## **Supporting Information**

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**Fig. S1.** Gating strategy for analysis of human monocytes. PBMC samples were stained with mAbs against human CD3/CD19/CD20/CD56/Vivid live/dead stain (dump channel), HLA-DR, CD14, CD16, CD11b, CD11c, and CD163. After gating on small and large cells in the SS-A/FS-A (A) and doublets (B and C), lineage-positive (FITC-dump) and dead cells (D) were excluded. From the live singlet cell population gated (D), FS-A was plotted against HLA-DR (E) and gates were drawn for monocytic lin-HLA-DR<sup>+</sup> cells based on isotype control (G). These cells were then analyzed for the expression of CD14 and CD16 (F) based on the isotype control (H). Each monocyte subset was then tested for expression of CD11b, CD11c, and CD163 relative to the isotype control (Fig. 1B).



Fig. 52. No difference in baseline peripheral CD3<sup>-</sup>NKp46<sup>+</sup>CD16<sup>+</sup> NK cell frequencies in responding vs. nonresponding patients with melanoma. PBMC samples were stained with mAbs against human CD19/CD20/Vivid live/dead stain (dump channel) and CD3, CD335 (NKp46), HLA-DR, CD14, and CD16. At baseline, there was no significant difference in peripheral frequencies of CD3<sup>-</sup>NKp46<sup>+</sup>CD16<sup>+</sup> NK cells in responding vs. nonresponding patients with melanoma.



**Fig. S3.** Gating strategy for sorting of human CD3<sup>+</sup>CD4<sup>+</sup> T cells according to CD25 expression. Before staining, CD4<sup>+</sup>CD3<sup>+</sup> T cells were enriched from PBMCs. T cells were then stained with mAbs against human CD3/CD4/CD25/AmCyan live/dead stain. After gating on SS-A/FS-A (A) and doublets (*B* and C), dead cells (*D*) were excluded. From the live singlet CD3<sup>+</sup>CD4<sup>+</sup> cell population gate (*D*), CD25 was plotted against CD4 and gates were drawn for CD4<sup>+</sup> cells with different expression level of CD25 (*E*). Based on this gating strategy, three different cell subpopulations were sorted: CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>bright</sup>, CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>int</sup>, and CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>int</sup>, and CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>int</sup>.



**Fig. S4.** CTLA4, Foxp3, and CD127 staining on sorted CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>bright</sup>, CD25<sup>int</sup>, or CD25<sup>neg</sup> T cells. Three sorted, live T-cell subpopulations with different expression levels of CD25—CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>bright</sup>, CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>int</sup>, and CD3<sup>+</sup>CD4<sup>+</sup>CD25<sup>neg</sup>—were stained with monoclonal antibodies against human CTLA-4, Foxp3, and CD127. After gating on cells in SS-A/FS-A (A), doublets (*B* and C), and CD3<sup>+</sup>CD4<sup>+</sup> (D), expression level of CD25 was checked once again (*E*, *H*, and *K*). Then, CTLA-4 or Foxp3 were plotted against CD127 (*F*, *G*, *I*, *J*, *L*, and *M*) and the expression of CTLA-4 (*P*), Foxp3 (*R*), and CD127 (*S*) was analyzed based on the isotypes controls (*N* and *O*).