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**Supporting information for article:**

**Raster-scanning serial protein crystallography using micro and nano-focused synchrotron beams**

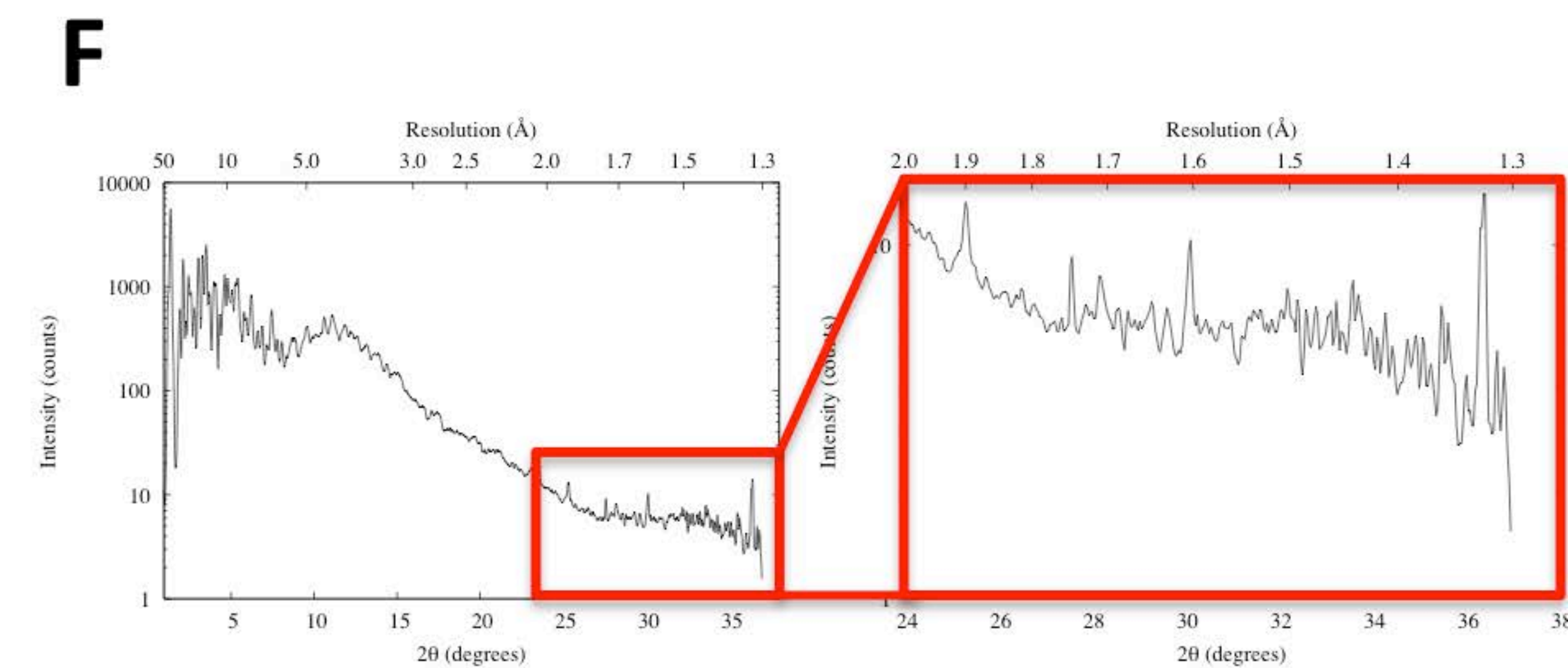
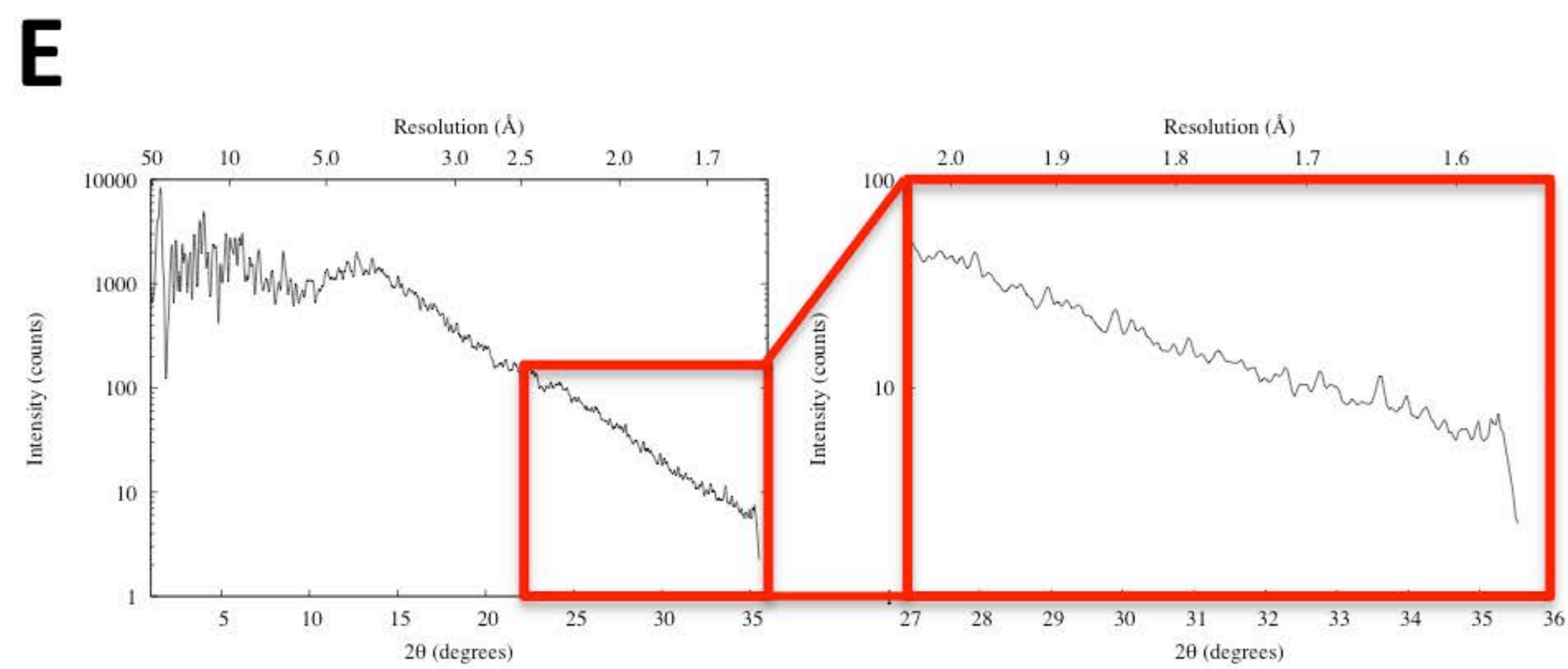
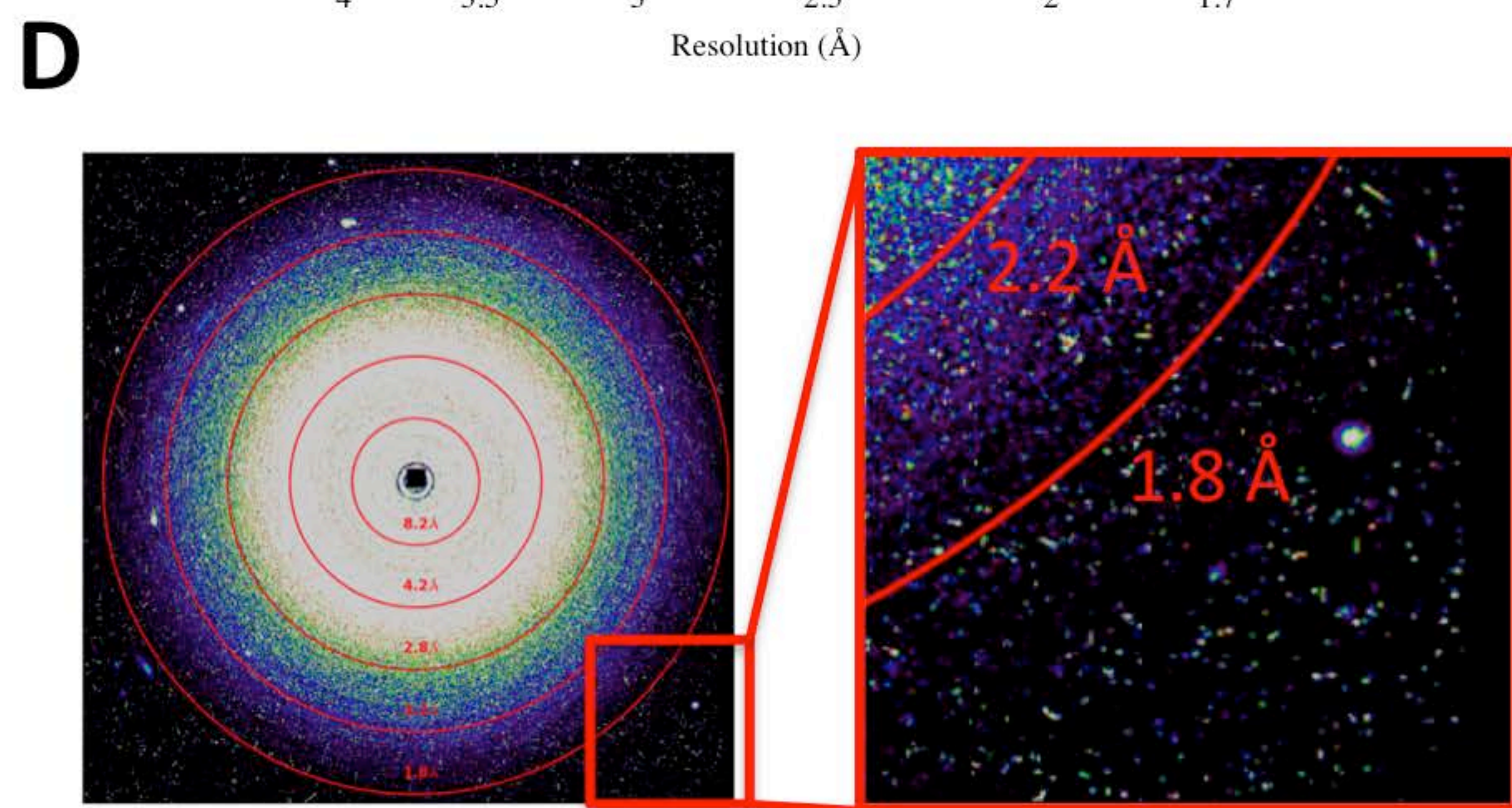
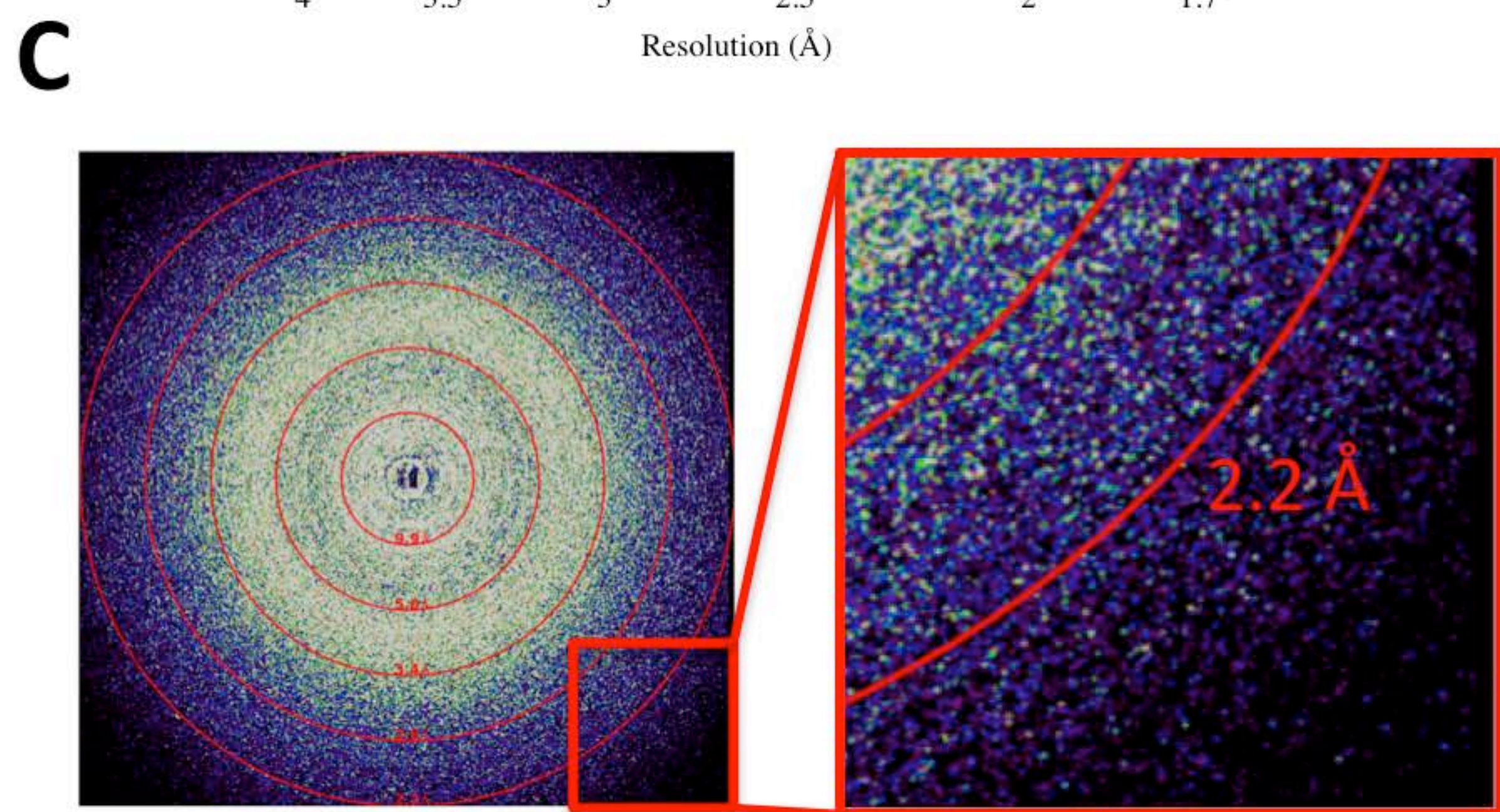
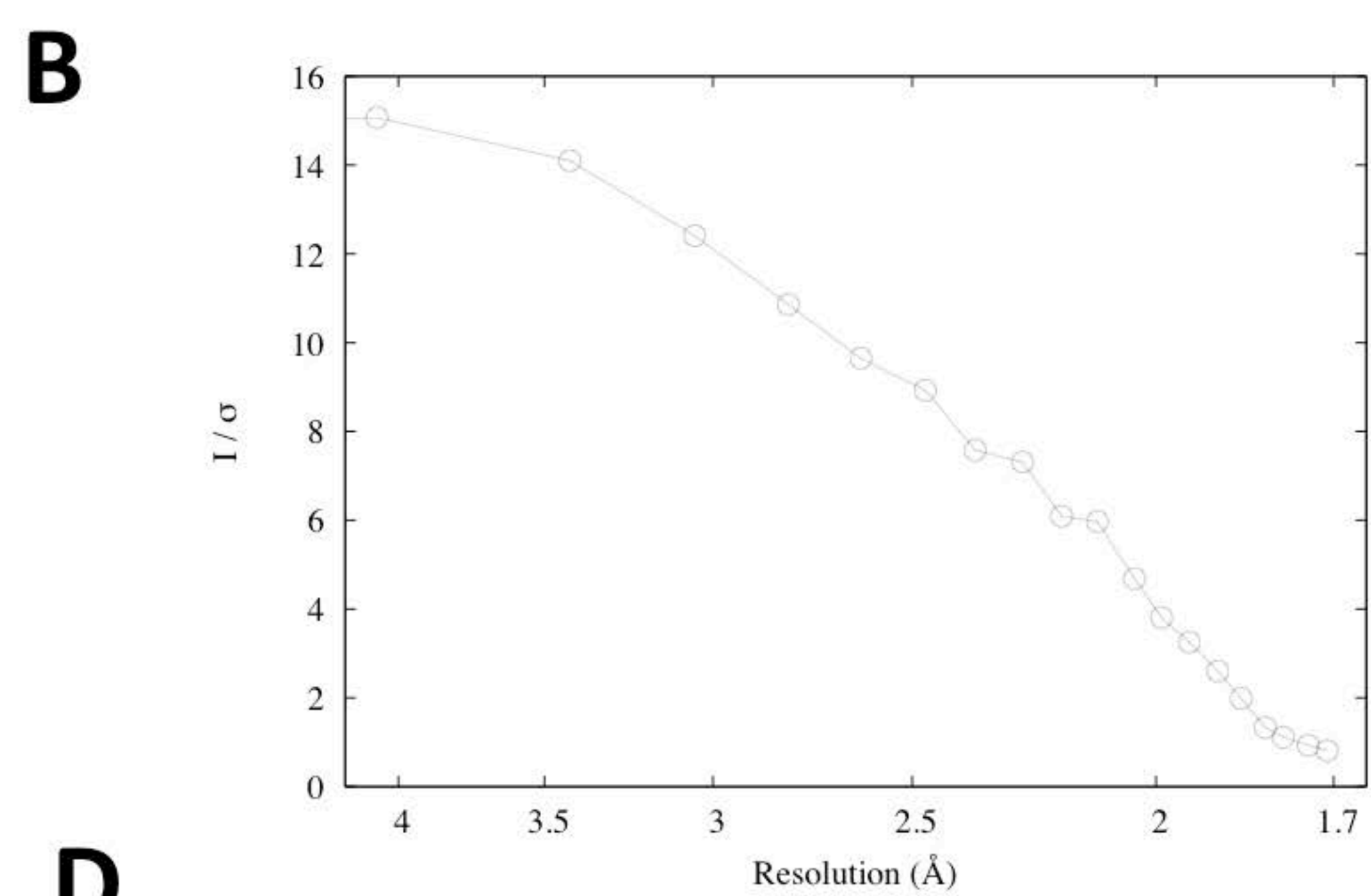
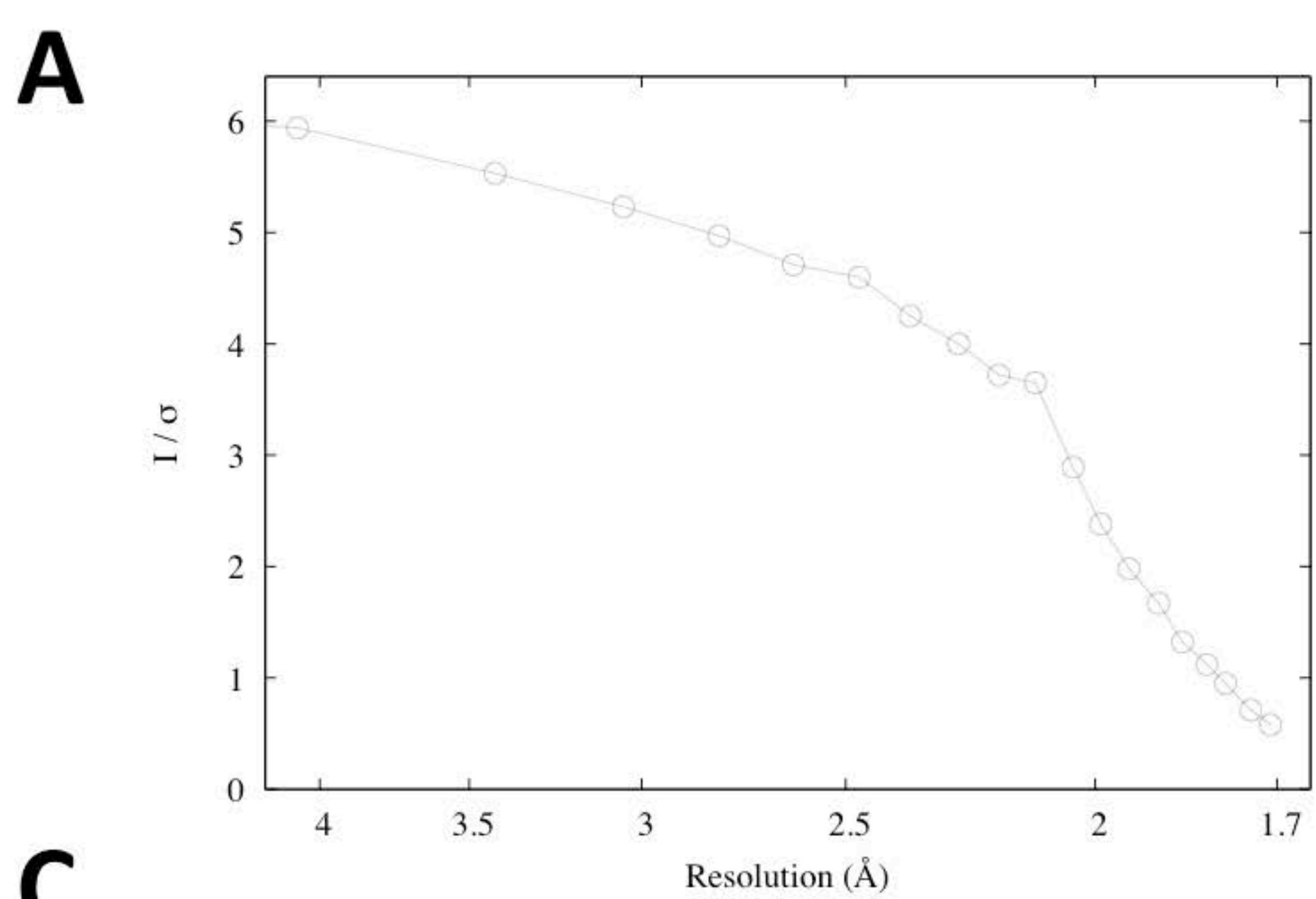
**Nicolas Coquelle, Aaron S. Brewster, Ulrike Kapp, Shilova Anastasya, Britta Weinhausen, Manfred Burghammer and Jacques-Philippe Colletier**

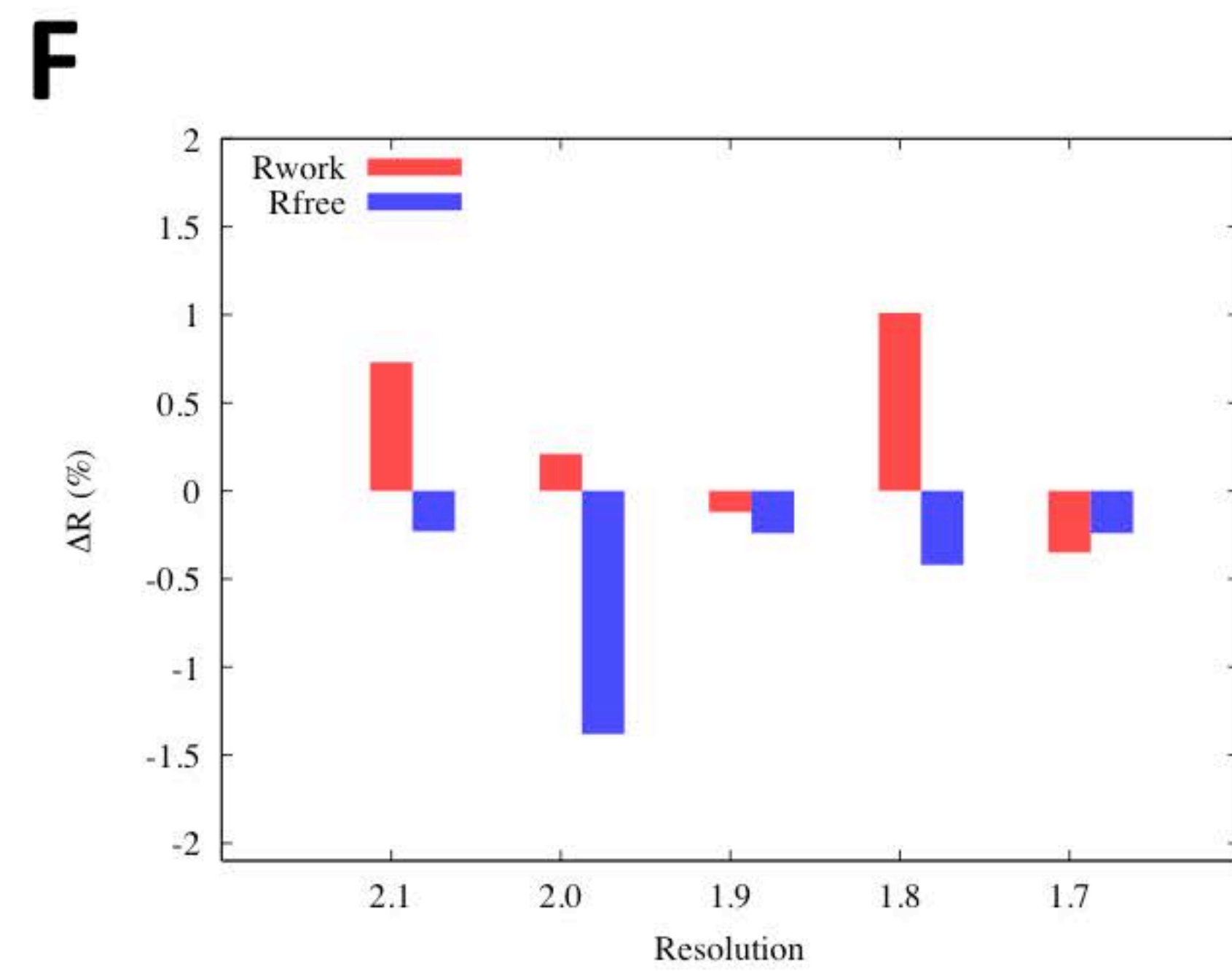
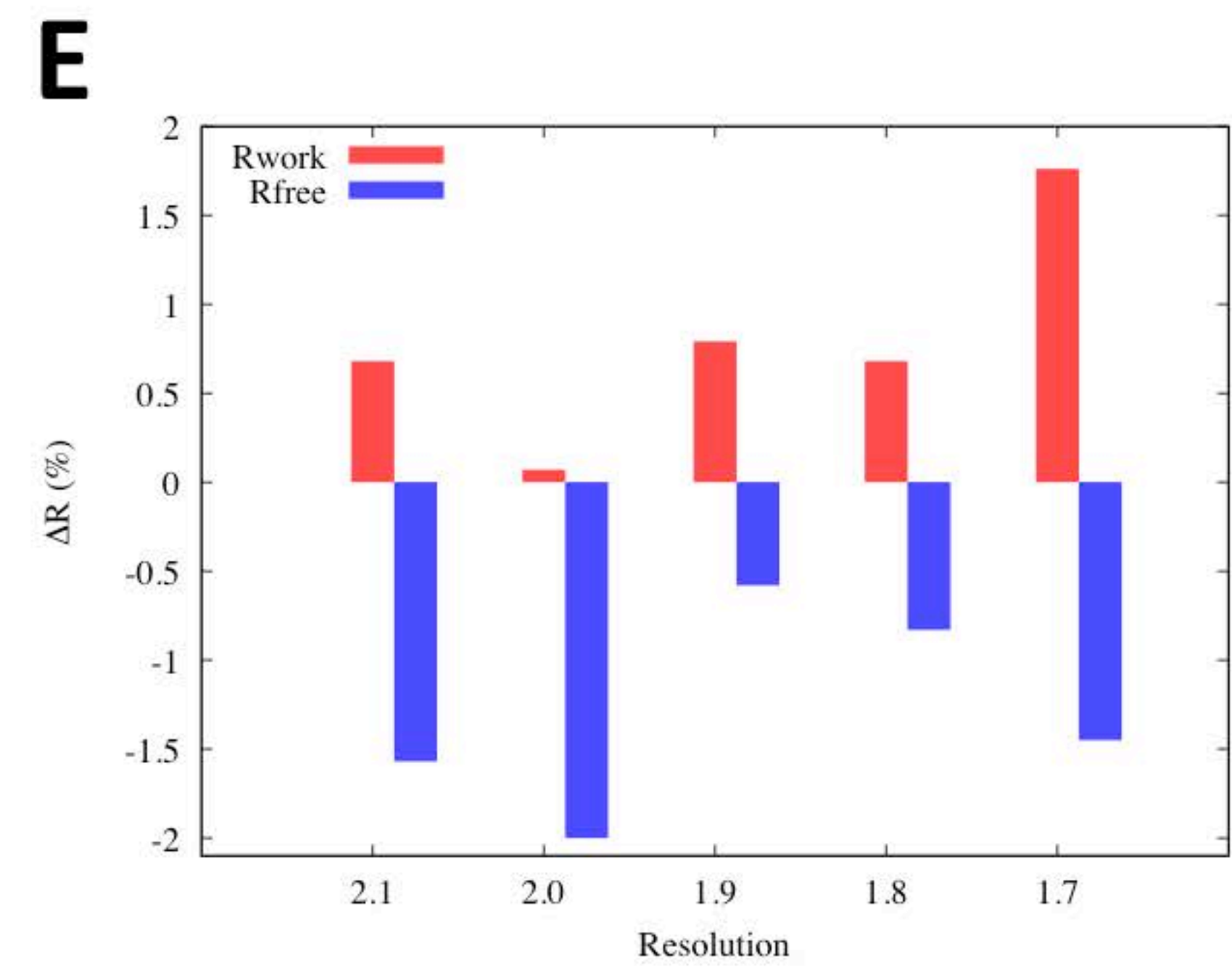
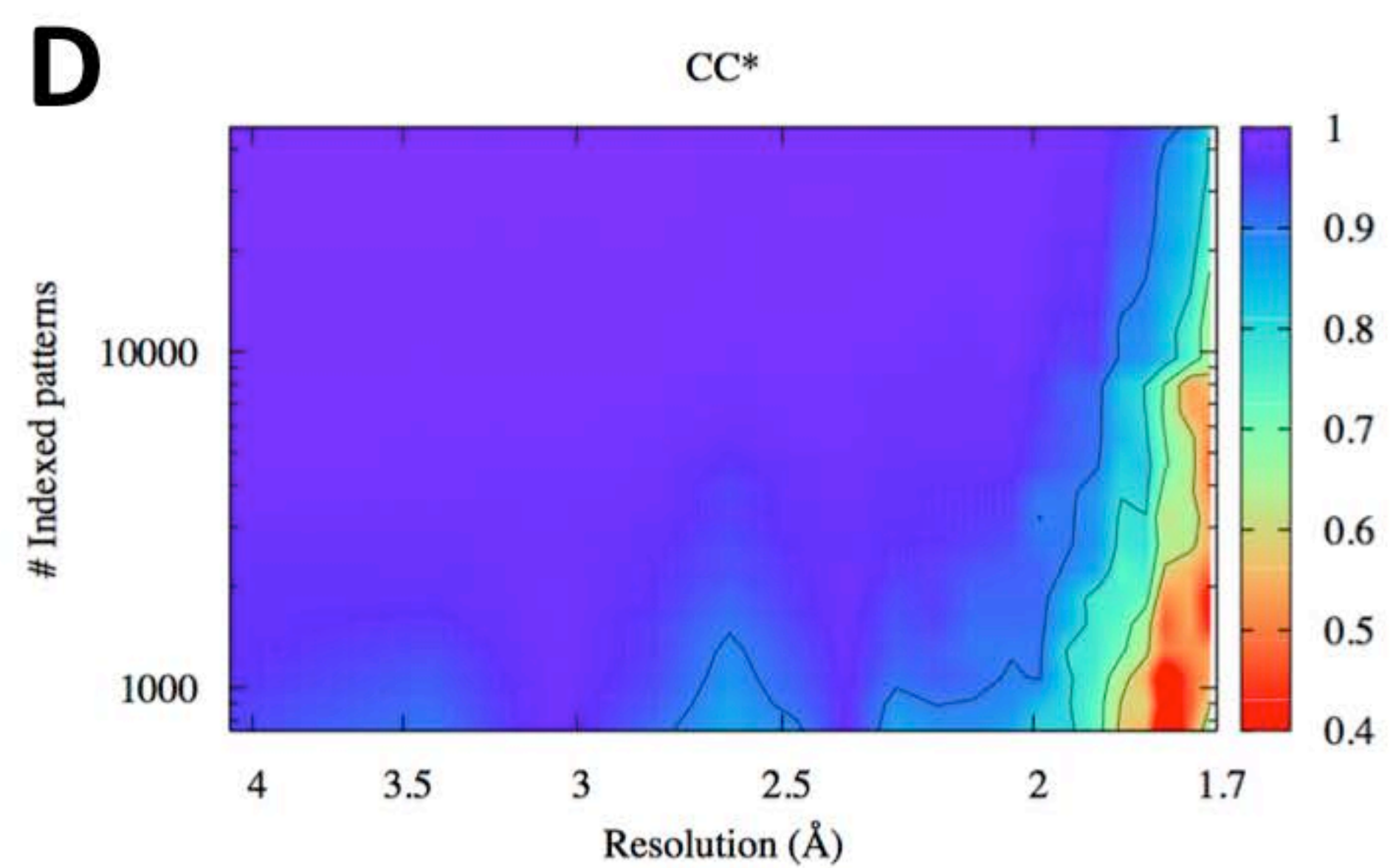
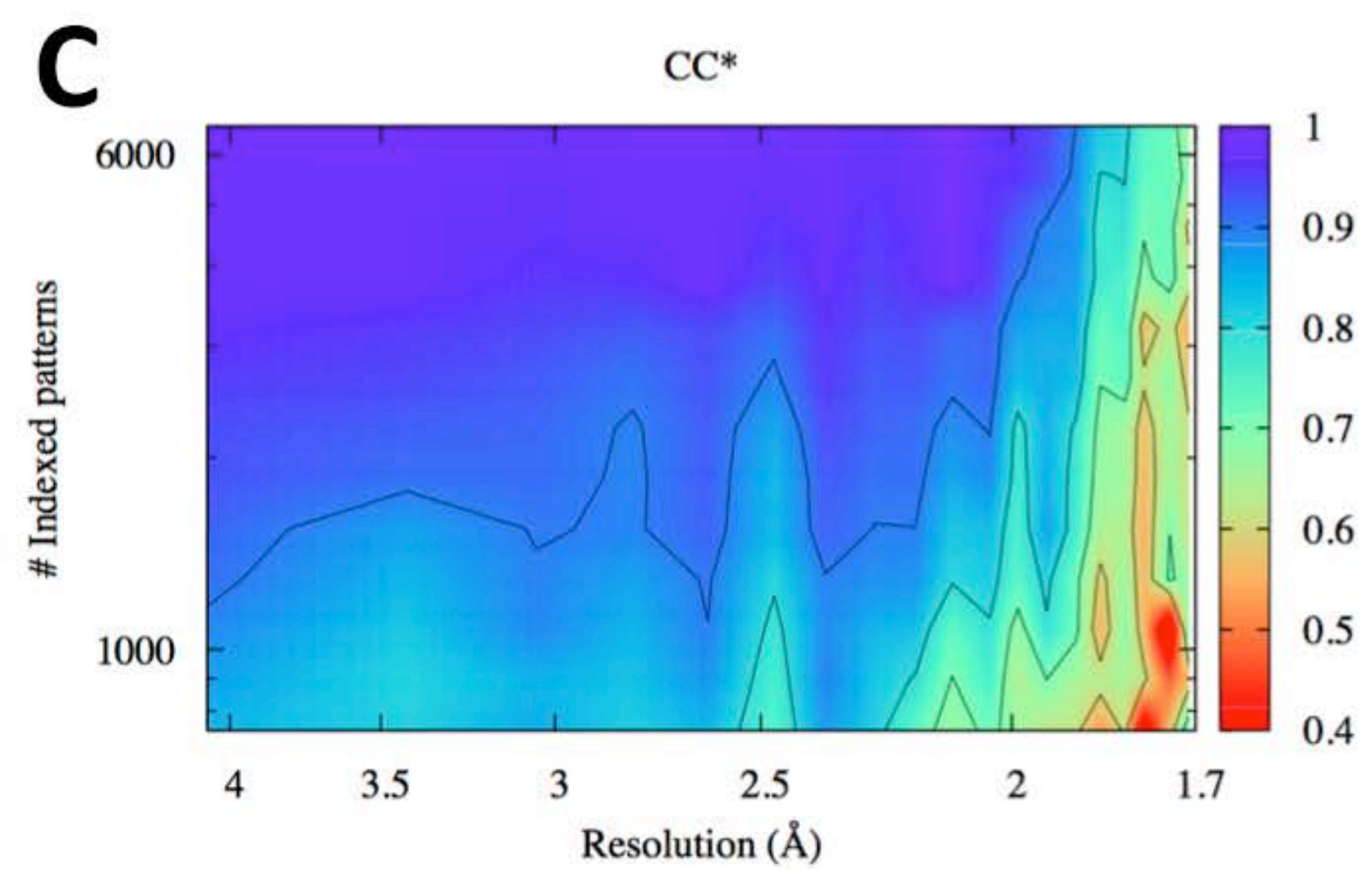
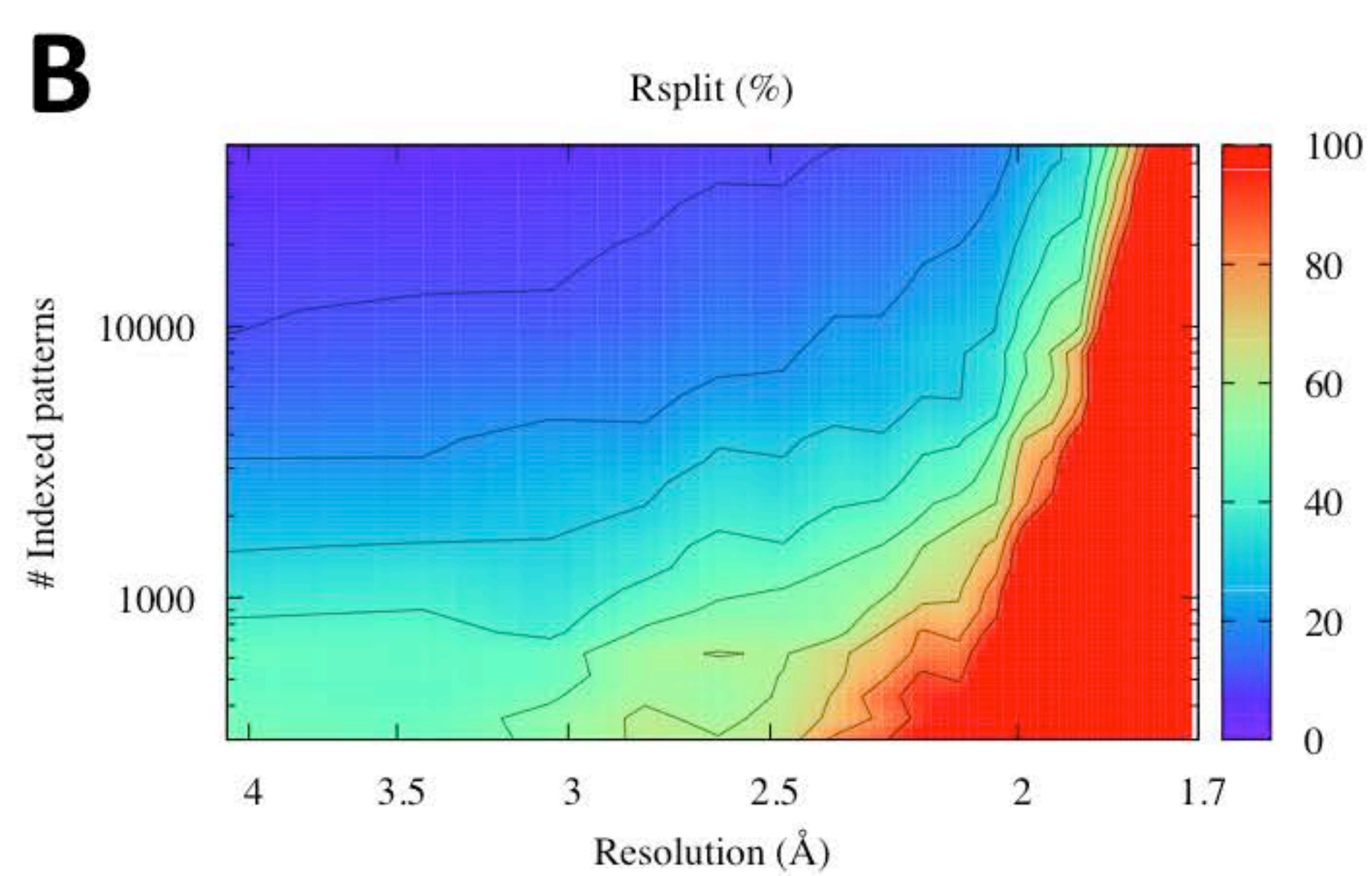
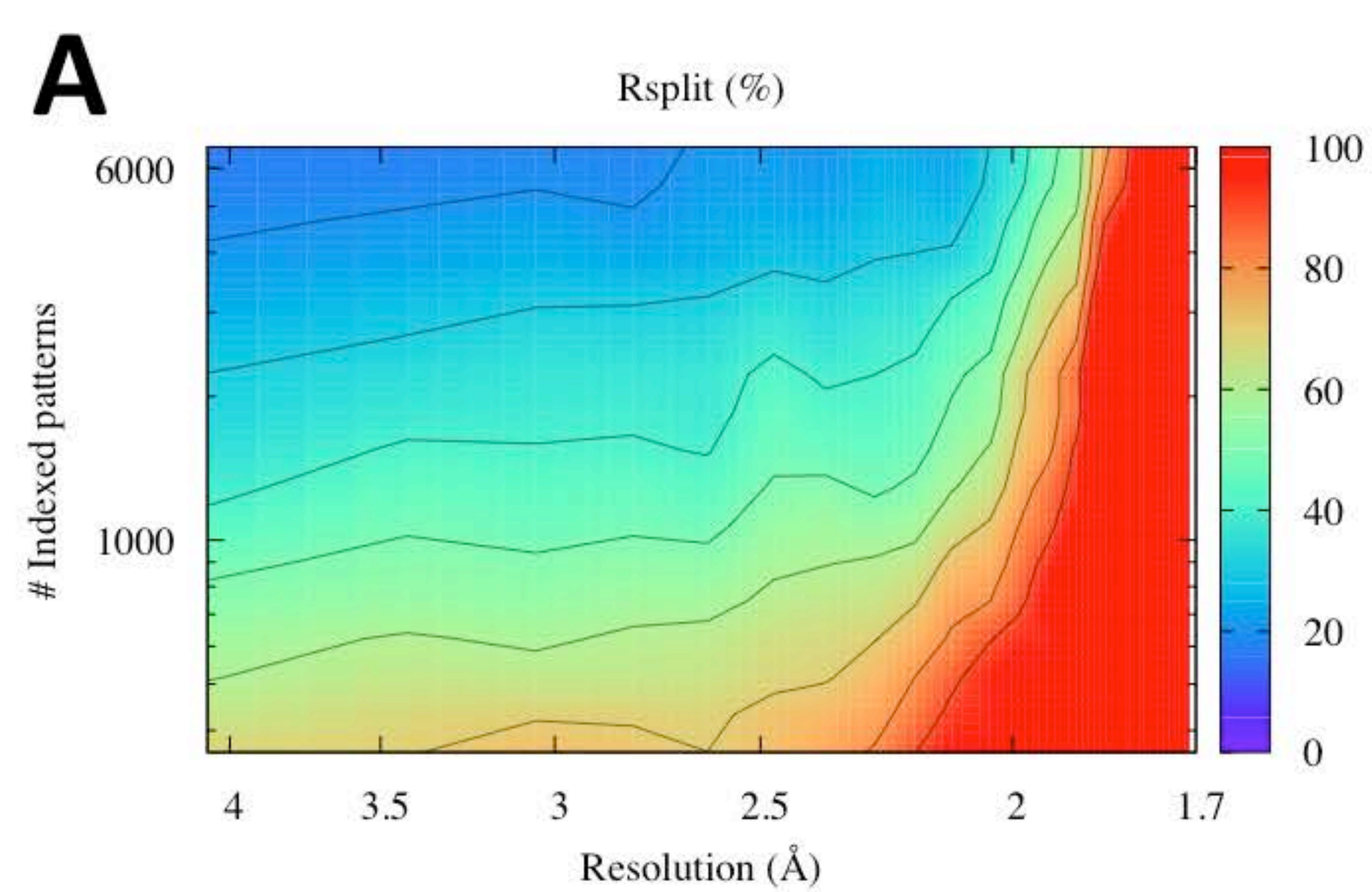
**Supplementary Figure 1: Overall quality and consistency of the progressive “micro” and “nano” dataset.** (A, B) Signal to noise ratio as a function of resolution in the progressive “micro” (A) and “nano” datasets (B), respectively. (C, D) Maximum projection of the patterns used to generate the “micro” (C) and “nano” datasets (D), respectively; for both, the right inset shows a zoom at high resolution. The detector resolution is 2.2/1.6 Å at the edge/corner for the “micro” (C), and 1.75/1.3 Å at the edge/corner for the “nano” (D). (E) 1D radial intensity profile of the “micro” maximum projection (C), integrated azimuthally with Fit2D (Hammersley *et al.*, 1996) over 360° and plotted as a function of 2theta (lower axis) and the resolution (upper axis); the inset shows a zoom in the 2.0-1.6 Å resolution range. (F) 1D radial intensity profile of the “nano” maximum projection (D), integrated azimuthally with Fit2D over 360° and plotted as a function of 2theta (lower axis) and the resolution (upper axis); the inset shows a zoom in the 2.0-1.3 Å resolution range.

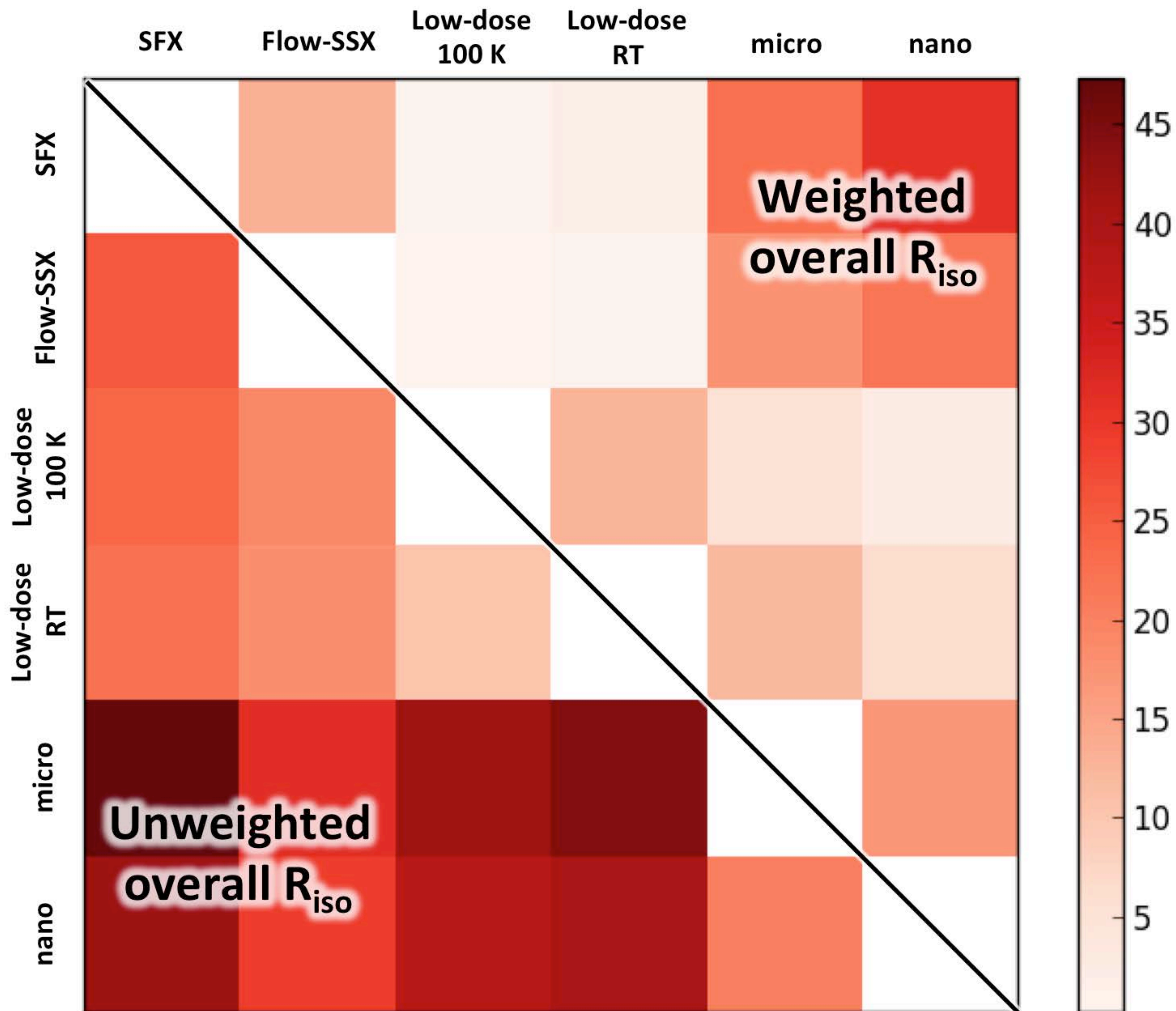
**Supplementary Figure 2: Determination of effective high-resolution cut-offs for the “micro” and “nano” dataset.** (A-D) Heat-maps of  $R_{\text{split}}$  (A, B) and  $CC^{1/2}$  (C, D) in the “micro” (A, C) and nano (B, D) datasets, respectively. These plots illustrate the convergence of our integrations. (E, F) Iterative-pair refinement was performed to validate the 1.7 Å resolution cut-offs of the progressive “micro” (E) and “nano” (F) datasets. In brief, the final model was refined at various resolutions ranging from 2.2 to 1.7 Å, with incremental steps of 0.1 Å. For each increment from resolution A to B, the difference in R-work and R-free values computed at resolution A, for models refined at resolution A and B; the differences in R-work and Rfree were plotted.

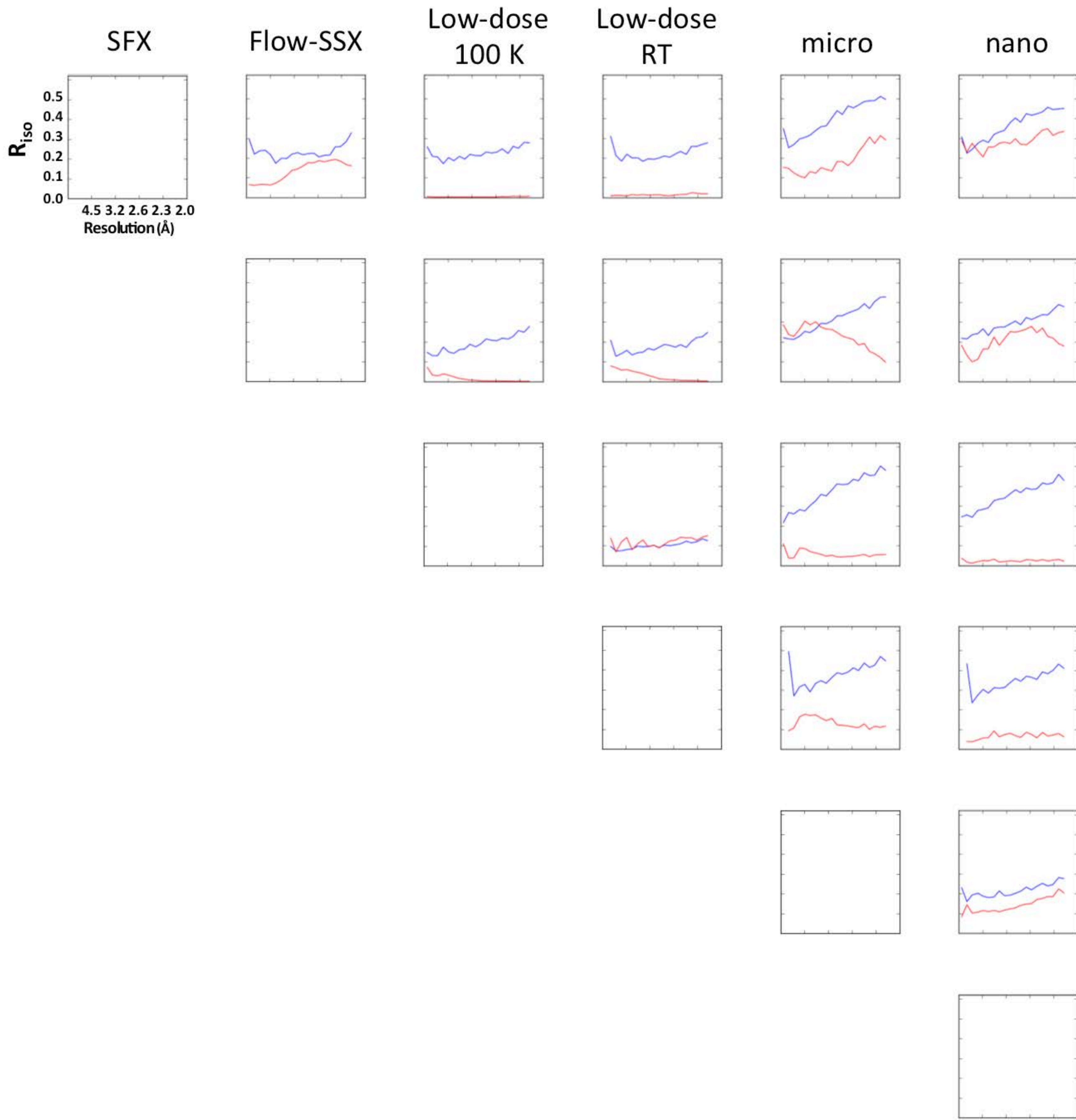
**Supplementary Figure 3: Overall  $R_{\text{iso}}$  between various HEWL datasets.** Unweighted and variance-weighted  $R_{\text{iso}}$  are shown in the lower left and upper right halves of the heat-map, respectively. Data were scaled using SCALEIT (Bailey *et al.*, 1994).

**Supplementary Figure 4:  $R_{\text{iso}}$  between various HEWL datasets plotted as a function of resolution.** Unweighted and variance-weighted  $R_{\text{iso}}$  are shown in the blue left and red, respectively. Data were scaled using SCALEIT (Bailey *et al.*, 1994).

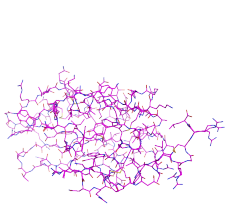
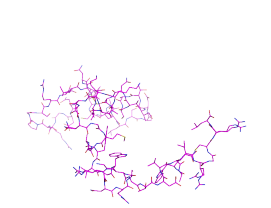
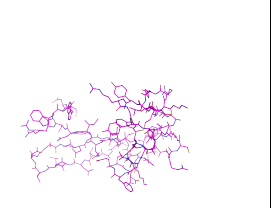
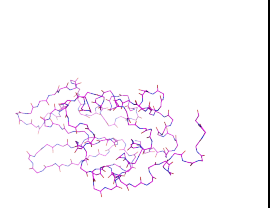
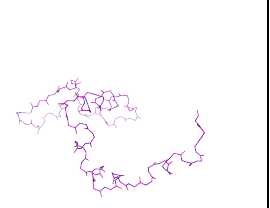
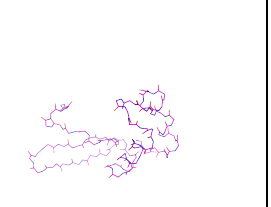








**Supplementary Table 1: Automated fitting of the HEWL sequence into experimental maps obtained by MR phasing of the “progressive nano” dataset with various starting models.**

	<b>Main and side chain atoms of residues 1-127</b>	<b>Main and side chain atoms of residues 1-65</b>	<b>Main and side chain atoms of residues 66-127</b>	<b>Main chain atoms of residues 1-127</b>	<b>Main chain atoms of residues 1-65</b>	<b>Main chain atoms of residues 65-127</b>
<b>Molecular Replacement model</b>						
<b>No. of mainchain / sidechain (of total 129)</b>	<b>126 / 126</b>	<b>117 / 117</b>	<b>125 / 125</b>	<b>125 / 125</b>	<b>127 / 117</b>	<b>126 / 126</b>
<b>R<sub>free</sub> / R<sub>work</sub></b>	<b>0.29 / 0.24</b>	<b>0.30 / 0.28</b>	<b>0.31 / 0.26</b>	<b>0.30 / 0.26</b>	<b>0.29 / 0.25</b>	<b>0.29 / 0.25</b>
<b>Map-to-model correlation coefficient</b>	<b>0.82</b>	<b>0.80</b>	<b>0.80</b>	<b>0.82</b>	<b>0.81</b>	<b>0.82</b>