

Supplemental Figure S1: Bronchial epithelial cell polarity and integrity is maintained in *Adamtsl2*^{-/-} bronchi. (A-E) Staining for selected markers of bronchial cell differentiation and intracellular compartments is shown in *Adamtsl2*^{-/-} (-/-) and WT mice. Comparable smooth muscle actin (SMA, from n = 5 per genotype) (A) and pan-cytokeratin (pCK, from n = 2 per genotype) (B) are indicative of an intact smooth muscle cell layer and epithelial cell identity, respectively, in the bronchi of *Adamtsl2*^{-/-} lungs. The occluding vesicles are not positive for lysosomes (Lamp2, from n = 2 per genotype) (C), or the Golgi apparatus marker GM130 (E, green, n = 3 per genotype), but stained for the ER marker KDEL (from n = 3 per genotype) (D). Staining for ciliated cells using β -tubulin (red) shows no differences between WT and *Adamtsl2*^{-/-} lungs. (F-I) Markers for spatial relationships of bronchial epithelium show that despite profound epithelial dysplasia in *Adamtsl2*^{-/-} bronchi, the overall bronchial epithelial identity and orientation were maintained (from n = 2 per genotype), although *Adamtsl2*^{-/-} bronchial epithelium shows reduced staining in H and I. Collagen IV staining indicates an intact basement membrane (F, arrows point to the epithelial/SMC junction), cortical actin, stained with phalloidin-FITC, is distributed apically and laterally in WT and *Adamtsl2*^{-/-} bronchial epithelium (G), PKC ζ mainly stains the luminal side of the bronchial epithelium (H). The zona occludens marker ZO-2, even though weaker in the *Adamtsl2*^{-/-} lungs, indicates tight junctions between bronchial epithelial cells (I). Bronchi are outlined with dashed line. Scale bars = 25 μ m, if not indicated otherwise. WT, wild-type, -/-, *Adamtsl2*^{-/-}; B, bronchi; BV, blood vessel.

Supplemental Figure S2: No alteration in cell proliferation or apoptosis in the bronchial epithelium of *Adamtsl2*^{-/-} lungs. Cell proliferation (top panels) was analyzed by staining with an antibody against phospho-histone H3, which marks proliferating cells (from n = 2 per

genotype). Positive (proliferating) cells are indicated with arrows. Apoptosis (bottom panels) was assessed with TUNEL staining (from n = 2 per genotype). TUNEL positive cells are indicated with arrows. Scale bars = 25 μ m. B, bronchi.

Supplemental Figure S3: Immunolocalization of FBN1 in paraffin-embedded lung sections.

FBN1 predominantly localizes to the blood vessel walls and only a minor amount is found in the bronchial epithelium at E14.5 (from n = 3 per genotype), E17.5 (from n = 3 per genotype), and P0 (from n > 6 per genotype). Nuclei are counterstained with DAPI (blue). Quantification of mean integrated density of fluorescence signal for FBN1 showed no statistically significant difference between the WT and *Adamtsl2*^{-/-} lungs. Scale bars = 25 μ m. B, bronchi; asterisk, blood vessel.

Supplemental Figure S4: Non-canonical TGF β -signaling pathways are not altered in

Adamtsl2^{-/-} lungs. (A) Western blots are shown for pErk1/2 or pp38 and total Erk1 or p38, respectively, using total protein extracted from *Adamtsl2*^{-/-} or WT lungs at E17.5 (left, n = 1 for WT, n = 4 for *Adamtsl2*^{-/-}; n = 4 for *Adamtsl2*^{+/-}) and P0 (right, n = 3 for WT and *Adamtsl2*^{-/-}; n = 2 for *Adamtsl2*^{+/-}). Equal amounts of protein were loaded and the mean intensity of the pErk1/2 or pp38 bands was quantified using ImageJ and normalized to the amount of total Erk1 or total p38, respectively. (B) Absence of nuclear pSmad2 staining in NAB treated lungs compared to IgG treated lungs is indicative of the efficiency of the NAB for blocking TGF β signaling (from n = 2 per genotype). Scale bars = 25 μ m.

Supplemental Table S1. Frequencies of genotypes arising from *Adamtsl2*+/- intercrosses¹

Stage	Genotype		
	<i>Adamtsl2</i> +/+	<i>Adamtsl2</i> +/-	<i>Adamtsl2</i> -/-
E14.5 – E18.5	33 (35.1)	34 (36.2)	27 (28.7)
Postnatal day 0 - 1	30 (33.3)	35 (38.9)	25 (27.8) ²
Postnatal day 7	60 (37.5)	95 (59.4) ⁴	5 (3.1) ³

Shown are the number of mice (percentage) at the indicated ages

¹Numbers are combined from intercrosses arising from *Adamtsl2*+/- mice before and after *Neo* deletion.

²8/21 (38.1%) were found gasping and/or cyanotic, others were found dead.

³These mice were small, with tight skin and joint contractures, and died before 14 days of age.

⁴These mice were viable, fertile and most were indistinguishable from WT littermates. A few *Adamtsl2*+/- mice were small, with variably tight skin and excess collagen deposition around bronchi.

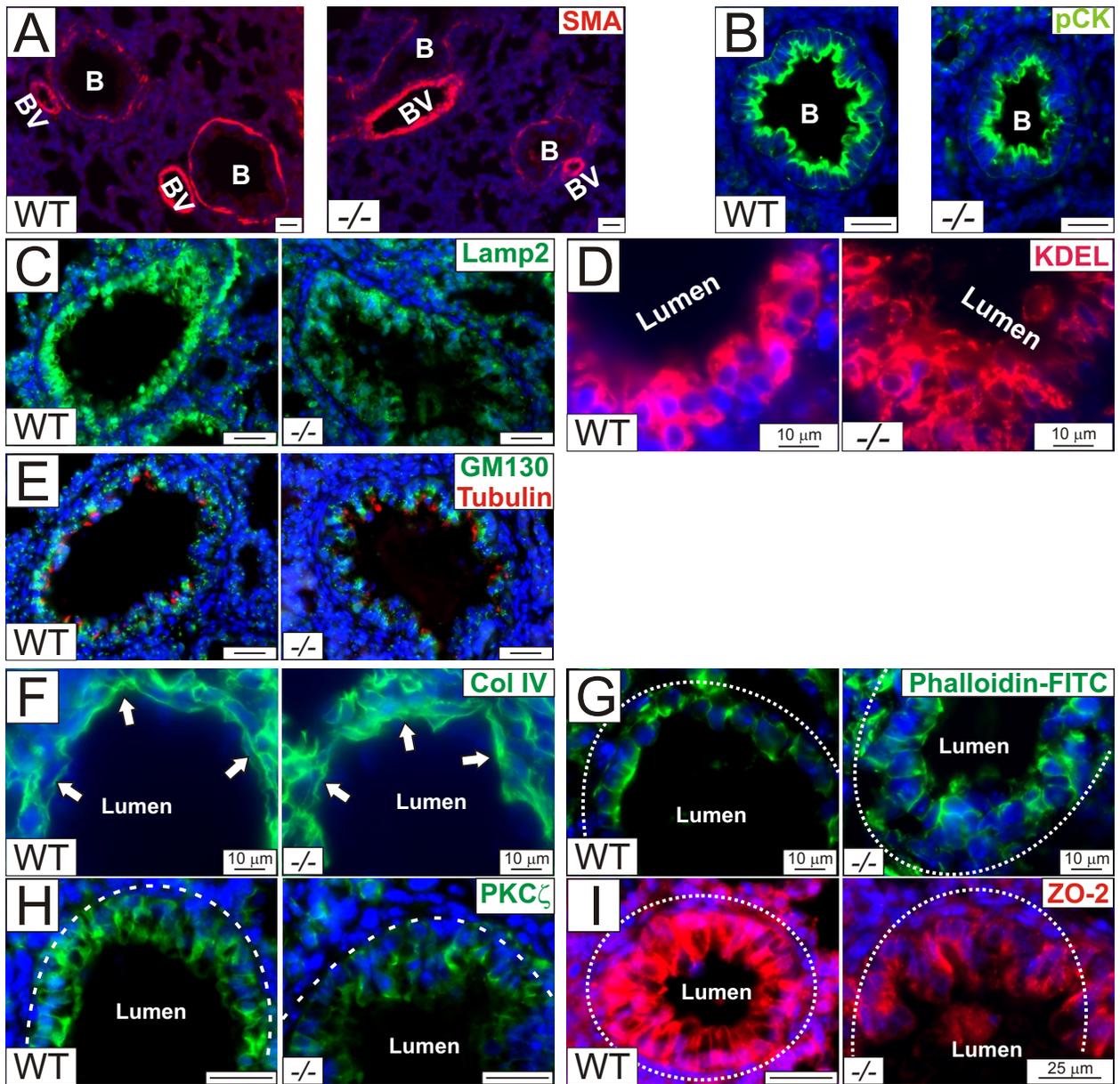
Supplemental Table S2. Primers used for genotyping and qRT-PCR

Primer	Sequence (5'-3')	Application
<i>Adamtsl2</i> WT F	GTACCAGCTCTGCAGAGTGC	Genotyping
<i>Adamtsl2</i> WT R	AAGCTCCTCCCATCCGGTGG	Genotyping
<i>Adamtsl2</i> KO F	AGCTGCGTGTTGTCTCCCC	Genotyping
<i>Adamtsl2</i> KO R	CACTGAGTCTCTGGCATCTC	Genotyping
<i>Adamtsl2</i> Exon2/3 F	GCTGTAGCAGTTGTGGCT	qRT-PCR
<i>Adamtsl2</i> Exon2/3 R	CCTCTAGGCTGTTGGATGTG	qRT-PCR
<i>Adamtsl2</i> Exon 8/9 F	CCAGATTGTGGAGAGGAAGAAG	qRT-PCR
<i>Adamtsl2</i> Exon 8/9 R	GTCCACTTTGTAGTTGCCATTG	qRT-PCR
<i>Fbn1</i> F	GCCAGAAAGGGTACATCGG	qRT-PCR
<i>Fbn1</i> R	ACACACCTCCCTCCGTT	qRT-PCR
<i>Fbn2</i> F	GTGAAACCACACAGAAATGTGAA	qRT-PCR
<i>Fbn2</i> R	GAACAGTCGCCAGTCTCAC	qRT-PCR
<i>Fn</i> F	GTCTAGGCGAAGGCAATGG	qRT-PCR
<i>Fn</i> R	CCTATAGGATGTCCGGGTGT	qRT-PCR
<i>Magp1</i> F	CATCCACAAGCCTTGCAAAC	qRT-PCR
<i>Magp1</i> R	CAGACAGTGCGGACACATATT	qRT-PCR
<i>Ltbp1</i> F	GGTTATTTGCCATCTTCCGTGTA	qRT-PCR
<i>Ltbp1</i> R	GAAATTTGGAGGGCACTGACA	qRT-PCR
<i>mHprt1</i> F	GATCCATTCTATGACTGTAGAT	qRT-PCR
<i>mHprt1</i> R	AGATCATCTCCACCAATAACTT	qRT-PCR
<i>mGapd</i> F	GTGCTGAGTATGTCGTGGAG	qRT-PCR
<i>mGapd</i> R	GCGGAGATGATGACCCTTT	qRT-PCR

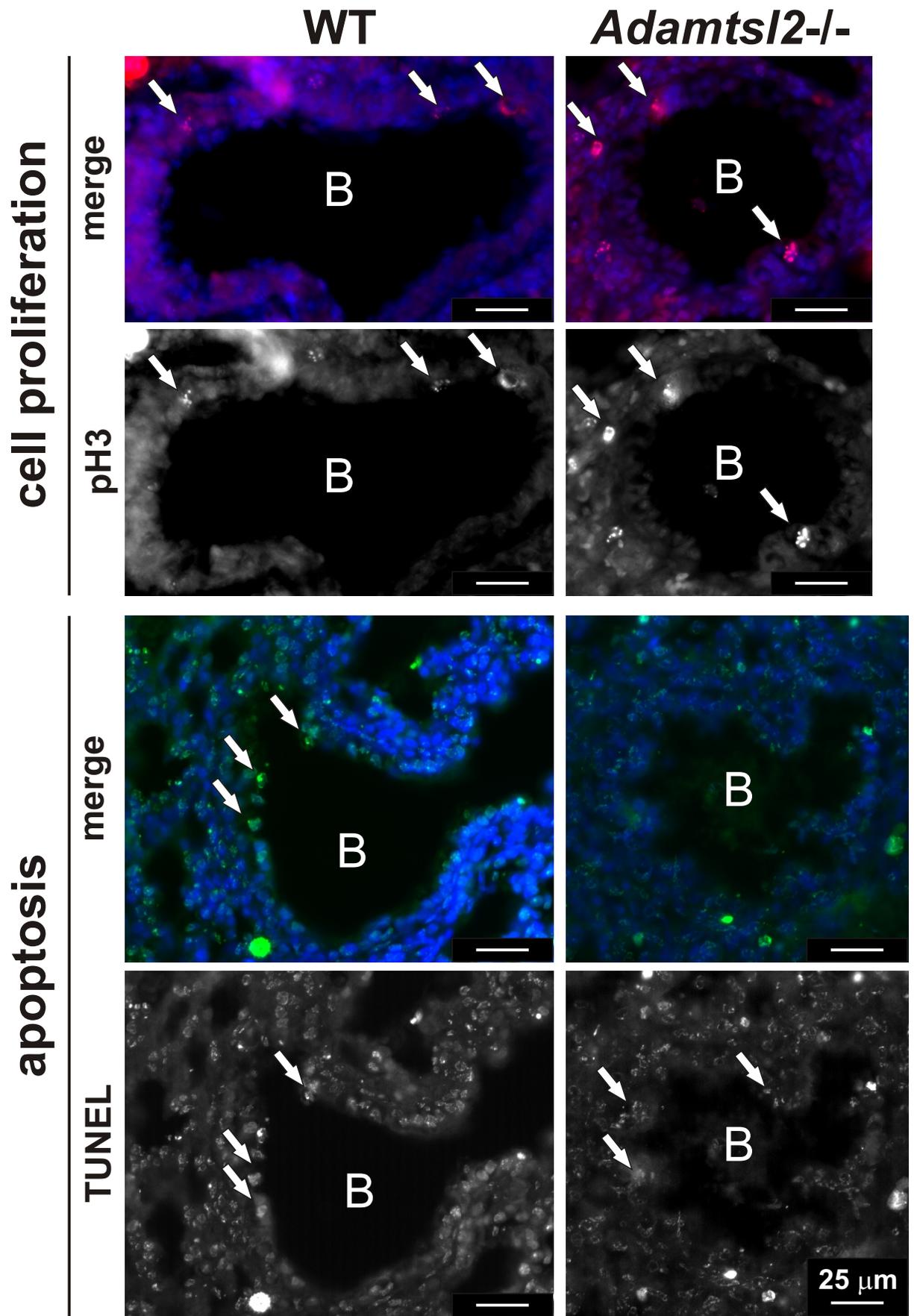
Supplemental Table S3. Antibodies used for immunostaining and western blotting

<i>Antibody</i>	<i>Host</i>	<i>Supplier</i>	<i>Product number</i>	<i>Dilution</i>	<i>Application (Embedding)</i>	<i>Antigen retrieval</i>
α -smooth muscle actin (SMA)	Ms	Sigma	A2547	1:300	IF (P)	-
pan-cytokeratin	Rab	Dako	Z0622	1:200	IF (P)	H/E
KDEL	Ms	ENZO	ADI-SPA-827	1:200	IF (P)	H/E
Magp1	Rab	-	-	1:200	IF (P)	H/E
fibronectin	Rab	Millipore	AB2033	1:200	IF (P)	H/E
fibrillin-1	Rab	-	-	1:250	IF (P, F)	H/E
fibrillin-2	Rab	-	-	1:300	IF (P, F)	H/E
Lamp2	R	Abcam	ABL-93	1:200	IF (F)	-
Ltp1-K	Rab	-	-	1:250	IF (F)	-
GM130	Rab	Abcam	ab52649	1:250	IF (P)	H/E
β -tubulin-IV	Ms	BioGENEX	MU178-US	1:500	IF (P)	- and H/E
Col IV	Rab	Rockland	600-401-106-.01	1:500	IF (P)	proteinase K
Phalloidin-FITC	-	Life Technologies	F432	1:500	IF (F)	-
PKC ζ	Ms	Santa Cruz	sc-17781	1:200	IF (P)	-
ZO-2	Rab	Santa Cruz	sc-11448	1:200	IF (P)	-
pSmad2	Rab	Cell Signaling	3101	1:600	IHC (P)	H/E
phospho-histone H3	Rab	Millipore	06-570	1:250	IF	H/E
pSmad2	Rab	Cell Signaling	3101	1:1000	WB	-
Smad2/3	Rab	Cell Signaling	3102	1:1000	WB	-
pSmad1/5/8	Rab	Millipore	AB3848	1:500	WB	-
Smad1	Ms	Millipore	04-1100	1:1000	WB	-
pErk1/2	Rab	Cell Signaling	4376	1:1000	WB	-
Erk1	Rab	Santa Cruz	sc-94	1:1000	WB	-
pp38	Rab	Cell Signaling	4511	1:500	WB	-
p38	Rab	Cell Signaling	9212	1:1000	WB	-
ADAMTSL2 (C3)	Rab	GeneTex	GTX102069	1:500	WB	-

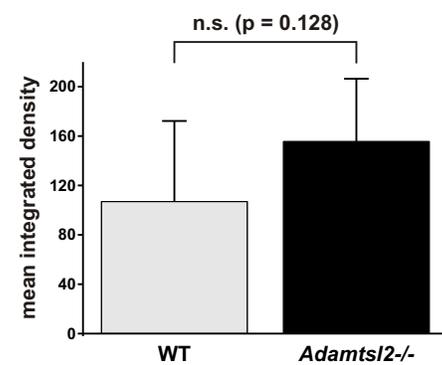
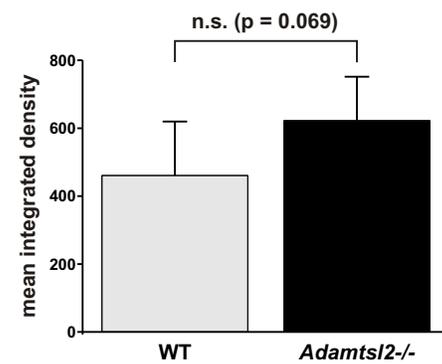
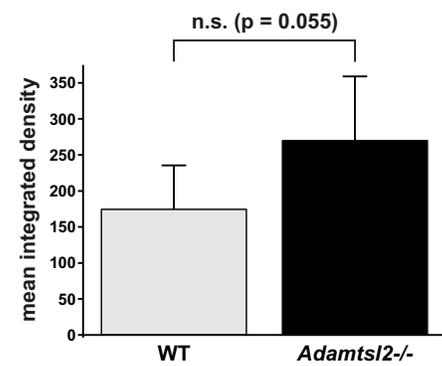
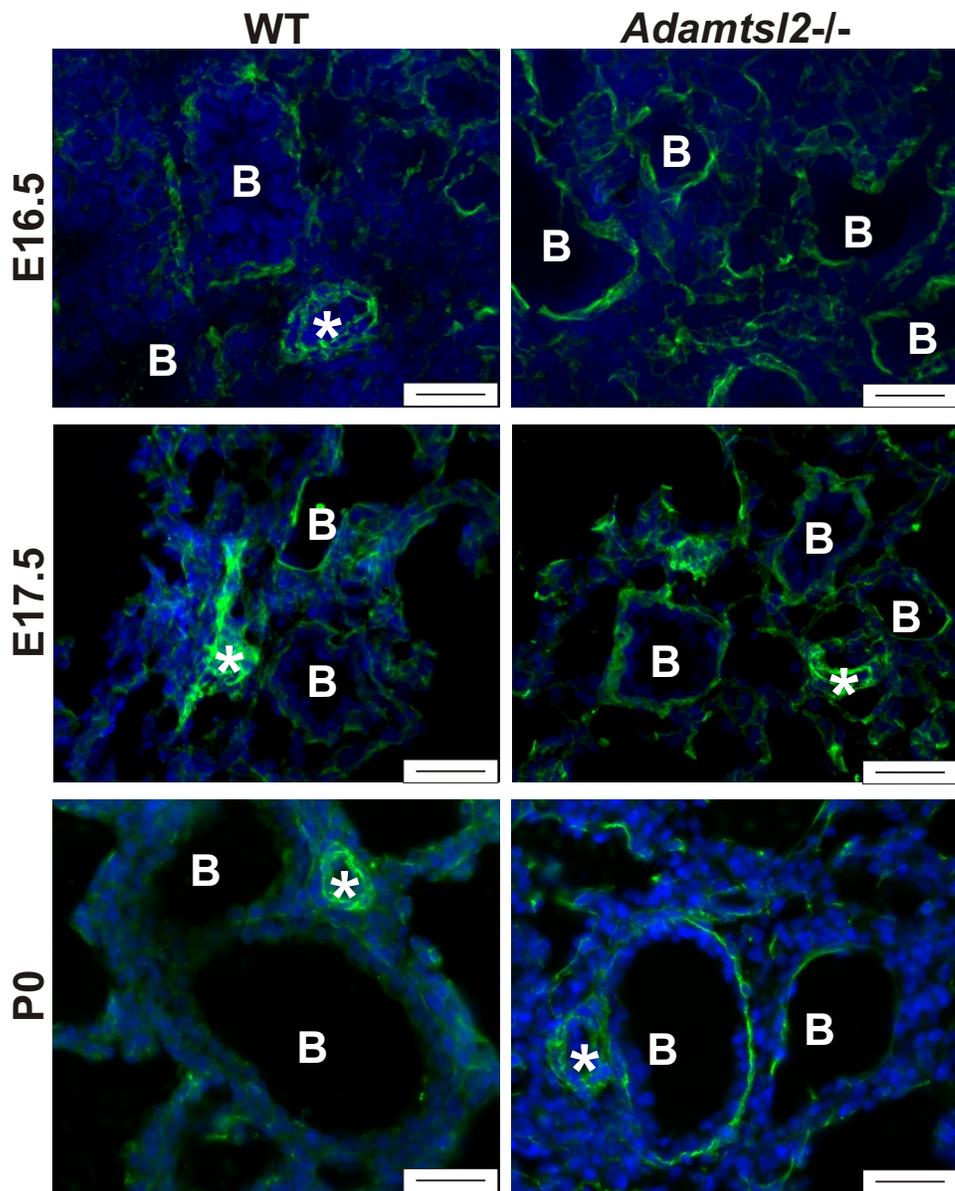
Ms: mouse; *Rab*: rabbit; *R*: rat; *IF*: immunofluorescence, paraffin (P) or frozen (F) sections; *IHC*, immunohistochemistry; *ICC*: immunocytochemistry; *WB*: western blotting; *H/E*: 4 x 1.5 min at 50% power with 30 sec intermission in 10 mM citric acid, 2 mM EDTA, 0.05% Tween-20, pH 6.2, in a microwave oven; *Proteinase K*: 2 min incubation with 20 mg/ml proteinase K solution at room temperature



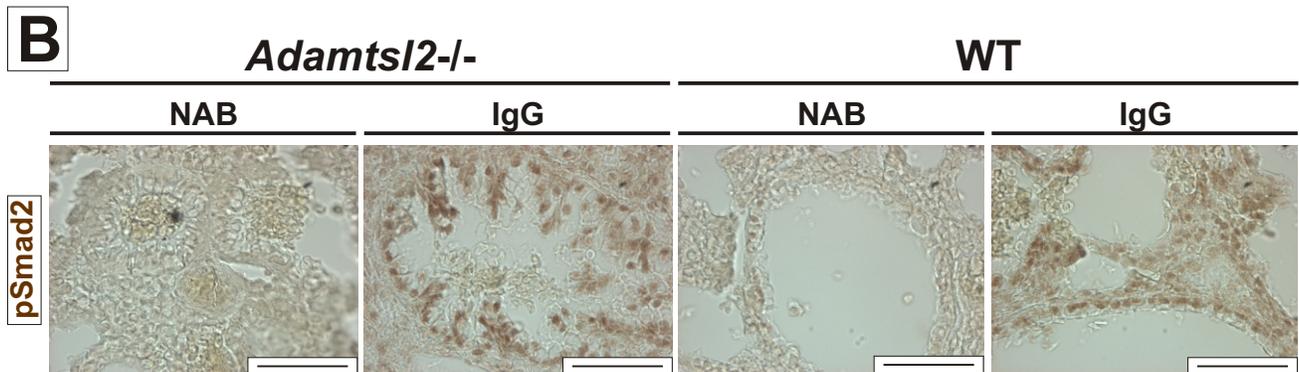
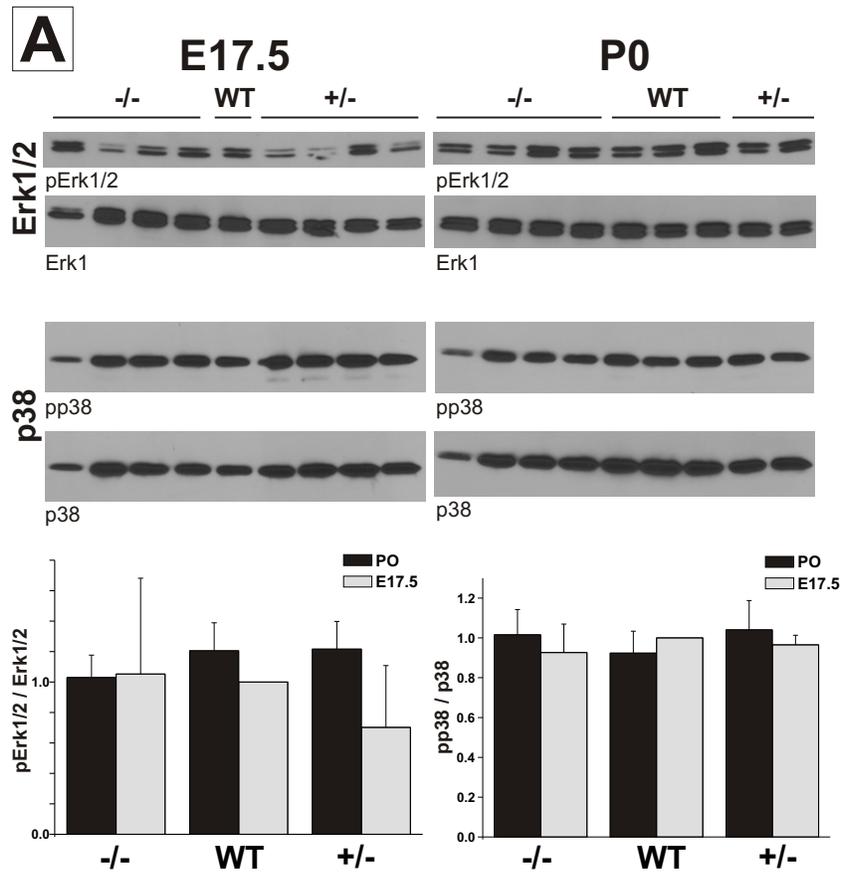
Supplemental Figure S1



Supplemental Figure S2



Supplemental Figure S3



Supplemental Figure S4