

Supplemental Table 1. Focus scores of samples used.

Assay	Status	Focus Score	Age
WB	Non-SS	0	53
		0	68
		0	35
	SS	1.2	59
		9.5	48
		1.6	63
IF	Non-SS	0	36
		0	61
		0	58
		0	71
		0	76
		0	57
	SS	4.7	55
		5.9	55
		1.6	47
		1	48
		1	57
		3.7	46

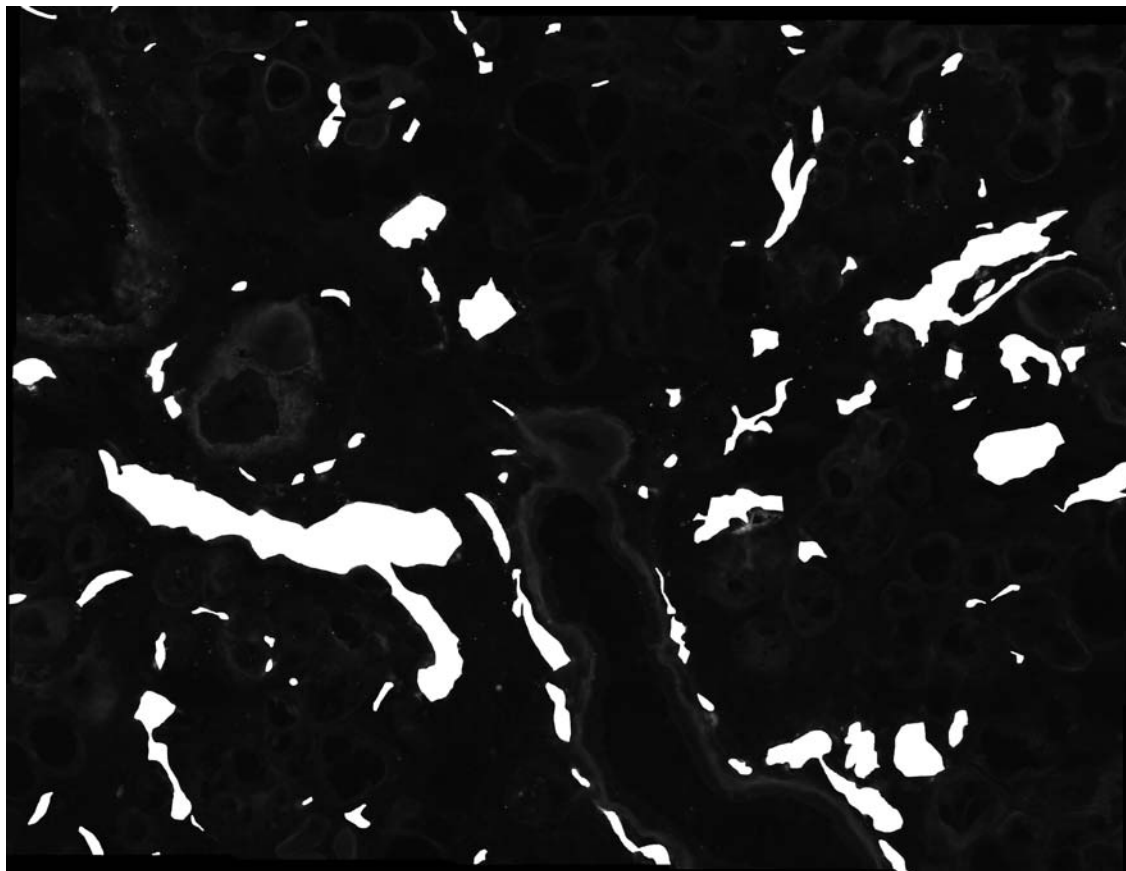
This table lists the focus scores (number of lymphocyte foci with >50 lymphocytes per 4 mm² of salivary tissue) and patient age for each sample used in this study. No individual sample was used for both Western blotting and immunofluorescence. WB: Western Blot; IF: Immunofluorescence; SS: Sjögren's Syndrome.

Supplemental Table 2. Antibodies used for Immunofluorescent Studies.

Antibody	Company	Dilution	Application
Rabbit anti- human vWF	Dako, Carpinteria, CA	1:500	IF Primary Ab (Blood)
Mouse anti-human PECAM-1	Millipore, Billerica, MA	1:250	IF Primary Ab (Blood, Lymph)
Goat anti-LYVE-1	R&D Systems, Minneapolis, MN	1:150	IF Primary Ab (Lymph)
Alexa Fluor 488 goat anti-rabbit	Invitrogen, Carlsbad, CA	1:500	IF Secondary Ab (Blood)
Alexa Fluor 568 goat anti-mouse	Invitrogen	1:500	IF Secondary Ab (Blood)
Alexa Fluor 488 donkey anti-goat	Invitrogen	1:500	IF Secondary Ab (Lymph)
Alexa Fluor 568 donkey anti-mouse	Invitrogen	1:500	IF Secondary Ab (Lymph)

This table lists the antibodies that were used for immunofluorescent staining of the human minor salivary gland tissue sections. IF: Immunofluorescence; Ab: Antibody.

**Supplemental Figure 1. Processed blood vessel
image.**



An example of a blood vessel image, processed as described in Materials and Methods. Expression of platelet endothelial cell adhesion molecule-1 (PECAM-1) and von Willebrand factor (vWF) was detected using immunofluorescent microscopy. Blood vessels (PECAM-1⁺ and vWF⁺) were highlighted, and the image was analyzed using the script described in Supplemental Text 1.

JHC—Journal of Histochemistry & Cytochemistry
DOI: 10.1369/ 0022155415573323
Authors: McCall and Baker
Copyright 2015

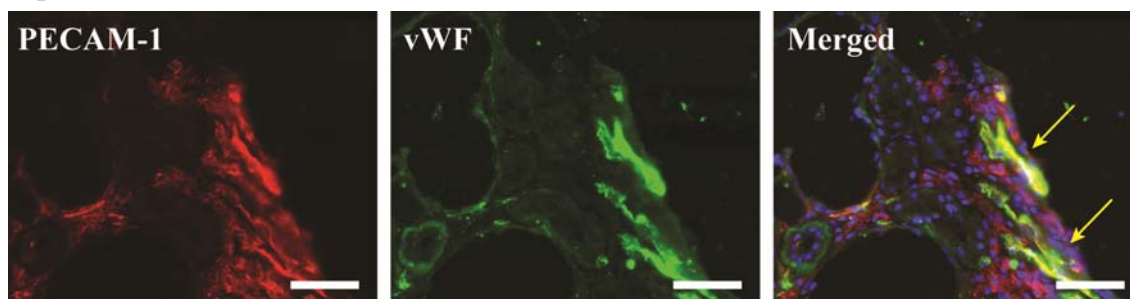
Supplemental Text 1. ImageJ script used for volume fraction analyses.

```
x = getWidth();           ¥¥ Stores the width of the image in variable 'x'.  
  
y = getHeight();         ¥¥ Stores the height of the image in variable 'y'.  
  
highlighted_pixel_count = 0; ¥¥ Initializes the counter for highlighted pixels.  
  
black_pixel_count= 0;     ¥¥ Initializes the counter for black pixels.  
  
for (i=0; i<= x; ++i){    ¥¥ These 'for' statements cycle through every pixel of  
the image.  
  
    for (j=0; j<=y; ++j){  
  
        if (getPixel(i,j)==4095){           ¥¥ If the pixel is highlighted, a counter is  
incremented.  
  
            highlighted_pixel_count += 1;  
  
        }  
  
        if (getPixel(i,j)==0){             ¥¥ If the pixel is black , a counter is incremented.  
  
            black_pixel_count += 1;  
  
        }  
  
    }  
  
}
```

```
}  
  
print (highlighted_pixel_count /((x*y)- black_pixel_count));           ¥¥ Returns the  
  
volume fraction ratio.
```

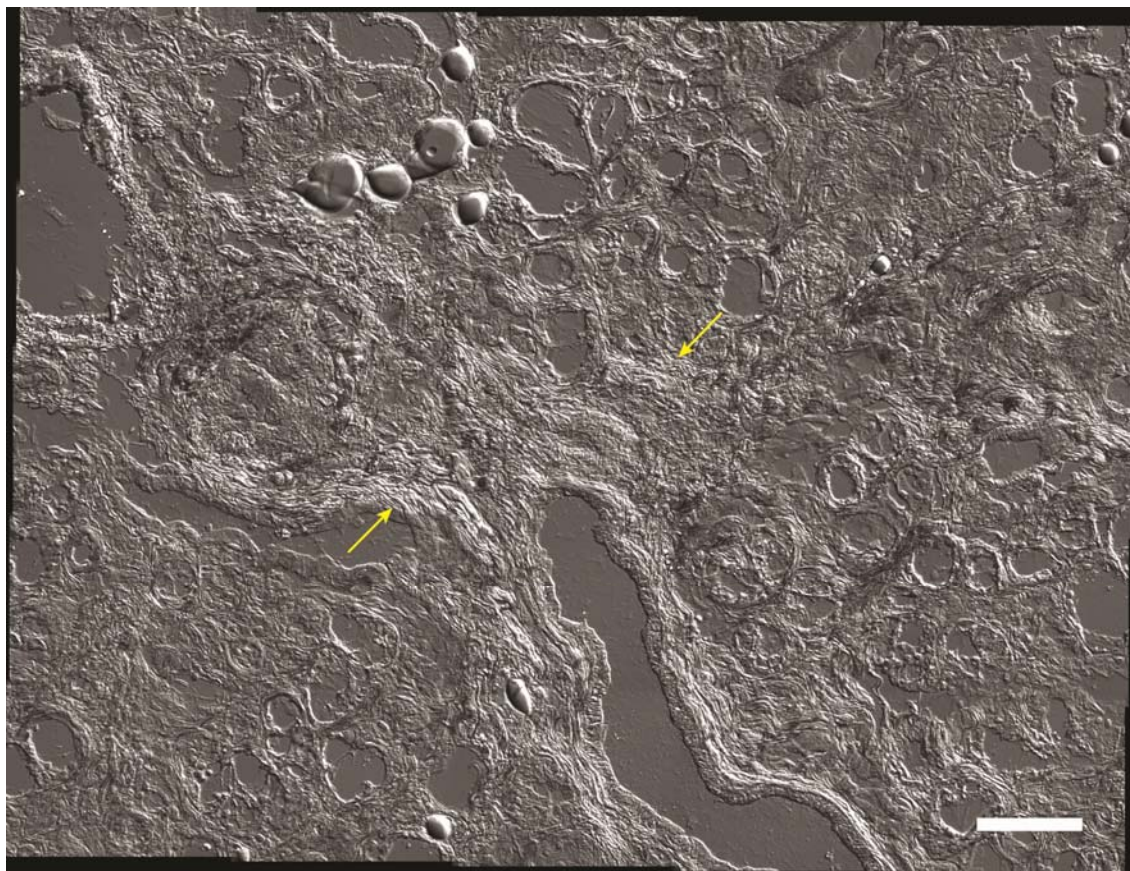
This ImageJ script returns the ratio of highlighted pixels (those with maximum intensity) to total imaged area. It accounts for any regions that were erased from the image (regions of no tissue or non-salivary tissue) by removing any black pixels (zero intensity) from the calculated image area. Any statements between '¥¥' and the end of the line are explanatory comments and not part of the executed program.

Supplemental Figure 2. Blood vessels in the minor salivary gland capsule.



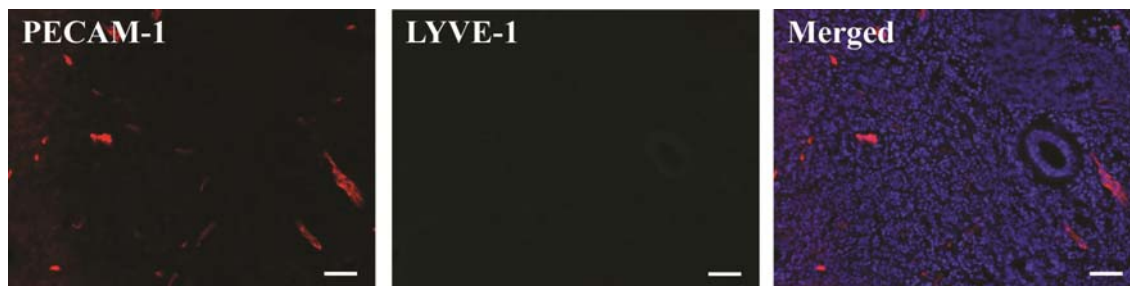
Blood vessels were observed in the capsules encasing the minor salivary glands from both Sjögren's syndrome and non-Sjögren's syndrome patients (the images provided are from a Sjögren's syndrome patient). Five μm frozen human minor salivary gland tissue sections were fixed as described in Materials and Methods. Expression of PECAM-1 and von Willebrand Factor was detected using immunofluorescent microscopy as follows: mouse-anti-PECAM-1 (red); rabbit-anti-von Willebrand Factor (green) and **Hoechst** nuclear stain (blue). Stained structures (red and green) correspond to salivary gland vasculature. Yellow arrows indicate blood vessels in the capsule of the salivary gland. The *xy* cross section images were obtained using a Leica DMI6000B inverted fluorescent microscope. Images are representative of $n = 6$ experiments. All scale bars represent 50 μm .

Supplemental Figure 3. Identification of Connective Tissue.



Connective Tissue (yellow arrows) was identified in minor salivary glands using DIC imaging (the image provided is from a non-Sjögren's syndrome patient). Five μm frozen human minor salivary gland tissue sections were fixed as described in Materials and Methods. The xy cross section images were obtained using a Leica DMI6000B inverted fluorescent microscope. Images are representative of $n = 6$ experiments. Scale bar represents 100 μm .

Supplemental Figure 4. Absence of Lymph vessels in Regions of Infiltration.



No lymph vessels were observed in regions of infiltration in minor salivary glands with Sjögren's syndrome. Five μm frozen human minor salivary gland tissue sections were fixed as described in Materials and Methods. Expression of PECAM-1 and LYVE-1 was detected using immunofluorescent microscopy as follows: mouse-anti-PECAM-1 (red); goat-anti-LYVE-1 (green) and **Hoechst** nuclear stain (blue). Stained structures (red) correspond to salivary gland vasculature. The xy cross section images were obtained using a Leica DMI6000B inverted fluorescent microscope. Images are representative of $n = 6$ experiments. All scale bars represent 100 μm .