

## SI APPENDIX

**Table S1.** Explanatory variables used in this study. Descriptors in bold were retained for statistical analyses after collinearity analyses. Data were downloaded from the Climatic Research Unit (CRU) at the British Atmospheric Data Centre (<http://badc.nerc.ac.uk/>), NASA Earth Observations (<http://neo.sci.gsfc.nasa.gov/>), NASA Socioeconomic Data and Applications Center (<http://sedac.ciesin.columbia.edu/>), World Soil Information (<http://www.isric.org/>), National Agricultural Statistics Service of the United States Department of Agriculture (<http://www.nass.usda.gov/>), the IUCN Red List (<http://www.iucnredlist.org/>), and Natural Earth (<http://www.naturalearthdata.com/>).

**Table S2.** Bacterial indicator taxa identified. For each indicator taxa, the number of samples where it was present is indicated.

**Figure S1.** Map of sampling locations (**A**) and information on the number of samples per state (**B**). Points indicate sampling locations. Color intensity indicates overlapping points.

**Figure S2.** Comparison between normalization and rarefaction pre-processing for bacterial communities for (**A**) richness ( $r = 0.84$ ,  $P < 0.001$ ) and (**B**) community similarity as NMDS axis ( $r = 0.95$ ,  $P < 0.001$ ).

**Figure S3.** Sample-based species accumulation curves (mean and 95% confidence interval).

**Figure S4.** Proportions of sequences representing bacterial taxa across samples at the phylum level (top) and within the Proteobacteria phylum (bottom). Samples are ordered along the bacterial NMDS ordination axis.

**Figure S5.** Proportions of sequences representing fungal taxa across samples at the phylum level (top) and within the Dothideomycetes class (bottom). Samples are ordered along the fungal NMDS ordination axis.

**Figure S6.** Mean relative abundances (where present) and occupancy relationships for (A) bacteria and (B) fungi.

**Figure S7.** Mantel correlogram. Solid points indicate significant ( $P < 0.05$  using progressive correction of multiple tests) correlations, while empty points indicate non-significant ( $P > 0.05$ ) correlations.

**Figure S8.** Geographical distribution maps for some bacterial sea indicator taxa: (A) *Prochlorococcus*, (B) *Synechococcus*, (C) Pelagibacteraceae and (D) *Pseudoalteromonas*. Points represent sampling locations.

**Figure S9.** Effects of urbanization on richness ( $P > 0.05$ , Mann-Whitney test) and community similarity ( $R^2 < 0.005$ , PERMANOVA) for (A) bacterial and (B) fungal communities.

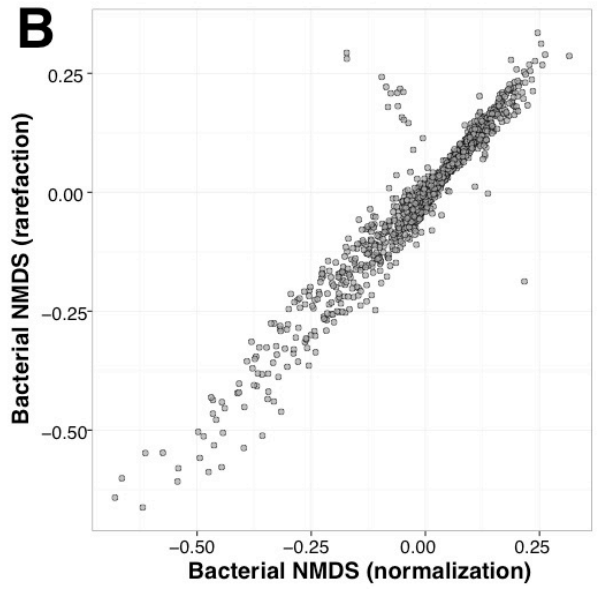
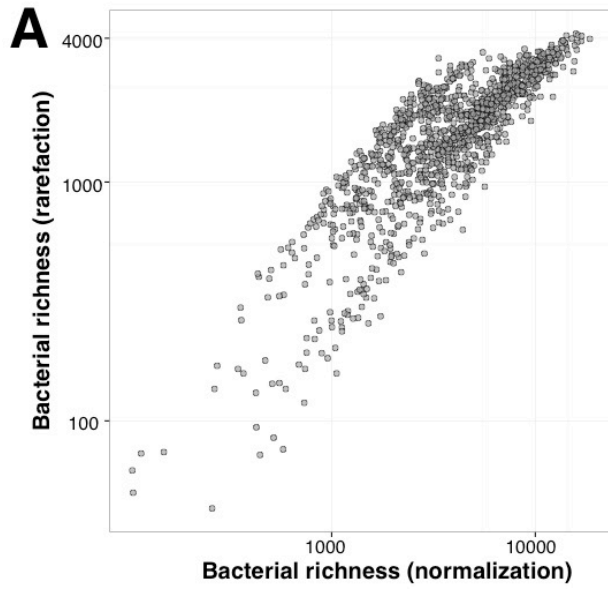
Class	Descriptors
Climate	<b>Mean annual temperature (MAT)</b> , temperature variability, <b>mean annual precipitation (MAP)</b> , precipitation variability, cloud fraction, days of frost during a year, net radiation.
Soil	<b>pH</b> , moisture, <b>dust deposition</b> , clay content.
Plant productivity	<b>Net primary productivity (NPP)</b> , normalized difference vegetation index (NDVI).
Organisms	<b>Human population density</b> , <b>cattle density</b> , chickens density, <b>hogs density</b> , <b>vascular plant diversity</b> , mammal diversity, bird diversity.
Geographic isolation	<b>Elevation</b> , <b>distance to the coast</b> .
House	<b>Construction age</b> , house space, number of bedrooms, <b>number of people</b> , <b>carpeting</b> , <b>number of house plants</b> , <b>number of dogs</b> , <b>number of cats</b> , <b>self-reported presence of mold</b> .

**Table S1**

Source environment	Bacterial indicator taxa
Plants	Chloroplasts (n=1186)
Insects	<i>Wolbachia</i> (n=459), <i>Buchnera</i> (n=429), <i>Rickettsiella</i> (n=737), <i>Cardinium</i> (n=152), <i>Rickettsia</i> (n=489), <i>Rhabdochlamydia</i> (n=153), <i>Fritschea</i> (n=30), <i>Regiella</i> (n=21), <i>Hamiltonella</i> (n=48), <i>Blochmannia</i> (n=18), <i>Portiera</i> (n=109), <i>Tremblaya</i> (n=19), Endomicrobia (n=20), Entomoplasmatales (n=217), Bartonellaceae (n=386), <i>Blattabacterium</i> (n=48), <i>Sulcia</i> (n=59), <i>Thorsellia</i> (n=1), <i>Baumannia</i> (n=2)
Sea	Pelagibacteraceae (n=11), <i>Prochlorococcus</i> (n=8), <i>Synechococcus</i> (n=78), <i>Polaribacter</i> (n=29), <i>Pseudoalteromonas</i> (n=166), <i>Marinobacter</i> (n=141), <i>Nitrosopumilus</i> (n=9)
Skin	<i>Propionibacterium</i> (n=176), <i>Staphylococcus</i> (n=1121), <i>Corynebacterium</i> (n=1144), <i>Streptococcus</i> (n=1057), <i>Rothia</i> (n=356), <i>Micrococcus</i> (n=706), <i>Anaerococcus</i> (n=507), <i>Brevibacterium</i> (n=1026)
Soil	Solibacteraceae (n=990), Chloracidobacteria (n=1153), Koribacteraceae (n=790), Acidobacteria-iii1.15 (n=1156), Acidobacteriaceae (n=1132), <i>Rhizobium</i> (n=873), <i>Bradyrhizobium</i> (n=1061), <i>Mesorhizobium</i> (n=1072), <i>Rhodoplanes</i> (n=1092), <i>Chitinophaga</i> (n=581), <i>Solirubrobacter</i> (n=728), <i>Opitutus</i> (n=689)
Stool	<i>Bacteroides</i> (n=889), <i>Faecalibacterium</i> (n=763), <i>Lachnospira</i> (n=176), <i>Oscillospira</i> (n=811), <i>Roseburia</i> (n=610), <i>Coprococcus</i> (n=845), <i>Ruminococcus</i> (n=762), <i>Parabacteroides</i> (n=459), <i>Phascolarctobacterium</i> (n=535), <i>Sutterella</i> (n=88), <i>Blautia</i> (n=836)

**Table S2**





**Fig. S2**

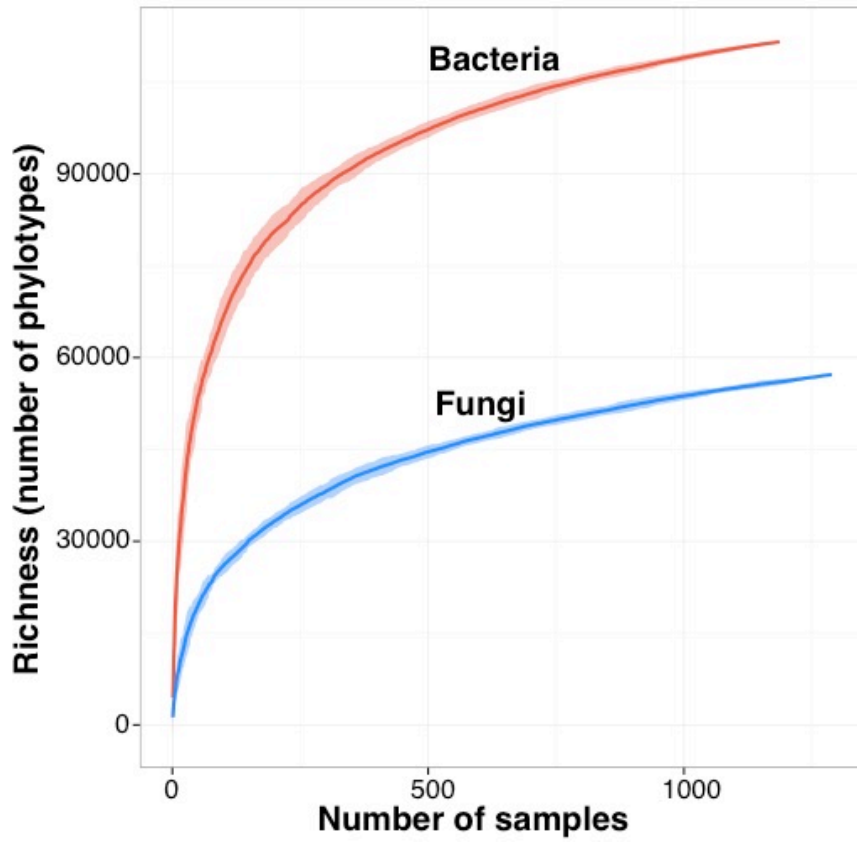
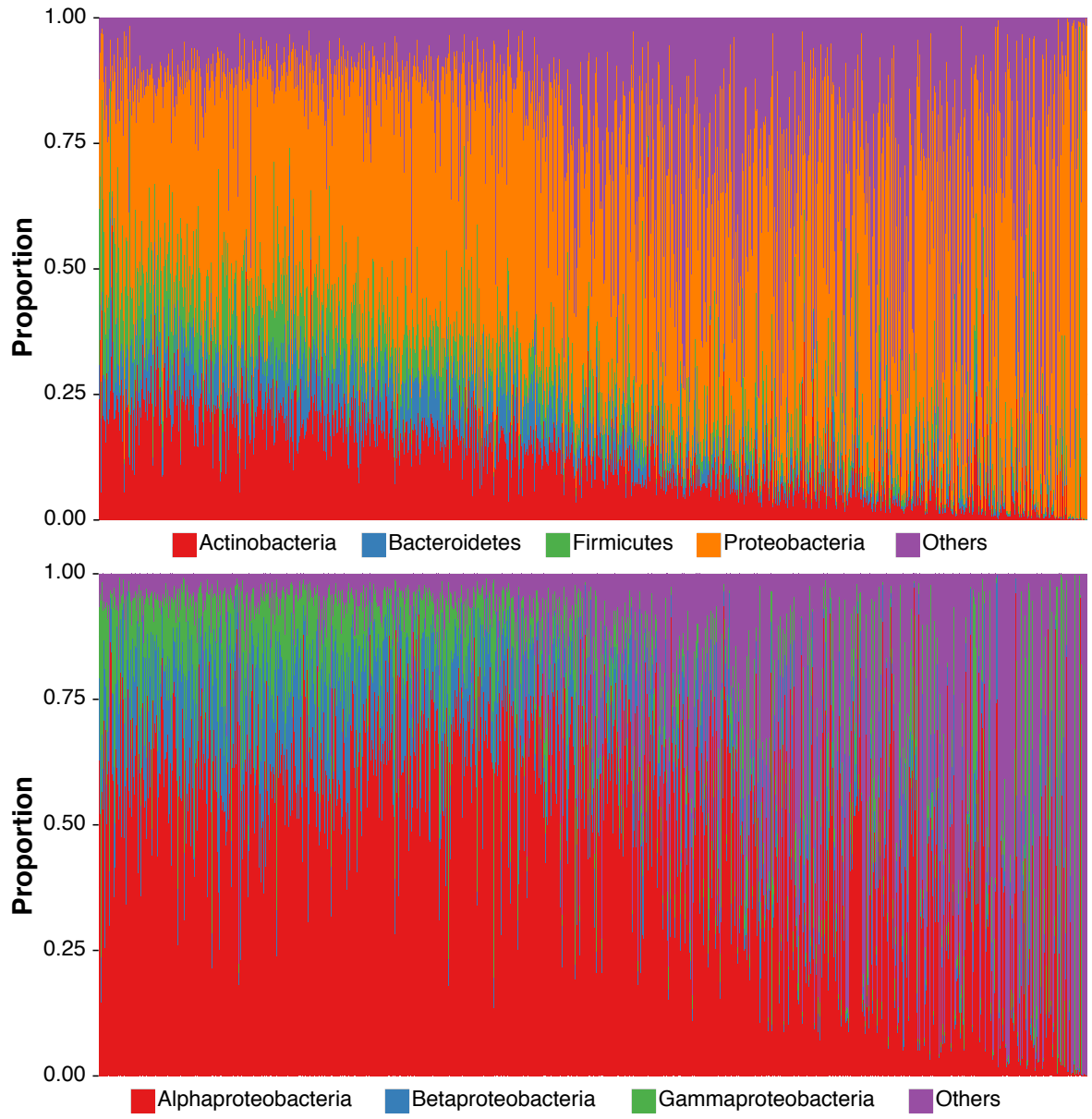
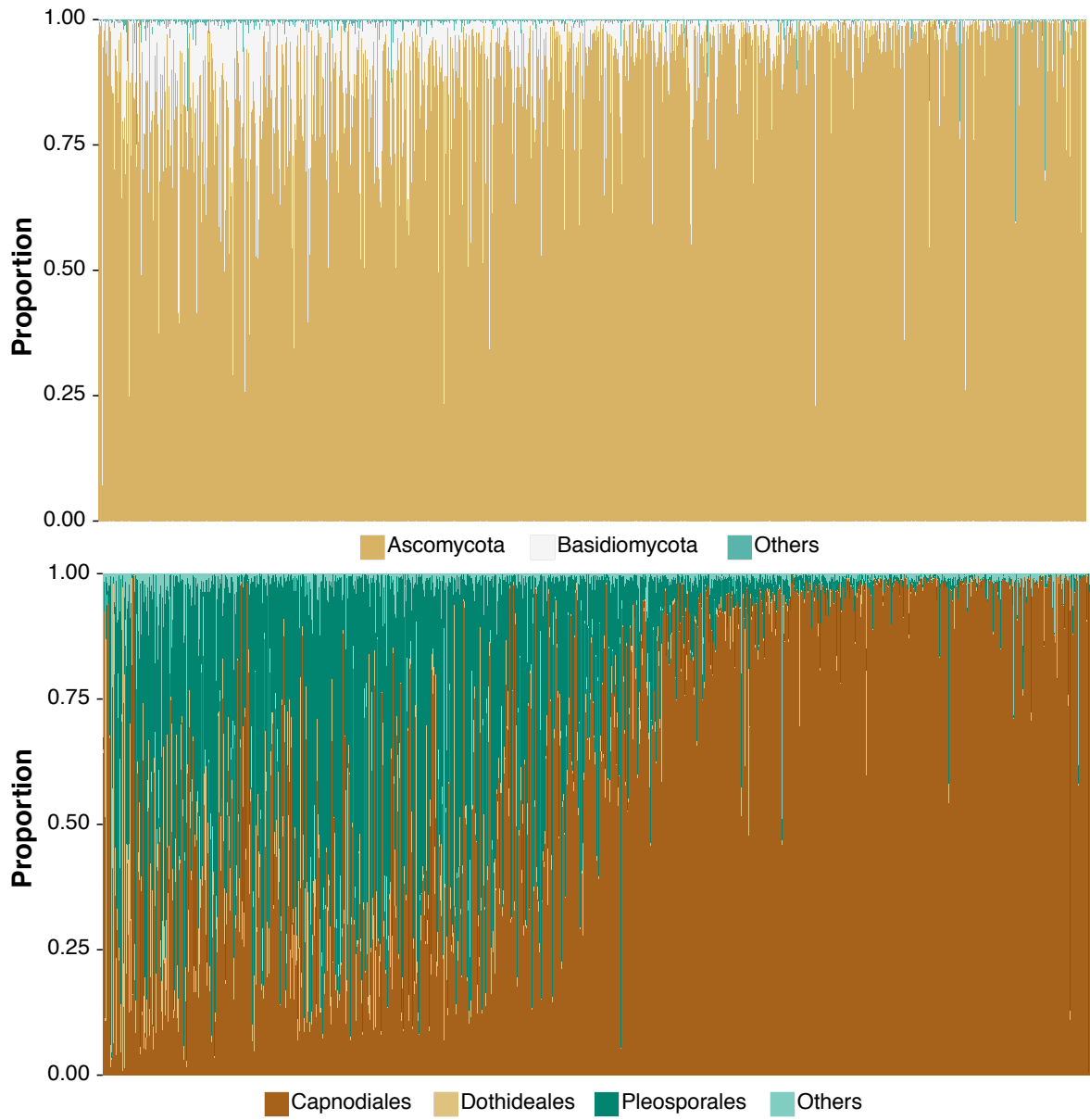


Fig. S3



**Fig. S4**





**Fig. S5**

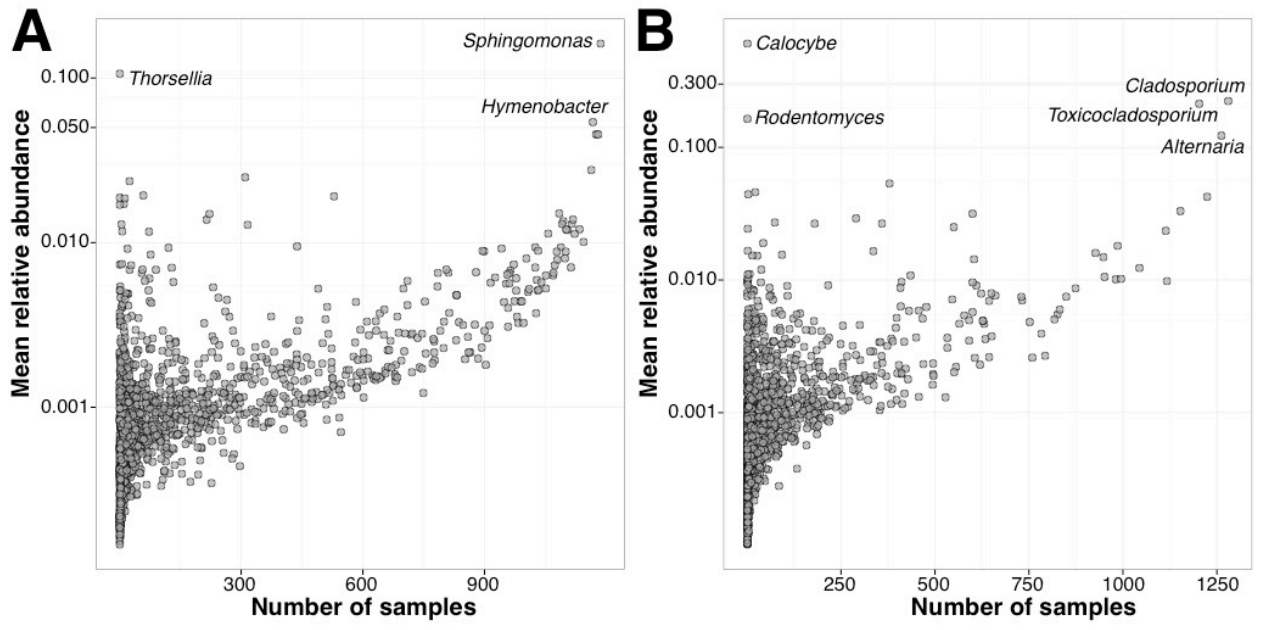


Fig. S6

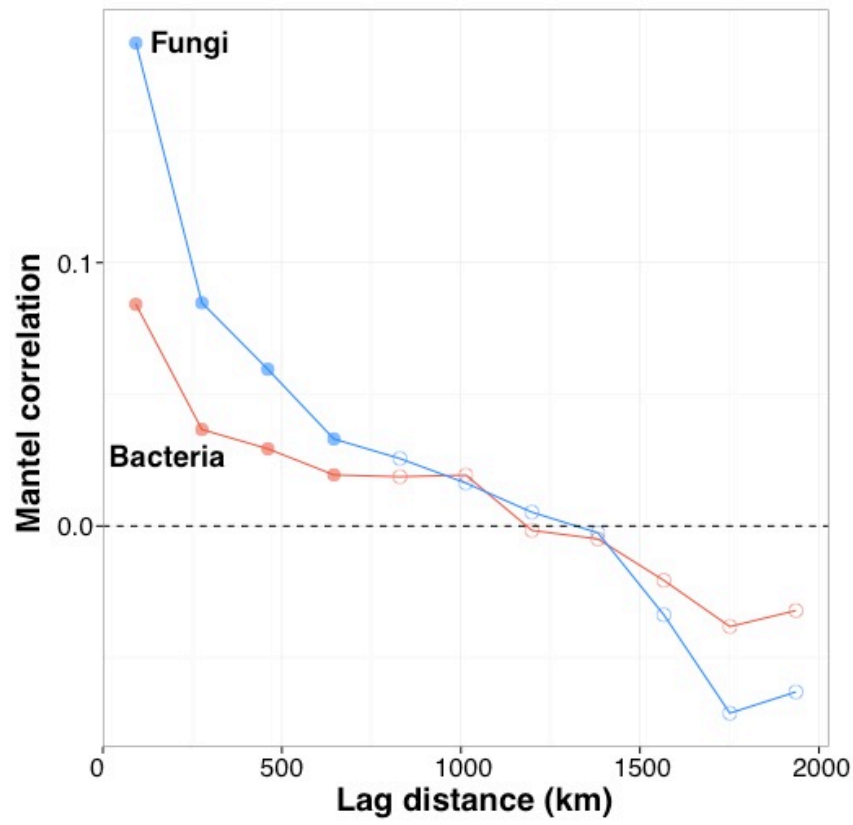


Fig. S7



**Fig. S8**

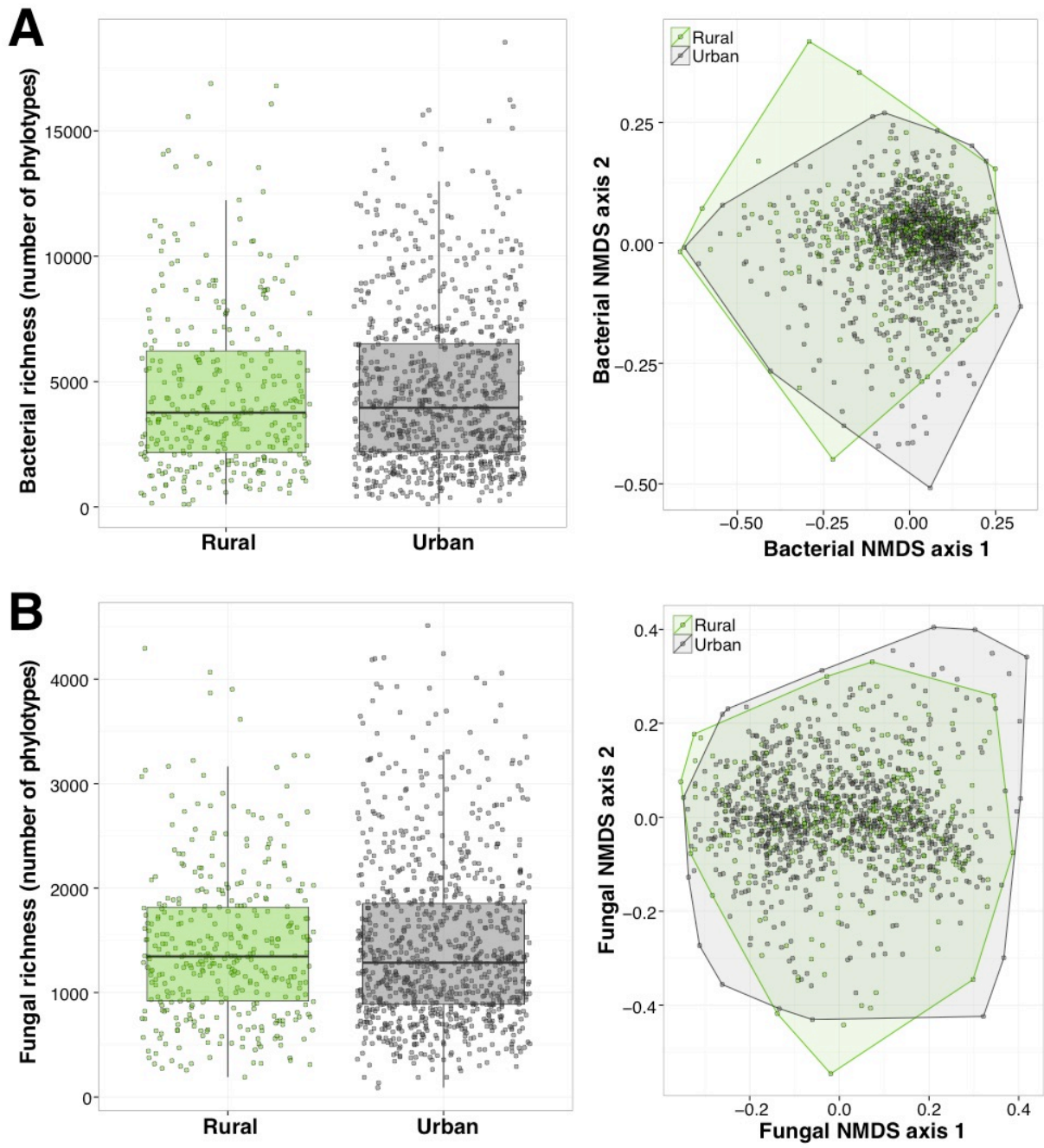


Fig. S9