Supplementary Figures

A novel caspase 8 selective small molecule potentiates TRAIL-induced cell death

Octavian Bucur^{1,4},*, Gabriel Gaidos^{2,*}, Achani Yatawara^{2,*}, Bodvael Pennarun¹, Chamila Rupasinghe², Jérémie Roux³, Stefan Andrei¹, Bingqian Guo², Alexandra Panaitiu², Maria Pellegrini², Dale F. Mierke^{2#}, Roya Khosravi-Far^{1,5#}

#Corresponding authors:

Roya Khosravi-Far, PhD, Department of Pathology, Beth Israel Deaconess Medical Center and Harvard Medical School, 330 Brookline Avenue, Boston, MA 02215, USA; E-mail: rkhosrav@gmail.com

Dale F. Mierke, PhD, Chair, Department of Chemistry, Dartmouth College, 6128 Burke Hall, Hanover, NH 03755, USA. E-mail: dale.mierke@dartmouth.edu

¹Department of Pathology, Beth Israel Deaconess Medical Center and Harvard Medical School, Boston, MA, USA;

²Department of Chemistry, Dartmouth College, Hanover, NH, USA;

³Department of Systems Biology, Harvard Medical School, Boston, MA, USA;

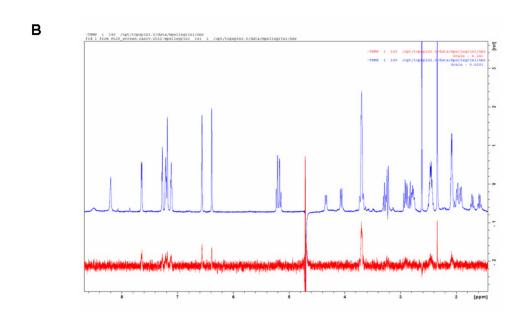
⁴Institute of Biochemistry of the Romanian Academy, Bucharest, Romania;

⁵Department of Biological and Biomedical Sciences Program, Harvard Medical School, Boston, MA, USA

^{*}These authors contributed equally to this work

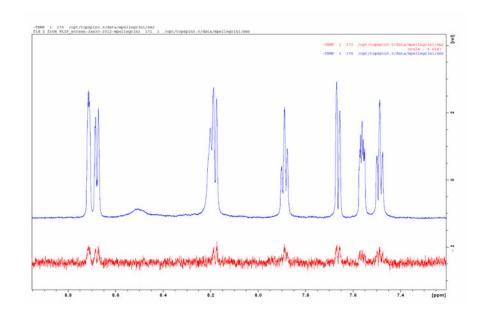
STD-NMR binding results

| Target | Compound 1 | Compound 2 | Compound 3 | Compound 4 |
|------------|--------------------------------|-------------------------------------|--------------|--------------------------------|
| | 800 | orong | and | |
| Caspase 8 | + v. small | broadening (see Figure 2) | - | - |
| cFLIP | - | - | - | - |
| DR5 | + small (see Suppl. Fig 1B) | - | + v.v. small | + small (see Suppl. Fig 1C) |
| FADD 4.8µM | t - . | - | - | N/A |
| FADD 5.7µM | - | + v.v. small | = | + almost nothing |
| GST (Ctrl) | - | - | N/A | - |



DR5 + C1
Blue: reference spectrum, C1 + DR5
Red: STD difference spectrum





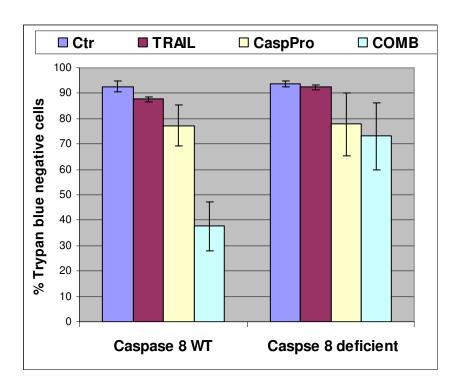
DR5 + C4
Blue: reference spectrum, C4 + DR5
Red: STD difference spectrum

Supplementary Figure 1. Binding potential of C1-C4 small molecules to Caspase 8, DR5, FADD and cFLIP

A. A summary of the binding potential of C1-C4 compounds to Caspase 8, DR5, FADD and cFLIP is presented. STD-NMR experiments were performed as described in the Material and Methods section of the manuscript. Saturation transfer from the protein to the bound small molecules causes a decrease in the intensity of the small molecule resonances, which is detected as residual signal in a difference spectrum. These experiments confirm that small molecule compound 2 directly binds caspase 8, as determined by the STD-NMR experiments, while compound 1 may also bind caspase 8, but with low affinity.

B.&C. Compounds C1 and C4 have some affinity towards binding DR5. Line broadening is not observed in the control experiments in the presence of GST or buffer alone;

"+ v. small" – very small broadening; "v. v. small" – very very small broadening; "-" – no broadening (no binding)



Supplementary Figure 2. Caspase 8 is required for CaspPro (C2, compound 2) - mediated potentiation of TRAIL-induced cell death

Caspase 8 WT and caspase 8 deficient Jurkat leukemic cells were treated with 7.5 ng/ml TRAIL, 80 μ M CaspPro and the combination for 18h in 1% FBS. Viability was measured by Trypan Blue dye exclusion method, using a TC10 Automated Cell Counter (Biorad, USA). The results represent the mean +/standard deviations (SDs) of 2 independent experiments, each one with multiple replicates. The results show that caspase 8 is required for CaspPro-mediated sensitization to TRAIL-induced cell death; We have previously confirmed the lack of expression of caspase 8 in caspase 8 deficient Jurkat cell line 8 .