

Fission yeast mitochondria are distributed by dynamic microtubules in a motor-independent manner

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Supplementary Information includes

Supplementary figure legends S1-3

Supplementary imaging conditions

Supplementary tables S1 and S2

Supplementary figure S1-3

Supplementary figure legends

Figure S1: TagRFP tagged ase1p microtubule binding domain localizes to microtubules, and TagRFP tagged tom22p cannot rescue mitochondria aggregation defects in mmb1 Δ cells.

(a) Schematic diagram depicting the subcellular localization of TagRFP tagged ase1p microtubule binding domain (ase1(MtB)-RFP) and TagRFP tagged tom22p (RFP-tom22). Domain structures of the two fusion proteins are illustrated.

(b) Maximum projection images of wild type cells expressing GFP-cox4p and TagRFP-tom22p. The cells were treated either with DMSO or with MBC for 10 min before imaging. TagRFP-tom22 colocalized with mitochondria in the absence or presence of microtubules. Scale bar, 5 μ m.

(c) Quantification of the indicated patterns of mitochondria distribution in WT, WT TagRFP-tom22, mmb1 Δ , and mmb1 Δ TagRFP-tom22 cells. Note that TagRFP-tom22 did not cause mitochondria aggregation in WT cells, nor did it rescue mitochondria aggregation defects in mmb1 Δ cells.

(d) Maximum projection images of wild type cells expressing GFP-atb2p and ase1-TagRFP. The cells were treated either with DMSO or with MBC for 10 min before imaging. As expected, ase1-TagRFP localized to microtubules. Scale bar, 5 μ m.

(e) Quantification of the indicated patterns of mitochondria distribution in WT, WT ase1(MtB)-TagRFP, mmb1 Δ , and mmb1 Δ ase1(MtB)-TagRFP cells. Note that ase1(MtB)-TagRFP did not cause mitochondria aggregation in WT cells, nor did it rescue mitochondria aggregation defects in mmb1 Δ cells.

Figure S2: Ectopic expression of TagRFP tagged tom22p has no effect on mitochondria aggregation.

(a) Schematic diagram depicting the subcellular localization of TagRFP tagged klp3 motor domain (GFP-klp3(MDo)).

(b) Maximum projection images of wild type cells expressing RFP-cox4p and GFP-klp3(MDo). Note that GFP-klp3(MDo) was localized in the cytoplasm and its localization to microtubules became apparent during mitosis (the spindle marked by the green arrowhead). Scale bar, 5 μ m.

(c) Maximum projection images of wild type cells expressing mCherry-atb2p and GFP-klp3(MDo). Note that GFP-klp3(MDo) was localized in the cytoplasm and its localization to microtubules became apparent during mitosis (the spindle marked by the green arrowhead). Scale bar, 5 μ m.

(d) Quantification of the indicated patterns of mitochondria distribution in WT, WT GFP-klp3(MDo), mmb1 Δ , and mmb1 Δ GFP-klp3(MDo) cells. Note that GFP-klp3(MDo) did

not rescue mitochondria aggregation defects in *mmb1* Δ cells, nor did it cause apparent mitochondria aggression in WT cells.

Figure S3: TagRFP tagged *rng2* actin binding domain is localized to actin filaments.

(a) Schematic diagram depicting the subcellular localization of TagRFP tagged *rng2* actin binding domain (*rng2*(CHD)-RFP).

(b) Maximum projection images of wild type cells expressing GFP-cox4p and *rng2*(CHD)-RFP. Note that *rng2*(CHD)-RFP was localized to the actomyosin ring in mitosis and to actin patches and cables in interphase. Ectopic expression of *rng2*(CHD)-RFP had no effect on mitochondria distribution. Scale bar, 5 μ m.

(c) Maximum projection time-lapse images of *mmb1* Δ *rng2*(CHD)-TagRFP-tom22 and *mmb1* Δ *rng2*(CHD)-TagRFP cells expressing mCherry-*atb2* and GFP-cox4. Mitochondria (yellow arrow) were forced to position at the actomyosin ring by *rng2*(CHD)-TagRFP-tom22 but not *rng2*(CHD)-TagRFP. Scale bar, 5 μ m.

Supplementary Imaging conditions

Figure 1b-d and Figure S1b & S1d, Maximum projection Z-stack images: 21 optical planes, 0.25 μ m step size, 500 ms mCherry exposure, 500 ms GFP exposure, 3X EM gain.

Figure 2c & 2d and Figure S2b & S2c, Maximum projection Z-stack images: 21 optical planes, 0.25 μ m step size, 500 ms mCherry exposure, 500 ms GFP exposure, 3X EM gain.

Figure 2e & 2f, live-cell single-plane images: 1 optical plane, 2 Sec intervals, 500 ms mCherry exposure, 500 ms GFP exposure, 3X EM gain.

Figure 3a, Maximum projection Z-stack images: 21 optical planes, 0.25 μ m step size, 500 ms mCherry exposure, 500 ms GFP exposure, 3X EM gain.

Figure 3c, Maximum projection live-cell Z-stack images: 3 optical plane, 10 Sec intervals, 500 ms mCherry exposure, 500 ms GFP exposure, 3X EM gain.

Figure 4b-d and Figure S3b, Maximum projection Z-stack images: 21 optical planes, 0.25 μ m step size, 500 ms mCherry exposure, 500 ms GFP exposure, 3X EM gain.

Figure 4c and Figure S3c, Maximum projection live-cell Z-stack images: 11 optical plane, 0.5 μ m step size, 1 min intervals, 500 ms mCherry exposure, 500 ms GFP exposure, 3X EM gain.

Supplementary Tables

Table S1. Yeast strains

Strain	Genotype	Source
<u>Figure 1</u>		
CF.2172	GFP-atb2:KanR Leu1-32:ase1P*-ase1(291-731)-TagRFP-tom22 leu1-32 ura4-D18 h-	This study
CF.2310	nmt1P*-GFP-cox4:LEU+ Ura4-294:ase1P*-ase1(291-731)-TagRFP-tom22 ade6-m210? leu1-32 ura4-294 h-	This study
CF.2379	nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-294 h+	This study
CF.2256	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-294 h?	This study
CF.2480	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR Ura4-294:ase1P*-ase1(291-731)-TagRFP-tom22 ade6-m210? leu1-32 ura4-294 h?	This study
CF.3066	nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR Ura4-294:nmt1P*-ase1(291-731)-TagRFP ade6-m210? leu1-32 ura4-294 h+	This study
CF.3067	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR [pSGP572a-nmt1P*-ase1(291-731)-TagRFP:URA4+] ade6-m210? leu1-32 ura4-294 h-	This study
<u>Figure 2</u>		
CF.2379	nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-294 h+	This study
CF.2256	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-294 h?	This study
CF.389	mal3Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 h+	This study
CF.379	klp2Δ:URA4+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 h+	This study
CF.380	klp3Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 h+	This study
CF.381	tea2Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 h+	This study
CF.382	klp5Δ:URA4+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 his3.D1? h-	This study
CF.383	klp6Δ:URA4+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 his3.D1? h-	This study
CF.384	klp8Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 h-	This study
CF.384	DHC1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 h+	This study
CF.3085	klp2Δ:URA4+ DHC1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 h-	This study
CF.3060	klp3Δ:KanR+ klp5Δ:URA4+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 his3D1? h+	This study
CF.3062	klp3Δ:KanR+ klp5Δ:URA4+ tea2Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-D18 his3D1? h?	This study
CF.2173	GFP-atb2:KanR Leu1-32:ase1P*-klp3(1-335)-TagRFP-tom22 leu1-32 ura4-D18 h-	This study
CF.2311	nmt1P*-GFP-cox4:LEU+ Ura4-294:ase1P*-klp3(1-335)-TagRFP-tom22 ade6-m210? leu1-32 ura4-294 h-	This study
<u>Figure 3</u>		
CF.2481	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR Ura4-	This study

CF.2657	294:ase1P*-klp3(1-335)-TagRFP-tom22 ade6-m210? leu1-32 ura4-294 h? nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR Ura4-294:ase1P*-klp3(1-335)- TagRFP-tom22 ade6-m210? leu1-32 ura4-294 h+	This study
<u>Figure 4</u>		
CF.2492	nmt1P*-GFP-cox4:LEU+ Ura4-294:ase1P*-rng2(1-189)-TagRFP-tom22 ade6- m210? leu1-32 ura4-294 h?	This study
CF.2379	nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-294 h+	This study
CF.2256	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR ade6-m210? leu1-32 ura4-294 h?	This study
CF.2493	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR Ura4- 294:ase1P*-rng2(1-189)-TagRFP-tom22 ade6-m210? leu1-32 ura4-294 h?	This study
CF.3075	mmb1Δ:KanR+ nmt1P*-GFP-cox4:LEU+ mCherry-atb2:HygR [pSGP572a- nmt1P*- rng2(1-189)-TagRFP:URA4+] ade6-m210? leu1-32? ura4-294 h-	This study
<u>Figure S1</u>		
CF.3148	GFP-cox4:KanMX Leu1-32:ase1P*-TagRFP-tom22 ade6-m210 leu1-32 ura4- 294 h-	This study
CF.3150	GFP-cox4:KanMX leu1-32:ase1P*-TagRFP-tom22 mCherry-atb2:HygR ade6- m210? leu1-32 ura4-294 h-	This study
CF.3151	mmb1Δ:KanR+ GFP-cox4:KanMX leu1-32:ase1P*-TagRFP-tom22 mCherry- atb2:HygR ade6-m210? leu1-32 ura4-294 h?	This study
CF.3065	GFP-atb2:KanMX ura4-294: ase1P*-ase1(291-731)-TagRFP leu1-32 ura4-294 h?	This study
<u>Figure S2</u>		
CF.3068	nmt1P*-RFP-cox4:LEU+ [pSGP573-nmt1P*-GFP-klp3(1-335):URA4+] ade6- m210 leu1-32 ura4-294 h-	This study
CF.3070	nmt1P*-RFP-cox4:LEU+ GFP-atb2:KanMX [pSGP573-nmt1P*-GFP-klp3(1- 335):URA4+] ade6-m210 leu1-32 ura4-294 h+	This study
CF.3071	mmb1Δ:KanR+ nmt1P*-RFP-cox4:LEU+ GFP-atb2:KanMX [pSGP573- nmt1P*-GFP-klp3(1-335):URA4+] ade6-m210? leu1-32 ura4-294 h-	This study
<u>Figure S3</u>		
CF.2492	nmt1P*-GFP-cox4:LEU+ [pSGP572a- nmt1P*-rng2(1-189)-TagRFP:URA4+] ade6-m210? leu1-32 ura4-294 h?	This study

Table S2. Plasmids

Plasmid	Genotype	Source
<u>Figure 1</u>		
pCF.1693	pJK210- ase1P*-ase1(291-731)-TagRFP-tom22	This study
pCF.2022	pSGP572a- nmt1P*-ase1(291-731)-TagRFP	This study
<u>Figure 2</u>		
pCF.1676	pJK148- ase1P*-klp3(1-335)-TagRFP-tom22	This study
pCF.1694	pJK210- ase1P*-klp3(1-335)-TagRFP-tom22	This study
<u>Figure 3</u>		
pCF.1694	pJK210- ase1P*-klp3(1-335)-TagRFP-tom22	This study
<u>Figure 4</u>		
pCF.1828	pJK210- ase1P*-rng2(1-189)-TagRFP-tom22	This study
pCF.2024	pSGP572a- nmt1P*-rng2(1-189)-TagRFP	This study

Figure S1

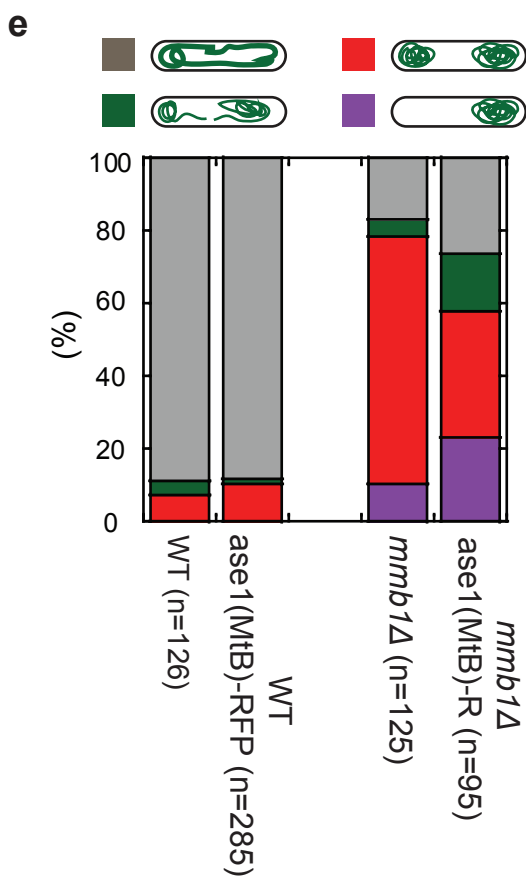
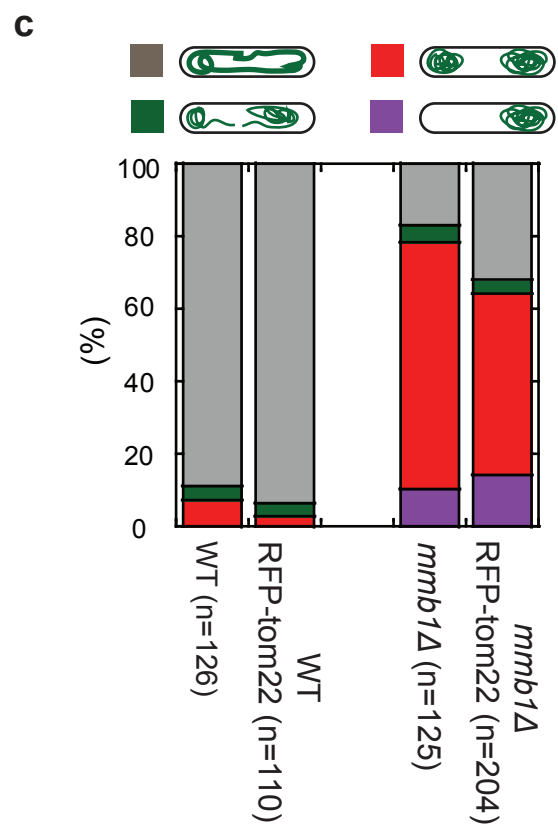
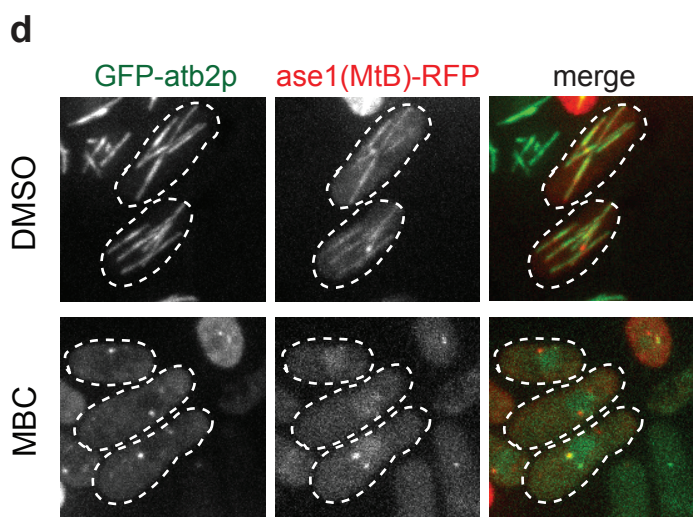
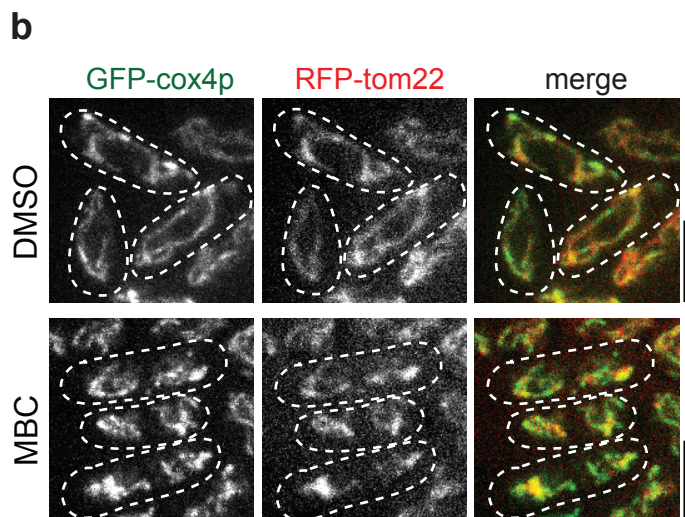
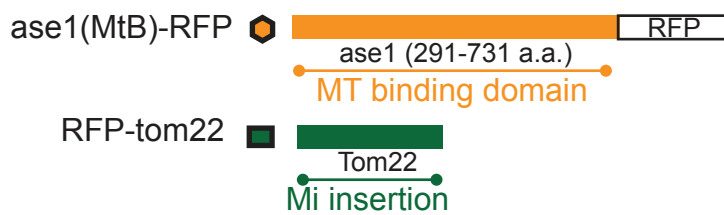
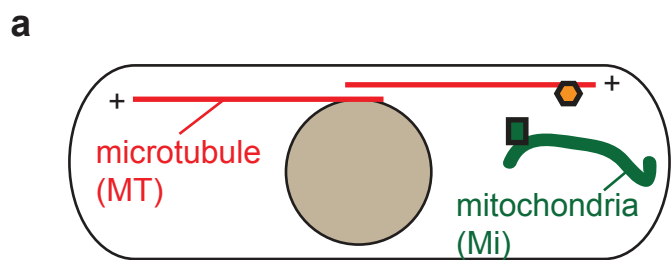
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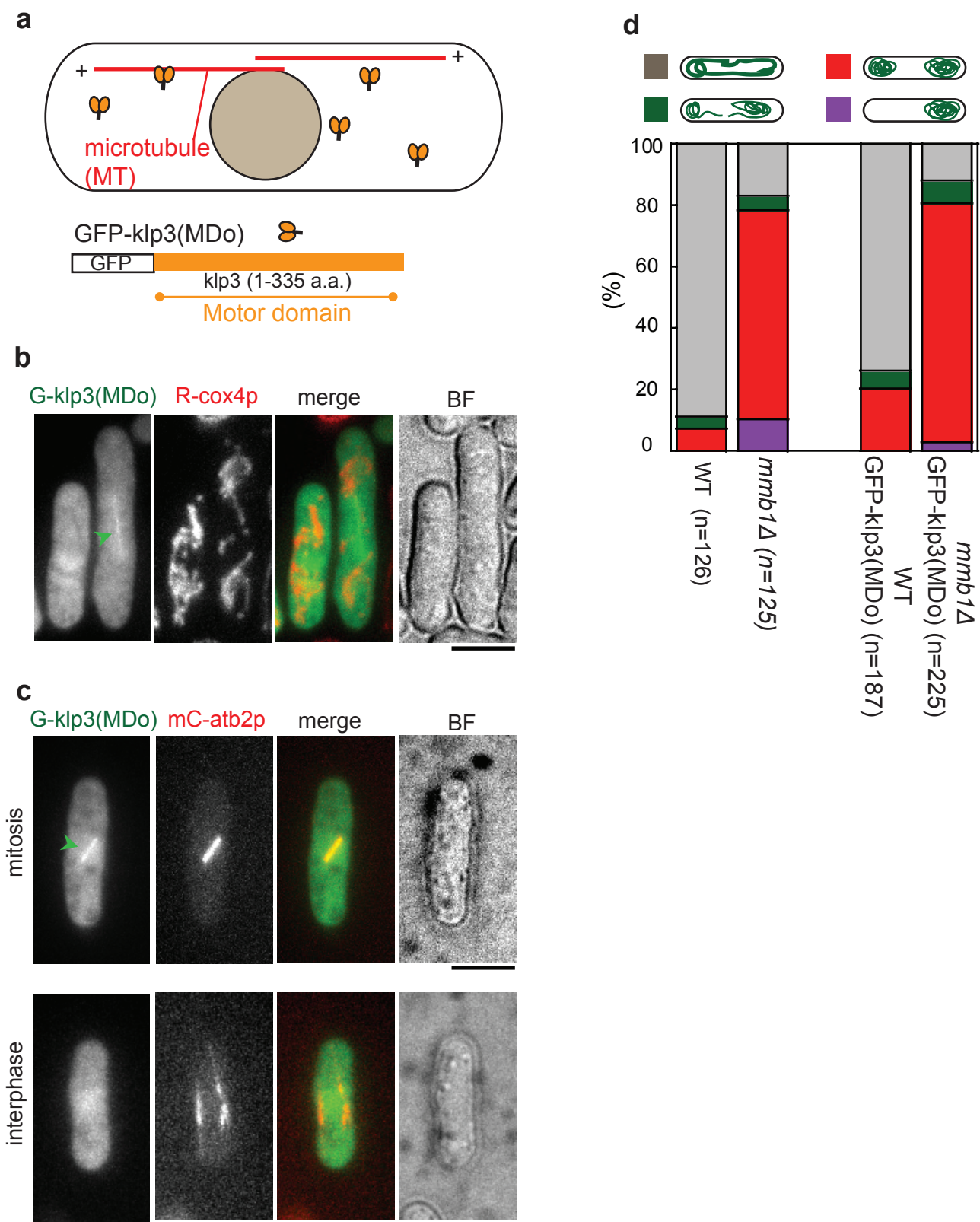
This study

Figure S2

pCF.2026 pSGP573-nmt1P*-GFP-klp3(1-335)

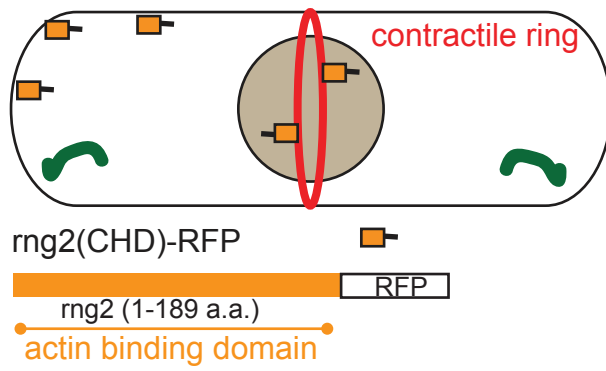
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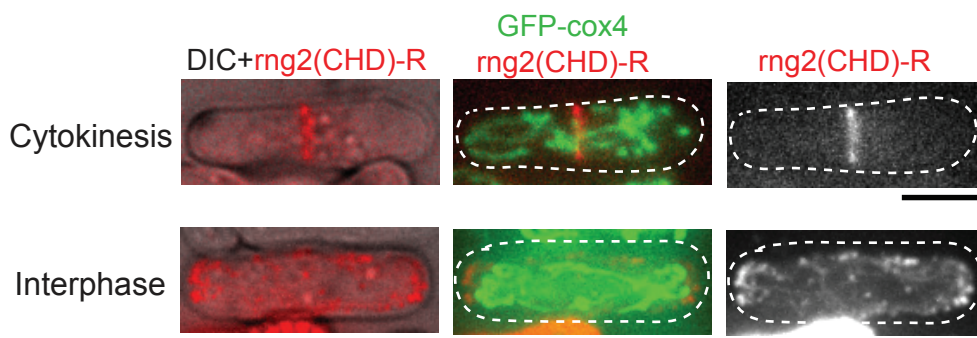


Supplementary Figure S3 Li et al.

a



b



c

