

Supplemental Materials and Methods, Figures and Figure Legends

Materials and Methods

Cells. Human G361, A375, Malme3M, SKMEL2, SKMEL5, SKMEL28, UACC62, UACC257, M14, MDAMB435, 501mel, MeWo, FEMX and LOX and A2058 melanoma cells and mouse B16 F10 cells were maintained in RPMI 1640/10%FBS/1% Penicillin-Streptomycin (Invitrogen, Inc.; Carlsbad, CA) and validated for pigment and differentiation gene markers; MCAM, MITF, melanocyte antigen recognized by T cells (MART-1), tyrosinase and/or SOX10. Human T cells from PBMC isolates were activated and expanded in X-VIVO15™ (Lonza, Inc.; Hopkinton, MA) and IL-2 as described (Cedeno-Laurent *et al.*, 2012). Short-term cultures of clinical melanoma specimens from (3) donors were isolated from surgical metastases in accordance with IRB approval and cultured in RPMI 1640/10% FBS/1% penicillin/streptomycin as described (Schatton *et al.*, 2008). Early passage (P2), primary normal human epidermal melanocytes (HEM) were purchased from PromoCell, Inc. (Heidelberg, Germany). Patient consent was not necessary and all human tissues were obtained according to institutional review board approval.

Silencing of MCAM. Lentiviral shRNA constructs in pLKO.puro against MCAM or scrambled (SCr) controls were purchased from the Mission collection (Sigma-Aldrich; St. Louis, MO). Retroviral supernatants were generated by co-transfection of pN8e-GagPolΔ8.1 and pNE8e-VSV/G in the packaging HEK293t cell line. Melanoma cells were transduced in the presence of 8μg/ml polybrene and selected with 1μg/ml puromycin to generate stable knockdown cell lines. Knockdown of human and mouse MCAM was achieved with the shRNA target sequence - AGTTGAAGTTAACAGATAA.

Overexpression of ST6GalNAc2. The following sequences were used for real-time qRT-PCR to validate ST6GalNAc2 levels: forward 5'-CTCGTCTCCTACTGGAATCTGG-3' and reverse 5'-CGATCTCAGCATCACATAGTCGC-3' as described (Barthel *et al.*, 2008; Barthel *et al.*, 2013). For overexpression, human ST6GalNAc2 cDNA was amplified by KOD polymerase PCR from reverse transcribed melanocyte RNA using forward: 5'-TTCTGCCTGGGACGTCAGCGGACG-3' and reverse: 5'-TCAGCGCTGGTACAGCTGAAGGA-3' primers. This ST6GalNAc2 cDNA

product was then cloned into pDONR223 by KOD amplification using forward: 5'-GGGGACAACTTGTACAAAAAAGTTGGCTCCACCATGGGCTCCCGCGCGG-3' and reverse: 5'-GGGGACAACTTGTACAAGAAAGTTGGTAGCGCTGGTACAGCTGAAGGA TGCC-3' attB-containing primers. Subsequent LR clonase (Invitrogen)-mediated recombination into pLenti6.3-DEST (Invitrogen) yielded vector pLenti6.3-ST6GalNAc2. Cells were transduced with lentiviral supernatants and selected for stable expression with blasticidin.

Flow Cytometry. Flow cytometric analysis using primary antibodies, including polyclonal goat anti-human Gal-1 (R&D Systems), anti-human Gal-3 (M3/38) (BioLegend), anti-human Gal-9 (9M1-3) Abs (BioLegend), human Gal-1 – hFc chimera (hGal-1) (generously provided by Dr. Kuo-I. Lin; Academia Sinica, Taiwan) (Tsai *et al.*, 2008; Tsai *et al.*, 2011), Gal-1hFc, dmGal-1hFc, isotype control Abs, and respective fluorophore-conjugated secondary Abs was performed as described (Cedeno-Laurent *et al.*, 2012). Where indicated, cells were either treated with the broadly-active protease, bromelain (Sigma), at 0.1U/ml for 1hr at 37°C to digest all cell surface glycoproteins or with kifunensine (Sigma) between 1-10µg/ml for 48hr to inhibit *de novo* synthesis of complex N-glycans and then stained with Gal-1hFc or with FITC-PHA-L (1µg/ml). Cells were also stained with biotinylated LEA (0.1µg/ml) and APC-avidin (1:500). Secondary Ab, APC-avidin or 50mM lactose-containing Gal-1hFc groups were included to control for background fluorescence and carbohydrate-dependence of Gal-1hFc, PHA-L or LEA. Experiments were performed at least three-times.

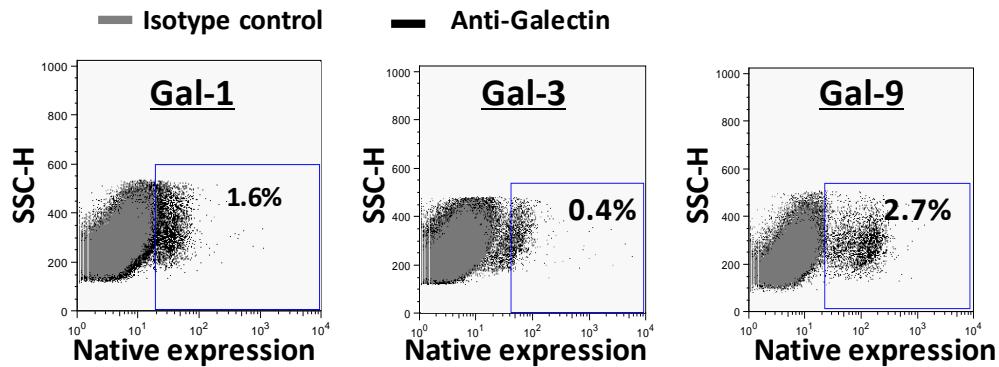
In vitro Melanoma Cell Proliferation Assay. We assayed for cell growth by CSFE-dilution assay over a 3-day period in the presence Gal-1hFc, hFc or diluent controls. Prior to assays, cells were cultured for 24h in RPMI 1640/10%FBS/1% Pen/Strep with 50mM lactose to recapitulate pretreatment conditions in cell migration assays. Cells were harvested with 1mM EDTA, washed in PBS, loaded with 2µM CFSE in PBS for 10 min at 37°C. Cells were then plated in RPMI 1640/10% FBS with diluent control, hFc or Gal-1hFc at either 5 or 20µg/ml in triplicate at 5x10⁴/well in 6-well plates. After 1, 2 and 3 days, cells were harvested and assayed by flow cytometry.

Statistical Analysis. Statistical significant comparisons were ascertained by two-tailed Student's *t*-test, paired *t*-test, one-way ANOVA with Dunnett's post test, or contingency table on GraphPad Prism (GraphPad Software, La Jolla, CA).

Supplemental Figure 1 and Legend

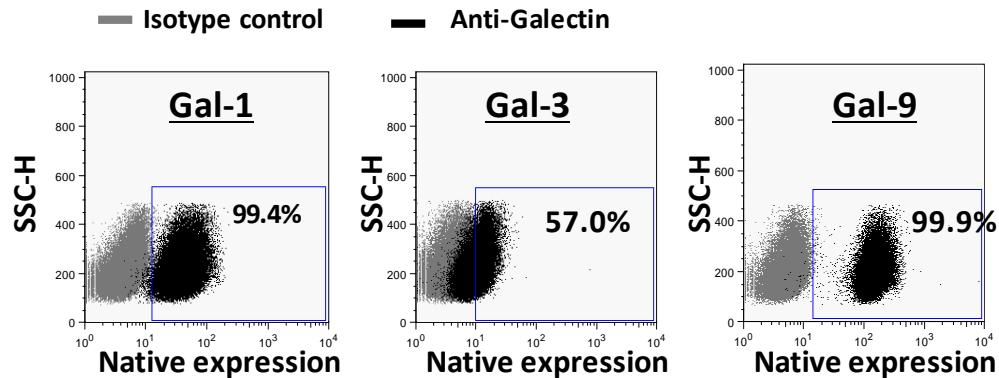
a

FACS Analysis of Surface (Non-permeabilized) Gal-1, -3 and -9



b

FACS Analysis of Intracellular (Permeabilized) Gal-1, -3 and -9

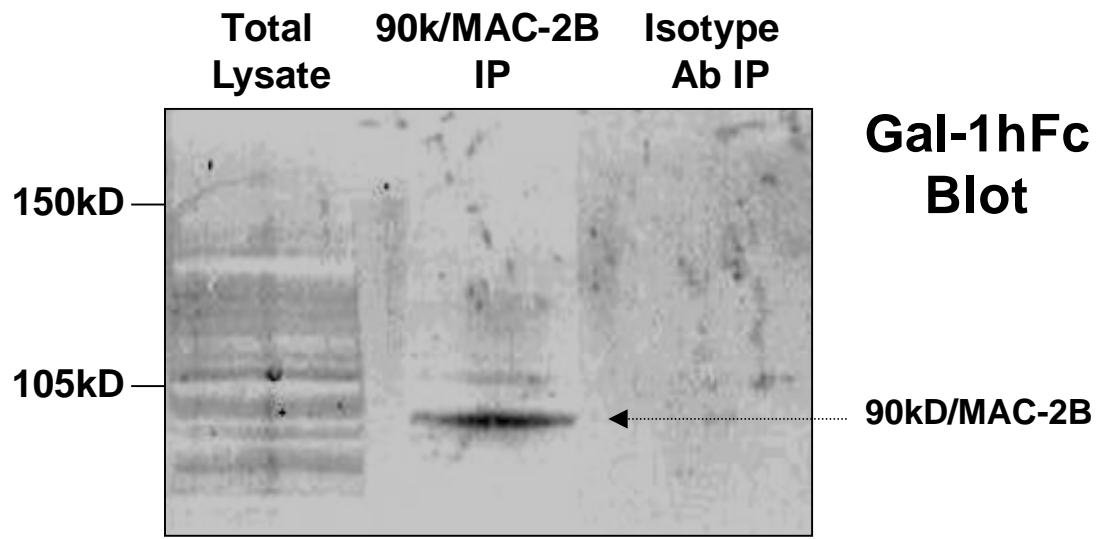


Supplemental Figure 1. Melanoma cell Gal-1, -3 and -9 are principally expressed and detected inside of the cell and not bound to the cell surface. Surface (Non-permeabilized) (a) and intracellular (permeabilized) (b) human A375 melanoma cells were FACS stained with anti-human Gal-1 , -3, or -9 Abs or respective fluorophore-conjugated secondary Ab. Cells were analyzed for fluorescence intensity (x-axis) and designated as percent positive (black dots) compared with cells stained with control Ab alone (gray dots). Data are representative of triplicate experiments.

Supplemental Figure 2 and Legend

Identified Protein	Accession Number	MW	# Peptide matches		
			Negative Control	Gal-1 Eluate	
Ig gamma-1 chain/region Osmoregulatory protein GN=IGG1 PE=1 SV=1	P02786[ALBU_HUMAN]	20 kDa	22	33	136
Small albumin-binding protein GN=ALB PE=1 SV=2	P02786[ALBU_HUMAN]	69 kDa	0	0	33
ATP-citrate synthase OS=Homo sapiens GN=ACLY_HUMAN	P53396[ACLY_HUMAN]	121 kDa	0	51	51
Cell surface glycoprotein MUC18 OS=Homo sapiens GN=MCAM PE=1 SV=2	P43121[MUC18_HUMAN]	72 kDa	0	40	40
Isocitrate dehydrogenase OS=Homo sapiens GN=IDH1_HUMAN (+3)	Q86881[IDH1_HUMAN (+3)]	150 kDa	0	36	36
Ribosome-binding protein 1 OS=Homo sapiens GN=RBP1 PE=1 SV=4	Q9P2E5[RBP1_HUMAN]	152 kDa	0	35	35
Galectin-3-binding protein OS=Homo sapiens GN=LGALS3BP PE=1 SV=1	Q93830[LGALS3BP_HUMAN]	65 kDa	3	32	32
Isoform 2 of Ig kappa chain C-region OS=Homo sapiens GN=IGKC PE=1 SV=3	P02787[IGKC_HUMAN (+2)]	112 kDa	0	35	35
DNA damage-binding protein 1 OS=Homo sapiens GN=DDR1 PE=1 SV=1	Q16531[DDR1_HUMAN]	127 kDa	0	34	34
Syntaphaglin-like protein 4 OS=Homo sapiens GN=SYTL4 PE=4 SV=1	F8W9B#F8W9B99_HUMAN (+2)	76 kDa	18	13	13
Isoform 1 of Vinculin OS=Homo sapiens GN=VCL_HUMAN	P18206[2[VINC_HUMAN (+1)]	117 kDa	0	30	30
Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	Q9P2E5[PKCepsilon_HUMAN]	99 kDa	0	30	30
Ig gamma-3 chain/region Osmoregulatory protein GN=HYOU1 PE=1 SV=1	Q9Y4L1[HYOU1_HUMAN]	111 kDa	0	25	25
Ubiquitin-like modifier-activating enzyme 1 OS=Homo sapiens GN=UBA1 PE=1 SV=3	P22341[UBA1_HUMAN]	118 kDa	0	27	27
Ig gamma-3 chain/region Osmoregulatory protein GN=HYOU1 PE=1 SV=2	P17171[IMPR1_HUMAN]	41 kDa	17	58	58
Cation-independent mannose 6-phosphate receptor OS=Homo sapiens GN=IGF2R PE=1 SV=3	B4E5Q1[B4E5Q1_HUMAN (+1)]	274 kDa	0	22	22
Calsenitin-1 OS=Homo sapiens GN=CLSTN1 PE=2 SV=1	P21091[CLSTN1_HUMAN]	108 kDa	0	23	23
Collagen alpha-1(VI) chain OS=Homo sapiens GN=COL6A1 PE=1 SV=3	Q14682[COL6A1_HUMAN]	109 kDa	0	22	22
Isoform 1 of Galectin-1 OS=Homo sapiens GN=GANAB_HUMAN	P09382[ILEG1_HUMAN]	15 kDa	4	17	17
Galectin-1 OS=Homo sapiens GN=LGALS1 PE=1 SV=2	F8VVB#F8VVB_HUMAN (+4)	86 kDa	0	21	21
Protein kinase C epsilon-binding protein NELL2 OS=Homo sapiens GN=NELL2 PE=4 SV=1	Q5T7D1[NELL2_HUMAN (+1)]	258 kDa	0	21	21
Isoform 2 of Fibronectin type-III domain-containing protein 3A OS=Homo sapiens GN=FNDC3A PE=26S proteasome non-ATPase regulatory subunit 1 OS=Homo sapiens GN=PSMD1 PE=1 SV=2	Q9Y2Z6[2[FIND3A_HUMAN (+1)]	126 kDa	0	21	21
Hexokinase-1 OS=Homo sapiens GN=PE=3 SV=1	Q99460[PSMD1_HUMAN]	106 kDa	0	20	20
Urokinase-type plasminogen activator 7 OS=Homo sapiens GN=USP7 PE=1 SV=2	E7ENR1[E7ENR4_HUMAN (+4)]	106 kDa	0	18	18
Nodal modulator 3 OS=Homo sapiens GN=NOMO3 PE=4 SV=1	Q3K936[J3K936_HUMAN (+4)]	123 kDa	0	19	19
Rat GTP-Activating-like protein IQAP1 OS=Homo sapiens GN=IQGP1_HUMAN PE=1 SV=1	P46940[IQGP1_HUMAN]	139 kDa	0	18	18
Staphylococcal nucleolysin domain-containing protein OS=Homo sapiens GN=SNL_HUMAN PE=1 SV=1	Q7P2X1[SNL_HUMAN]	189 kDa	0	18	18
Lysosomal membrane glycoprotein 1 OS=Homo sapiens GN=LAMP2 PE=2 SV=1	B4E5ST[B4E5ST2_HUMAN (+1)]	102 kDa	0	16	16
Lysozyme-associated membrane glycoprotein 1 OS=Homo sapiens GN=LAMP1 PE=1 SV=3	P11279[LAMP1_HUMAN]	40 kDa	0	16	16
Isoform 5 of fractin tyrosine-protein kinase 2 OS=Homo sapiens GN=PTK7	Q13308[5[PTK7_HUMAN (+2)]	45 kDa	0	16	16
Isoform 1 of Integrin alpha-1 OS=Homo sapiens GN=GNTM1_HUMAN	P05507[ITGA1_HUMAN (+4)]	87 kDa	0	16	16
Chondroitin sulfate proteoglycan 4 OS=Homo sapiens GN=CSPG4 PE=1 SV=2	Q6UVK1[CSPG4_HUMAN]	251 kDa	0	16	16
Histone shock cognate 71 kDa protein OS=Homo sapiens GN=HSPPA8 PE=3 SV=1	E8PK3#E8PK3E_HUMAN (+1)	69 kDa	0	12	12
Isoform 4 of Phosphoglycerate kinase OS=Homo sapiens GN=PGK_HUMAN	P27959[PGK_HUMAN (+1)]	84 kDa	0	12	12
Isoform 1 of Integrin alpha-1 OS=Homo sapiens GN=ITGA1_HUMAN	P22924[ITGA1_HUMAN (+6)]	119 kDa	0	12	12
Heat shock 70 kDa protein 4 OS=Homo sapiens GN=HSP40 PE=1 SV=4	P34932[HSP74_HUMAN]	94 kDa	0	15	15
Stress-70 protein, mitochondrial OS=Homo sapiens GN=HSP90_HUMAN PE=1 SV=2	P38646[GRP75_HUMAN]	74 kDa	4	10	10
Isoform 1 of Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	Q9P2E5[ITGB3_HUMAN]	124 kDa	0	12	12
Neurogranin OS=Homo sapiens GN=NEURO_HUMAN	P14618[KPYM_HUMAN]	58 kDa	0	13	13
Pyruvate kinase PMK OS=Homo sapiens GN=PKM PE=1 SV=4	P17607[MA2A1_HUMAN]	131 kDa	0	14	14
Alpha-mannosidase 2 OS=Homo sapiens GN=MAN2A1 PE=1 SV=2	Q3T1J1[MAN2A1_HUMAN]	144 kDa	0	14	14
Isoform 3 of Mannosidase 2 OS=Homo sapiens GN=MAN2B1_HUMAN	P05023[3[MAN2B1_HUMAN (+2)]	110 kDa	1	12	12
N-acetylglucosaminidase 1 OS=Homo sapiens GN=GNATG1_HUMAN	F6Y097[FEY097_HUMAN (+2)]	63 kDa	0	12	12
Isoform 2 of Mannosidase superfamily member 3 OS=Homo sapiens GN=GNPTAB PE=1 SV=1	Q75054[2[IGSF3_HUMAN]	138 kDa	0	12	12
Isoform 2 of Mannosidase superfamily member 3 OS=Homo sapiens GN=IGSF3_HUMAN	Q9P2E5[IGSF3_HUMAN]	100 kDa	0	11	11
Isoform 3 of Mannosidase superfamily member 3 OS=Homo sapiens GN=IGSF3_HUMAN	P06733[ENOA_HUMAN]	47 kDa	2	9	9
Isoform 1 of Integrin alpha-2 OS=Homo sapiens GN=ITGA2_HUMAN	P11047[ITGA2_HUMAN]	178 kDa	1	10	10
Tyrosine kinase receptor OS=Homo sapiens GN=TKR1_HUMAN PE=3 SV=3	C0E2X1[CK2AX1_HUMAN (+1)]	155 kDa	0	10	10
Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	Q7P2X1[PKCepsilon_HUMAN]	500 kDa	0	10	10
Stress-70 protein, mitochondrial OS=Homo sapiens GN=HSP90_HUMAN PE=1 SV=2	P10516[3[ITB3_HUMAN]	87 kDa	0	8	8
Isoform 4 of Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	F5H025[5[GH025_HUMAN]	140 kDa	0	8	8
Transmembrane endoplasmic reticulum ATPase OS=Homo sapiens GN=LRP1 PE=4 SV=1	P02896[2[GLSG1_HUMAN (+2)]	137 kDa	0	9	9
Neurogranin OS=Homo sapiens GN=NEURO_HUMAN	P10193[NEURO_HUMAN]	36 kDa	0	45	45
Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	P04406[2[G3P_HUMAN]	32 kDa	5	3	3
Actin, cytoplasmic 1 OS=Homo sapiens GN=ACTB PE=1 SV=1	P670709[ACTB_HUMAN]	42 kDa	2	5	5
Isoform 2 of Presenilin protease, mitochondrial OS=Homo sapiens GN=PITRM1_HUMAN	P07935[PITRM1_HUMAN]	100 kDa	0	11	11
Alpha-endopeptidase OS=Homo sapiens GN=ENO1 PE=1 SV=2	B1A4H1[ENO1_HUMAN]	78 kDa	0	10	10
CD276 antigen OS=Homo sapiens GN=CD276 PE=1 SV=1	Q5ZPR3[CD276_HUMAN]	57 kDa	0	10	10
Isoform 2 of Filamin-1 OS=Homo sapiens GN=FLN_HUMAN	P21333[2[FLN_HUMAN]	280 kDa	0	8	8
Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	P10536[ITGB3_HUMAN]	93 kDa	0	8	8
Neurogranin OS=Homo sapiens GN=NEURO_HUMAN	P05106[3[ITB3_HUMAN]	87 kDa	0	8	8
Neurogranin OS=Homo sapiens GN=NEURO_HUMAN	F5H025[5[GH025_HUMAN]	140 kDa	0	8	8
Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	P50572[2[TERA_HUMAN]	86 kDa	0	7	7
Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	Q9N660[2[AHNK_HUMAN]	178 kDa	0	7	7
Transmembrane endoplasmic reticulum ATPase OS=Homo sapiens GN=LRP1 PE=4 SV=1	B4D8ER1[B4D8ER1_HUMAN]	629 kDa	0	5	5
Leucine-rich repeat transmembrane protein OS=Homo sapiens GN=LRRK2_HUMAN	A4DER1[A4DER1_HUMAN]	131 kDa	1	5	5
Large conductance potassium channel OS=Homo sapiens GN=VGK_HUMAN	Q174764[MVP_HUMAN]	99 kDa	0	5	5
Actin, cytoskeletal OS=Homo sapiens GN=ACTB_HUMAN	K7E0Q5[ACTB_HUMAN]	100 kDa	0	5	5
Neurogranin OS=Homo sapiens GN=NEURO_HUMAN	MOR165[MOR165_HUMAN (+2)]	83 kDa	0	5	5
Phosphotyrosyl-protein phosphatase OS=Homo sapiens GN=PTPNS_HUMAN	P15067[PTPNS_HUMAN]	145 kDa	0	6	6
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Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	P12270[TPR_HUMAN]	61 kDa	0	6	6
Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	P13639[EF2_HUMAN]	267 kDa	0	6	6
Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	P10809[CH60_HUMAN]	95 kDa	0	6	6
Integrin beta-3 OS=Homo sapiens GN=ITGB3_HUMAN	P04932[IFAS_HUMAN]	58 kDa	0	6	6
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Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	Q9N660[2[AHNK_HUMAN]	178 kDa	0	7	7
Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	A4DER1[A4DER1_HUMAN]	629 kDa	0	5	5
Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	P07935[2[CD276_HUMAN]	100 kDa	0	5	5
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Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	P07935[2[CD276_HUMAN]	302 kDa	0	2	2
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Protein kinase C epsilon OS=Homo sapiens GN=PKCepsilon_HUMAN	P07935[2[CD276_HUMAN]	374 kDa	0	2	2
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Supplemental Figure 3 and Legend

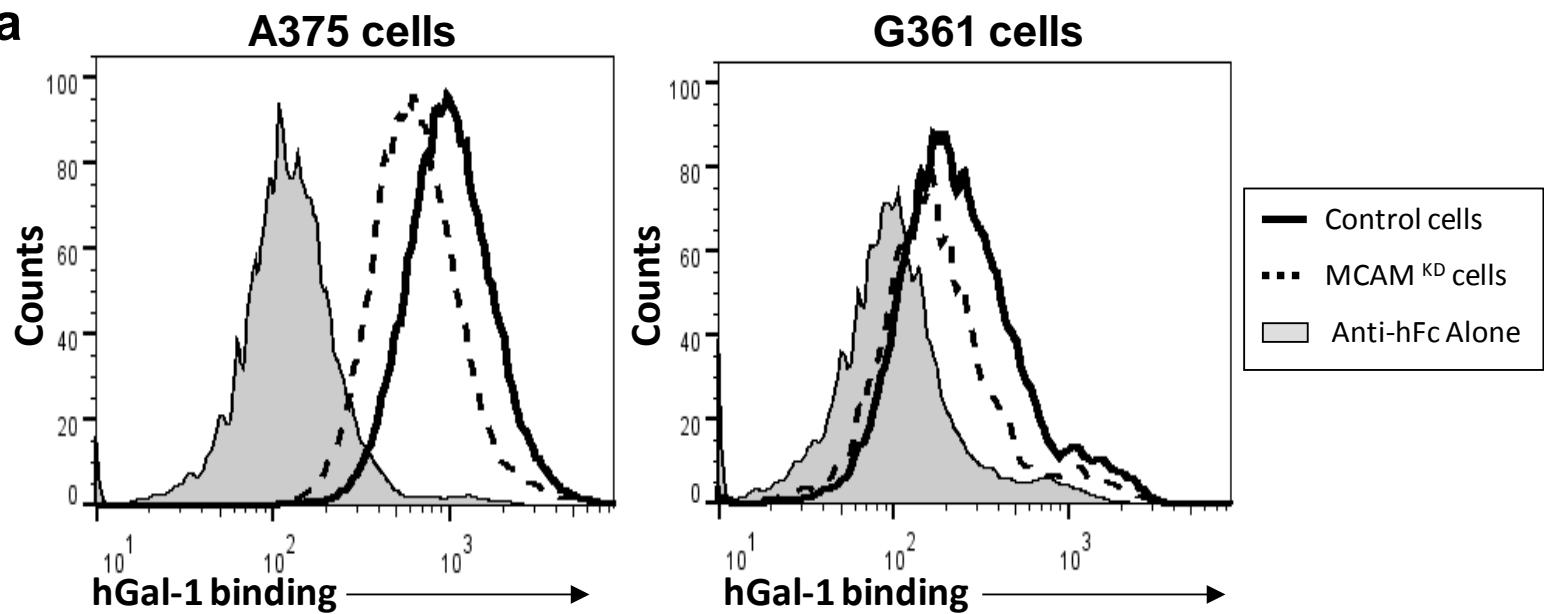


Supplemental Figure 3. Detection of Gal-3-binding Protein (90k/MAC-2B) immunoprecipitated from human melanoma cells with Gal-1hFc. Gal-3-binding protein (90k/MAC-2B) was immunoprecipitated with anti-90k moAb SP-2 or with isotype control Ab from human A375 melanoma cell lysate and blotted with Gal-1hFc. Total lysate, anti-90k immunoprecipitate (IP) and isotype control Ab IP were separated on reducing 4-20% gradient SDS-PAGE gels and blotted with Gal-1hFc. Data was representative from replicate experiments.

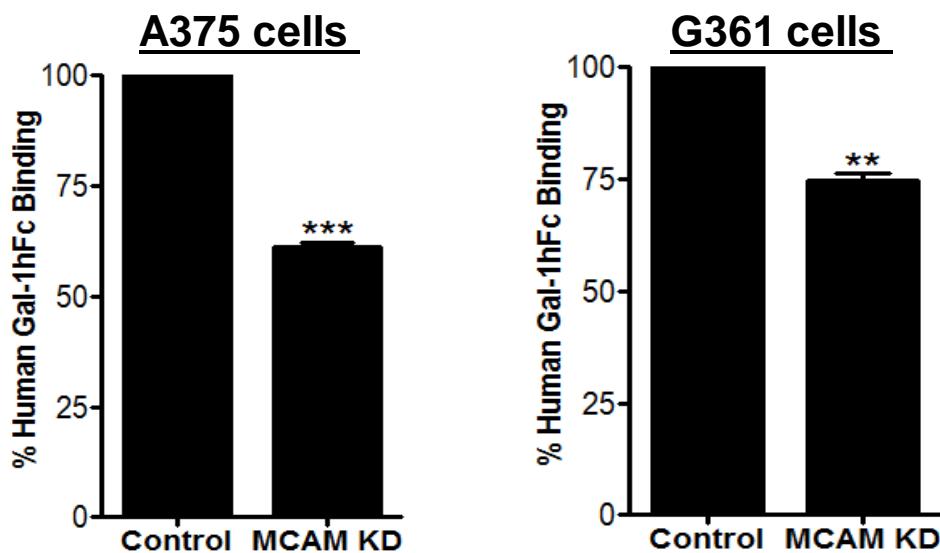
Supplemental Figure 4 and Legend

Human Gal-1 binding to melanoma cell Gal-1 ligands is dependent on MCAM.

a

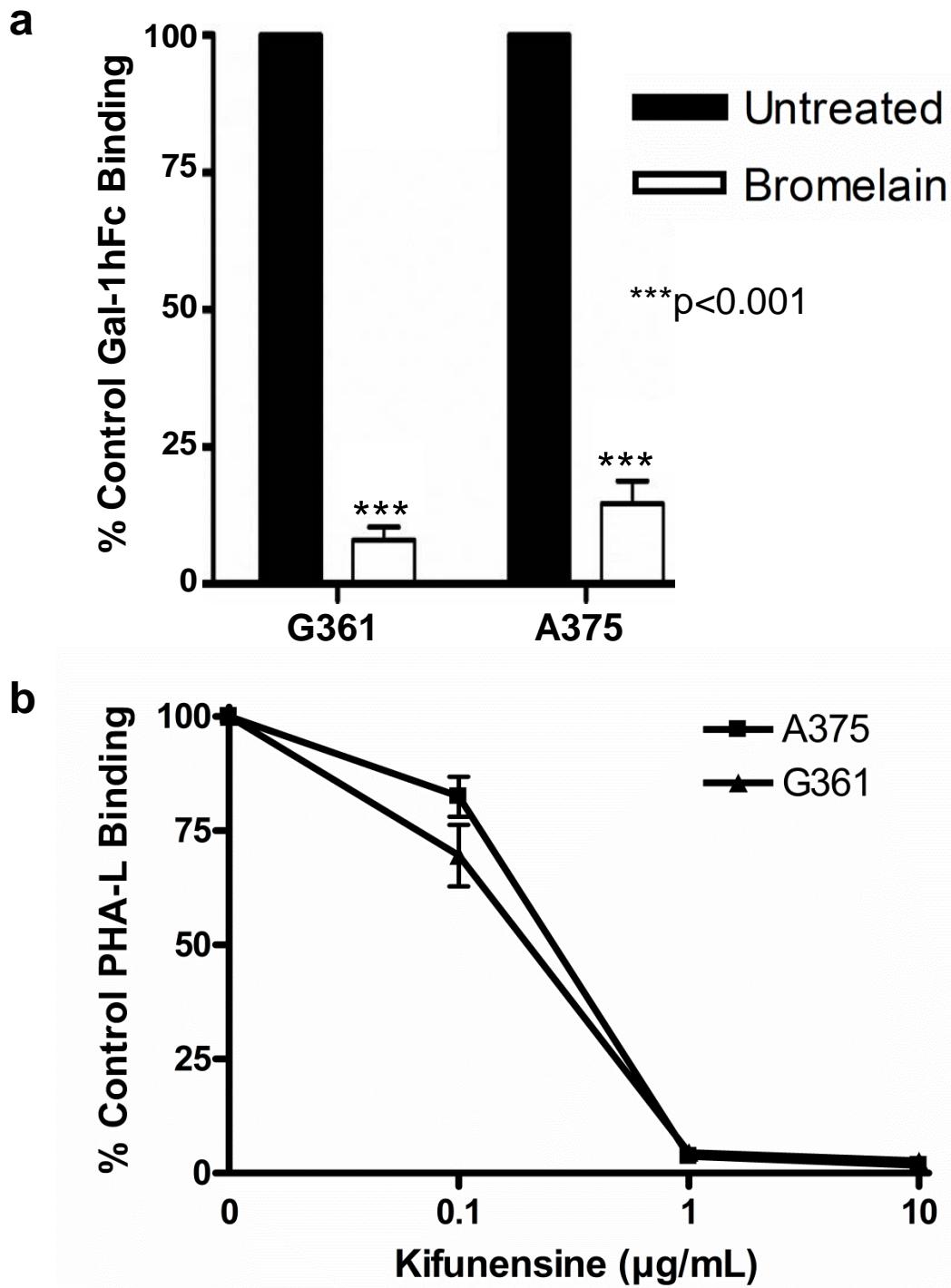


b



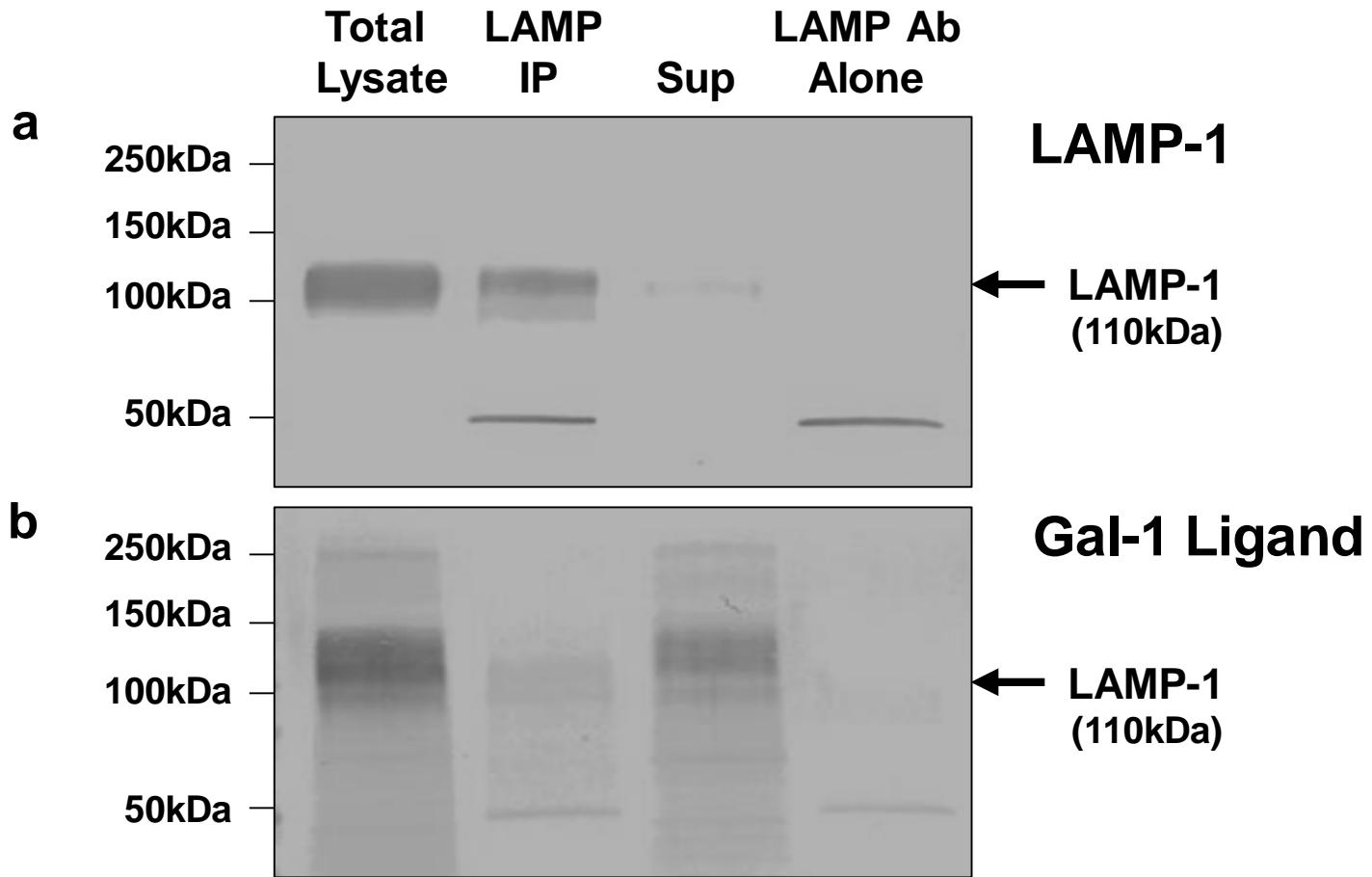
Supplemental Figure 4. Human Gal-1 binds melanoma cell Gal-1 ligands in a MCAM-dependent manner. (a) Representative histograms are shown for human scrambled control or MCAM^{KD} A375 and G361 melanoma cells FACS analyzed with human Gal-1 - hFc chimera (hGal-1) (45 μ g/ml) and anti-hFc Abs or with anti-hFc alone. **(b)** Relative binding of hGal-1 was quantified by mean fluorescence intensity after subtracting the hFc negative control. (Statistically significant difference; ***p=0.0002; **, p=0.0082). Data are representative of triplicate repeats.

Supplemental Figure 5 and Legend



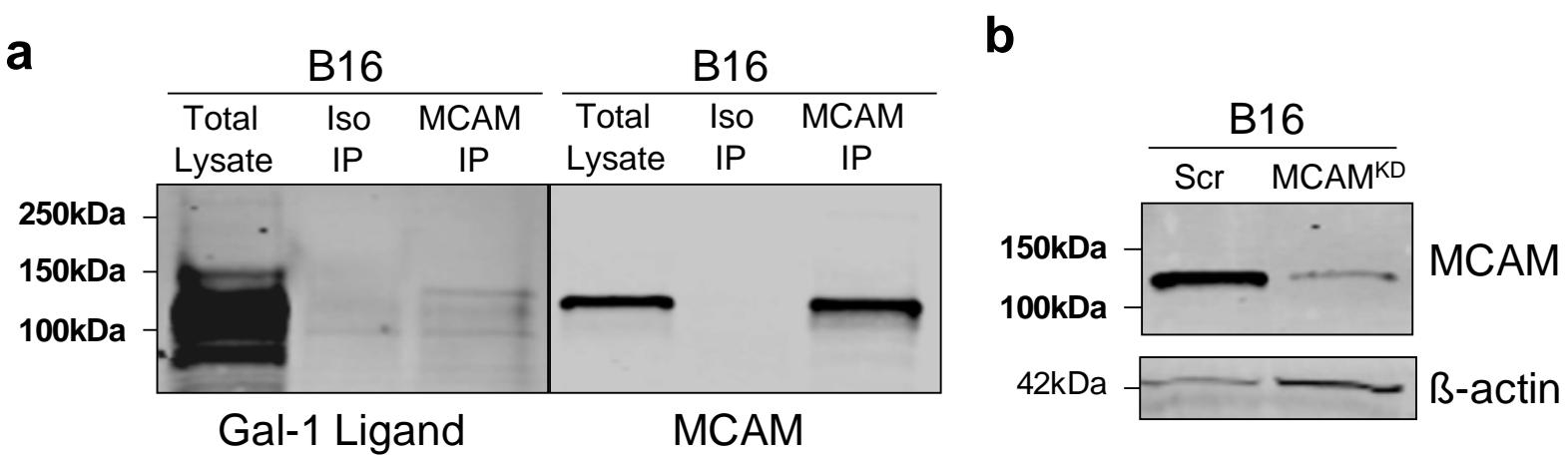
Supplemental Figure 5. The broadly-active protease, bromelain, digests the majority of melanoma cell Gal-1 ligand activity, and kifunensine effectively removes all complex N-glycans on melanoma cells. Prior to FACS assaying with Gal-1hFc, A375 and G361 melanoma cells were first treated for 1hr at 37°C with 0.1U/ml bromelain or buffer control (a). Alternatively, A375 and G361 cells were treated with 0-10µg/ml kifunensine for 48hr then FACS assayed for PHA-L-binding activity (b). Data are representative of triplicate experiments.

Supplemental Figure 6 and Legend



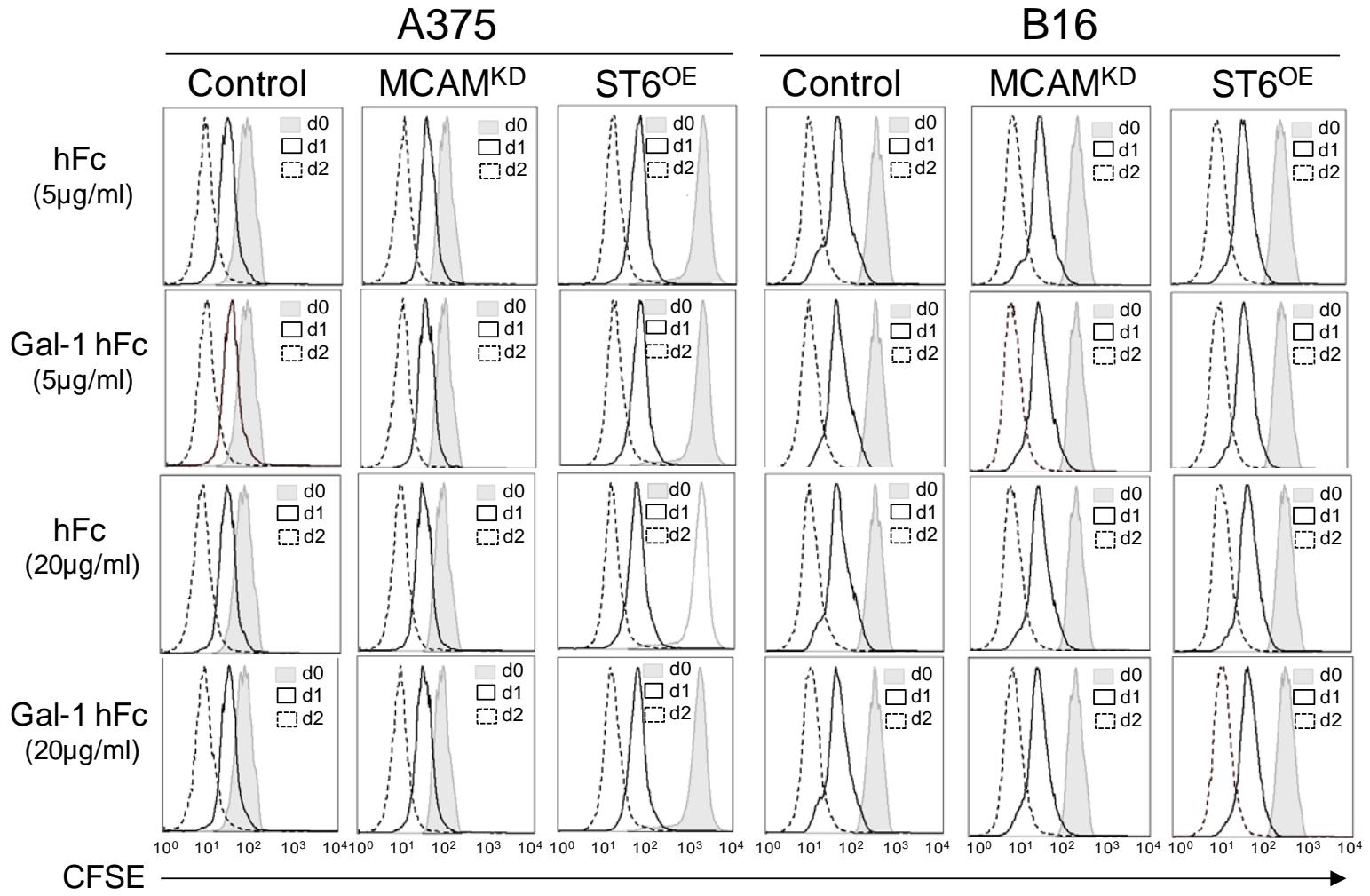
Supplemental Figure 6. LAMP-1 functions as a Gal-1 ligand on human melanoma cells.
Western blot analysis of LAMP-1 was performed on human G361 melanoma cells. Total lysate, anti-LAMP-1 (clone H4A3) immunoprecipitate (LAMP IP), residual supernatant (Sup) or H4A3 Ab alone (LAMP Ab Alone) were separated on reducing 4-20% gradient SDS-PAGE gels and blotted with either **(a)** anti-LAMP (H4A3) Ab or **(b)** Gal-1hFc to identify LAMP-1 polypeptide and LAMP Gal-1 ligand activity, respectively. All experiments were performed 3-times.

Supplemental Figure 7 and Legend



Supplemental Figure 7. Mouse melanoma B16 cells express Gal-1 ligand MCAM, and MCAM is silenced in MCAM^{KD} B16 cell variants. In (a), lysate or anti-MCAM immunoprecipitate (IP) from mouse B16 melanoma cells was blotted with Gal-1 hFc or anti-MCAM Ab. In (b), lysate from MCAM^{KD} B16 cells was blotted with anti-MCAM Ab or control anti-β-actin Ab. Data are representative of triplicate experiments.

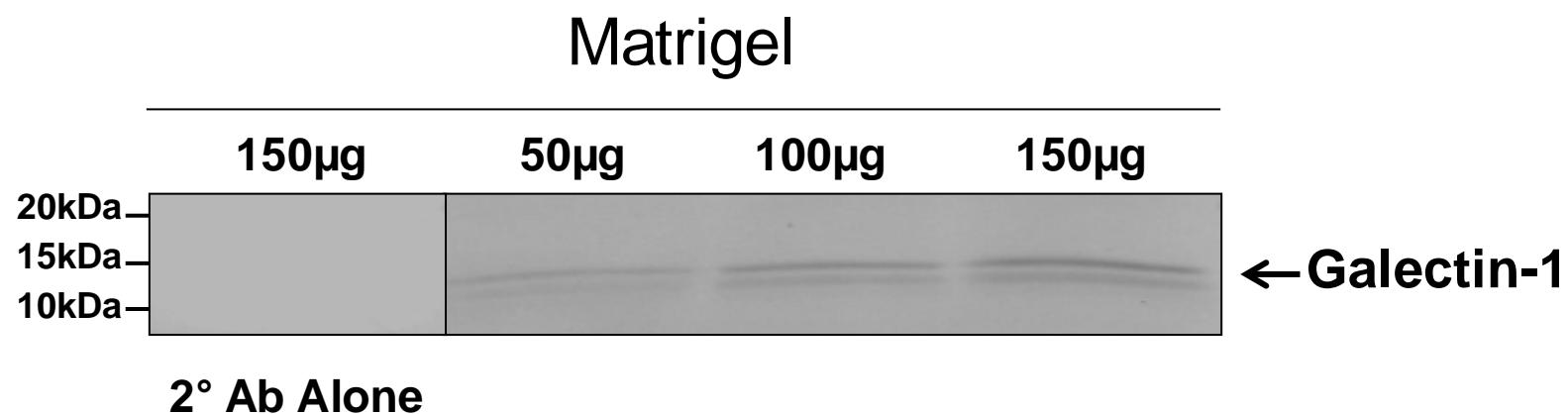
Supplemental Figure 8 and Legend



Supplemental Figure 8. Gal-1hFc incubation does not influence melanoma cell

proliferation. Prior to assays, cells were first cultured for 24h with 50mM lactose to recapitulate pretreatment conditions in cell migration assays. Control, MCAM^{KD} or ST6^{OE} A375 and B16 melanoma cells were then loaded with CFSE and then incubated with Gal-1hFc or hFc controls at 5 or 20 μ g/ml for 48hr. Cells were analyzed for fluorescence intensity by flow cytometry at day 0 (grey fill), day 1 (open fill), or day 2 (dashed line). As shown, there was no difference in diluted fluorescence intensity in Gal-1hFc-treated cells when compared with cells treated with hFc. Data are representative of replicate experiments.

Supplemental Figure 9 and Legend



Supplemental Figure 9. Gal-1 is contained in the Matrigel. Matrigel (50, 100 and 150µg protein lane) was mixed with Laemmli reducing sample buffer and Western blotted with anti-Gal-1 antibody and doublet was stained at 14kDa. A control blot stained with AP-secondary Ab alone showed no staining. Western blots were repeated 3-times.

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