# A Light-Driven Therapy of Pancreatic Adenocarcinoma Using Gold Nanorods-Based Nanocarriers for Co-Delivery of Doxorubicin and siRNA

Feng Yin,<sup>a#</sup> Chengbin Yang,<sup>a#</sup> Qianqian Wang,<sup>c</sup> Shuwen Zeng,<sup>a,f</sup> Rui Hu,<sup>a</sup> Guimiao Lin,<sup>e</sup> Jinglin Tian,<sup>e</sup> Siyi Hu,<sup>g</sup> Rong Feng Lan,<sup>h</sup> Ho Sup Yoon,<sup>b,i</sup> Fei Lu,<sup>c\*</sup> Kuan Wang,<sup>d\*</sup> and Ken-Tye Yong<sup>a\*</sup>

<sup>*a*</sup> School of Electrical and Electronic Engineering, Nanyang Technological University, Singapore 639798, Singapore Email: <u>ktyong@ntu.edu.sg</u>

<sup>b</sup> Division of Structural Biology & Biochemistry, School of Biological Sciences, Nanyang Technological University, Singapore 639798, Singapore

<sup>c</sup> Laboratory of Chemical Genetics, School of Chemical Biology and Biotechnology, Peking University Shenzhen Graduate School, Shenzhen, 518055, China Email: <u>lufei@pkusz.edu.cn</u>

<sup>d</sup> Nanomedicine Program and Institute of Biological Chemistry, Academia Sinica, Nankang, Taipei 115, Taiwan Email:<u>wangk@gate.sinica.edu.tw</u>

<sup>e</sup> The key lab of Biomedical Engineering and Research Institute of Uropoiesis and Reproduction, School of Medical Sciences, Shenzhen University, Shenzhen, 518060, China

<sup>*f*</sup> CINTRA CNRS/NTU/THALES, UMI 3288, Research Techno Plaza, 50 Nanyang Drive, Border X Block, Singapore, 637553

<sup>g</sup> School of Science, Changchun University of Science and Technology, Changchun, 130022, China

<sup>*h*</sup> Institute of Research and Continuing Education, Hong Kong Baptist University (Shenzhen), Shenzhen 518057, China

<sup>*i*</sup> Department of Genetic Engineering, College of Life Sciences, Kyung Hee University, Yongin-si Gyeonggi-do, 446-701, Republic of Korea

<sup>#</sup> These authors contributed equally to this work

## **Supplementary Information**

### **Materials and Methods**

#### Paraffin section and histology

For histological experiments, tumor tissues were collected on Day 25 and the samples are fixed with 4% buffered formalin-saline at room temperature for 24 hours. Next, these fixed tissue samples were embedded in paraffin blocks and they were sliced into 4  $\mu$ m-thick sections using a microtome. The paraffin sections were then mounted on glass slides for hematoxylin and eosin (H&E) staining. The H&E staining slices were examined using an Olympus BX51 light microscope.

## **Figures and Figure legends**



Supplementary Figure S1: The exclusion of the off-target effect of siRNA. Panc-1 cells were treated with Blank, AuNRs, AuNRs/scramble siRNA, AuNRs/K-Ras siRNA, Lipo2000, Lipo-scramble siRNA and Lipo-K-Ras siRNA. (A) After 48 hours treatment, mRNA relative expression levels detected by RT-PCR. (B) Cell viability test of Panc-1 cells after 12, 24, 48 and 72 hours treatment. Data are presented as the mean±SEM of triplicate experiments.\*, P < 0.05, \*\*, P < 0.01 vs blank, AuNRs and Lipofectamine.



Supplementary Figure S2: Statistical data of cell cycle analysis. The change of S phase were monitored by fluorescence activated cell sorting (FACS) analysis and quantified by GraphPad Prism (version 5) software. Data are presented as the mean $\pm$ SEM of triplicate experiments. \*\*, P < 0.01 vs blank; \*, P < 0.05, the comparison between single delivery group and co-delivery group.



**Supplementary Figure S3:** Temperature dependence of AuNRs/DOX/siRNA nanoplex under 665 nm light irradiation. AuNRs/DOX/siRNA nanoplex was dispersed in PBS and the temperatures of the samples were measured by Thermometer (T208, Digitron). Within the first 30 minutes of the 665 nm light exposure, a 7 °C increment was observed for the AuNRs/DOX/siRNA nanoplex (from 26 °C to 33 °C). On the other hand, only 1 °C increment was observed for PBS buffer solution (from 26°C to 27 °C). Error bars represent SEMs for triplicate experiments.



В



**Supplementary Figure S4**: Comparison of the antitumor activity of AuNRs and AuNRs/DOX/siRNA nanoplex in a Panc-1 xenograft animal model. In both cases, the experimental subjects were exposed to 665 nm light. (A) Representative images of mouse and tumor tissues treated with (1) PBS, (2) free AuNRs, (3) combination of AuNRs and 665 nm light irradiation or (4) combination of AuNRs/DOX/siRNA and

665 nm light irradiation. The mice treated with combination of AuNRs/DOX/siRNA and 665 nm light exhibits the strongest tumor inhibition rate when compare to the other 3 control groups. (**B**) Relative changes in tumor volume versus time of mice treated by PBS, free AuNRs, free AuNRs and 665 nm light or AuNRs/DOX/siRNA and 665 nm light, respectively. Relative tumor volume was defined as  $(V-V_0)/V_0$ , where V and V<sub>0</sub> indicate the tumor volume on a particular day and day 0, respectively. Error bars represent SEMs for triplicate data. Mean tumor volumes were analyzed using one-way ANOVA. \*, P < 0.05, \*\*, P < 0.01 (n=5-7 tumors).



mRNA level of K-Ras in tumors

**Supplementary Figure S5:** K-Ras mRNA relative expression levels in pancreatic tumors were detected by RT-PCR. Values are means  $\pm$  SEM, n = 3; \*\*, P < 0.01 vs Control, AuNRs and siRNA.



**Supplementary Figure S6:** Representative histological images from the major tumors of the mice treated by AuNRs/DOX/siRNA with or without 665 nm light, comparing with the untreated control at day 25. The arrows indicate the AuNRs nanoplex.



**Supplementary Figure S7:** The body weights of nude mice were measured every other day treated by PBS, AuNRs, AuNRs with 665 nm light, DOX, AuNRs/DOX, AuNRs/siRNA, AuNRs/DOX/siRNA or AuNRs/DOX/siRNA with 665 nm light. Values are means  $\pm$  SEM, n = 5.

Name of nanoplex	Grafted	Zeta	Hydrodynamic	
	polymers	potential	size	
AuNRs		+33.33 mV	24.90±2.15 nm	
AuNRs/PSS	PSS	-33.92 mV	29.50±1.83 nm	
AuNRs/PSS/PAH	PSS & PAH	+35.50 mV	34.52±1.67 nm	
AuNRs/PSS/DOX/PAH	PSS & PAH	+37.88 mV	59.20±1.78 nm	
AuNRs/PSS/PAH/siRNA	PSS & PAH	+29.90 mV	44.73±3.57 nm	
AuNRs/PSS/DOX/PAH/siRNA	PSS & PAH	+31.34 mV	67.59±1.85 nm	

**Supplementary Table S1:** Zeta potential and hydrodynamic size of different AuNRs-based nanoplex.

Supplementary Table S2: Transfection efficiency and average fluorescence intensity.

Name of reagent	Transfection		Average		SSC signal	FSC signal
	efficiency		fluorescence		intensity	intensity
	(DOX	and	intensity (DOX a	and	$(10^4)$	$(10^4)$
	siRNA <sup>FAM</sup> )		siRNA <sup>FAM</sup> )			
Blank	0.35%	and	$0.05 \times 10^3$ a	and	6.61	5.08
	0.69%		$0.15 \times 10^{3}$			
AuNRs	0.72%	and	$0.15 \times 10^3$ a	und	7.34	6.02
	0.54%		$0.08 \times 10^{3}$			
siRNA <sup>FAM</sup>	0.51%	and	$0.13 \times 10^3$ at	and	6.69	5.12
	3.30%		$0.12 \times 10^{3}$			
DOX	94.6%	and	$8.93 \times 10^3$ at	and	7.02	5.43
	2.57%		$0.09 \times 10^{3}$			
AuNRs/DOX/	94.3%	and	$8.31 \times 10^3$ a	und	7.65	6.09
siRNA <sup>FAM</sup>	85.8%		$3.13 \times 10^{3}$			
Lipo- siRNA <sup>FAM</sup>	3.52%	and	$0.56 \times 10^3$ a	und	7.57	5.91
	87.2%		$3.20 \times 10^{3}$			