

Supplemental Materials and Methods, Figures and Figure Legends

Materials and Methods

Cells. Human G361, A375, Malme3M, SKMEL2, SKMEL5, SKMEL28, UACC62, UACC257, M14, MDAMB435, 501mel, MeWo, FEMX and LOX and A2058 melanoma cells and mouse B16 F10 cells were maintained in RPMI 1640/10%FBS/1% Penicillin-Streptomycin (Invitrogen, Inc.; Carlsbad, CA) and validated for pigment and differentiation gene markers; MCAM, MITF, melanocyte antigen recognized by T cells (MART-1), tyrosinase and/or SOX10. Human T cells from PBMC isolates were activated and expanded in X-VIVO15™ (Lonza, Inc.; Hopkinton, MA) and IL-2 as described (Cedeno-Laurent *et al.*, 2012). Short-term cultures of clinical melanoma specimens from (3) donors were isolated from surgical metastases in accordance with IRB approval and cultured in RPMI 1640/10% FBS/1% penicillin/streptomycin as described (Schatton *et al.*, 2008). Early passage (P2), primary normal human epidermal melanocytes (HEM) were purchased from PromoCell, Inc. (Heidelberg, Germany). Patient consent was not necessary and all human tissues were obtained according to institutional review board approval.

Silencing of MCAM. Lentiviral shRNA constructs in pLKO.puro against MCAM or scrambled (SCr) controls were purchased from the Mission collection (Sigma-Aldrich; St. Louis, MO). Retroviral supernatants were generated by co-transfection of pN8e-GagPolΔ8.1 and pNE8e-VSV/G in the packaging HEK293t cell line. Melanoma cells were transduced in the presence of 8µg/ml polybrene and selected with 1µg/ml puromycin to generate stable knockdown cell lines. Knockdown of human and mouse MCAM was achieved with the shRNA target sequence - AGTTGAAGTTAAGTCAGATAA.

Overexpression of ST6GalNAc2. The following sequences were used for real-time qRT-PCR to validate ST6GalNAc2 levels: forward 5'-CTCGTCTCCTACTGGAATCTGG-3' and reverse 5'-CGATCTCAGCATCACATAGTCGC-3' as described (Barthel *et al.*, 2008; Barthel *et al.*, 2013). For overexpression, human ST6GalNAc2 cDNA was amplified by KOD polymerase PCR from reverse transcribed melanocyte RNA using forward: 5'-TTCTGCCTGGGACGTCAGCGGACG-3' and reverse: 5'-TCAGCGCTGGTACAGCTGAAGGA-3' primers. This ST6GalNAc2 cDNA

product was then cloned into pDONR223 by KOD amplification using forward: 5'-GGGGACAACCTTTGTACAAAAAAGTTGGCTCCACCATGGGGCTCCCGCGCGG-3' and reverse: 5'-GGGGACAACCTTTGTACAAGAAAGTTGGGTAGCGCTGGTACAGCTGAAGGATGCC-3' attB-containing primers. Subsequent LR clonase (Invitrogen)-mediated recombination into pLenti6.3-DEST (Invitrogen) yielded vector pLenti6.3-ST6GalNAc2. Cells were transduced with lentiviral supernatants and selected for stable expression with blasticidin.

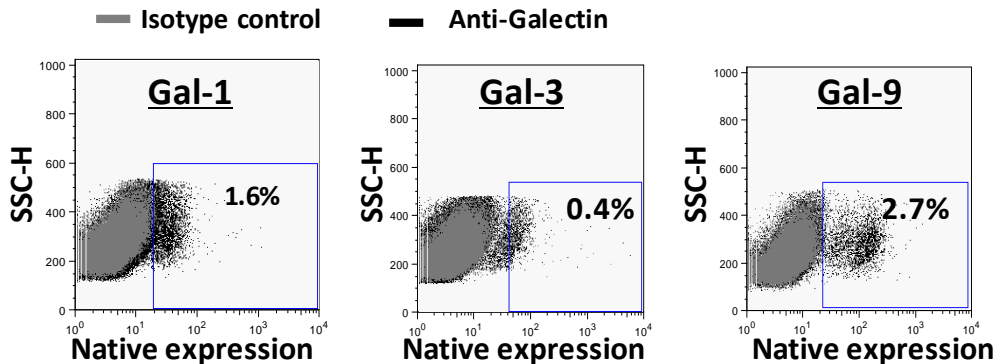
Flow Cytometry. Flow cytometric analysis using primary antibodies, including polyclonal goat anti-human Gal-1 (R&D Systems), anti-human Gal-3 (M3/38) (BioLegend), anti-human Gal-9 (9M1-3) Abs (BioLegend), human Gal-1 – hFc chimera (hGal-1) (generously provided by Dr. Kuo-I. Lin; Academia Sinica, Taiwan) (Tsai *et al.*, 2008; Tsai *et al.*, 2011), Gal-1hFc, dmGal-1hFc, isotype control Abs, and respective fluorophore-conjugated secondary Abs was performed as described (Cedeno-Laurent *et al.*, 2012). Where indicated, cells were either treated with the broadly-active protease, bromelain (Sigma), at 0.1U/ml for 1hr at 37°C to digest all cell surface glycoproteins or with kifunensine (Sigma) between 1-10µg/ml for 48hr to inhibit *de novo* synthesis of complex N-glycans and then stained with Gal-1hFc or with FITC-PHA-L (1µg/ml). Cells were also stained with biotinylated LEA (0.1µg/ml) and APC-avidin (1:500). Secondary Ab, APC-avidin or 50mM lactose-containing Gal-1hFc groups were included to control for background fluorescence and carbohydrate-dependence of Gal-1hFc, PHA-L or LEA. Experiments were performed at least three-times.

In vitro Melanoma Cell Proliferation Assay. We assayed for cell growth by CFSE-dilution assay over a 3-day period in the presence Gal-1hFc, hFc or diluent controls. Prior to assays, cells were cultured for 24h in RPMI 1640/10%FBS/1% Pen/Strep with 50mM lactose to recapitulate pretreatment conditions in cell migration assays. Cells were harvested with 1mM EDTA, washed in PBS, loaded with 2µM CFSE in PBS for 10 min at 37°C. Cells were then plated in RPMI 1640/10% FBS with diluent control, hFc or Gal-1hFc at either 5 or 20µg/ml in triplicate at 5×10^4 /well in 6-well plates. After 1, 2 and 3 days, cells were harvested and assayed by flow cytometry.

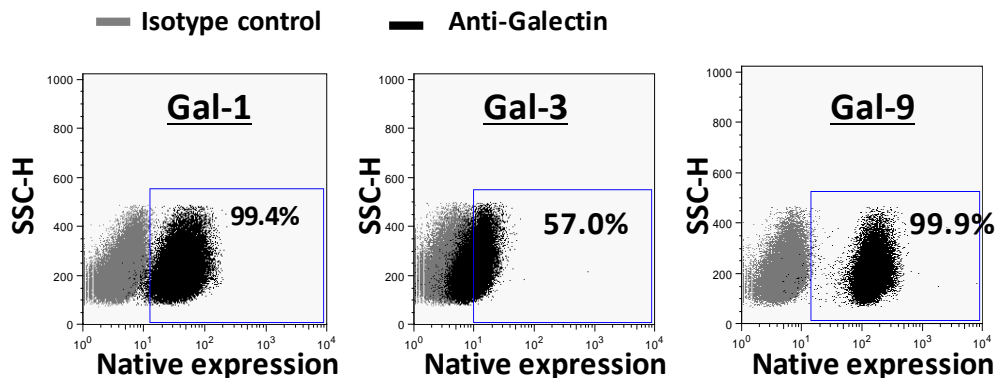
Statistical Analysis. Statistical significant comparisons were ascertained by two-tailed Student's *t*-test, paired *t*-test, one-way ANOVA with Dunnett's post test, or contingency table on GraphPad Prism (GraphPad Software, La Jolla, CA).

Supplemental Figure 1 and Legend

a FACS Analysis of Surface (Non-permeabilized) Gal-1, -3 and -9



b FACS Analysis of Intracellular (Permeabilized) Gal-1, -3 and -9



Supplemental Figure 1. Melanoma cell Gal-1, -3 and -9 are principally expressed and detected inside of the cell and not bound to the cell surface. Surface (Non-permeabilized) (a) and intracellular (permeabilized) (b) human A375 melanoma cells were FACS stained with anti-human Gal-1, -3, or -9 Abs or respective fluorophore-conjugated secondary Ab. Cells were analyzed for fluorescence intensity (x-axis) and designated as percent positive (black dots) compared with cells stained with control Ab alone (gray dots). Data are representative of triplicate experiments.

Identified Protein

Accession Number

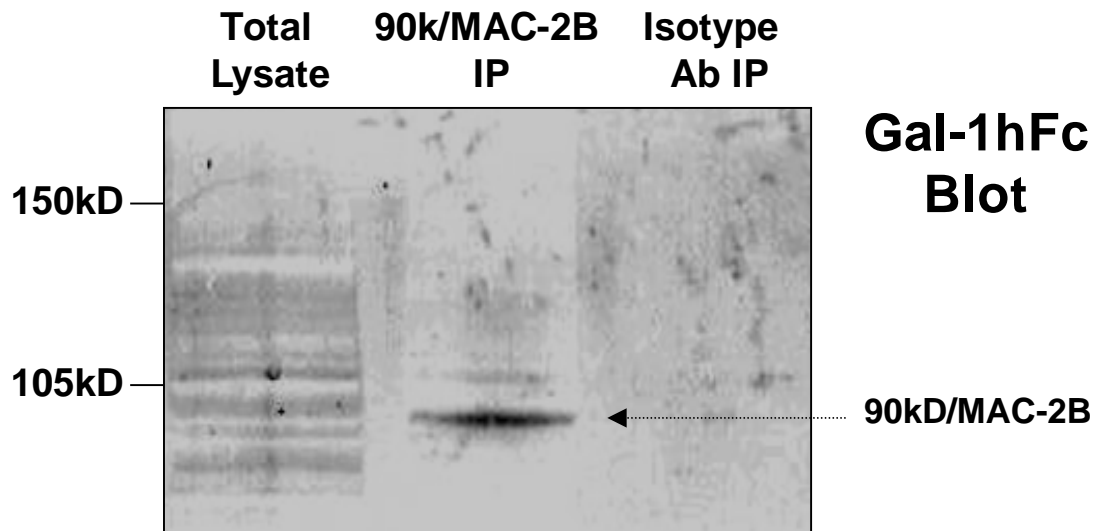
MW

Negative Control Gal-1 Eluate

Table with columns: Identified Protein, Accession Number, MW, Negative Control, Gal-1 Eluate. Lists various proteins and their corresponding accession numbers and molecular weights, with peptide match counts for negative control and Gal-1 eluate.

Supplementa Figure 2. Identification of Gal-1-binding proteins by tandem MS/MS. The following table lists human membrane proteins and their accession number identified in eluates from protein G chromatography of human G361 cell lysate using Gal-1hFc or negative control dmGal-1hFc as probes. Serine/threonine phosphorylation spectral counts (or # Peptide matches) correspond with respective predicted proteins.

Supplemental Figure 3 and Legend

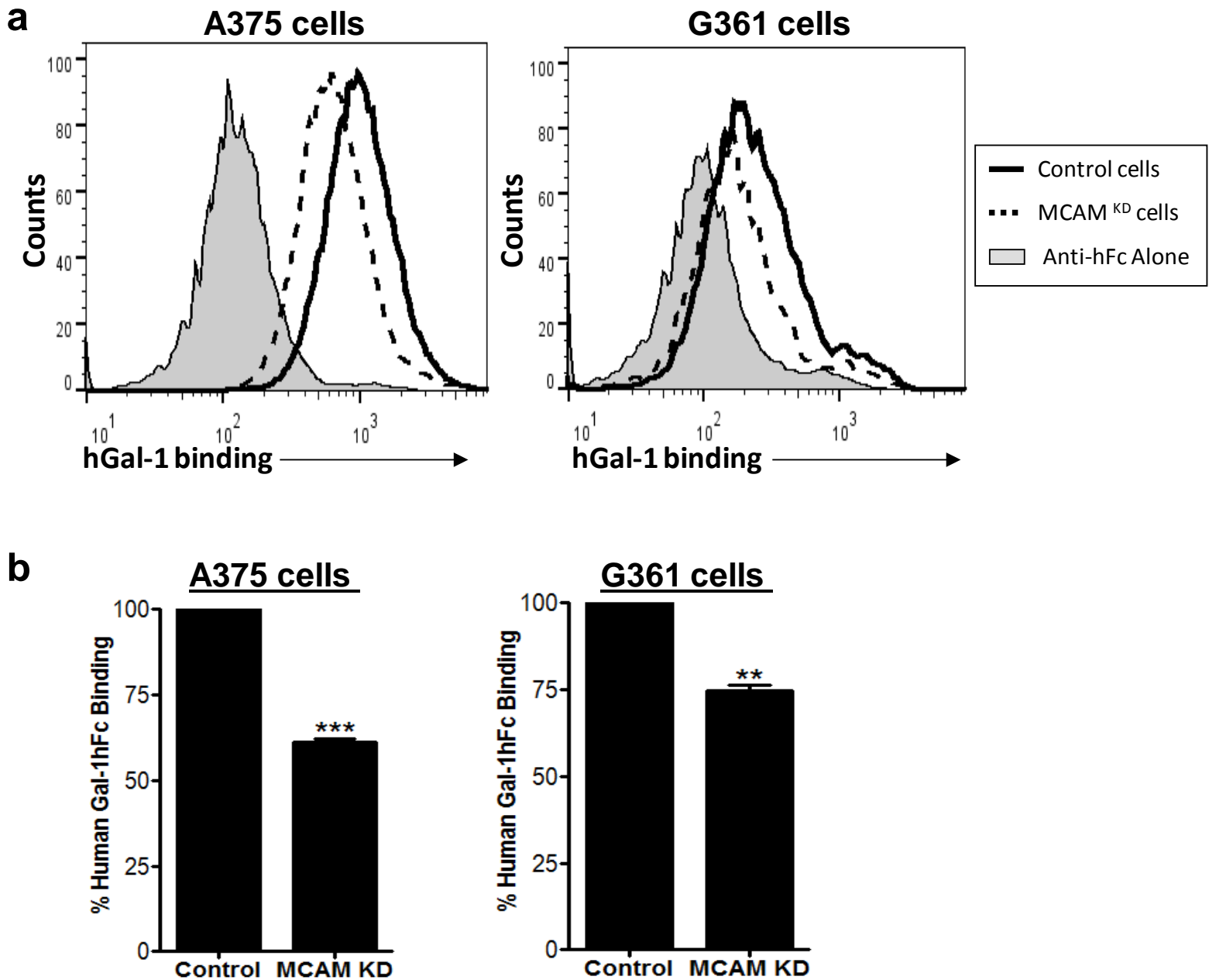


Supplemental Figure 3. Detection of Gal-3-binding Protein (90k/MAC-2B)

immunoprecipitated from human melanoma cells with Gal-1hFc. Gal-3-binding protein (90k/MAC-2B) was immunoprecipitated with anti-90k moAb SP-2 or with isotype control Ab from human A375 melanoma cell lysate and blotted with Gal-1hFc. Total lysate, anti-90k immunoprecipitate (IP) and isotype control Ab IP were separated on reducing 4-20% gradient SDS-PAGE gels and blotted with Gal-1hFc. Data was representative from replicate experiments.

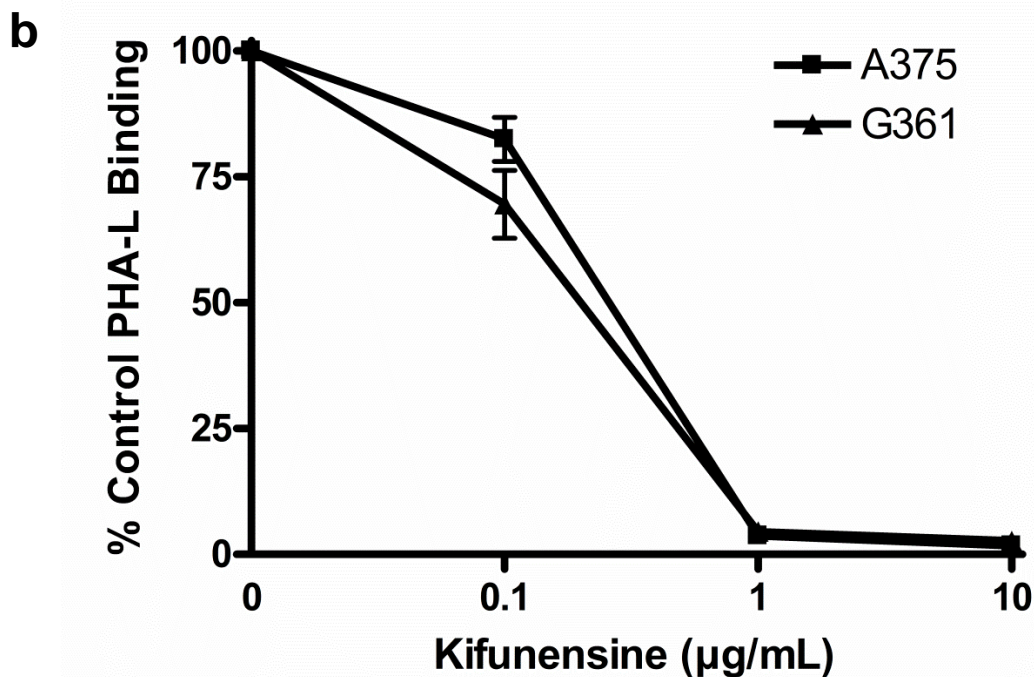
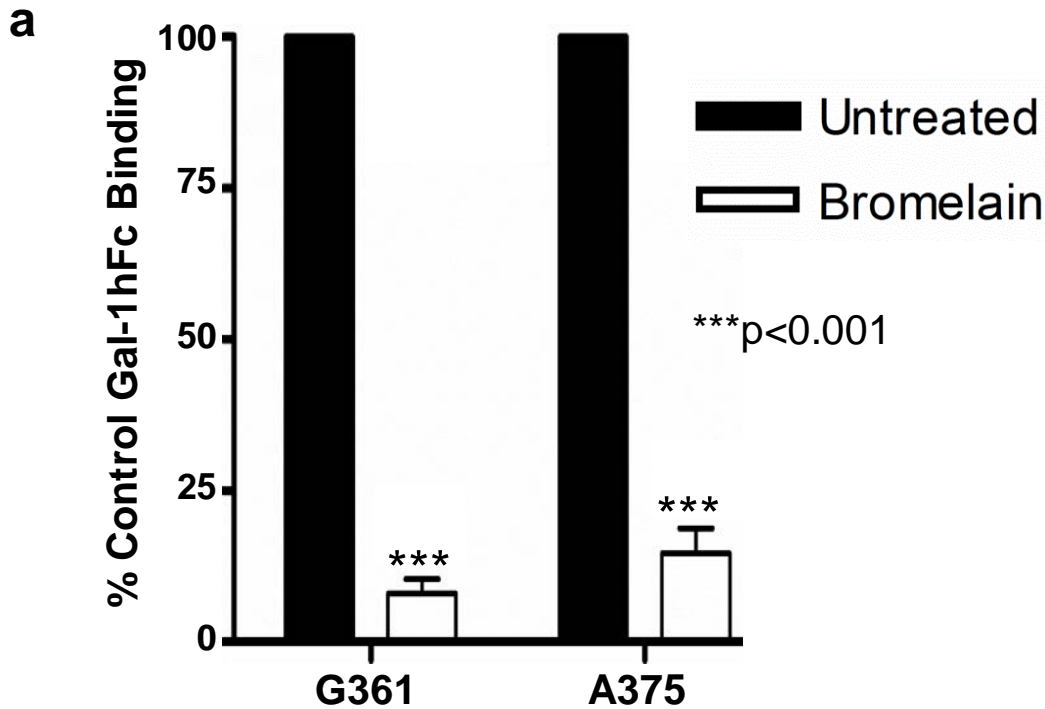
Supplemental Figure 4 and Legend

Human Gal-1 binding to melanoma cell Gal-1 ligands is dependent on MCAM.



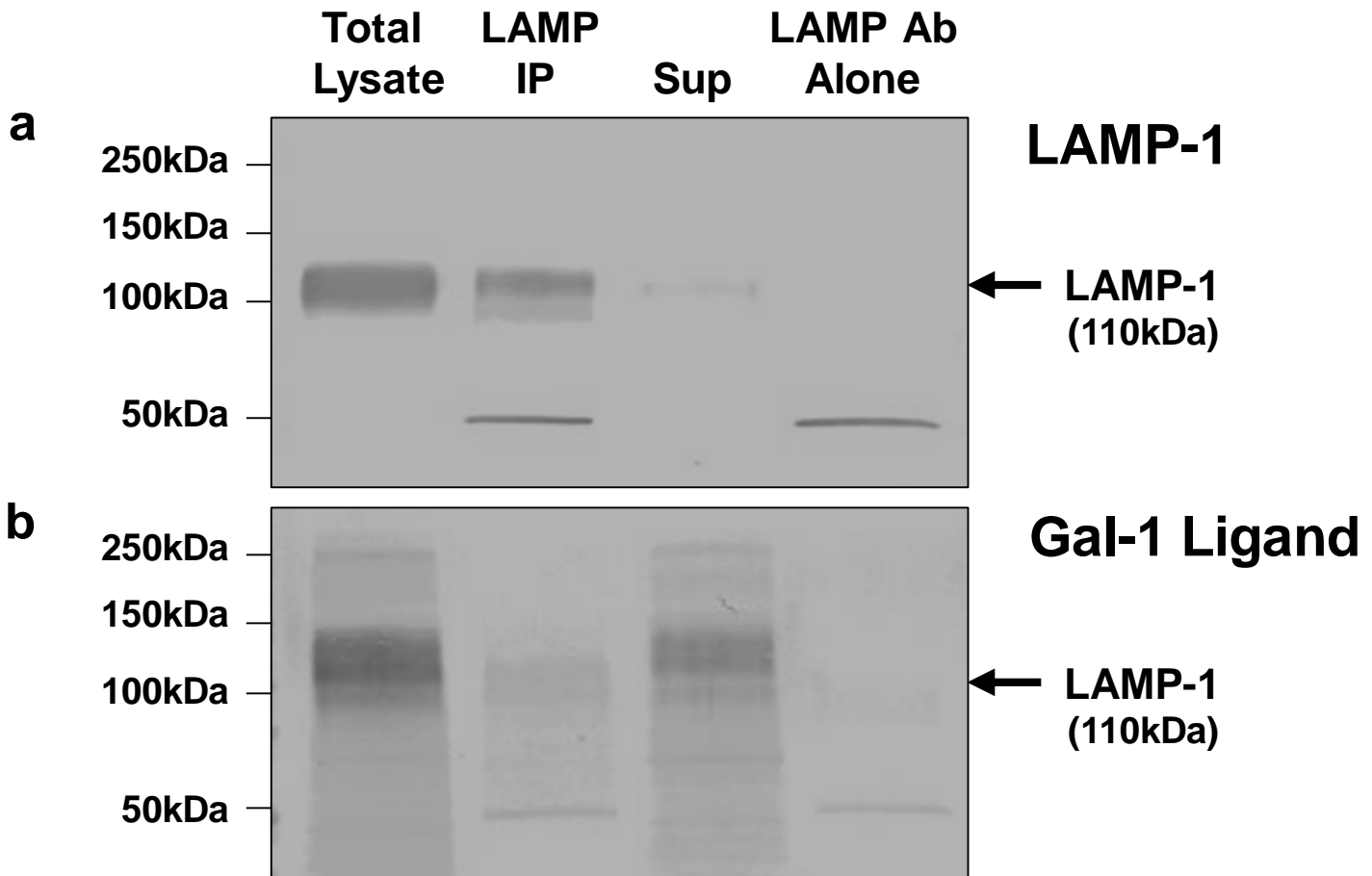
Supplemental Figure 4. Human Gal-1 binds melanoma cell Gal-1 ligands in a MCAM-dependent manner. (a) Representative histograms are shown for human scrambled control or MCAM^{KD} A375 and G361 melanoma cells FACS analyzed with human Gal-1 - hFc chimera (hGal-1) (45µg/ml) and anti-hFc Abs or with anti-hFc alone. (b) Relative binding of hGal-1 was quantified by mean fluorescence intensity after subtracting the hFc negative control. (Statistically significant difference; ***p=0.0002; **, p=0.0082). Data are representative of triplicate repeats.

Supplemental Figure 5 and Legend



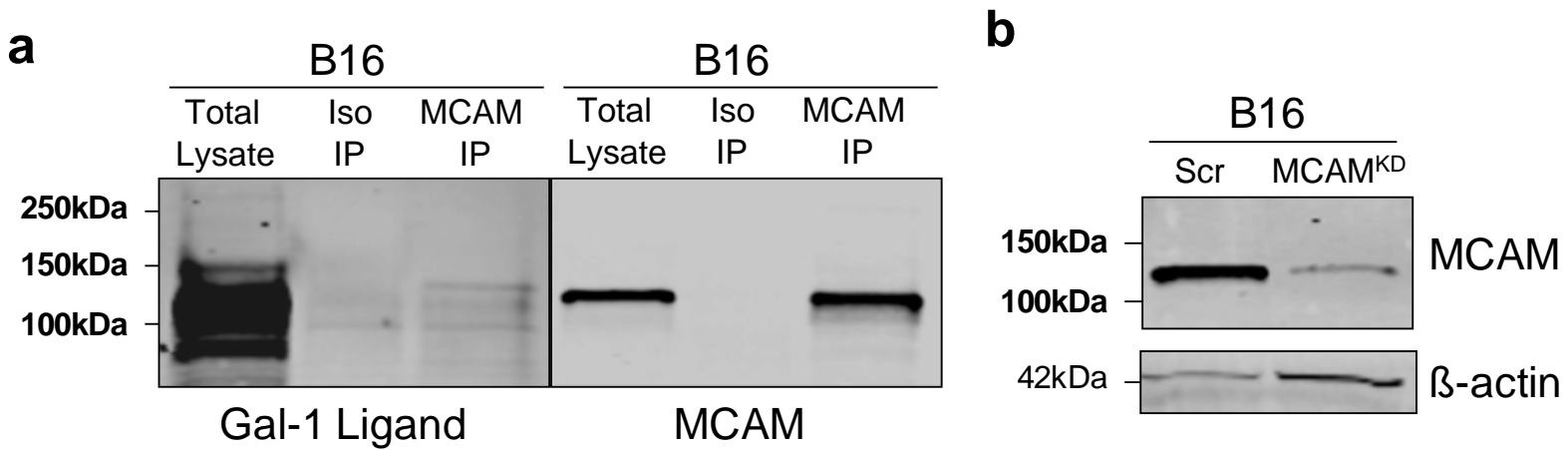
Supplemental Figure 5. The broadly-active protease, bromelain, digests the majority of melanoma cell Gal-1 ligand activity, and kifunensine effectively removes all complex N-glycans on melanoma cells. Prior to FACS assaying with Gal-1hFc, A375 and G361 melanoma cells were first treated for 1hr at 37°C with 0.1U/ml bromelain or buffer control (**a**). Alternatively, A375 and G361 cells were treated with 0-10µg/ml kifunensine for 48hr then FACS assayed for PHA-L-binding activity (**b**). Data are representative of triplicate experiments.

Supplemental Figure 6 and Legend



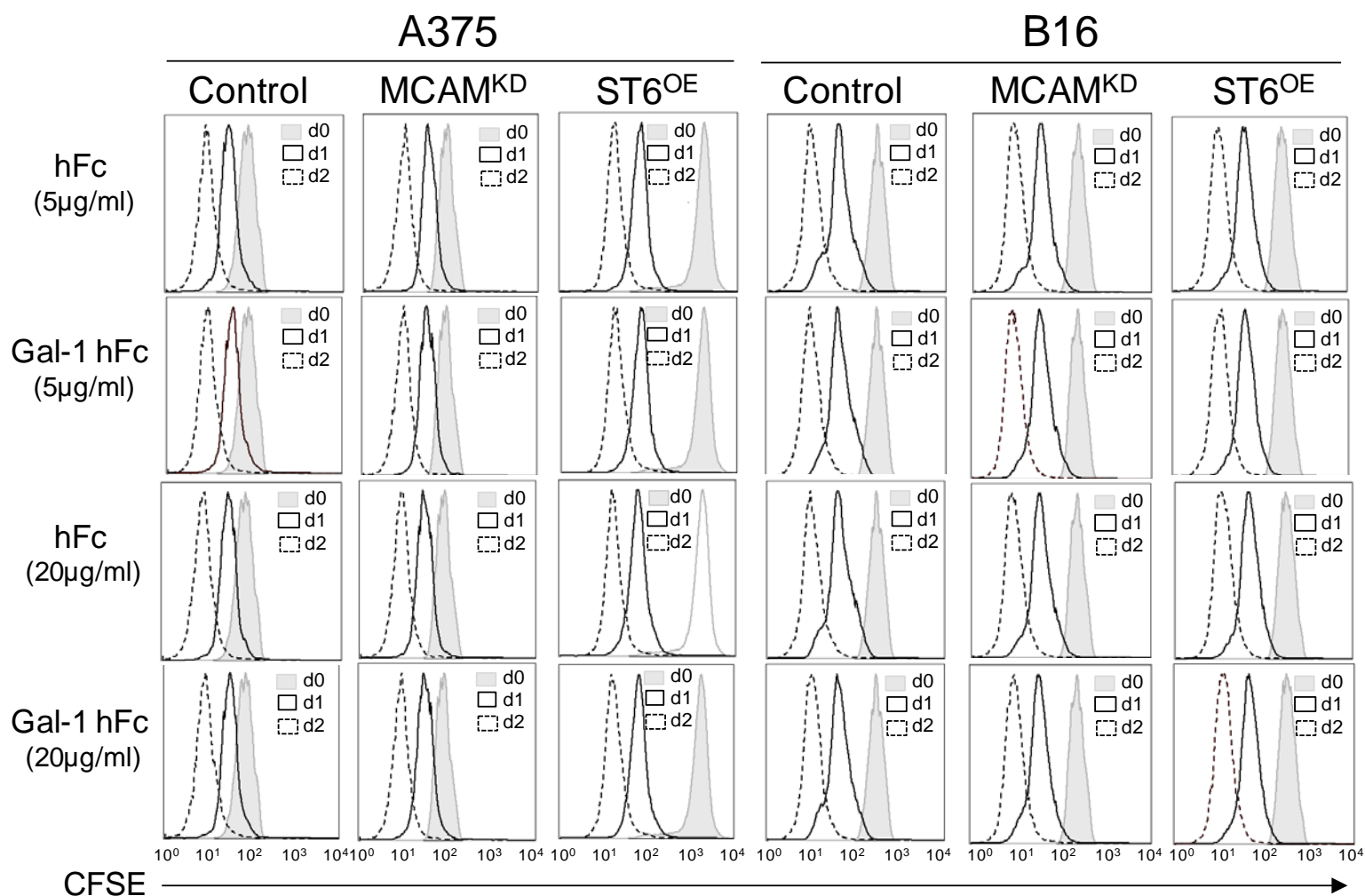
Supplemental Figure 6. LAMP-1 functions as a Gal-1 ligand on human melanoma cells. Western blot analysis of LAMP-1 was performed on human G361 melanoma cells. Total lysate, anti-LAMP-1 (clone H4A3) immunoprecipitate (LAMP IP), residual supernatant (Sup) or H4A3 Ab alone (LAMP Ab Alone) were separated on reducing 4-20% gradient SDS-PAGE gels and blotted with either **(a)** anti-LAMP (H4A3) Ab or **(b)** Gal-1hFc to identify LAMP-1 polypeptide and LAMP Gal-1 ligand activity, respectively. All experiments were performed 3-times.

Supplemental Figure 7 and Legend



Supplemental Figure 7. Mouse melanoma B16 cells express Gal-1 ligand MCAM, and MCAM is silenced in MCAM^{KD} B16 cell variants. In **(a)**, lysate or anti-MCAM immunoprecipitate (IP) from mouse B16 melanoma cells was blotted with Gal-1 hFc or anti-MCAM Ab. In **(b)**, lysate from MCAM^{KD} B16 cells was blotted with anti-MCAM Ab or control anti-β-actin Ab. Data are representative of triplicate experiments.

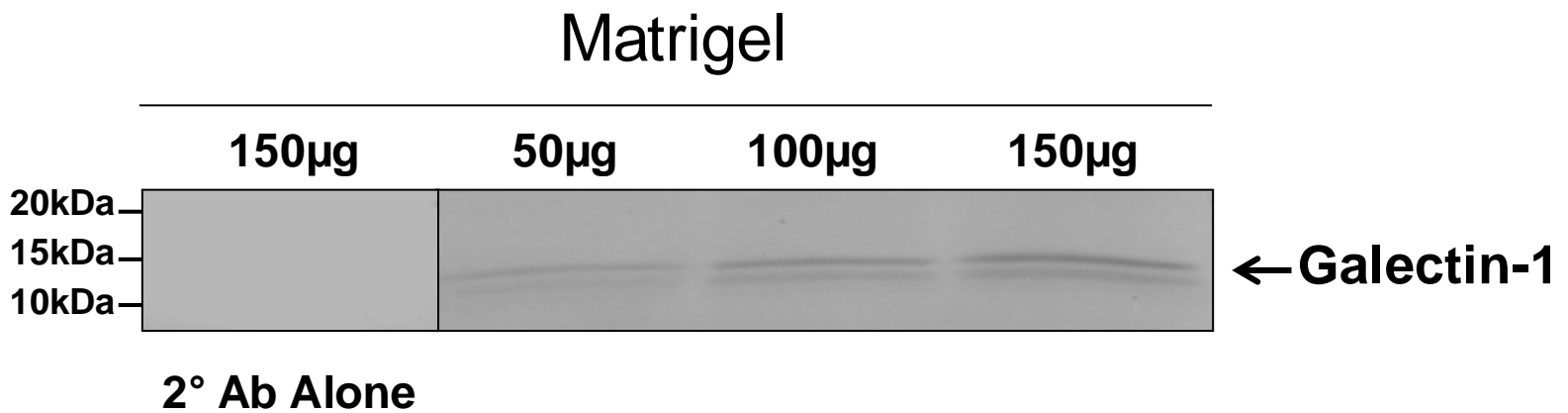
Supplemental Figure 8 and Legend



Supplemental Figure 8. Gal-1hFc incubation does not influence melanoma cell

proliferation. Prior to assays, cells were first cultured for 24h with 50mM lactose to recapitulate pretreatment conditions in cell migration assays. Control, MCAM^{KD} or ST6^{OE} A375 and B16 melanoma cells were then loaded with CFSE and then incubated with Gal-1hFc or hFc controls at 5 or 20µg/ml for 48hr. Cells were analyzed for fluorescence intensity by flow cytometry at day 0 (grey fill), day 1 (open fill), or day 2 (dashed line). As shown, there was no difference in diluted fluorescence intensity in Gal-1hFc-treated cells when compared with cells treated with hFc. Data are representative of replicate experiments.

Supplemental Figure 9 and Legend



Supplemental Figure 9. Gal-1 is contained in the Matrigel. Matrigel (50, 100 and 150 μ g protein lane) was mixed with Laemmli reducing sample buffer and Western blotted with anti-Gal-1 antibody and doublet was stained at 14kDa. A control blot stained with AP-secondary Ab alone showed no staining. Western blots were repeated 3-times.

References

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