

Supplementary Information

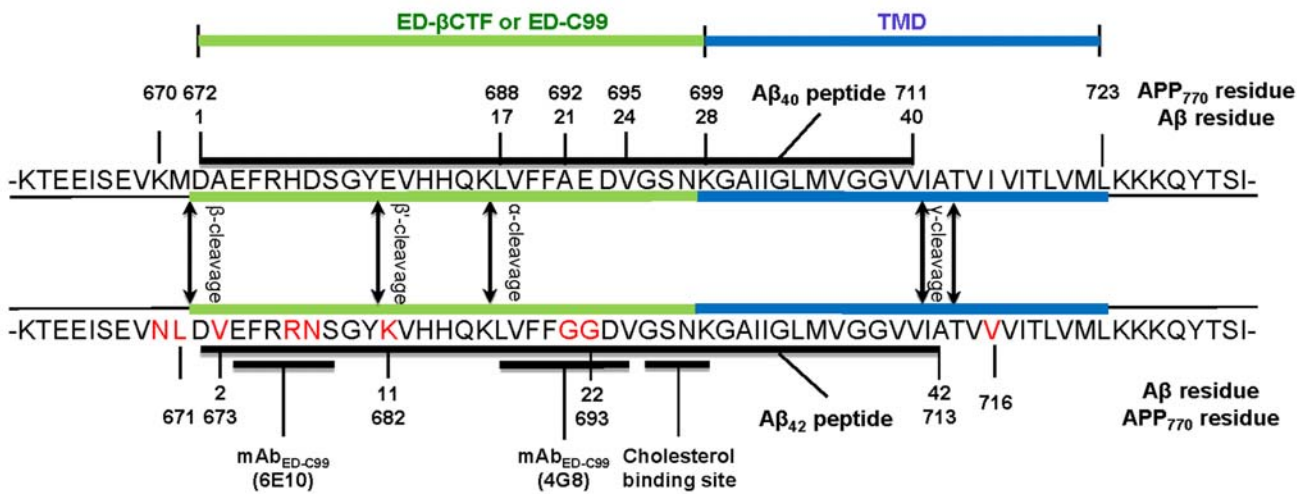
Specific antibody binding to the APP₆₇₂₋₆₉₉ region shifts APP processing from α - to β -cleavage

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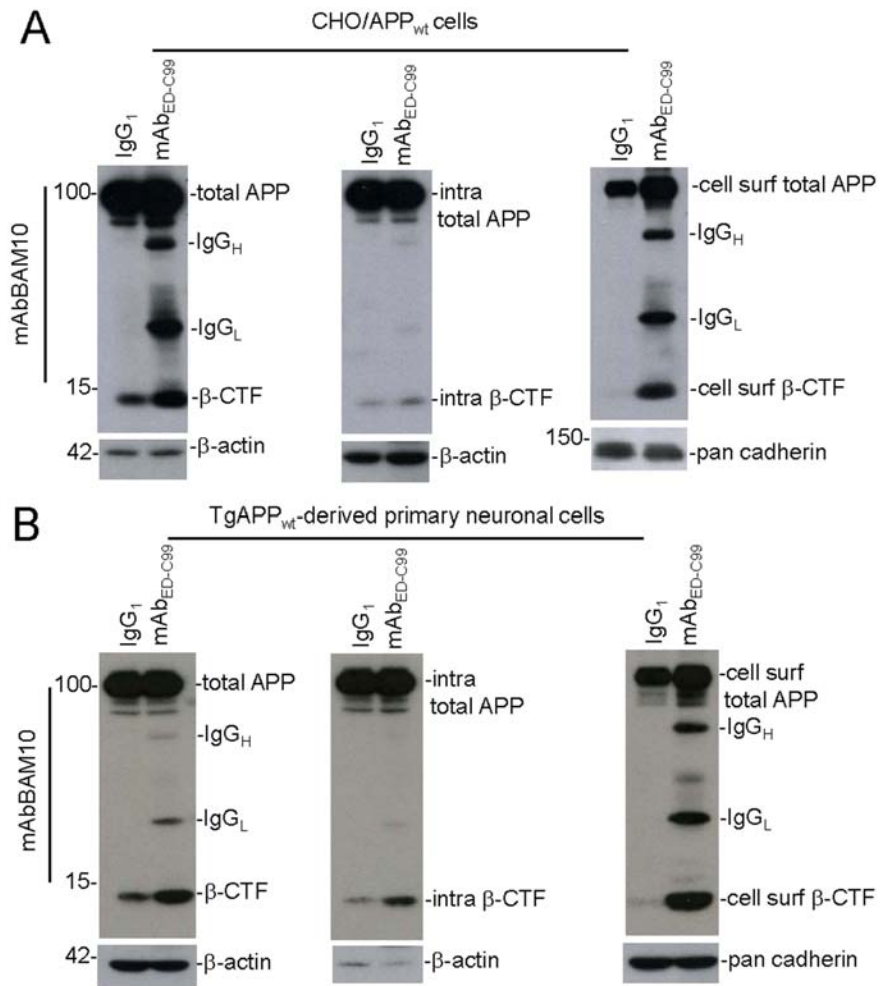
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Running title: Binding on APP₆₇₂₋₆₉₉ promotes APP β -cleavage

⁸These authors contributed equally to this work.

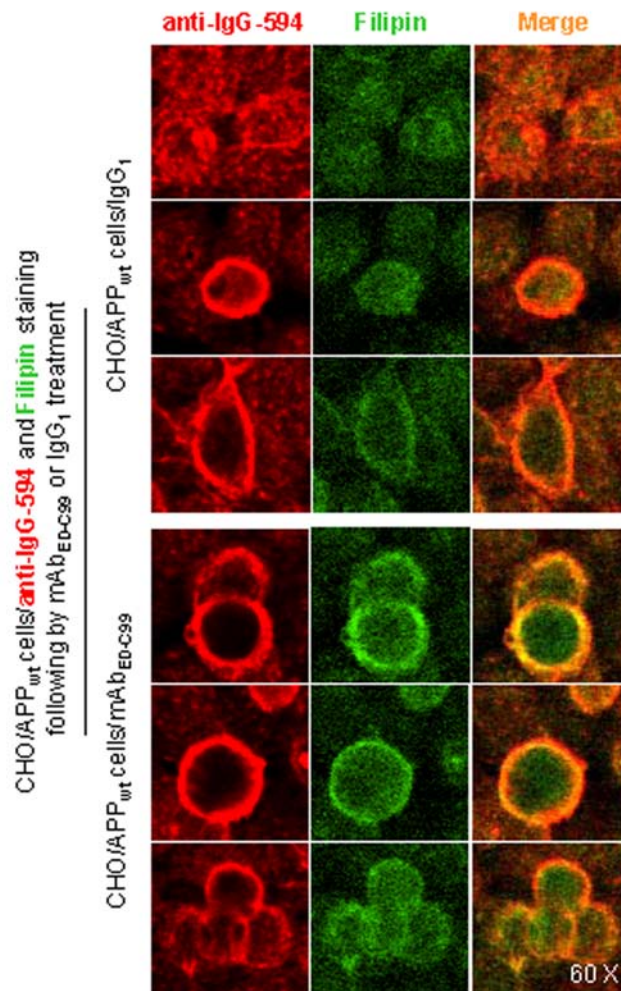


Supplemental Figure S1 A schematic diagram of the β -amyloid precursor protein (APP) proteolytic cleavage sites and mutations. The sites of α -, β - and γ -secretase-mediated cleavage are indicated with arrows. The trans-membrane domain (TMD) of APP is highlighted in blue, and ectodomain (ECD) of APP in green. γ -cleavage produces a pool of A β fragments that vary in length and hydrophobicity. The mutations (indicated in red) around α - and β -cleavage sites either increase the total A β production (670/671, 682, 692), alter A β biophysical properties (677, 678, 693), or affect the A β spectrum in both quantitative and qualitative ways (673, 716).



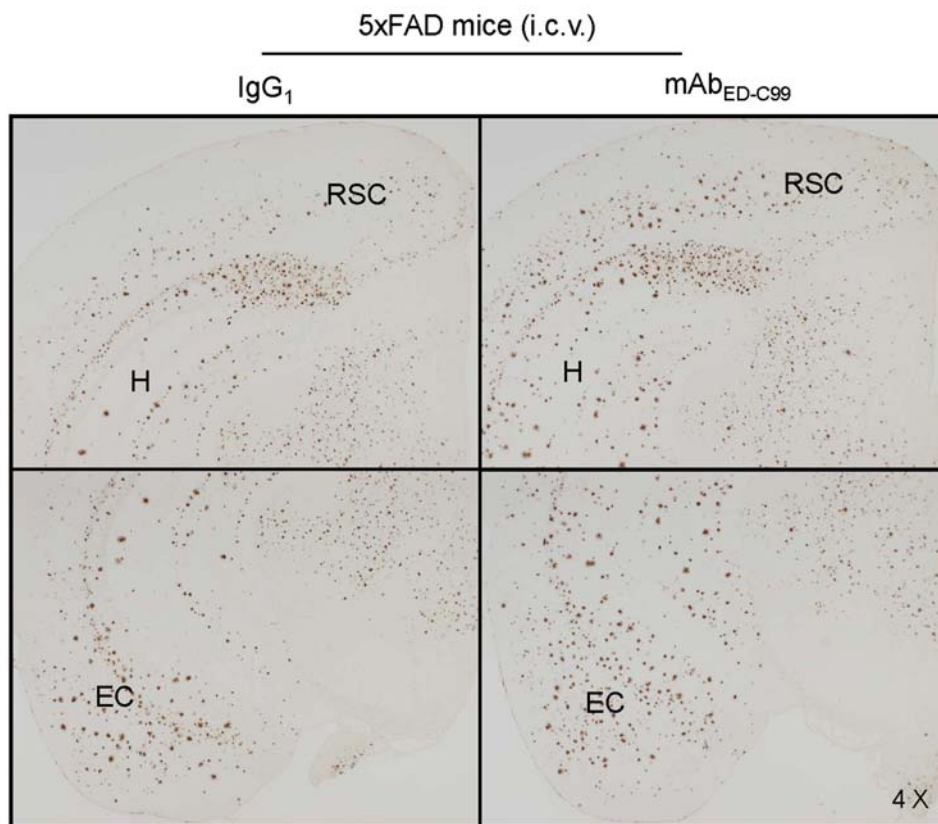
Supplemental Figure S2 Increased cell surface β-CTF is further confirmed by a β-CTF specific antibody. (A) CHO/APP_{wt} cells in 24-well plates (5×10^5 /well) were treated with mAb_{ED-C99} or IgG₁ isotype control at 1.25 μg/ml for 2 h, washed three times with PBS containing CaCl₂ and MgSO₄ (PBS-CM), and then cell lysate portions of these cells were directly subjected to WB analysis using a β-CTF specific antibody, mAbBAM10. The remaining cells were biotinylated with Sulfo-NHS-LC-Biotin dissolved in ice-cold borate buffer and quenched with NH₄Cl-PBS-CM and lysed. These cell lysates were then immunoprecipitated (IP) using Neutravidin beads. The intracellular proteins obtained by IP/Neutravidin depletion (middle panels) and the cell surface (cell surf) proteins obtained by IP/Neutravidin precipitation (right panels) were subjected to WB analysis using mAbBAM10. (B) TgAPP_{wt}-derived primary

neuronal cells were cultured from cortical tissues of one-day-old TgAPP_{wt} mouse pups and replated in 24-well plate at 2×10^5 /well overnight. These primary cultured neuronal cells were treated with mAb_{ED-C99} or IgG₁ isotype control at 1.25 μ g/ml for 2 h, washed three times with PBS-CM, and then cell lysates were directly subjected to WB analysis using mAbBAM10. The remaining cells were biotinylated, immunoprecipitated with Neutravidin beads and subjected to WB analysis using mAbBAM10. These WB data are representative of four independent experiments with similar results.



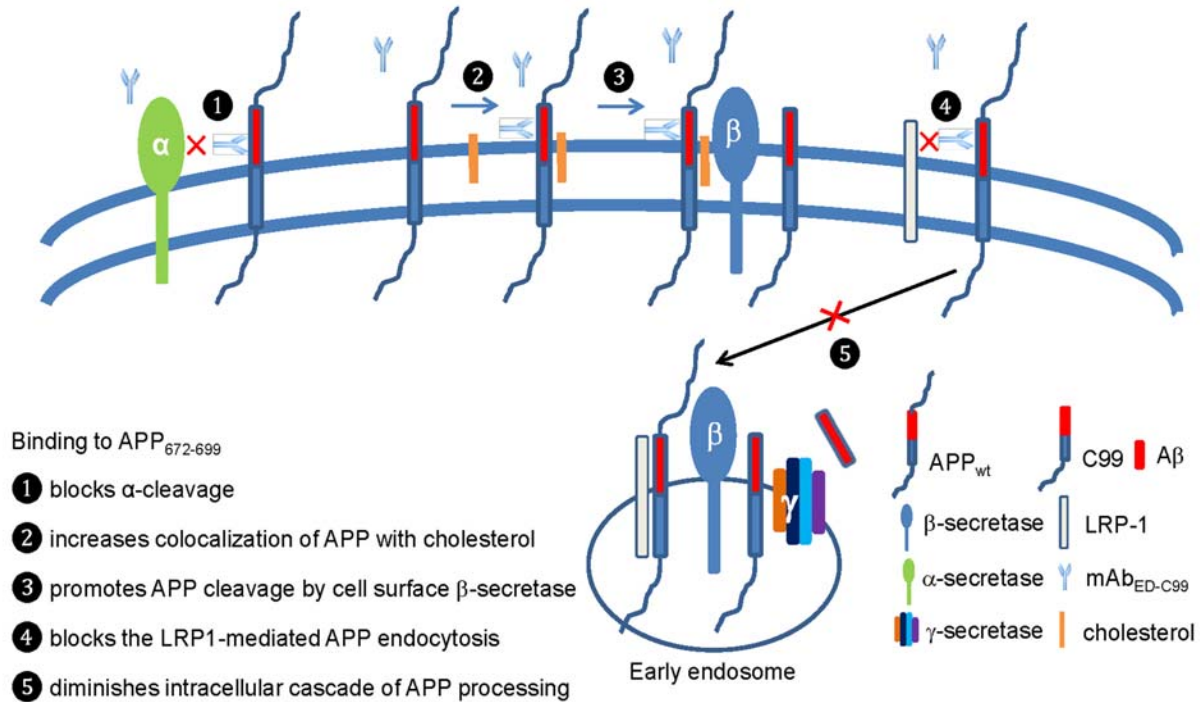
Supplemental Figure S3 Increased co-localization of APP with cholesterol is cell surface specific. CHO/APP_{wt} cells were plated to 8-well slide chamber and then, after overnight

incubation, the cells were treated with mAb_{ED-C99} or IgG₁ control at 1.25 µg/ml for 2 h. Two hours after treatment, these cells were stained by filipin, permeabilized and counterstained with rabbit anti-APP-C-terminal antibody 4°C for overnight, as described for Figure 5. Alexa Fluor® 594 Donkey anti-rabbit IgG was used to detect APP signal and images taken with a confocal microscopy.



Supplemental Figure S4 Intracerebroventricular (i.c.v.) injection of mAb_{ED-C99} yields no increase Aβ deposits. Eight-month-old 5xFAD female mice (n = 3) were treated with mAb_{ED-C99} or isotype IgG₁ as negative control *via* i.c.v. injection daily for 5 days (5 µg/mouse). The immunohistochemical staining using an anti-Aβ₁₇₋₂₆ antibody (4G8) indicated no changes of β-

amyloid plaques in retrosplenial cortex (RSC), entorhinal cortex (EC), and hippocampus (H) regions of 5×FAD mouse brain.



Supplemental Figure S5. Illustration of effects of mAb_{ED-C99} binding to APP₆₇₂₋₆₉₉ on wild-type APP processing. APP is primarily processed by α -secretase on the cell membrane surface leading to sAPP α release, or processed while co-localized with cholesterol on the cell membrane surface by β -secretase, leading to C99 production. In addition, APP can be endocytosed for β -secretase cleavage and endocytosed C99 is further processed by γ -secretase to yield A β . As shown, mAb_{ED-C99} can (1) directly block α -secretase activity, (2) increase co-localization of APP with cholesterol, (3) promote APP to be cleaved by cell surface β -secretase, (4) block LRP1-mediated APP endocytosis, and thereby, (5) diminish intracellular cascade of APP processing.