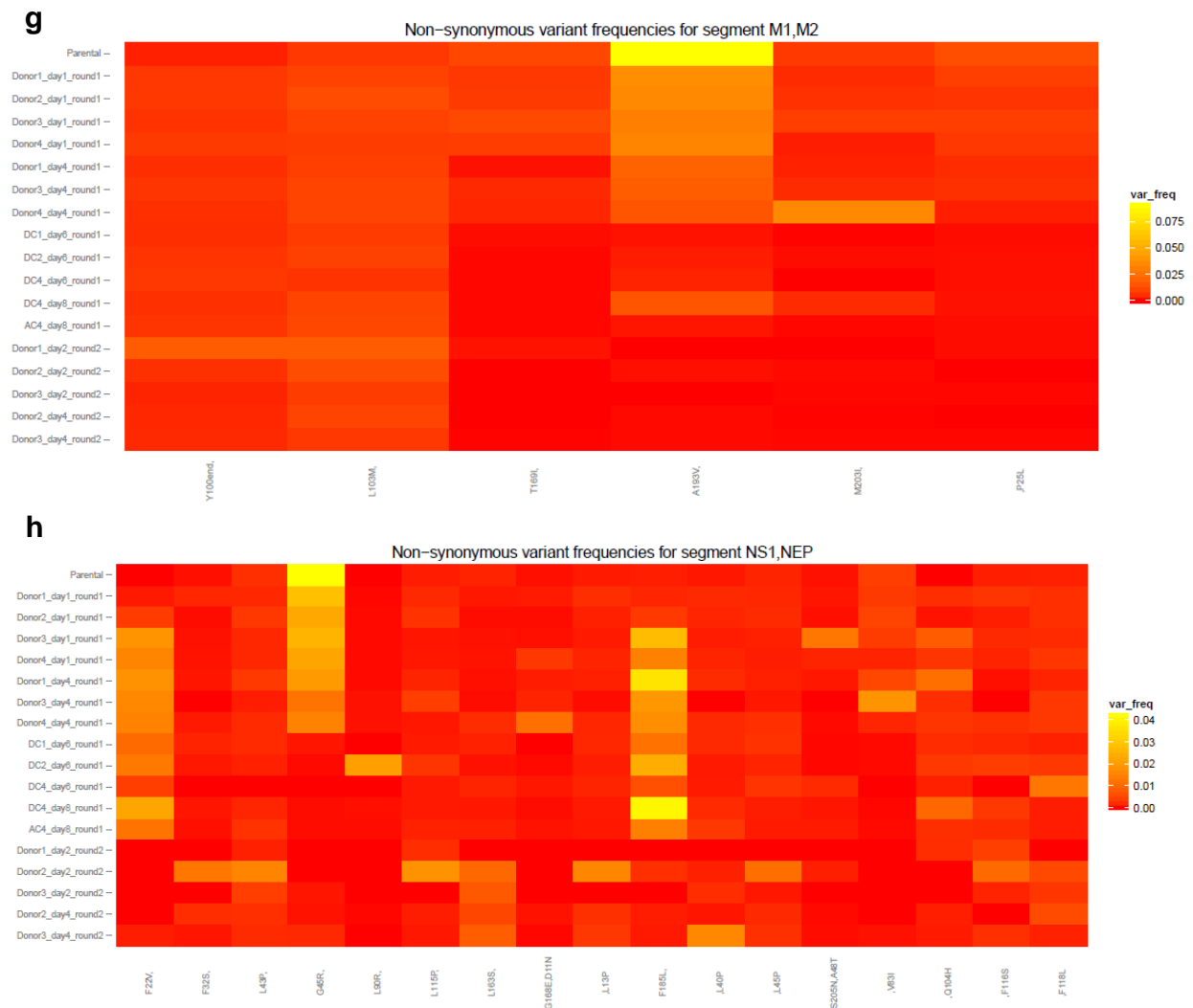


Supplementary Figure 1 | Weight and body temperature of ferrets inoculated with A/Anhui/1/2013 (H7N9) influenza virus. (a) Body temperature and (b) weight change of ferrets after intranasal inoculation with 10^6 TCID₅₀ of A/Anhui/1/2013 (H7N9) influenza virus, direct contact, and aerosol contact ($n = 4$ per group). Body temperature is shown for only 3 direct contact ferrets, as one temperature probe was faulty.

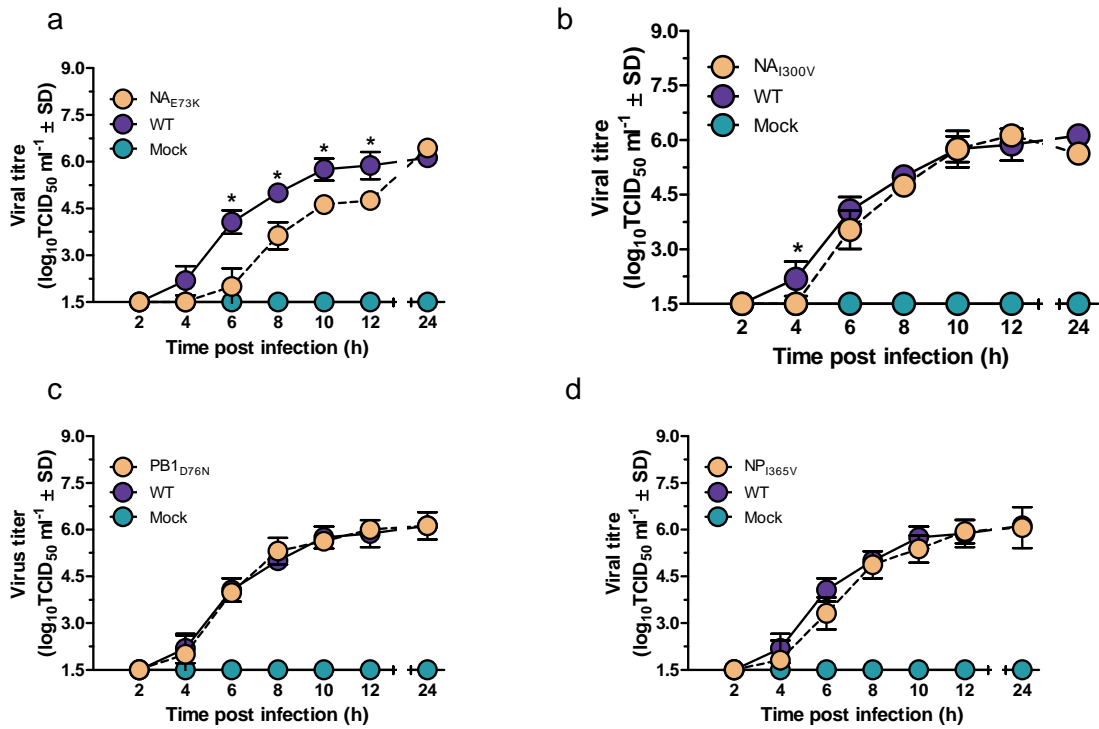


Supplementary Figure 2 | Mean variation frequencies of HA and NA genes after virus

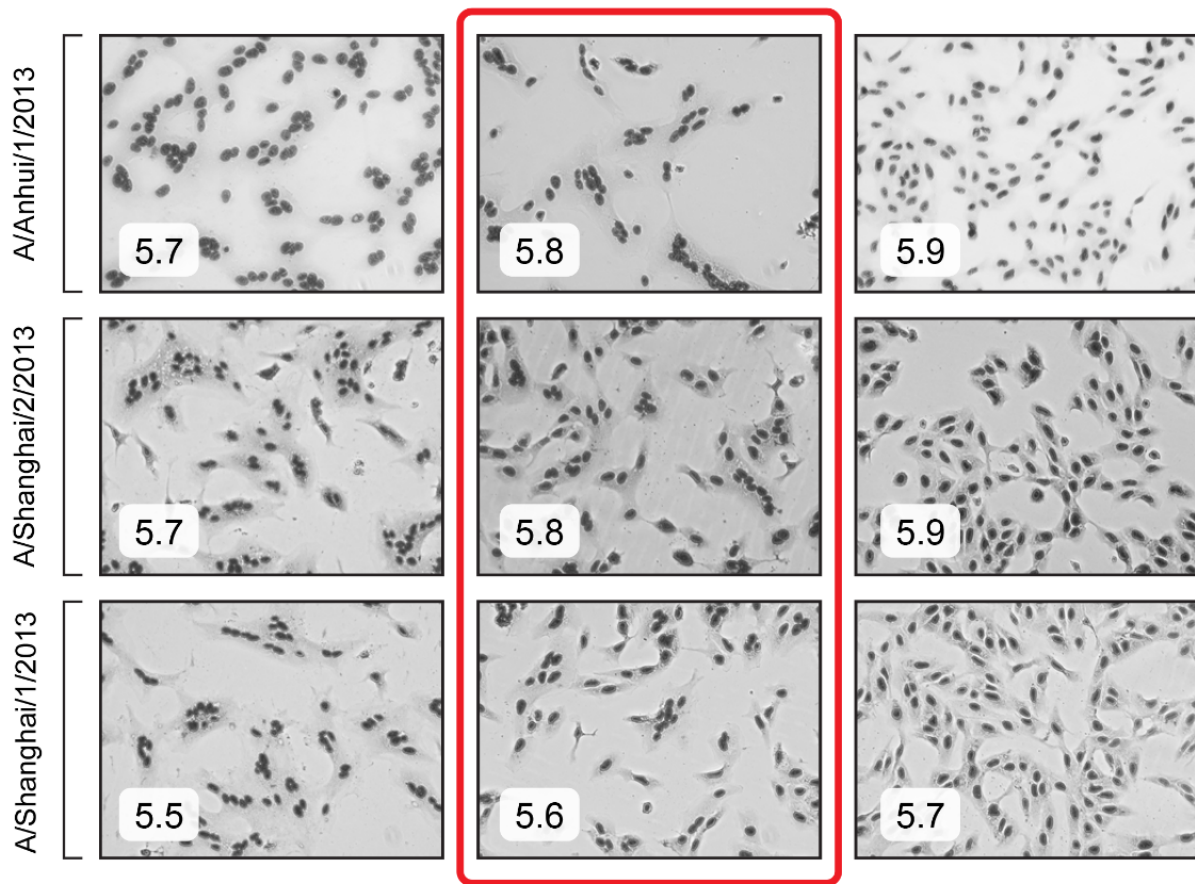
replication in chickens and ferrets. “Round 1 ferrets” were inoculated with parental A/Anhui/1/13 (H7N9) influenza virus; “all ferrets” include donor and contact ferrets in rounds 1 and 2. Statistical comparisons were performed by using two-tailed Mann-Whitney U test, $*P < 0.05$.



Supplementary Figure 3 | Heat map of variations in the whole genome of A/Anhui/1/2013 (H7N9) influenza virus in ferrets. Non-synonymous variant sequences for segments PB2 (a); PB1, PB1-F2 (b); PA, PA-X (c); HA (d); NP (e); NA (f); M1, M2 (g); and NS1, NEP (h) are shown. Parental virus is compared with viruses obtained in experimental rounds 1 and 2. Rows represent viruses from donor, direct contact (DC) and aerosol contact (AC) ferrets. Columns represent amino acid positions (counted from the start codon). Mutations in PA, M1, and NS1 genes are shown in black while those in PA-X, M2, and NS2 are in purple. Mutations with frequency > 0.01 are shown.



Supplementary Figure 4 | Effect of identified amino acid substitutions in the NA, PB1, and NP on replication kinetics of A/Anhui/1/2013 (H7N9) influenza viruses *in vitro*. Growth curves of the wild-type and the NA_{E73K} (**a**), NA_{I300V} (**b**), PB1_{D76N} (**c**), and NP_{I365V} (**d**) rg-A/Anhui/1/2013 (H7N9) influenza viruses using MOI 1 TCID₅₀ per cell in MDCK cells at 37°C. Values are mean and SD from two independent experiments (n= 4). *<0.05 compared to WT virus by 2-way ANOVA.



Supplementary Figure 5 | Activation pH of the HA proteins of H7N9 influenza viruses.

Vero cells were inoculated with the indicated viruses at an MOI of 5 TCID₅₀/cell. Cells were then treated with pH buffers at the indicated pH values, and syncytium formation was monitored. The highest pH at which syncytia were observed (boxed in red) was recorded as the pH of HA activation. Data is representative of two independent experiments.

1 **Supplementary Table 1 | Homology sequence analysis of A/Anhui/1/2013 (H7N9) influenza virus in comparison with H7N9**
 2 **influenza viruses from birds and environment isolated during the first wave of the outbreak in China, 2013.**
 3

H7N9 virus (isolated form birds and environment)	Date of isolation	Segment ^a	Accession No. ^b	% Nucleotide sequence homology with A/Anhui/1/2013 ^c	% Amino acid sequence homology with A/Anhui/1/2013
A/Chicken/Shanghai/S1053/2013	2013-04-03	PB2	EPI440682	99.8	99.8
		PB1	EPI440683	100.0	100.0
		PA	EPI440681	99.8	99.6
		HA	EPI440685	99.3	98.7
		NP	EPI440678	100.0	100.0
		NA	EPI440684	99.8	100.0
		MP	EPI440680	99.9	100.0
		NS	EPI440679	99.9	100.0
A/Chicken/Zhejiang/SD033/2013	2013-04-11	PB2	EPI457746	99.9	99.7
		PB1	EPI457747	100.0	100.0
		PA	EPI457745	100.0	100.0
		HA	EPI457749	99.9	100.0
		NP	EPI457742	100.0	100.0
		NA	EPI457748	99.9	99.8
		MP	EPI457744	98.3	99.3
		NS	EPI457743	99.9	100.
A/Environment/Shanghai/S1088/2013	2013-04-03	PB2	EPI440690	99.9	99.8
		PB1	EPI440691	100.0	100.0
		PA	EPI440689	99.4	98.7
		HA	EPI440693	99.4	99.1
		NP	EPI440686	100.0	100.
		NA	EPI440692	99.9	100.0
		MP	EPI440688	99.7	99.8
		NS	EPI440687	99.9	100.0
A/Pigeon/Shanghai/S1069/2013	2013-04-02	PB2	EPI440698	99.8	99.7
		PB1	EPI440699	100.0	100
		PA	EPI440697	99.8	99.6
		HA	EPI440701	99.8	100.0

A/Pigeon/Shanghai/S1423/2013	2013-04-03	NP	EPI440694	98.8	99.7
		NA	EPI440700	99.8	99.8
		MP	EPI440696	99.8	100
		NS	EPI440695	96.9	99.4
		PB2	EPI457626	99.9	99.8
		PB1	EPI457627	100.0	100.0
		PA	EPI457625	97.8	98.7
		HA	EPI457629	99.8	99.7
		NP	EPI457622	100.0	100.0
		NA	EPI457628	99.8	100.0
MP	EPI457624	99.9	99.8		
NS	EPI457623	99.9	100.0		

4

5 ^a PB2: RNA polymerase basic subunit 2; PB1: RNA polymerase basic subunit 1; PA: RNA polymerase acidic subunit; HA:

6 haemagglutinin; NP: nucleoprotein; NA: neuraminidase; MP: matrix gene; NS: non-structural gene.

7 ^b Sequence data used in this table were downloaded from GISAID, Global Initiative on Sharing Avian Influenza Data

8 (<http://platform.gisaid.org>)

9 ^c Homology analysis was performed using BioEdit software (<http://www.mbio.ncsu.edu/bioedit/bioedit.html>) Sequence Identity

10 Matrix tool.

11

12 **Supplementary Table 2 | Genetic diversity of A/Anhui/1/2013 (H7N9) influenza viruses in chickens.**

13

Animal number	Dpi	Gene segment							
		PB2	PB1	PA	HA	NP	NA	M	NS
		— ^b	—	—	N141D (0.4211), A143T (0.1250), N167D (0.5824), L235Q (0.2182), Y501N (0.1454), Y501C (0.1457)	A373T (0.1277)	T10I (0.2283)	—	—
(Parental virus)	N/A								
1	3	N100D (0.4203), K627R (0.1687), L665P (0.125530625)	N346S (0.125), V421A (0.3125), V609A (0.1355)	T210N (0.4737), A255G (0.3125), E377G (0.100)	N141D (0.6923), N167D (0.7945), Q444L (0.1477), Y501N (0.1874), Y501C (0.1884)	R26G (0.4809), R98K (0.1094), A284T (0.1975), A428T (0.1407)	T50K (0.1250), K139N (0.1677)	— ^a	—
	5	R70G (0.1102)	—	P28L (0.1070), S184N (0.7272), Q654stop (0.1698), R663G (0.1709)	N141D (0.4201), A143T (0.1203), G151E (0.1761), N167D (0.4350), Y501N (0.1935), Y501C (0.1935)	R99K (0.2261), A373T (0.1207), R446K (0.1058)	—	—	—

2	3	K48R (0.1020), W98stop (0.72), D275E (0.125)	—	E623K (0.3325)	N141D (0.4225), A143T (0.1509), N167D (0.5347), N208D (0.1339), A310T (0.1724), G475D (0.36951), Y501N (0.1598), Y501C (0.1598)	—	—	—	—
	5	—	—	—	N141D (0.6972), N167D (0.5634), Y501Y (0.1596), Y501C (0.1586)	—	T10I (0.2241)	M165I (0.1438), M192I (0.1846), A193V (0.1969)	—
3	3	M90T (0.2526)	K54E (0.6257), M523I (0.6470), Q569K (0.1161)	Y110H (0.1724)	N141D (0.7217), N167D (0.6614), L235Q (0.8971), Y501N (0.2537), Y501C (0.2529), G520V (0.1451)	A284T (0.1267),	—	—	—

5	—	F94L (0.1726), L108F (0.1274)	—	N141D (0.2786), N167D (0.3247), L235Q (0.1111), K414R (0.1111), Y501N (0.46939), Y501C (0.4694)	A373T (0.1265)	S119P (0.1097)	—	—
---	---	-------------------------------------	---	--	-------------------	-------------------	---	---

14 Three 6-week-old, specific pathogen-free chickens were inoculated with 10^6 EID₅₀ of virus in 0.5 ml PBS via natural routes (nares,
15 trachea, eyes). RNA was extracted from oropharyngeal swabs on the indicated days post-inoculation (dpi) and influenza virus genes
16 were amplified and sequenced. Shown are all amino acid variations that differed from the consensus parental sequence and were
17 present at a frequency ≥ 0.1 .
18 —, no nonsynonymous mutations were observed at a frequency ≥ 0.1 .

19 **Supplementary Table 3 | Ferret serum hemagglutination inhibition titers.**

A/Anhui/1/2013 (H7N9) virus	Serum HI titer (HAU 50 μl^{-1})											
	After inoculation				After direct contact				After aerosol contact			
	D1	D2	D3	D4	DC1	DC2	DC3	DC4	AC1	AC2	AC3	AC4
Parental	160	80	40	160	160	320	160	40	<	<	10	40
AC	80	160	<	ND	ND	ND	ND	ND	<	<	<	ND

20 Titers are reciprocal values. HAU, hemagglutinating units. D, donor; DC, direct contact; AC aerosol contact; <, below the lower limit of detection
 21 (10 HAU/50 μl); ND, not determined.

22

23 **Supplementary Table 4 | Variation frequency of A/Anhui/1/2013 (H7N9) influenza virus after inoculation of ferrets and**
 24 **chickens.**
 25

Animal model	Experiment round	Group	Animal no.	DPI	Total variation frequency (log)	HA variation frequency (log)	NA variation frequency (log)		
N/A	N/A	Parental virus	N/A	N/A	-2.905440162	-2.338603516	-2.972306897		
Ferrets	1	Donor	D1	1	-2.806927772	-2.457879004	-2.833868038		
				4	-2.742696441	-2.453907496	-2.805941954		
			D2	1	-2.816327184	-2.387386966	-2.783279964		
				4	-2.769769832	-2.394822726	-2.81441596		
			D3	1	-2.769769832	-2.394822726	-2.81441596		
				4	-2.776612542	-2.292597553	-2.807442559		
			D4	1	-2.802506048	-2.24905896	-2.832470569		
				4	-2.774978978	-2.313737707	-2.839594171		
			Direct Contact	6	DC1	6	-2.772057833	-2.438035302	-2.756583119
						6	-2.748001965	-2.439238638	-2.773588496
						6	-2.841064242	-2.811259698	-2.637607146
			Aerosol Contact	8	DC4	8	-2.779612706	-2.787387123	-2.696347657
						8	-2.837547661	-2.349990787	-2.809618722
						8	-2.837547661	-2.349990787	-2.809618722
			Donor	2	D1-2	2	-2.832660393	-2.617216784	-2.660533406
2	-2.846847365	-2.575942667				-2.687619251			
4	-2.847458857	-2.694908903				-2.673835236			
2	-2.851934700	-2.801427075				-2.681704602			

				4	-2.841801531	-2.799644372	-2.647563697
Chicken	N/A	N/A	1	3	-2.634408252	-2.233465376	-2.841465687
s				5	-2.675060293	-2.127884799	-2.891189157
			3	3	-2.67762697	-2.209500848	-2.697862833
				5	-2.736429542	-2.342424772	-2.822763484
			4	3	-2.659243758	-2.335177799	-2.695671659
				5	-2.764034125	-2.266838626	-2.789758423

26 Three- to 4-month-old male ferrets were inoculated intranasally with 10^6 TCID₅₀ of virus. Four 6-week-old, specific-pathogen-free
27 chickens were inoculated with 10^6 50% egg infectious doses (EID₅₀). Viral genes were amplified and sequenced from RNA extracted
28 from nasal washes (ferrets) or oropharyngeal swabs (chickens) on the indicated day pi. Mean variation frequency for each sample was
29 calculated using positions that are variable in at least one of the examined samples. If a site has a minor variant frequency above zero
30 in any of the samples this was included in the nominator and the denominator of the proportion was the total number of variable sites
31 that was variable in all samples. Variation frequency was expressed as the negative log.
32 DPI, day post-inoculation when mutation was detected; N/A, not applicable.
33

		DC4	6	—	D76N (0.5316)	—	N141D (0.1818), N167D (0.4178), A228E (0.1173), L235Q (0.1737)	L193M (0.1111), M352I (0.1887)	R103K (0.9576) T10I (0.2404), I26M (0.1266), N340I (0.1511)	—	—
			8	—	D76N (0.2793)	—	N141D (0.2464), N167D (0.5005), A228E (0.1004), L235Q (0.1174)	—	—	—	—
	Aerosol Contact	AC4	8	—	D76N (0.4320)	—	—	I365V (0.6387)	E73K (0.4058), I300V (0.4103)	—	—
2	Donor	D1-2	2	—	D76N (0.3177)	—	K414E (0.1563), Y501N (0.3182), Y501C (0.3182)	I365V (0.6163)	E73K (0.3941), I300V (0.3451)	—	—
	Donor	D2-2	2	—	D76N (0.3887)	—	Y501N (0.3137), Y501C (0.3137)	I365V (0.6236)	E73K (0.4348), I300V (0.3739)	—	—
			4	—	D76N (0.2604)	—	—	I365V (0.4976)	E73K (0.2933), I300V (0.2848)	—	—
		D3-2	2	—	D76N (0.2993)	—	K414E (0.1455), Y501N (0.1783), Y501C (0.1719)	I365V (0.6174)	E73K (0.3309), I300V (0.4053)	—	—
			4	—	D76N (0.2016)	—	Y501N (0.1449), Y501C (0.1439)	I365V (0.5547)	E73K (0.4471), I300V (0.4426)	—	—

37 Three to 4-month-old male ferrets were inoculated intranasally with 10^6 TCID₅₀ of virus. Viral genes were amplified and sequenced
38 from RNA extracted from nasal washes on the indicated day pi. Shown are all amino acid variations that differed from the consensus
39 parental sequence and were present at a frequency of ≥ 0.1 . Numbers in parentheses indicate the frequency at which each mutation
40 occurred under each condition.
41 DPI, day post inoculation when mutation was detected; NA, not applicable; –, no nonsynonymous mutations were detected at a
42 frequency ≥ 0.1 .
43
44

45 **Supplementary Table 6 | List of primers used in the study.**

Primer	Description	Sequence (5' → 3')
N9 E73K F	E73K site-directed mutagenesis	CAAACATCACCAACATCCAAATGAAAGAGAGAACAAG
N9 E73K R	E73K site-directed mutagenesis	GAAATTCCTGCTTGTTCTCTCTTTCATTTGGATGTTG
N9 I300V F	I300V site-directed mutagenesis	GCAGGGCTCAAATAGACCAGTGGTTCAGATAGACCC
N9 I300V R	I300V site-directed mutagenesis	GTGTCATTGCTACTGGGTCTATCTGAACCACTGGTCTATTTG
N9 ClaI/- F	ClaI site deletion	GTGGACCAGCAATAGTATAGTTTCGATGTGTTCCAGTACAG
N9 ClaI/- R	ClaI site deletion	CCAGGAATTCTGTACTGGAACACATCGAAACTATACTATTGC
PB2 K627E F	K627E site-directed mutagenesis	CCATTTGCAGCAGCCCCGCCGAGCAGAGTAGG
PB2 K627E R	K627E site-directed mutagenesis	CGGCGGGGCTGCTGCAAATGGTAATAGCTT
PB1 D76N F	D76N site-directed mutagenesis	ATTGATGGACCATTACCTGAGAACAACGAGCCGA
PB1 D76N R	D76N site-directed mutagenesis	CTCAGGTAATGGTCCATCAATTGGATTGAGT
I365V F	I365V site-directed mutagenesis	GCTATCCACTAGAGGGGTTCAAGTTGCTTCAAAT
I365V R	I365V site-directed mutagenesis	TTGAACCCCTCTAGTGGATAGCTGTCCTCTT
N9 ClaI F	N-terminal ClaI restriction site insertion for pCAAGS cloning	TTACCATCGATATGAATCCAAATCAGAAGATTC
N9 NheI HA-tag R	C-terminal NheI restriction site and HA-tag insertion	TTTCGCGCTAGCTTAAGCGTAATCTGGAACATCGTATGGGTAGAGGAAGTACTCTATTTTAGCC
N9 NheI R	C-terminal NheI restriction site insertion	TTTCGCGCTAGCTTAGAGGAAGTACTCTATTTTAGCC

47 **Supplementary Table 7 | List of sialic acid-containing glycans used in glycan arrays**

48

Chart number	Name	NeuAc α -	Type	Sulfated
1	Gal β (1-4)-GlcNAc β -ethyl-NH ₂	—	Type 2	
2	Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	—	N	
3	NeuAc α (2-3)-Gal β (1-4)-6-O-sulfo-GlcNAc β -propyl-NH ₂	3	Type 2	6S-GlcNAc
4	NeuAc α (2-3)-Gal β (1-4)-[Fuca(1-3)]-6-O-sulfo-GlcNAc β -propyl-NH ₂	3	Type 2	6S-GlcNAc
5	NeuAc α (2-3)-6-O-sulfo-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	3	Type 2	6S-Gal
6	NeuAc α (2-3)-6-O-sulfo-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β -propyl-NH ₂	3	Type 2	6S-Gal
7	NeuAc α (2-3)-Gal β (1-3)-6-O-sulfo-GlcNAc β -propyl-NH ₂	3	Type 1	6S-GlcNAc
8	NeuAc α (2-3)-Gal β (1-4)-Glc β -ethyl-NH ₂	3	Lacto	
9	NeuAc α (2-3)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	3	Type 2	
10	NeuAc α (2-3)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	3	Type 2	
11	NeuAc α (2-3)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	3	Type 2	
12	NeuAc α (2-3)-GalNAc β (1-4)-GlcNAc β -ethyl-NH ₂	3	LacDiNAc	
13	NeuAc α (2-3)-Gal β (1-3)-GlcNAc β -ethyl-NH ₂	3	Type 1	
14	NeuAc α (2-3)-Gal β (1-3)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	3	Type 2	

15	NeuAca(2-3)-Galβ(1-3)-GlcNAcβ(1-3)-Galβ(1-3)-GlcNAcβ-ethyl-NH ₂	3	Type 1
16	NeuAca(2-3)-Galβ(1-3)-GalNAcβ(1-3)-Gala(1-4)-Galβ(1-4)-Glcβ-ethyl-NH ₂	3	Lacto
17	NeuAca(2-3)-Galβ(1-3)-GalNAcα-Thr-NH ₂	3	Core 1
18	NeuAca(2-3)-Galβ(1-3)-[GlcNAcβ(1-6)]-GalNAcα-Thr-NH ₂	3	Core 2
19	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-6)-[Galβ(1-3)]-GalNAcα-Thr-NH ₂	3	Core 2
20	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-6)-[Galβ(1-3)]-GalNAcα-Thr-NH ₂	3	Core 2
21	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-GalNAcα-Thr-NH ₂	3	Core 3
22	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-3)-GalNAcα-Thr-NH ₂	3	Core 3
23	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-[NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-6)]-GalNAcα-Thr-NH ₂	3	Core 4
24	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-3)-[NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-6)]-GalNAcα-Thr-NH ₂	3	Core 4
25	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-2)-Manα(1-3)-[NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-2)-Manα(1-6)]-Manβ(1-4)-GlcNAcβ(1-4)-GlcNAcβ-Asn-NH ₂	3	N
26	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-2)-Manα(1-3)-[NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-2)-Manα(1-6)]-Manβ(1-4)-GlcNAcβ(1-4)-GlcNAcβ-Asn-NH ₂	3	N
27	NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-2)-Manα(1-3)-[NeuAca(2-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-3)-Galβ(1-4)-GlcNAcβ(1-2)-Manα(1-6)]-Manβ(1-4)-GlcNAcβ(1-4)-GlcNAcβ-Asn-NH ₂	3	N

28	NeuAca(2-3)-[GalNAc β (1-4)]-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	3	Type 2	
29	NeuAca(2-3)-[GalNAc β (1-4)]-Gal β (1-4)-Glc β -ethyl-NH ₂	3	Ganglio	
30	Gal β (1-3)-GalNAc β (1-4)-[NeuAca(2-3)]-Gal β (1-4)-Glc β -ethyl-NH ₂	3	Ganglio	
31	NeuAca(2-3)-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β -propyl-NH ₂	3	Type 2	
32	NeuAca(2-3)-Gal β (1-3)-[Fuca(1-4)]-GlcNAc β (1-3)-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β -ethyl-NH ₂	3	Type 2	
33	NeuAca(2-3)-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β (1-3)-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β -ethyl-NH ₂	3	Type 2	
34	NeuAca(2-3)-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β (1-3)-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β (1-3)-Gal β (1-4)-[Fuca(1-3)]-GlcNAc β -ethyl-NH ₂	3	Type 2	
35	NeuGca(2-3)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	3-Gc	Type 2	
36	NeuAca(2-6)-Gal β (1-4)-6-O-sulfo-GlcNAc β -propyl-NH ₂	6	Type 2	6S-GlcNAc
37	NeuAca(2-6)-Gal β (1-4)-Glc β -ethyl-NH ₂	6	Lacto	
38	NeuAca(2-6)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	6	Type 2	
39	NeuAca(2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	6	Type 2	
40	NeuAca(2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	6	Type 2	
41	NeuAca(2-6)-Gal β (1-4)-GlcNAc β (1-3)-[NeuAca(2-6)]-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	6	Type 2	
42	NeuAca(2-6)-GalNAc β (1-4)-GlcNAc β -ethyl-NH ₂	6	LacDiNAc	

43	NeuAc α (2-6)-[Gal β (1-3)]-GalNAc α -Thr-NH ₂	6	Core 1
44	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-6)-[Gal β (1-3)]-GalNAc α -Thr-NH ₂	6	Core 2
45	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-6)-[Gal β (1-3)]-GalNAc α -Thr-NH ₂	6	Core 2
46	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-GalNAc α -Thr-NH ₂	6	Core 3
47	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-3)-GalNAc α -Thr-NH ₂	6	Core 3
48	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-6)]-GalNAc α -Thr-NH ₂	6	Core 4
49	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-6)]-GalNAc α -Thr-NH ₂	6	Core 4
50	Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	6	N
51	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	6	N
52	GlcNAc β (1-2)-Man α (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	6	N
53	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	6	N
54	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	6	N

55	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-3)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	6	N
56	NeuGc α (2-6)-Gal β (1-4)-GlcNAc β -ethyl-NH ₂	6-Gc	Type 2
57	NeuAc α (2-3)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	3/6	N
58	NeuAc α (2-6)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-3)-[NeuAc α (2-3)-Gal β (1-4)-GlcNAc β (1-2)-Man α (1-6)]-Man β (1-4)-GlcNAc β (1-4)-GlcNAc β -Asn-NH ₂	3/6	N
