474 Figure 4. Immunoblot analysis of SDS solubilised total bacterial protein probed with 475 monoclonal anti-multiple banded antigen antibody. Of 26 individual transformation 476 experiments only one strain (U6) showed altered mobility of the MBA following transposon 477 mutagenesis (+Tn). Representative gel of several experiments is shown showing 3 strains of 478 serovar (SV) 3 and 2 strains of serovar 6. 479 Figure 5. Growth kinetics for *U. parvum* parent strain (HPA5) compared to membrane 480 protein disruption (UU390::mTn) or DEADbox RNA-helicase gene disruption (UU582::mTn) 481 when incubated at 37°C (A.), 33°C (B.), or 25°C (C.). Strains were titrated out in a 10-fold 482 dilution series and growth measured at time points indicated by urease conversion of urea to 483 ammonium ions. Ureaplasma growth is shown as colour (pH indicator) changing units per 484 ml. Mean and standard deviation of dilutions performed in triplicate. Results were consistent 485 through three repeated experiments. 486 Figure 6. Serum killing (A) and immunoblot analysis using human high titre seropositive 487 serum (B) for parental U. urealyticum strain W11 and following successful transposon 488 mutagenesis (+Tn). Serum killing of increases significantly following 1 h challenge with 489 human serum containing anti-Ureaplasma antibodies and analysis of this serum shows the 490 serum-sensitive transposon mutated strain has lost a 41 kDa band that was immunoreactive 491 with the challenging serum. Bar graph shows mean +/- SEM of experiments performed in 492 triplicate. Representative immunoblot from three repeat experiments shown. 493 494 Supplementary Figure 1. Further PCR mapping of successful transposon-mutagenised 495 strains of Ureaplasma parvum and W11 (Ureaplasma urealyticum) using the methods outlined 496 for Figure 1. Presence of the gentamicin resistance gene (primers 1518F to 1898R) was only 497 found in transposon mutated strains. PCR probing identified one of three DFK1 serovar 1 498 strains (three different experiments) and HPA78 to have some of the transposase (Tnase) gene

499 integrated into the genome. Expected amplicon size is indicated to the right of the figure.

500 Supplementary Figure 2. PCR amplification of U. parvum genes (as designated by ATCC 501 strain 700970 nomenclature) UU390, UU450, UU520, and UU582 using primers designed 502 within the coding region of these genes. An additional primer set amplifying the end of gene 503 UU187 and the beginning of UU188 (including the 31 bp promoter region) was also included. 504 Non-mutagenised HPA5 and U6 serovar 3 U. parvum are included as controls. Single clones 505 with disrupted genes are shown for each primer set as well as 7 additional clones successfully 506 transformed with the mini-transposon to show specificity. 507 Supplementary figure 3: Growth of HPA5 in triplicate at 25, 33, and 37°C as measured by 508 colour change of Ureaplasma selective medium from Mycoplasma Experience ltd. 180 µl 509 were placed in each well and 20 µl of prototype laboratory U. parvum strain HPA5

510 was inoculated into the first well (in triplicate). Filter tips were changed between each

511 dilution (transfer of 20 μ l) across the plate. The plates were then incubated in separate

512 incubators at the listed temperatures and the growth documented at various time points.

513 These images were taken at the 48 hr time point for one of the repeats of the experiment.

514 Supplementary Figure 4. Sequence and detail of the pMT85 vector.

515



Supplementary Figure 1. Further PCR mapping of successful transposon-mutagenised strains *of Ureaplasma parvum* and W11 (*Ureaplasma urealyticum*) using the methods outlined for Figure 1. Presence of the gentamicin resistance gene (primers 1518F to 1898R) was only found in transposon mutated strains. PCR probing identified one of three DFK1 serovar 1 strains (three different experiments) and HPA78 to have some of the transposase (Tnase) gene integrated into the genome.. Expected amplicon size is indicated to the right of the figure.

supplementary figure 2



Supplementary Figure 2. PCR amplification of U.parvum genes (as designated by ATCC strain 700970, nomenclature) UU390, UU450, UU520, and UU582 using primers designed within the coding region of these genes. An additional primer set applifying the end of gene UU187 and the beginning of UU188 (including the 32 bp intragenic region) was also included. Non-mutagenised HPA5 and U6 serovar 3 U.parvum are included as controls. Single clones with disrupted genes are shown for each primer set as well as 7 additional clones successfully transformed with the mini-transposon to show specificity.

supplementary figure 3



Supplementary figure 3: Growth of HPA5 in triplicate at 25, 33, and 37oC as measured by colour change of Ureaplasma selective medium from Mycoplasma Experience ltd. 180 μl were placed in each well and 20 μl of prototype laboratory U. parvum strain HPA5 was inoculated into the first well (in triplicate). Filter tips were changed between each dilution (transfer of 20 μl) across the plate. The plates were then incubated in separate incubators at the listed temperatures and the growth documented at various time points. These images were taken at the 48 hr time point for one of the repeats of the experiment.

Supplementary Figure 4. Sequence and detail of the pMT85 vector

Legend: Inverted repeat 1 : 1-26 Inverted repeat 2: 3412-3438

Gentamicin resistance: 1036 - 2475 479 a.a. bifunctional AAC/APH (AAC(6'): 6'-aminoglycoside N-acetyltransferase and APH(2'): 2''-aminoglycoside phosphotransferase [synthetic Mycoplasma genitalium JCVI-1.0] Sequence ID: <u>gi|166079093|gb|ABY79711.1|</u>Length: 495 (100% match)

transposase 3538-4710 390 a.a.

IS256, transposase [Enterococcus faecalis V583] Sequence ID: <u>gi|29374776|ref|NP_813928.1|</u>Length: 390 (100% match)

>pMT85gen

GATAAAGTCCGTATAATTGTGTAAAAACCCATAGCTTTGGACACACACTAGTACGGA TCCACCCGCAATTACTGTGAGTTAGCTCACTCATTAGGCACCCCAGGCTTTACACTT TATACTTCCGGCTCGTATATTGTGTGGGAATTGTGAGCGGATAACAATTTCACACAGG AAACAGCTATGACCTTGATTACGGAATTCACGGCCGGGGGGGCCACCCCACCAATTG ACGCGGCCGCAACTCTAGAGGATTCATCGGCCGTCGTTGCCTGGTTTCCGGCACCAG AAGCGGTGCCGGAAAGCTGGCTGGAGTGCGATCTTCCTGAGGCCGATACTGTCGTCG TCCCCTCAAACTGGCAGATGCACGGTTACGATGCGCCCATCTACACCAACGTGACCT ATCCCATTACGGTCAATCCGCCGTTTGTTCCCACGGAGAATCCGACGGGTTGTTACT GCGCTCACATTTATATGCTGACTGAAAGCTGGCTACAGGAAGGCCAGACGCGAATTA TTTTTGATGGCTAGAAGCTTTAGGATGAATGGATTTATTCTTCAAGAAAATACATCA ATTTTGATAAGTAGAAATGGTAAAAACATTGTATAGCATTTTACACAGGAGTCTGGA CTTGACTGAGTTTATGGAAGAAGTTTTAATTGATGATAATATGGTTTTTGATATTGA TAATTTAAAAGGATTTCTTAATGATACCAGTTCAATTTGGGTTTATAGCTAAAGAAA ATAATAAAATTATAGGATTTGCATATTGCTATACACTTTTAAGACCTGATGGAAAAA CAATGTTTTATTTACACTCAATAGGAATGTTACCTAACTATCAAGACAAAGGTTATG **GTTCAAAATTATTATCTTTTATTAAGGAATATTCTAAAGAGATTGGTTGTTCTGAAA** TGTTTTTAATAACTGATAAAGGTAATCCTAGAGCTTGCCATGTATATGAAAAATTAG **GTGGTAAAAATGATTATAAAGATGAAATAGTATATGTATATGATTATGAAAAAGGTA GATAAATAA**ATGAATATAGTTGAAAATGAAATATGTATAAGAACTTTAATAGATGAT GATTTTCCTTTGATGTTAAAATGGTTAACTGATGAAAGAGTATTAGAATTTTATGGT **GGTAGAGATAAAAATATACATTAGAATCATTAAAAAAACATTATACAGAGCCTTGG** GAAGATGAAGTTTTTAGAGTAATTATTGAATATAACAATGTTCCTATTGGATATGGA CAAATATATAAAATGTATGATGAGTTATATACTGATTATCCATTATCCAAAAACTGAT GAGATAGTCTATGGTATGGATCAATTTATAGGAGAGCCAAATTATTGGAGTAAAGGA ATTGGTACAAGATATATTAAATTGATTTTTGAATTTTTGAAAAAAGAAAGAAATGCT AATGCAGTTATTTTAGACCCTCATAAAAATAATCCAAGAGCAATAAGGGCATACCAA AAATCTGGTTTTAGAATTATTGAAGATTTGCCAGAACATGAATTACACGAGGGCAAA AAAGAAGATTGTTATTTAATGGAATATAGATATGATGATAATGCCACAAATGTTAAG GCAATGAAATATTTAATTGAGCATTACTTTGATAATTTCAAAGTAGATAGTATTGAA ATAATCGGTAGTGGTTATGATAGTGTGGCATATTTAGTTAATAATGAATACATTTTT AAAACAAAATTTAGTACTAATAAGAAAAAAGGTTATGCAAAAGAAAAAGCAATATAT AATTTTTTAAATACAAATTTAGAAACTAATGTAAAAATTCCTAATATTGAATATTCG TATATTAGTGATGAATTATCTATACTAGGTTATAAAGAAATTAAAGGAACTTTTTTA ACACCAGAAATTTATTCTACTATGTCAGAAGAAGAACAAAATTTGTTAAAACGAGAT