## Supplemental material



Figure S1. Motion dynamics of keratinocytes seeded at clonal density in various cell culture media. (A) Full dataset of Fig. 1 B. Correlation between rotational speed in the two-cell colony stage (D1) and proliferative capacity (number of cell doublings) after 6 d of cultivation with a 3T3 feeder layer and serum-containing medium (Spearman's  $\rho = 0.489$ , P = 6.04 × 10<sup>-9</sup>, and n = 126 colonies). P-value was calculated by Student's *t* test. (B) The generation of two-cell colonies from single keratinocytes in various conditions. The time when two-cell colonies were generated after cell division was considered 0 min. Bar, 50 µm.

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Figure S2. **Collective motion dynamics of keratinocytes in a progressively growing colony.** (A) Motion analysis of individual keratinocytes in a progressive growing colony. Top panel shows location of traced cells in the colony. Bottom panel shows changes in locomotion speed of traced cells. The data shown were obtained from a single representative experiment out of multiple observations. (B) Tracing of cell locomotion in the growing colony presented in A and in Fig. 2 A. Bar, 50 µm. (C) Combination experiments of time-lapse imaging and clonal analysis were performed independently five times. Each experiment, except for an experiment on 20091007, revealed a negative correlation between mean speed of cell locomotion in the colony and the ratio of terminal colonies by clonal analysis.



Figure S3. Fractionation of cultured keratinocytes based on  $\alpha$ 6 integrin and IVL expression. (A) Expression of IVL in progressively growing and growtharrested terminal colonies. Growing colonies were constituted by IVL-negative keratinocytes. Bar, 100 µm. (B) Distribution of  $\alpha$ 6 integrin expression in cultured human keratinocytes. The cells were fractionated into four subpopulations (P2–P5) by flow cytometry. The data shown were obtained from a single representative experiment out of triplicate repeats. (C) Immunofluorescent images of sorted keratinocytes in each subpopulation. P5 contained a lot of large cells. Bar, 10 µm. (D) Flow cytometric signatures of each subpopulation. The data shown were obtained from a single representative experiment out of triplicate repeats. (E) Flow cytometric analysis of IVL expression in each fraction. Geometrical means of IVL expression are indicated. IVL-negative keratinocytes were predominantly comprised in P2 and P3 subpopulations. The data shown were obtained from a single representative experiment out of triplicate repeats. (F) Detailed results of Fig. 4 C showing colony-forming efficiency of each fraction during serial cultivation. Bar, 10 mm.



Figure S4. **Characterization and validation of functional shRNA targeted to ITGA6.** (A) Structure of doxycycline-inducible shRNA lentiviral vector (See Materials and methods). (B) Lentiviruses (Lv 1–4) carrying shRNA targeted to *ITGA6* under the control of doxycyclin-inducible promoter were infected into HaCaT keratinocytes. 3 d later, shRNA was induced by doxycyclin treatment, and HaCaT cells were maintained for an additional 3 d to then be fixed and immunostained with an  $\alpha$ 6 integrin antibody (GoH3). Bar, 100 µm. (C) A shRNA targeted to *ITGA6* (Lv 2) efficiently decreased  $\alpha$ 6 integrin expression in normal human keratinocytes. Bar, 50 µm. (D) Transduction of shRNA targeted to *ITGA6* (Lv 2) into the two-cell colony efficiently decreased expression of  $\alpha$ 6 integrin. Bar, 20 µm.



Figure S5.  $\alpha \delta \beta 4$  integrin is involved in collective motion of keratinocytes. (A) Movement of human epidermal keratinocytes (top) and distribution of  $\alpha \delta \beta 4$  integrin (bottom) in two-dimensional cell aggregates. Bars, 20 µm. (B) Secondary antibodies control. Goat antibodies against rat (conjugated with Cy3) and mouse (conjugated with FITC) antibodies did not recognize mouse and rat antibodies, respectively. Bars, 20 µm. (C) Applied antibodies recognized  $\alpha \delta \beta 4$  integrin and remained on cell surface after overnight cultivation. Bars, 20 µm.



Video 1. A time-lapse image of a fast-rotating two-cell colony of normal human keratinocytes. Human epidermal keratinocytes were seeded at clonal density in a 35-mm cell culture dish with mitomycin C-treated 3T3-J2 cells and maintained at 37°C and 10% CO<sub>2</sub>, in a chamber mounted on a microscope (Axiovert 200M; Carl Zeiss). Images were collected at 5-min intervals for 30 min without any stimulation.



Video 2. A time-lapse image of a nonrotating two-cell colony of normal human keratinocytes. Human epidermal keratinocytes were seeded at clonal density in a 35-mm cell culture dish with mitomycin C-treated 3T3-J2 cells and maintained at  $37^{\circ}$ C and 10% CO<sub>2</sub>, in a chamber mounted on a microscope (Axiovert 200M; Carl Zeiss). Images were collected at 5-min intervals for 30 min without any stimulation.



Video 3. A time-lapse imagie of motion dynamics of normal human keratinocytes at high density culture in CnT-PR medium. Human epidermal keratinocytes were seeded into a 35-mm cell culture dish and maintained at 37°C and 10% CO<sub>2</sub>, in a chamber mounted on a microscope (Axiovert 200M; Carl Zeiss). Images were collected at 5-min intervals for 60 min without any stimulation.



Video 4. A time-lapse image of motion dynamics of normal human keratinocytes at high density culture in EpiLife medium with supplement S7. Human epidermal keratinocytes were seeded into a 35-mm cell culture dish and maintained at 37°C and 10% CO<sub>2</sub>, in a chamber mounted on a microscope (Axiovert 200M; Carl Zeiss). Images were collected at 5-min intervals for 60 min without any stimulation.



Video 5. A time-lapse image of motion dynamics of normal human keratinocytes at high density culture in MCDB153 medium containing bovine pituitary extract. Human epidermal keratinocytes were seeded into a 35-mm cell culture dish and maintained at 37°C and 10% CO<sub>2</sub>, in a chamber mounted on a microscope (Axiovert 200M; Carl Zeiss). Images were collected at 5-min intervals for 60 min without any stimulation.



Video 6. A time-lapse image of a growing colony of normal human keratinocytes was collected at 5-min intervals for 180 min without any stimulation. Human epidermal keratinocytes were seeded at clonal density in a 35-mm cell culture dish with mitomycin C-treated 3T3-J2 cells and maintained at 37°C and 10% CO<sub>2</sub>, in a chamber mounted on a microscope (Axiovert 200M; Carl Zeiss).

The Fortran program used for the simulation experiments is provided online as an RTF file.