

Supplemental Figure S1. Coding sequence of p16 TALE-DNMT3a-3L.

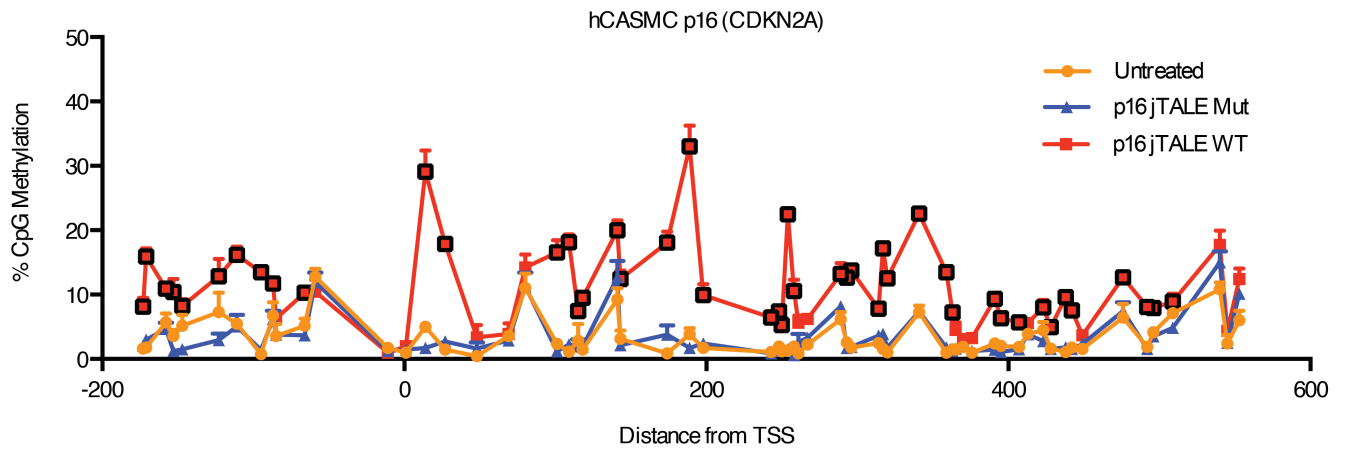
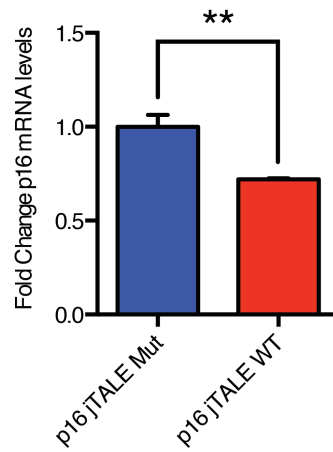
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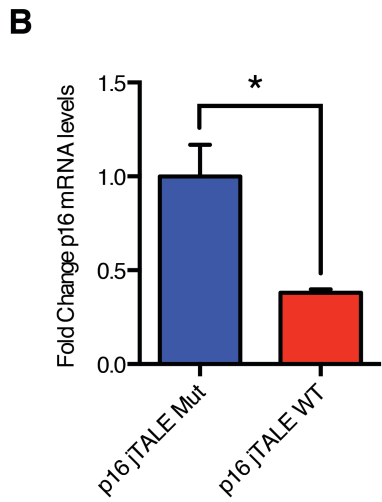
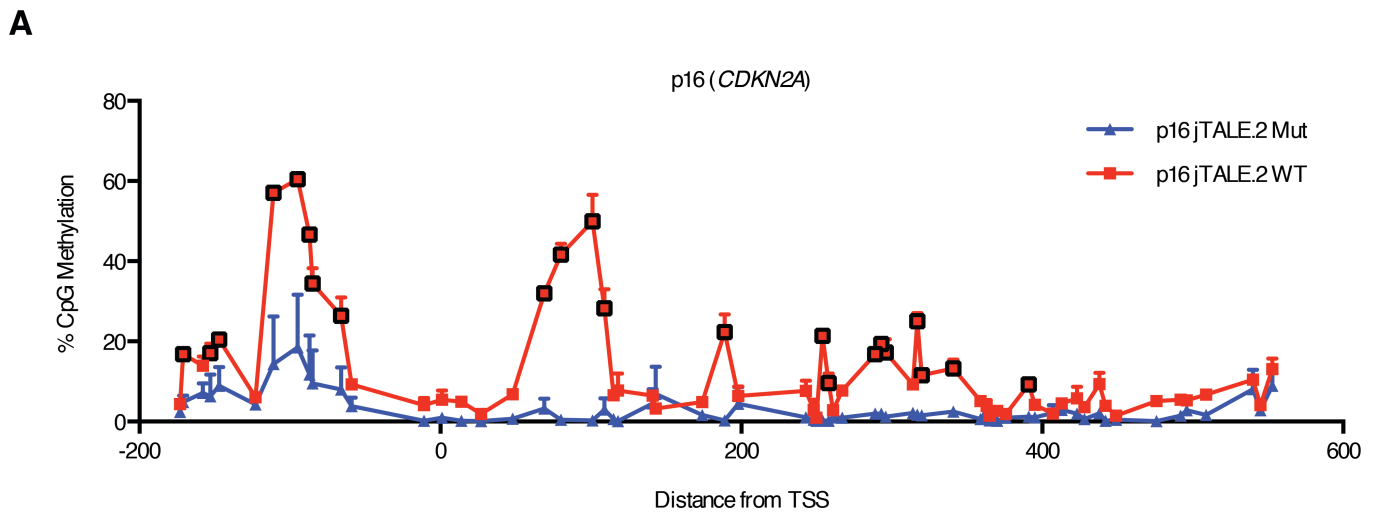
- TALE N-Terminus and C-Terminus
- p16 TALE repeat domain
- Dnmt3a-3L
- 2A GFP

Supplemental Figure S2. Jumbled p16 TALE repeat domain.

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A**B**

Supplemental Figure S3. p16 TALE-DNMT strategy can be employed in primary human coronary artery smooth muscle cells to decrease p16 expression. Primary human coronary artery smooth muscle cells (hCASCs) were infected with p16 jTALE-DNMT wild-type or mutant lentivirus. After 4 days of infection, cells were harvested and assessed for DNA methylation (**A**) and p16 (*CDKN2A*) expression (**B**) as described in the main text.



Supplemental Figure S4. Multiple p16 TALE-DNMT constructs can be designed to decrease p16 expression. An additional TALE-DNMT (p16 jTALE-DNMT.2) was designed to target the p16 (*CDKN2A*) promoter region 118 to 139 base pairs upstream of the transcription start site. Wild-type and mutant infected with p16 jTALE-DNMT.2 were evaluated for DNA methylation (**A**) and p16 (*CDKN2A*) gene expression (**B**) as described in prior human fibroblast experiments.

Supplemental Table 1. PCR primers for amplification of bisulfite converted genomic DNA. Primers were designed to PCR amplify regions of interest from bisulfite converted genomic DNA. Each primer pair amplifies an approximately 250-300 base pair region within the CpG island closest to the transcription start site of the gene analyzed. In instances where there was not a CpG island near the gene, the promoter was evaluated. Amplicons were subsequently used to prepare DNA sequencing libraries for DNA methylation analysis.

Locus	Coordinates	Forward (5' to 3')	Reverse (5' to 3')
<i>CDKN2A_1</i>	chr9:21,975,075-21,975,330	ATTTGGTAGTTAGGAAGGTTGTA	CCCTAAAATCCCCCAAATTAATCTCC
<i>CDKN2A_2</i>	chr9:21,974,909-21,975,178	GGTGGGGAGGAGTTTAGT	AACCAACCCCTCCTCTTT
<i>CDKN2A_3</i>	chr9:21,974,597-21,974,918	GGGGTTGGTTGGTTATTAGAG	CCTAATTCCAATTCCCCTACAACTT
<i>CDKN2A_4</i>	chr9:21,974,558-21,974,863	AGAGGGGGAGAGTAGGTA	CCAAAAACCTCCCCTTTTTCC
<i>CDKN2A CGI2</i>	chr9:21,970,930-21,971,229	GTATGGTTATTGTTTTGGTGT	ACCATTCTATTCTCTCTAACAAATCA
<i>CDKN2A CGI3</i>	chr9:21,968,280-21,968,569	AGGTAAAGATGTGTGGTATATT	ACAACCACTAAAACCTATTATATAAC
<i>CDKN2A CGI3_2</i>	chr9:21,968,551-21,968,805	AATGAGTTTTAGTGGTTGTTTATAAT	AACACATAAATAAATAAACATCCATT
<i>ACTB</i>	chr7:5,570,376-5,570,636	TGGGAGTTTAGTGTTAAGAGATGTTTA	CCCTCTCCCCTCCTTTTTAC
<i>CDKN2B_1</i>	chr9:22,009,012-22,009,290	AGGAGGGGTAGTGAGGATT	ACACTCTCCCTTCTTTCC
<i>CDKN2B_2</i>	chr9:22,008,863-22,009,088	GGGTAATGAAGTTGAGTTTAGG	CACCTTCTCCACTAATCCC
<i>p14ARF</i>	chr9:21,995,629-21,995,928	TGGGAATAGTAAAAGTAGGGTAAGG	ACACCCACCCACTCAAAC
<i>MTAP</i>	chr9:21,802,505-21,802,762	GGAAAGGTTTGAAAAGGG	AATACCAAAAACCATATCTACAC
<i>IFNE</i>	chr9:21,481,642-21,481,862	GAAAAGGGAATTTGAAAATTTAATGT	AATAAAAACCAACAACACCAACACA
<i>CDKN2C</i>	chr1:51,435,514-51,435,751	GGAGGATTTGTTTTGTAGTTTTG	CACAAACACACATACATTCTAATT
<i>CDKN2D</i>	chr19:10,679,809-10,680,058	TTTATAGGGTTTGTATAATTAGTGG	CTACCTCCCTTCCCTCAAA
<i>CDKN1A</i>	chr6:36,651,742-36,651,971	GATTAGTTGGAAGGAGTGAGAGA	ACTCAACTACTCCCTATCCACTAAAC
<i>CDKN1B</i>	chr12:12,869,846-12,870,092	GGGTAGAGTTGGGGGTAG	ACAAACCTACTCTAACTAACCT
<i>CDKN1C</i>	chr11:2,907,681-2,907,919	GGTTTGGTTAAGGTTGAGAAG	AACCCTCACACACTACT
<i>RB1</i>	chr13:48,877,360-48,877,609	GTGATTTTAAAAGGTTAGTAAGTG	AAACCTCATCCCTATCCC
<i>CDK4</i>	chr12:58,145,866-58,146,110	GGTGAGGGGGTTTTTTTAGT	ACCCTTCCATAACCAACTC
<i>CDK6</i>	chr7:92,462,912-92,463,146	GGTTTGGGAGAGGGTAGG	ATCCCTCCTCTTCCCTCC
<i>CCND1</i>	chr11:69,455,880-69,456,104	ATTATAGGGGAGTTTTGTTGAAGTT	CAACACAAAAACTAATATTCCATAAC

Supplemental Table 2. qPCR primers for gene expression analysis

Locus	Forward (5' to 3')	Reverse (5' to 3')
<i>HPRT1</i>	GCAGACTTTGCTTTCCTTGG	AACACTTCGTGGGGTCCTTT
<i>CDKN2A</i> (p16)	CCCAACGCACCGAATAGTT	AGCAGCAGCTCCGCCACT
<i>CDKN2A</i> (p14ARF)	AGAACATGGTGCGCAGGT	GGGATGTGAACCACGAAAAC
<i>CDKN2B</i>	GCTGTTTCATCAGCAGCCTA	CCACAATGGAGCTAGAAGCA
<i>MTAP</i>	CTACCTCAGGGCATGGTTGT	TGCTGGCTAAATGCTTGTTG
<i>IFNE</i>	ACCCTTGAACGACATGAAGC	AAGAATCATGTCTGGAGAAGTCC
<i>CDKN2C</i>	GTGATTTGGCCAGGCTCTAT	AGCCCTCCCCACGTTTATT
<i>CDKN2D</i>	CCAAGGGCAGAGCATTAAAG	AAGCAACGTGCACACTTCAG
<i>CDKN1A</i>	GATTAGCAGCGGAACAAGGA	CAACTACTCCCAGCCCCATA
<i>CDKN1B</i>	GAGGTGCTTGGGAGTTTTGA	TGTTTACACAGCCCGAAGTG
<i>CDKN1C</i>	CAGCGTTCGGTTTTGTTTTT	GGGACCAGTGTACCTTCTCG
<i>RB1</i>	AGGGCTTACTATTTCTGGGTCTT	CAATACACGATCTCTGAAGTTCC
<i>CDK4</i>	CGAGCTCTGCAGCACTCTTA	AGAAGGGAAATGGCAGCTTT
<i>CDK6</i>	TGTCTATCTCCCGGCACTTC	TGAAAGCAAGCAAACAGGTG
<i>CCND1</i>	AGCGCTGTTTTTGTGTGTG	CTTGCCTCAAAGTCCTGCTT