

Figure S3. Analysis of the clades representing the cathepsin L cysteine proteases. (a) Protein alignment using representative sequences for each clade of cathepsin L protease. The genomic organisation of these genes is conserved across all the cathepsin L genes identified within the *F. hepatica* genome, regardless of which clade the proteases belong to. Intron-exon borders, resulting in four exons are indicated by the black arrows. The signal peptide and pro-segment domain are highlighted in green and blue, respectively, with the short blue arrow indicating the position of the pro-protein cleavage site. The residues in bold and shaded in grev represent the active site residues. that comprise the S1 binding subsite. The residues highlighted in red represent those residues that comprise the S2 binding subsite. The particular S2 residues that confer substrate specificity are highlighted by the red *. (b) Comparison of the residues from the S1 and S2 binding subsites across the cathepsin L clades. Variability across the sequences represented by the different genes within each clade is shown. (c) Phylogenetic analysis of the F. hepatica cathepsin L gene family, based on the genes identified within the *F. hepatica* genome. A maximum likelihood tree was constructed with the nucleotide sequence corresponding to the prosegment of the protein including the catalytic domain. For three genes the sequences are represented by two gene models (*). The tree is drawn to scale and branch lengths are measured in number of substitutions per site using MEGA v 5.05 (Tamura et al., 2011). Bootstrap values >70% from 100 iterations are shown. The tree is rooted using cathepsin L sequences from the closely related trematode species, Schistosoma mansoni and Schistosoma japonicum (SjCL: U38476; SmCL1; Z32529; SjCL: U38475).