STUDIES OF THE VENTILATION-PERFUSION RELATIONSHIPS IN THE LUNGS OF SUBJECTS WITH CHRONIC PULMONARY EMPHYSEMA, FOLLOWING A SINGLE INTRAVENOUS INJECTION OF RADIOACTIVE KRYPTON (KR⁸⁵). I. PRESENTATION AND VALIDATION OF A THEORETICAL MODEL *

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The study of the elimination of an inert gas from the lungs of human individuals has proven a useful tool in the determination of ventilation patterns (1, 2). In particular it could be demonstrated that the lungs of emphysematous patients can be conceived as being composed of at least two different compartments, i.e., a relatively small space with good ventilation and a relatively large space with poor ventilation (3).

In an attempt to establish the ventilationperfusion relationship in the different lung compartments of patients suffering from pulmonary emphysema, Briscoe (4, 5) correlated the ventilation and volume patterns with the oxygen saturation of mixed arterial blood: a very large and poorly ventilated air compartment in the emphysematous lung was shown to be relatively underperfused with blood (6).

In order to check on these findings it was decided to measure the distribution of ventilation and perfusion (especially the latter) in emphysema with an independent technique. Dissolved radioactive krypton (Kr^{85}) was injected intravenously and its rate of disappearance was measured in both expired air and arterial blood. A preliminary report of this work has been published (7). The purpose of this paper is to describe the behavior of intravenously injected krypton in the emphysematous subject and to show how this behavior may be used to determine the distribution of perfusion in the lung. With this objective we present 1) a theoretical model of the lungs and tissues of the emphysematous man under these conditions; 2) a mathematical solution of the equations of the three-compartment analog thus depicted; 3) a comparison of the predicted theoretical curves with the experimental curves which are actually obtained; 4) three methods for interpreting experimental data in terms of the dimensions, ventilation and blood perfusion of several lung compartments: a) an exact method, which might be applicable under optimal conditions to experiments with sampling at three sites, and b) two approximate methods, applicable to data limited as to time and number of sampling sites as they usually are in practice.

I. THEORETICAL MODEL: THREE COMPARTMENT ANALOG OF THE BODY IN EMPHYSEMA

The sequence of events which take place after the rapid injection of a dissolved tracer gas of low solubility into a systemic vein of a human subject is best understood if the behavior of a model, essentially analogous to the experimental situation, is studied. The experiment begins with the rapid injection of radioactive Kr⁸⁵ dissolved in saline (8). The usual site of injection is a large vein as near as possible to the heart; the optimal site is the right heart itself which was used in the studies reported here. As the slug of dissolved Kr⁸⁵ passes through the pulmonary capillaries almost all of it diffuses into the alveolar gas spaces because of the relative insolubility of krypton (8). In emphysema these spaces vary greatly in their degree of ventilation. A small part of the lung is well ventilated. That fraction of the injected Kr⁸⁵ which is carried in

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FIG. 1. MODEL SIMULATING THE WASHOUT OF INTRA-VENOUSLY INJECTED KR⁸⁶ IN PATIENTS WITH PULMONARY EMPHYSEMA. For explanation of symbols see text. 1A shows the three chambers under consideration and the flows between them; 1B the fractional rate constants, indicated by arrows, are used in the solution of the differential equations 2, 3 and 4.

the blood to, and deposited in the well ventilated air spaces, is rapidly eliminated in 2 to 5 minutes. A much larger part of the lung is homogeneously very poorly ventilated. The krypton deposited in these air spaces at the time of injection is still being eliminated in appreciable amounts in the gas expired from them 20 minutes later. The end capillary blood leaving both groups of alveoli is considered at all times to contain krypton at the same partial pressure as the alveolar gas. The quantities of krypton in solution in blood are relatively small, compared to the quantities in the gas phase $\lceil \lambda = 0.051 \ (9) \rceil$. The end capillary blood from the differently ventilated lung spaces mixes in the arterial blood stream which then passes to the tissues. In the first few minutes after injection, arterial blood contains more Kr⁸⁵ than the tissues, and the tissue Kr⁸⁵ concentration rises. As krypton is ventilated out of the lung spaces, arterial Kr⁸⁵ concentration falls. After reaching a peak a few minutes after injection, tissue Kr⁸⁵ concentration falls. Venous Kr⁸⁵ concentration is considered at all times to be equal to that in the tissues. Krypton from the tissues is carried in venous blood to the lungs, redeposited into alveolar gas, and eliminated in ventilation, until eventually no Kr⁸⁵ remains in the body.

The simplest model, therefore, for the quantitative considerations of this situation consists of three reservoirs or mixing chambers, exchanging krypton by means of the blood stream, and with two of these reservoirs eliminating Kr⁸⁵ through ventilatory channels into an infinite space. This model is schematically depicted in Figures 1A and 1B. The three mixing chambers are the well ventilated small lung compartment (L_1) , the poorly ventilated large lung compartment (L_2) and the tissue space (T). Three-compartment systems, both closed (10, 11) and open (12-14), have previously been described; the algebraic treatment is therefore abbreviated here. It is noteworthy that in the present model direct access is limited to one compartment (T). The output from the two lung compartments (L_1 and L_2) cannot be measured individually; it is possible, however, to measure their combined output which is reflected in arterial blood and expired air. Arterial blood is the mixture of the blood equilibrated in the two lung compartments, weighted by their respective flows. Mixed expired alveolar gas is the mixture of the gas in the two lung compartments, weighted by their respective ventilations. Expired gas is mixed expired alveolar gas with its krypton diluted by inspired gas from the dead space. In the model calculations the dead space is assumed to be zero, in which case mixed expired alveolar and expired gas are the same. Another specific feature of this model is the combination of a liquid and a gaseous carrier phase. This required knowledge of a) the solubility characteristics of the tracer

gas, and b) the difference in counting efficiency of liquid and gaseous specimens containing tracer (9).

The description of the emphysematous patient's body in terms of only three compartments did not originate *de novo* but from studies of their behavior. Although there are probably differences among the well ventilated alveoli (L_1) and among the various tissues included in the tissue space (T), it appears from nitrogen washout studies that in severe emphysema there is often a surprising lack of variation among the alveoli in the poorly ventilated group L_2 (6). This group of alveoli constitutes the major reservoir for krypton, and has a dominant influence on the behavior of injected krypton; so it is justifiable, and a very useful simplification, to ignore the variation in L_1 and T and to consider the body as if it consisted of only three compartments.

II. MATHEMATICAL DESCRIPTION OF THE BEHAVIOR OF THE MODEL

Having described the model in physiological terms which characterize the emphysematous patient, our next aim is to construct theoretical curves for comparison with the data actually obtained in patients. Furthermore, these theoretical curves will be analyzed by various approximate methods easily applicable to patients in order to determine the error in these methods. The construction of the curves necessitates some rather lengthy calculations, and before proceeding to the comparison between predicted and observed behavior it is necessary to indicate in outline the assumptions (Section IIA), the algebraic principles (Section IIC), and the numerical values of the coefficients in our examples (Section IID).

A. Assumptions

The theoretical model is applied to experimental situations with the following assumptions:

1. Blood and gas leaving a mixing chamber are in complete equilibrium with this chamber. Deviations from this are believed to be insignificant as regards inert gas exchange in the lung (15, 16) with which this study is mainly concerned. The quantities exchanged in the tissues are of secondary importance and if this assumption does not apply here it has no important effect on our conclusions.

2. Ventilation and perfusion are continuous rather than periodic. There is no rebreathing of Kr⁸⁵ by one lung compartment from the respiratory dead space of gas expired from a differently ventilated lung compartment. This assumption has no important effect on the conclusions when there are large differences between the ventilation of the different lung compartments, as is the case in emphysema. Expired gas is mixed expired alveolar gas diluted by a constant flow of inspired gas, the dead space ventilation. Since this dead space ventilation has no effect on the system except to reduced expired concentrations to a constant fraction of mixed expired alveolar concentrations, it has been considered to be zero in our theoretical cases; as explained later it must be allowed for in practice.

3. The solubility of $\mathrm{Kr}^{85}\left(\lambda\right)$ is the same for blood and body tissues.

4. The amount of Kr⁸⁵ in the blood is negligible.

B. Explanation of symbols

Some of the symbols have already been referred to. It is convenient at this point to give a complete list of the symbols used subsequently, with units of each indicated.

Gas volumes (liters): L_1 in well ventilated alveoli; L_2 in poorly ventilated alveoli, $L_T = L_1 + L_2$. Tissue volume (liters): T. Gas flows (liters per minute): \dot{V}_1 in well ventilated alveoli; \dot{V}_2 in poorly ventilated alveoli; $\dot{V}_T = \dot{V}_1$ + \dot{V}_2 ; \dot{V}_E expired ventilation; \dot{V}_D dead space ventilation. Blood flows (liters per minute): \dot{Q}_1 in well ventilated alveoli; \dot{Q}_2 in poorly ventilated alveoli; $\dot{Q}_T = \dot{Q}_1 + \dot{Q}_2$.

Quantities of Kr^{85} (counts per minute): X in L₁, Y in L₂, I injected. Concentrations of krypton in gas phase (counts per minute per liter): F₁ in well ventilated alveoli; F₂ in poorly ventilated alveoli; F_E in expired air. F_A^e in mixed expired alveolar gas. Concentration of krypton in blood phases (counts per minute per liter): C₁ = F₁ λ in well ventilated alveoli, C₂ = F₂ λ in poorly ventilated alveoli; C_a in arterial blood; C₇ in mixed venous blood and tissue. C₇' in mixed venous blood during passage in injectate. Solubility coefficient at body temperature: λ . Time (minutes): t = time in general, u = time spent by blood in alveolar capillary; t_p = time of maximal concentration in mixed venous blood. The symbol (0) signifies boundary conditions at zero time.

With these assumptions and symbols in mind, the behavior of the analog model can be expressed in the following way.

C. Algebraic formulation

1. Rate of change of krypton concentration: differential equations. Considering compartment L_1 as an example, the following relationship holds for each instant:

$$\frac{dX}{dt} = \frac{d(L_1F_1)}{dt} = (\dot{Q}_1C_{\bar{v}} + \dot{V}_1 0) - (\dot{Q}_1F_1\lambda + V_1F_1).$$
 [1]

Rearranging and using rate constants in Figure 1B, Equation 1 and the similar equations for L_2 and T become:

$$\frac{\mathrm{d}F_1}{\mathrm{d}t} = \frac{\dot{Q}_1}{L_1}C_{\overline{v}} - \frac{\dot{Q}_1\lambda}{L_1}F_1 - \frac{\dot{V}_1}{L_1}F_1 \qquad [2]$$

$$\frac{dF_2}{dt} = \frac{\dot{Q}_2}{L_2}C_{\bar{v}} - \frac{\dot{Q}_2\lambda}{L_2}F_2 - \frac{\dot{V}_2}{L_1}F_2$$
[3]

$$\frac{\mathrm{d}C_{\bar{\mathbf{v}}}}{\mathrm{d}t} = \frac{\dot{Q}_1\lambda}{T} F_1 + \frac{\dot{Q}_2\lambda}{T} F_2 - \frac{\dot{Q}_1}{T} C_{\bar{\mathbf{v}}} - \frac{\dot{Q}_2}{T} C_{\bar{\mathbf{v}}}.$$
 [4]

This system of three simultaneous linear differential equations, each being homogeneous, of the first order, and

with constant coefficients, can be solved by classical methods to yield a solution in the following form (17):

$$F_{1} = {}^{1}C_{1}e^{m_{1}t} + {}^{1}C_{2}e^{m_{2}t} + {}^{1}C_{3}e^{m_{3}t}$$
[5]

$$F_2 = {}^{2}C_1 e^{m_1 t} + {}^{2}C_2 e^{m_2 t} + {}^{2}C_3 e^{m_3 t}$$
[6]

$$C_v = {}^{3}C_1 e^{m_1 t} + {}^{3}C_2 e^{m_2 t} + {}^{3}C_3 e^{m_3 t}.$$
[7]

The exponents m_1 , m_2 , m_3 are the three roots of a cubic equation and can all be computed (e.g., by Newton's method) from the numerical values of \dot{V}_1 , \dot{V}_2 , \dot{Q}_1 , \dot{Q}_2 , L_1 , L_2 , T and λ . To determine the values of the constants (C's) in Equations 5, 6 and 7, it is necessary to know also the initial values $F_1(0)$, $F_2(0)$ and $C_{\overline{v}}(0)$ of the krypton concentrations in the three compartments.

2. Initial krypton concentrations: boundary conditions, $F_1(0)$, $F_2(0)$. These were determined by the following approach. It is assumed that the injectate reaches the lungs as a homogeneous square front slug which takes u minutes to pass through the alveoli. The value of u was taken to be 0.2 minute. Its exact value has no important effect on our conclusions.

During this time the mixed venous concentration of krypton is constant at $C_{\overline{v}}'$, the prime indicating that we are dealing with the period during which boundary conditions are established.

Before the end of the passage of the injectate through the lungs, the tissue concentration of Kr^{s5} ($C_{\overline{v}}$) will have begun to rise. However, this rise is very small and it greatly simplifies the situation to regard it as negligible. In that case $C_{\overline{v}}(0) = 0$, after the passage of the injectate through the lungs.

The equations which apply during the passage of the injectate through the lungs are the same as Equations 2, 3 and 4, except that the variable $C_{\overline{v}}$ (from tissues) is replaced in all three equations by the constant $C_{\overline{v}'}$ (from injectate), so that Equation 4 becomes $dC_{\overline{v}'}/dt = 0$.

During the time u, F_1 and F_2 rise from zero to $F_1(u)$ and $F_2(u)$ at the end of the passage of the injectate through the lungs. By integration of Equations 2 and 3 under these conditions one has:

$$F_{j}(u) = \frac{Q_{j}C_{\overline{v}'}}{\dot{V}_{j} + \dot{Q}_{j}\lambda} \begin{bmatrix} 1 - e^{-u(\dot{V}_{j} + \dot{Q}_{j}\lambda)/L_{j}} \end{bmatrix}$$
[8]

where the subscript j is either 1 or 2. These concentrations $F_1(u)$ and $F_2(u)$ occurring at the end of the passage of the slug are the peak values of F_1 and F_2 , and are used as boundary conditions $F_1(0)$ and $F_2(0)$ in numerical examples discussed subsequently. The initial concentration in the tissues $C_{\overline{v}}(0)$ is zero.

It is relevant to our later discussion of the slow krypton method to consider what fraction (θ) of the krypton brought to the alveoli in the blood is lost in the expired gas and the blood which leaves during the passage of the injectate. The krypton remaining in the alveoli at the end of the passage of the injectate is $1 - \theta$.

$$\theta_{j} = \frac{\dot{Q}_{j}C_{\overline{v}}'u - F_{j}(u)L_{j}}{\dot{Q}_{j}C_{\overline{v}}'u} = 1 - \frac{1 - e^{-u(\dot{V}_{j} + \dot{Q}_{j}\lambda)/L_{j}}}{u(\dot{V}_{j} + \dot{Q}_{j}\lambda)/L_{j}} \quad [9]$$

in which $\dot{Q}_i C_{\bar{v}}' u$ is the quantity of Kr⁸⁵ brought to these alveoli by the blood, and $F_i(u) = F_i(0)$. Figure 2 illustrates the values of θ for alveoli of different quantities of



FIG. 2. THE RELATION BETWEEN ALVEOLAR VENTILA-TION, ALVEOLAR PERFUSION, LUNG VOLUME AND THE FRAC-TION OF THE INJECTED KRYPTON REMAINING IN THE ALVEOLI AFTER PASSAGE THROUGH THE LUNG OF THE INJECTATE. θ is the fraction of the injected Kr⁸⁵ which is lost from the alveoli in expired gas and end capillary blood during the passage of the injectate. This passage occupies time u, which is 0.2 minute in this case. The diagram shows how much θ changes with variation in blood perfusion per unit lung volume (\dot{Q}_e/L) and alveolar ventilation per unit lung volume (\dot{V}_A/L). The four blocks whose tops are shaded apply to conditions in L₂ and indicate that in this case θ = 1 to 3 per cent.

ventilation and perfusion per unit lung volume. In the case of the "slow" alveoli, θ is about 0.02. This concludes the discussion of the boundary conditions.

When I, \dot{Q}_1 , \dot{Q}_2 , L_1 , L_2 , \dot{V}_1 , \dot{V}_2 , T and u are known, then $F_1(0)$ and $F_2(0)$ can be determined.

3. Krypton concentrations resulting from admixture: mixing equations. Mixed expired alveolar gas is formed by admixture from L_1 and L_2 and its krypton concentration is:

$$F_{A^{e}} = \frac{\dot{V}_{1}F_{1} + \dot{V}_{2}F_{2}}{\dot{V}_{1} + \dot{V}_{2}}.$$
 [10]

Arterial blood is also formed by admixture,

$$Ca = \frac{\dot{Q}_1 \lambda F_1 + \dot{Q}_2 \lambda F_2}{\dot{Q}_1 + \dot{Q}_2}.$$
 [11]

		•	
 Numerical example	I	II	IIIa IIIb
 Lung space L_1 (<i>liters</i>) L_2	1 4	1 4	$\begin{array}{ccc}1&1\\4&4\end{array}$
Tissue space TVentilation \dot{V}_1 (L/min) \dot{V}_2 Perfusion \dot{Q}_1 (L/min) \dot{Q}_2	60 4.5 0.5 2 3	10 4.5 0.5 2 3	$\begin{array}{cccc} 60 & 60 \\ 4.5 & 4.5 \\ 0.5 & 0.5 \\ 1.4 & 1.4 \\ 3.6 & 3.6 \end{array}$
$ \begin{array}{l} F_1(0) \ (c \rho m/L) \\ F_2(0) \\ C_v^-(0) \end{array} $	131,000 74.000 0	131,000 74,000 0	$ \begin{bmatrix} 260.98 = F_1(10) & 15.61 = F_1(60) \end{bmatrix}^{\dagger} \\ \begin{bmatrix} 16,542 & = F_2(10) & 268.7 & = F_2(60) \end{bmatrix} \\ \begin{bmatrix} 598.8 & = C_{\overline{v}}(10) & 35.42 & = C_{\overline{v}}(60) \end{bmatrix} $
Amount of Kr ⁸⁵ injected at zero minus u (= 0.2) min (cpm)		500,000	

			TA	BLE	I				
Parameters and	boundary	conditions	used for	· the	calculation	of i	the three	numerical	examples*

 * The parameters are assumed round figures which are typical of patients with severe chronic pulmonary emphysema. The boundary conditions have been calculated from the amount of Kr⁸⁵ injected (I = 500,000 counts per minute), its passage time, u, through the pulmonary capillaries (u = 0.2 min), and the parameters for examples I and II.
 † In examples IIIa and IIIb the boundary conditions are concentrations, respectively, 10 or 60 minutes after injection.

These are indicated by the symbols F_1 (10), F_2 (10), $C_{\overline{v}}$ (10), and F_1 (60), F_2 (60), $C_{\overline{v}}$ (60).

Thus the behavior of the analog model, i.e., the changes with time of the Kr⁸⁵ concentrations in the three mixing chambers, in arterial blood and in mixed expired alveolar gas can be completely described in terms of its parameters (size of chambers, exchange rates of carrier between chambers) and the boundary conditions. Mixed expired gas is mixed expired alveolar gas diluted by the admixture of dead space ventilation. Therefore, the mixed expired krypton concentration is:

$$F_{E} = F_{A}^{e} \frac{\dot{V}_{1} + \dot{V}_{2}}{\dot{V}_{1} + \dot{V}_{2} + \dot{V}_{D}}$$
[12]

where $\dot{V}_1 + \dot{V}_2 + \dot{V}_D = \dot{V}_E$. \dot{V}_D is constant in any study and must be determined by other methods (18).

D. Numerical examples calculated from the analog model

Three numerical examples have been calculated in detail. *I*. It was of interest to know how the model behaves if the naturally occurring situation is imitated as closely as possible. The values chosen for the parameters $L_{1,2}$, $\dot{V}_{1,2}$ and $\dot{Q}_{1,2}$ are typical of those found by other methods in patients with severe chronic pulmonary emphysema (6). These parameters are listed in Table I, (numerical examples I and II).

2. The volume of tissues, however, which equilibrates with Kr^{85} in such patients is not known, even as an approximation. An attempt was, therefore, made to assess the magnitude of the possible effect of the tissue volume on the washout of Kr^{85} by considering two extreme situations:

TABLE II

Numerical values of the exponents, m, and the constants of integration, C, of Equations 6, 7 and 8 calculated from the figures listed in Table 1*

 Numerical example	I	II	IIIa	IIIb	
 m1	-4.6007	-4.6049	-4.5	5703	
m_2	-0.1814	-0.5260	-0.1	1943	
m ₃	-0.0636	-0.1315	-0.0)586	
۱C	130 996 7	130.960.0	77.215	4.5823	
${}^{1}C$	- 706 90	-1.232.488	-83.013	0.1307	
¹ C ² ₃	709.868	1,272.158	266.777	10.8969	
2 C	8 157	54,168	0.0040	0.00024	
${}^{2}C^{1}$	61 777 3	5.180.114	9.593.67	-15.1144	
${}^{2}\widetilde{C}_{3}^{2}$	12,214.4	68,765.82	6,948.32	283.814	
³C.	-48.273	- 320.85	-0.019	-0.00117	
³ C,	-1.561.73	-2,510.58	-259.455	0.4087	
³Č.	1,610.11	2,842.32	859.66	35.114	
	,	•			

* The units of the constants are counts per minute per liter, and of the exponents, minutes⁻¹.

in one numerical example (no. I) a value of 60 L was arbitrarily chosen for the tissue volume T, whereas in the second numerical example (no. II) T was assumed to be 10 L (see Table I).

3. It has been shown that the blood flow supplying the poorly ventilated lung space in emphysematous subjects increases by about 20 per cent when 100 per cent O_2 is breathed instead of room air (7). In order to confirm this finding, a number of studies was made in which the patient was switched from room air to pure oxygen as the inspired gas in the middle of a Kr⁸⁶ washout. It was expected that a 20 per cent change in the distribution of perfusion to the



FIG. 3. THE WASHOUT OF KRYPTON FROM THE MODEL IN EXAMPLE I WITH TISSUE VOLUME (T) 60 L. Symbols are the same in Figures 3 through 7. Circles, expired airconcentration, (F_E); squares, arterial blood concentration, (C_s); triangles, mixed venous blood, (C_{\overline{v}}). In Figures 3 and 4 the krypton concentration in the well ventilated (F_1) and poorly ventilated (F_2) alveoli are indicated by dotted lines. Ordinates in Figures 3, 4 and 5 are counts per minute per liter on a logarithmic scale. In these same figures the dead space ventilation (\dot{V}_D) is assumed to be zero, so that $F_E = F_A^{\bullet}$. When dead space ventilation is not zero all values of F_E are reduced: $F_E = F_A^e (\dot{V}_E - \dot{V}_D)/$ \dot{V}_E , and would lie in a line below but parallel to the F_E lines in Figures 3, 4 and 5. The curves for F_1 , F_2 , C_8 , $C_{\overline{v}}$ are not affected by the magnitude of the dead space ventilation.

lung would be reflected in a detectable alteration of the washout pattern. In the numerical example III, we study this in the analog model by assuming that an instantaneous 20 per cent increase of \dot{Q}_2/\dot{Q}_T occurs 10 minutes (example IIIa) and 1 hour (example IIIb), respectively, after the injection of Kr⁸⁵. The parameters which apply to this situation are again listed in Table I (numerical examples IIIa and IIIb). Using the figures listed in Table I the numerical values for the constants of integration and the



FIG. 4. WASHOUT OF KR^{85} FROM THE MODEL IN EX-AMPLE II, WITH TISSUE VOLUME (T) 10 L. The venous peak is earlier than in Figure 1 and compares with Figures 5 and 6.

exponents m_1 , m_2 and m_3 of Equations 5, 6 and 7 have been computed (Table II). Finally, the values ot F_1 , F_2 , $C_{\overline{v}}$, F_A^e , and C_a have been computed for times varying from 0 to 120 minutes after injection. The corresponding curves are shown in Figures 3, 4 and 5.



FIG. 5. WASHOUT OF KR⁸⁵ IN THE MODEL III IN WHICH THE DISTRIBUTION OF PERFUSION CHANGES ABRUPTLY AT 10 MINUTES (IIIA) OR AT 60 MINUTES (B). The modified curves of F_E , C_a and $C_{\overline{\nu}}$ resulting from this are indicated by dashed lines. The main effect of this change in the distribution of perfusion is on C_a .

III. COMPARISON OF THEORETICAL AND EXPERIMENTAL CURVES

A. Theoretical curves

As can be seen in Figures 3 and 4, both mixed expired air and mixed arterial blood behave similarly up to about 15 minutes. Thereafter, however, the washout in example I becomes much more delayed; this is due to the large size of the tissue compartment. The concentration of Kr⁸⁵ is highest at all times, except the first few seconds in the poorly ventilated alveoli (L_2) . In the first few seconds the concentration in the well ventilated alveoli (F_1) may exceed this if its blood flow (\dot{Q}_1) is large in relation to its volume (L_1) . F₁ falls steeply due to its large relative ventilation (\dot{V}_1/L_1) and then rises at about two minutes when Kr⁸⁵ returns in larger amounts from the tissues $(C_{\overline{v}})$. However, these events in F_1 occur at insignificant concentrations, and concern insignificant amounts of Kr⁸⁵, when compared to the much larger concentrations and quantities of Kr⁸⁵ in other chambers at the same time.

 F_E always exceeds C_a ; in Figures 3 and 4 these lines run approximately parallel courses. Both these parameters are formed by admixture, so F_E must always lie between F_1 and F_2 , and C_a between C_1 and C_2 .

The venous and tissue concentration $(C_{\overline{v}})$ is initially zero and rises rapidly to a peak. It then falls gradually back toward zero. The quantity of Kr⁸⁵ entering the tissues before the peak is equal to the total quantity leaving the tissues after the peak, i.e., $\dot{Q}_T \int_0^{t_p} (C_a - C_{\overline{v}}) \cdot dt$

 $=\dot{Q}_T \int_{t_p}^{\infty} (C_{\overline{v}} - C_a) \cdot dt, \text{ where } t_p \text{ is the time of}$ the peak, or more generally: $\int_0^{\infty} (C_a - C_{\overline{v}}) \cdot dt = 0$. The peak of venous concentration occurs at the crossover point where $C_a = C_{\overline{v}}$. The time at which this peak occurs depends on the tissue volume; it is earlier in example II with the small tissue volume.

In example I, with a large tissue volume, this dominates the rate of the latter stages of washout. $C_{\overline{v}}$ exceeds C_a by much more than it does in example II. There is an inflection in the lines in example I after 20 minutes which is not seen in example II. F_1 exceeds C_a in example I but not in example II, due to this cause. The partial pressure in the tissues exceeds that in L_2 by a considerable amount in the late stages of example I, but not in example II. (Equal partial pressures occur in Figures 3, 4 and 5 when $F_2 = 20 C_{v}$.)

B. Experimental curves

It is instructive to consider the situation which is found to occur in fact when Kr⁸⁵ is injected into patients with emphysema.

Most of the experimental data which have been obtained so far were based on curves of Kr85 decay in expired air and arterial blood after injection of $300\mu c$ (7). A few studies are available, however, with the more complete protocol which included mixed venous blood. Examples are given in Figures 6 and 7 which should be compared with Figures 3 and 4. In Case I, 1,000 µc of Kr⁸⁵ was injected through a cardiac catheter into the pulmonary artery.¹ Another cardiac catheter was used to sample venous blood. This catheter was in the superior vena cava. In Case II, 600 µc of Kr⁸⁵ was injected into the right ventricle through one catheter, while the other catheter sampled mixed venous blood in the right atrium. Samples were taken at the times indicated in Figures 6 and 7 from the brachial artery and from collections of mixed expired air, as well as from the cardiac catheter. The two subjects (Figures 6 and 7) resemble the model example I in the approximately parallel course taken by C_a and F_E . The wider separation of these in the subjects is due to the higher counting efficiency of our equipment for gases. which is not allowed for in the model curves. They also resemble the model in the times at which the various exponents dominate the F_E curve. The first rapid exponent has no apparent effect after about 5 minutes. The third exponent does not dominate until after 20 minutes have elapsed.

Subject I (Figure 6) shows resemblance both to example I (Figure 4) and example II (Figure 5). As in example I, he has a prolonged slow washout, presumably from tissues, which indi-

¹ The tracheal mucosa, which is the tissue most irradiated, receives a dose of 0.7 rad after the intravenous injection of 1,000 μ c of Kr⁸⁵. This small dose is calculated on the basis of a biological half-life of 10 minutes.

cates that some tissues which took up Kr⁸⁵ are perfused by a relatively small blood flow. Like example II, he shows an early venous concentration peak, but this peak does not coincide with the crossover point of the arterial and venous concentration curves. When mixed venous blood is sampled, it is mathematically inevitable that the peak coincides with the crossover point. The inconsistency here arises from the position of the sampling catheter in the superior vena cava. Case II is given here, although the smaller injected dose results in low and therefore



FIG. 6. WASHOUT OF KR⁸⁵ IN CASE I, WITH CHRONIC PULMONARY EMPHYSEMA. The venous sampling catheter was in the superior vena cava. The venous peak is at 2 minutes. The C_s and C_{$\bar{\nu}$} lines cross later (see text). In Figures 6 and 7 the ordinates are counts per minute on a logarithmic scale. In blood, 1 mµc per ml gives 70 cpm; in gas, 1 mµc per ml gives 6,200 cpm.

erratic counts later in the study, to show that when mixed venous blood is sampled inside the heart, the peak and crossover points do in fact coincide. Furthermore, the peak occurs early, as in example II, rather than late, as in example I.

These considerations suggest that when dealing with the earlier points of the curve (0 to 15 minutes), the model with the small effective tissue volume (example II) is more applicable, whereas in the late stages (60 minutes), the model with the large tissue volume (example I)



FIG. 7. WASHOUT OF KR⁸⁵ IN CASE II WITH CHRONIC PULMONARY EMPHYSEMA. The venous peak is at 3 minutes and coincides with the crossover point of the C_a and $C_{\overline{v}}$ lines. The venous sampling catheter was in the right auricle. The later C_a and $C_{\overline{v}}$ points are scattered because of the low activity after a relatively small injectate (see text).

is closer to the observed facts. At least two tissue compartments with different blood flows per unit volume would be needed to fit an early peak which occurred with a slow rate of late fall in $C_{\overline{v}}$ as is suggested by the lines drawn among the scattered points in Figure 7.

TABLE III

Effects of changes in the distribution of perfusion upon the ratio of Kr^{85} concentration between arterial blood (C_a) and the expired air $(F_E)^*$

Time	Q₂/Q́т unchanged	Q2/Qt increased by 20%		
injection	C _a /F _E	C_{a}/F_{E}	Increase	
min		A†	%	
10	0.265	0.318	20	
12	0.258	0.320	24	
50	0.205	0.273	33	
		B†		
60	0.204	0.241	18	
62	0.204	0.268	31	
100	0.204	0.273	34	

* Dead space ventilation is zero, so $F_E = F_{A^0}$. † A, at 10 minutes; B, at 60 minutes. Be that as it may, the data in Figures 6 and 7 show reasonable agreement with the threecompartment model and the imperfections and irregularities inevitable in data of this type do not warrant any more involved system of interpretation at present.

C. Influence of instantaneous changes in the distribution of perfusion upon the curves

The principal changes occurring in the washout pattern when the distribution of perfusion is altered instantaneously (increase of \dot{Q}_2/\dot{Q}_T by 20 per cent) are summarized in Table III. The most striking change takes place in the mixed arterial blood: its Kr85 concentration increases instantaneously by 20 per cent if the distribution of perfusion changes at 10 minutes and by 18 per cent if \dot{Q}_2/\dot{Q}_T rises at 60 minutes. An equally important change will occur in the ratio of the Kr⁸⁵ concentrations of simultaneously collected arterial blood and gas specimens. However, the changes in the F_E and $C_{\overline{y}}$ curves are slight, as can be seen in Figure 5. This illustrates the important point that changes in the distribution of perfusion can be detected only by the arterial blood composition, or by some ratio related to this, such as C_a/F_E .

IV. METHODS FOR THE ANALYSIS OF EXPERI-MENTAL CURVES TO DERIVE RELATIVE VENTI-LATION AND/OR PERFUSION OF COMPARTMENTS

So far we have been considering the behavior of a theoretical three-compartment model. We have chosen arbitrary values for its parameters and tried to predict the changes with time of Kr⁸⁵ in the three compartments. The ultimate aim of a washout study is, of course, reciprocal: we must analyze the experimental data and we want to determine from such an analysis as many parameters as possible (i.e., in our case, volumes, ventilation and perfusion).

Such an analysis will vary depending on whether the experimental protocol provides access to all three possible sampling sites—expired air, arterial and mixed venous blood (which entails sampling from the right heart)—or whether access is limited to expired air and arterial blood. First, a method is indicated which might be used under optimal conditions. Second, two approximation methods are described which have been widely used for the evaluation of actual experimental data. Finally, an attempt is made to assess the possible error introduced by the use of approximation formulae.

A. Exact analysis for curves obtained under optimal conditions: sampling from three sites— $C_{\overline{v}}, C_a$ and F_E

The task in this case is to analyze the three experimental curves for F_{E} , C_{a} and $C_{\overline{v}}$. Since the $C_{\overline{v}}$ curve represents the output from one single compartment, its conventional analysis (10) should readily yield the numerical values of its constants and exponents, i.e., ³C₁, ³C₂, ³C₃ and m_1 , m_2 and m_3 . This, however, can only yield useful values when the data show less scatter and continue for longer periods than is the case in Figures 6 and 7, since the first stage in the analysis is the exact determination of the slowest component of the washout in venous blood. The analysis of the F_E and C_a curves is complicated by the fact that they each represent a mixture of the contents of more than one compartment. Nevertheless, a method for deriving the values of \dot{Q}_1 , \dot{Q}_2 . \dot{V}_1 , \dot{V}_2 , L_1 , L_2 and T from the analysis of the curves has been developed.² It would be very laborious to use in practice. Furthermore, the values derived are most sensitive to the rate of washout from the tissues, and are dependent on the assumption that there is only one tissue compartment.

B. Sampling from two sites for a limited time— C_a and F_E : approximation methods

If catheterization of the right heart cannot be performed, no representative specimen of mixed venous blood can be obtained and sampling is confined to mixed arterial blood and mixed expired air. Furthermore, in practice, studies are limited in time. When 300 μ c, which was the usual dose, is injected C_a and F_E are too low for accurate counting with our equipment about 20 minutes after injection.

The slow krypton method for determining \dot{Q}_2/\dot{Q}_T . This depends on the notion that the fate of the injected Kr⁸⁵ is determined in the branches of

1088

 $^{^{2}}$ A description of this method of analysis, which was developed with the help of Mr. Boss, will be sent by the authors on request.

the pulmonary artery a few seconds after injection, and just prior to its passage through the Those Kr⁸⁵ molecules which are directed lung. to the well ventilated alveoli pass into the alveolar gas and are rapidly eliminated in the expired air in the next two or three minutes. Those molecules which are directed to the poorly ventilated alveoli remain much longer in alveolar They are slowly eliminated in the expired gas. air from the poorly ventilated alveoli and to a lesser extent from the well ventilated alveoli which they reach after transfer to and temporary storage in the tissues. An assessment of this quantity which is slowly eliminated is made as follows.

It can be seen from Figure 7 (experimental data) that, when data for F_E are available for only 20 minutes they resemble, within the limits of experimental error, a washout with only two exponents, m_4 and m_5 , and two constants, C_4 and C_5 :

$$F_{E} = C_4 e^{m_4 t} + C_5 e^{m_5 t}.$$
 [13]

The values for m_5 and C_5 can be determined by the slope and the intercept at zero time of a line drawn through the data between 5 and 20 minutes. These constants do not necessarily have any simple relationship to those numbered 1, 2 and 3. There is no need for such algebraic formulation since they are used merely to derive an integral: $\dot{V}_{\rm E} \cdot C_5 \int_0^\infty e^{m_5 t} \cdot dt$. This quantity of krypton which is slowly eliminated is considered as an approximation to consist of those molecules which were present in L₂ at zero time, i.e.,

$$F_2(0) L_2 = \dot{V}_E C_5 \int_0^\infty e^{m_5 t} \cdot dt.$$

Furthermore, it is shown in Figure 2 that θ is about 2 per cent in the slow alveoli, i.e., 98 per cent of the Kr⁸⁵ which reached these alveoli in the blood during the passage of the injectate is present in the alveolar gas at the end of this passage. Thus $L_2 \cdot F_2(0) = I \cdot \dot{Q}_2 / \dot{Q}_T$ where I is the total quantity of Kr injected. Therefore, \dot{Q}_2 / \dot{Q}_T can be estimated by the relationship:

$$\frac{\dot{Q}_2}{\dot{Q}_T} = \frac{\dot{V}_E C_5 \int_0^\infty e^{m_5 t} \cdot dt}{I} \qquad [14]$$

where I is the total quantity injected or the total quantity eliminated in expired gas in infinite time. This approximation was tested by analyzing the data over the period 5 to 20 minutes in examples I and II in each of which $\dot{Q}_2/\dot{Q}_T = 0.60$. In example I, \dot{Q}_2/\dot{Q}_T was estimated at 0.524, an error of -12.6 per cent. In example II the estimate was 0.609, an error of +1.5 per cent.

The partition coefficient method for determining \dot{Q}_2/\dot{Q}_T . Like the slow krypton method, this method of assessing \dot{Q}_2/\dot{Q}_T depends on data collected in studies limited in time to 15 or 20 min-In this case data for both C_a and F_E are utes. used. It is evident from study of examples I and II that in the period from between 5 and 20 minutes after injection, the Kr⁸⁵ which was initially present in the fast alveoli, $L_1F_1(0)$, has been eliminated. The only Kr^{85} in L_1 is that brought by the blood from the tissues. During this period F_1 is small compared to F_2 , and may be regarded as negligible. If F_1 is zero then the only source of Kr⁸⁵ in C_a and F_E in this period, from 5 to 20 minutes, is L_2 . In that case the following approximate relationship can be used to determine \dot{Q}_2/\dot{Q}_T :

$$\frac{\mathbf{F}_{\mathbf{E}}}{\mathbf{C}_{\mathbf{a}}} = \frac{\dot{\mathbf{V}}_{\mathbf{2}}}{\dot{\mathbf{V}}_{\mathbf{E}}\lambda} \div \frac{\dot{\mathbf{Q}}_{\mathbf{2}}}{\dot{\mathbf{Q}}_{\mathbf{T}}}$$
[15]

in which C_a and F_E are simultaneous measurements at a time between 5 and 20 minutes after injection, and \dot{V}_2/\dot{V}_E is known by use of another technique (N₂ washout curve). Applying this method to examples I and II in which \dot{Q}_2/\dot{Q}_T = 0.60 we have estimates of 0.543 and 0.524 for \dot{Q}_2/\dot{Q}_T . They are in error by -9.5 and -12.6 per cent, respectively.

Thus, these two arbitrary assessments of \dot{Q}_2/\dot{Q}_T , the slow krypton method and the partition coefficient method, have errors in these examples comparable to the errors of other physiological assessments in man.

SUMMARY

1. The behavior of intravenously injected Kr⁸⁵ in the emphysematous patient is considered in terms of a model composed of three compartments: well ventilated alveoli, poorly ventilated alveoli, and tissues.

2. The differential equations applicable in

this model have been solved. Using typical emphysematous values for the volume, ventilation and blood flows of the two lung compartments, the behavior of Kr⁸⁵ in the various gases and blood streams is predicted when the effective tissue volume is either 10 or 60 L. Further calculated examples show the predicted effect of changes, during a study, in the distribution of blood flow to the lungs.

3. Data are presented on the findings in two patients with emphysema in whom Kr^{85} concentrations were followed in expired air, arterial blood and mixed venous blood, after intracardiac injection of Kr^{85} . The behavior of the patients resembled that predicted for the model; the differences are discussed.

4. A method has been developed in which, by analysis of simultaneous expired, arterial and mixed venous concentration curves, the distribution of perfusion to the lung could be assessed. It is laborious in use and of very doubtful validity. Two approximate methods for determining the distribution of perfusion in the lung are presented. One depends only on expired gas analysis and the other on expired gas and arterial blood analysis. When tested on the predicted curves these appear to be in error by about 10 per cent.

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