

EXTENDED EXPERIMENTAL PROCEDURES

DNA sequences: Oligonucleotides (synthesized by IDT) for PCR amplification of templates. The name of the oligonucleotide corresponds to the labeling strand and position. Biosg and ibio refer to biotin, idSp refers to abasic site, iAmMC6T refers to amino-modified C6 dT linker for labeling with Cy3 or Cy5, 5Cy5 and 5Cy3 refer to end-labeled positions generated through phosphoramidite chemistry. Other abbreviations include: LH (left half), RH (right half), MF (middle fragment) RRH (right right handle), LL(left left), RTA (right TA (dinucleotide), A (for immobilization scheme A), B (for immobilization scheme B). The primer sequences are color-coded according to the sections of the 601 template (see below): extra-chromosomal handles (black); left outer quarter (magenta); left inner quarter (orange); right inner quarter (green); right outer quarter (blue).

- (1) I77 5'- /5Biosg/TATA CGCGG CCGCC CTGGAGAATC CCGGTGCCGA
GGCCGCTCAA TTGGTCGTAG ACAGCTCTAG CACCGCTTAA
ACGCACGTAC GCGCTG/iAmMC6T/CCC
- (2) J-12 5'- GG GCGGCGACCT /idSp/GGTCGCTG/iAmMC6T/T CAATACATGC
ACAGGAT GTATATATC
- (3) I68 5'-/5Biosg/TATA CGCGG CCGCC CTGGAGAATC CCGGTGCCGA
GGCCGCTCAA TTGGTCGTAG ACAGCTCTAG CACCGCTTAA
ACGCACG/iAmMC6T/AC G
- (4) J-1 5'- GG GCGGCGACCT /idSp/GGTCGCTGTT CAATACATG/iAmMC6T/
ACAGGAT GTATATA
- (5) J7 5'- GG GCGGCGACCT /idSp/GGTCGCTGTT CAATACATGC
ACAGGA/iAmMC6T/ GTATATA
- (6) I57 5'-/5Biosg/TATA CGCGG CCGCC CTGGAGAATC CCGGTGCCGA
GGCCGCTCAA TTGGTCGTAG ACAGCTCTAG
CACCGC/iAmMC6T/TAA
- (7) J15 5'- GG GCGGCGACCT /idSp/GGTCGCTGTT CAATACATGC ACAGGAT
GTATATA/iAmMC6T/CT G
- (8) I46 5'-/5Biosg/TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA
GGCCGCTCAA TTGGTCGTAG ACAGC/iAmMC6T/CTA
- (9) J24 5'- GG GCGGCGACCT /idSp/GGTCGCTGTT CAATACATGC ACAGGAT
GTATATATCT GACACG/iAmMC6T/GCC TGGA
- (10) I38 5'-/5Biosg/TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA
GGCCGCTCAA TTGGTCG/iAmMC6T/AG A
- (11) J28 5'-GG GCGGCGACCT /idSp/GGTCGCTGTT CAATACATGC ACAGGAT
GTATATATCT GACACGTGCC /iAmMC6T/GGA
- (12) I27 5' GG GCGGCGACCT /idSp/GGA CCCTATA CGCGG CCGCC
CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCG/iAmMC6T/AG A
- (13) J45 5'- /5Biosg/TGTT CAATACATGC ACAGGAT GTATATATCT
GACACGTGCC TGGAGACTAG GGAGTAA/iAmMC6T/CC C
- (14) I15 5'- GG GCGGCGACCT /idSp/ GGA CCC TATA CGCGG CCGCC
CTGGAGAATC CCGG/iAmMC6T/GCC
- (15) J57 5'- /5Biosg/TGTT CAATACATGC ACAGGAT GTATATATCT
GACACGTGCC TGGAGACTAG GGAGTAATCC
CCTTGCGGG/iAmMC6T/ TAA
- (16) I9-A 5'-/5Biosg/TATA CGCGG CCGCC CTGGAGAA/iAmMC6T/C CC
- (17) J58-A 5'- GG GCGGCGACCT /idSp/TGTT CAATACATGC ACAGGAT
GTATATATCT GACACGTGCC TGGAGACTAG GGAGTAATCC
CCTTGCGGGT /iAmMC6T/AAA

- (18) I9 5'- GG GCGGCGACCT /idSp/ TATA CGCGGCCGCC
CTGGAGAA/iAmMC6T/C CC
- (19) J58 5'- /5Biosg/TGTT CAATACATGC ACAGGAT GTATATATCT
GACACGTGCC TGGAGACTAG GGAGTAATCC CCTTGGCGGT
/iAmMC6T/AAA
- (20) I-1 5'-/5Biosg/TATA CGCGG CCGC/iAmMC6T/ CTGGAGAA/iAmMC6T/C
CCGGT
- (21) J79 5'- GG GCGGCGACCT /idSp/GGTCGCTGTT CAATACATGC ACAGGAT
GTATATATCT GACACGTGCC TGGAGACTAG GGAGTAATCC
CCTTGGCGGT TAAAACGCGG GGGACAGCGC G/iAmMC6T/AC G
- (22) I-12 5'-/5Biosg/ GGACCCTA/iAmMC6T/A CGCGGCCGCC
CTGGAGAA/iAmMC6T/C CCGGT
- (23) I58-MF 5'- /5Biosg/TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA
GGCCGCTCAA TTGGTCGGGA GTAATCCCCT
TGGCGGT/iAmMC6T/AA A
- (24) J58-MF 5'- /5Biosg/TGTT CAATACATGC ACAGGAT GTATATATCT
GACACGTGCC TGGAGACTAG TAGACAGCTC TAGCACCGCT
/iAmMC6T/AAA
- (25) J58-RTA 5'- /5Biosg/TGTT CAATACATGC ACAGGAT GTATATATCT
GACACGTGCC TGGAGACTAG TAAGTAATCC TATTGGCGGT
/iAmMC6T/AAA
- (26) I9-RRH-1-10 5'- GG GCGGCGACCT /idSp/ TATA CAATACATGC
CTGGAGAA/iAmMC6T/C CC
- (27) J58-LL8-24 5'- /5Biosg/TGTT CAATACATGC ACAGGAT ATCCCGGTGC
CGAGGCCGCC TGGAGACTAG GGAGTAATCC CCTTGGCGGT
/iAmMC6T/AAA
- (28) J7-LL8-24 5'- GG GCGGCGACCT /idSp/GGTCGCTGTT CAATACATGC
ACAGGA/iAmMC6T/ ATCCCG
- (29) DIG oligo 5'-AGGTCGCCGCCCT TT/digoxigenin/
- (30) 601-LH top /5Cy3/CAGAATCCGT CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCG
- (31) 601-LH bottom /5Cy5/ACGGATTCTG CGC GTACGTGCGT /iBiodT/TAAGCGGTG
CTAGAGCTGT CTACGACCAA TTGAGCGGCC TCGGCACCGG
GATTCTCCAG
- (32) 601-RH top /5Cy5/ACGGATTCTG TGTCCC CCGCGTT/iBiodT/TA ACCGCCAAGG
GGATTACTCC CTAGTCTCCA GGCACGTGTC AGATATATAC
ATCCTGT
- (33) 601-RH bottom /5Cy3/CAGAATCCGT ACAGGAT GTATATATCT GACACGTGCC
TGGAGACTAG GGAGTAATCC CCTTGGCGGT TAAAACGCGG
GGGACA
- (34) 601MF-LH top /5Cy3/CAGAATCCGT CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGGGA GTAATCCCCT TGGCGGTTAA AACGCGGGGG ACA
- (35) 601MF-LH bottom /5Cy5/ACGGATTCTG TGT CCCC CGCGTT /iBiodT/TAACCGCCA
AGGGGATTAC TCCGACCAA TTGAGCGGCC TCGGCACCGG
GATTCTCCAG
- (36) 601MF-RH top /5Cy5/ACGGATTCTG CGCGTA CGTGCGT/iBiodT/TA AGCGGTGCTA
GAGCTGTCTA CTAGTCTCCA GGCACGTGTC AGATATATAC
ATCCTGT
- (37) 601MF-RH bottom /5Cy3/CAGAATCCGT ACAGGAT GTATATATCT GACACGTGCC
TGGAGACTAG TAGACAGCTC TAGCACCGCT TAAACGCACG
TACCGC

- (38) 601RTA-RH top /5Cy5/ACGGATTCTG TGTCTA CCGCGTT/iBiodT/TA ACCGCCATA
GGATTACTTA CTAGTCTCCA GGCACGTGTC AGATATATAC
ATCCTGT
- (39) 601RTA-RH bottom /5Cy3/CAGAATCCGT ACAGGAT GTATATATCT GACACGTGCC
TGGAGACTAG TAAGTAATCC TATTGGCGGT TAAAACGCGG
TAGACA
- (40) LL8-24-RH top /5Cy5/ACGGATTCTG TGTCCC CCGCGTT/iBiodT/TA ACCGCCAAGG
GGATTACTCC CTAGTCTCCA GGCGGCCTCG GCACCGGGAT
ATCCTGT
- (41) LL8-24-RH bottom /5Cy3/CAGAATCCGT ACAGGAT ATCCCGGTGC CGAGGCCGCC
TGGAGACTAG GGAGTAATCC CCTTGGCGGT TAAAACGCGG
GGGACA

DNA templates and labeling schemes:

A: The templates of the 601 sequence and derivatives used in this investigation. The 601 template is Addgene Plasmid 26656: pGEM-3z/601 plasmid. The top and bottom strands of the 601 sequence are denoted by I and J strand, respectively (pdb file: 3MVD). The ‘left’ and ‘right’ sides in our templates correspond to the 5’ and 3’ ends of the I strand shown in the table. The templates 601 RRH1-10 and 601 LL8-24 were synthesized by PCR, while 601MF and 601RTA were synthesized by IDT. The 601 sequence is color coded: extra-chromosomal handles (black); left outer quarter (magenta); left inner quarter (orange); right inner quarter (green); right outer quarter (blue). The parts of the derivative sequences which are varied in comparison to the original 601 sequence are underlined and changed to the corresponding color portion of the 601 sequence.

B: Labeling schemes and names for all reconstituted nucleosomes using the original 601 sequence. The top and bottom strands of the 601 sequence are denoted by I and J strand, respectively. The I strand is shown in the table. The 601 sequence is color coded as described in panel A. Labeled positions are highlighted in red, and underlined. The sequence index starts from the 5’ end of the 601 on each strand. The specific primers (listed above) used to generate each template are listed next to the template name.

A

601 GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

RRH1-10 GGACCCTATA CAATACATGC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

LL8-24 GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACGCCAAGG GGATTACTCC CTAGTCTCCA GGCGGCCTCG
GCACCGGGAT ATCCTGT GCATGTATTG AACAGCGACC

601MF GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGGGA GTAATCCCT TGGCGGTAA AACGCGGGG ACACCGCGTA
CGTGCGTTTA AGCGGTGCTA GAGCTGTCTA CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

601RTA GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCTA

CCGCGTTTTA ACCGCCAATA GGATTACTTA CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

B

ED1-12 (I77,J-12) TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

ED1-1 (I68,J-1) TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT CATGTATTG AACAGCGACC

ED1 (I68,J7) TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

ED1.5 (I57,J15) TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

ED1.7 (I46,J24) TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

INT (I38,J28) TATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

ED2.8 (I27,J45) GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACA

ED2.5 (I15,J57) GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACA

ED2 (I9,J58) GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACA

ED2-1 (I-1,J79) GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACA

ED2-12 (I-12,J79) GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACA

IJ-12 (I-12,J-12) GGACCCTATA CGCGGCCGCC CTGGAGAATC CCGGTGCCGA GGCCGCTCAA
TTGGTCGTAG ACAGCTCTAG CACCGCTTAA ACGCACGTAC GCGCTGTCCC
CCGCGTTTTA ACCGCCAAGG GGATTACTCC CTAGTCTCCA GGCACGTGTC
AGATATATAC ATCCTGT GCATGTATTG AACAGCGACC

Monte Carlo Simulation Procedures

We adopted a continuum model of symmetric nucleosomal DNA unwrapping developed by Sudhanshu *et al.* (Sudhanshu et al., 2011) and extended it to a more general, asymmetric case. In this model, the nucleosomal DNA is modeled as a left handed helix with a radius of 4.18 nm and height per turn of 2.39 nm. The total free energy change ΔF upon binding of DNA to the nucleosome spool (the histone core) relative to the unbound state is calculated at forces ranging from 0 pN to 12 pN as was done in Figure 3 of Sudhanshu *et al.* (Sudhanshu et al., 2011). In a more general case of asymmetric unwrapping, m and n base pairs can be unwrapped from the weak and strong side, respectively, where m can be different from n . In the model of Sudhanshu et al, $m=n$. In our two-dimensional model, in order to calculate the free energy change ΔF for (m, n) values, we assumed that the free energy change ΔF for (m, n) is equal to the $(m + n)$ base pairs unwrapped case in the symmetric model. In addition, we reduced the binding energy of the inner quarter of the weak side to 1.5 pN nm / bp while all other quarters (two outer quarters and the inner quarter of the strong side) are assumed to have the binding energy of 3.5 pN nm / bp so that the average binding energy is 3 pN nm / bp as was used by Sudhanshu et al. With this energy function, we performed Monte Carlo simulations starting from 0.1 pN and increasing the force in 0.1 pN increments every 2000 time steps until 10 pN of force was reached. The initial position was $m=1$ and $n=1$.

Supplemental References

Sudhanshu, B., Mihardja, S., Koslover, E.F., Mehraeen, S., Bustamante, C., and Spakowitz, A.J. (2011). Tension-dependent structural deformation alters single-molecule transition kinetics. *Proceedings of the National Academy of Sciences of the United States of America* 108, 1885-1890.