

A novel male sterility-fertility restoration system in plants for hybrid seed production

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Supplementary Information

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Supplementary Table S1. List of primers used in this study

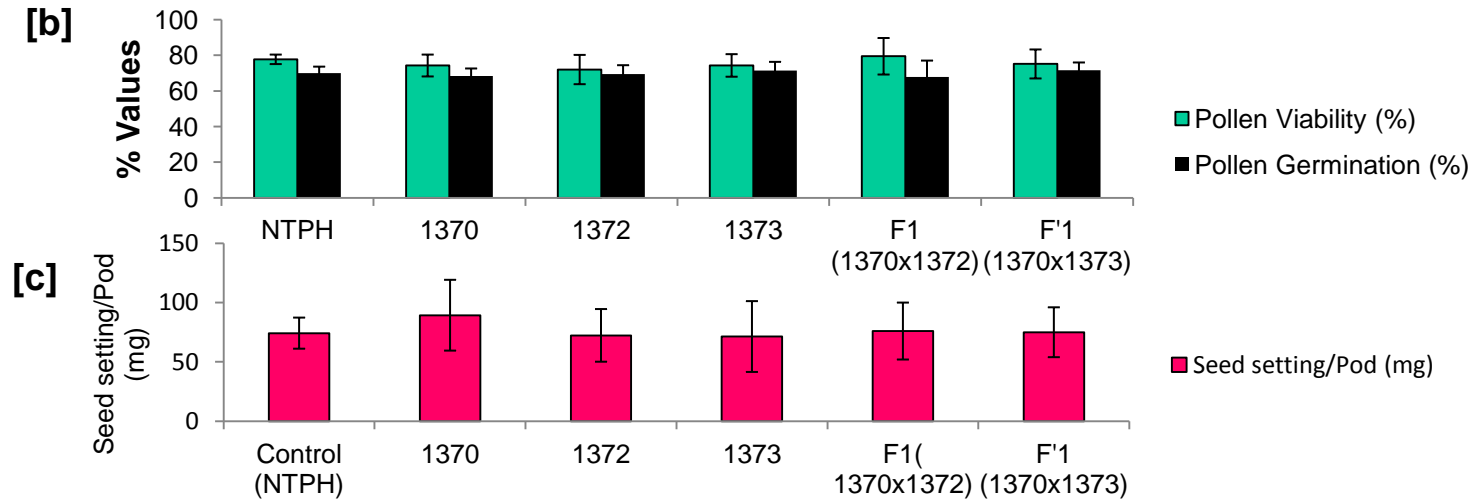
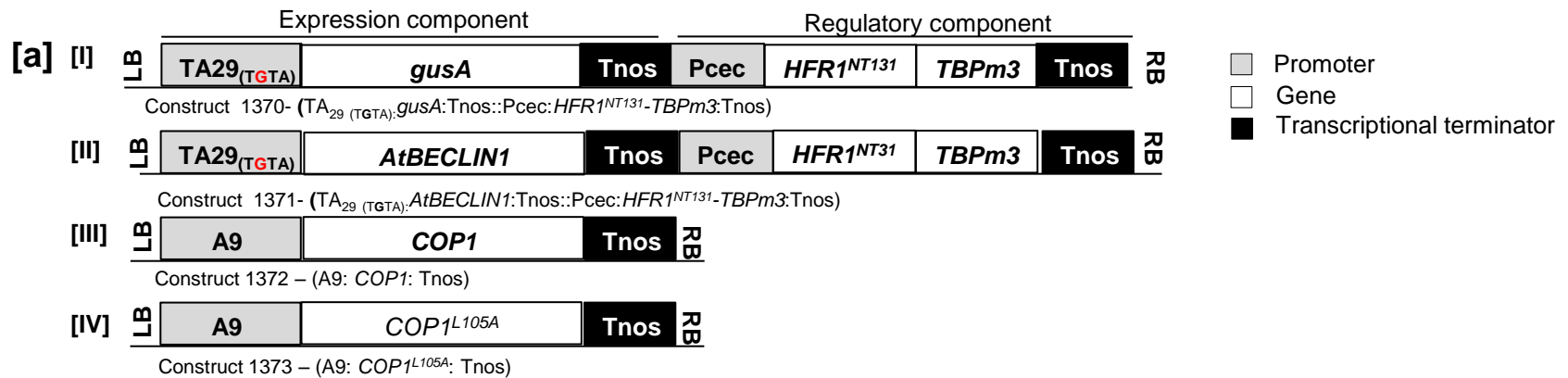
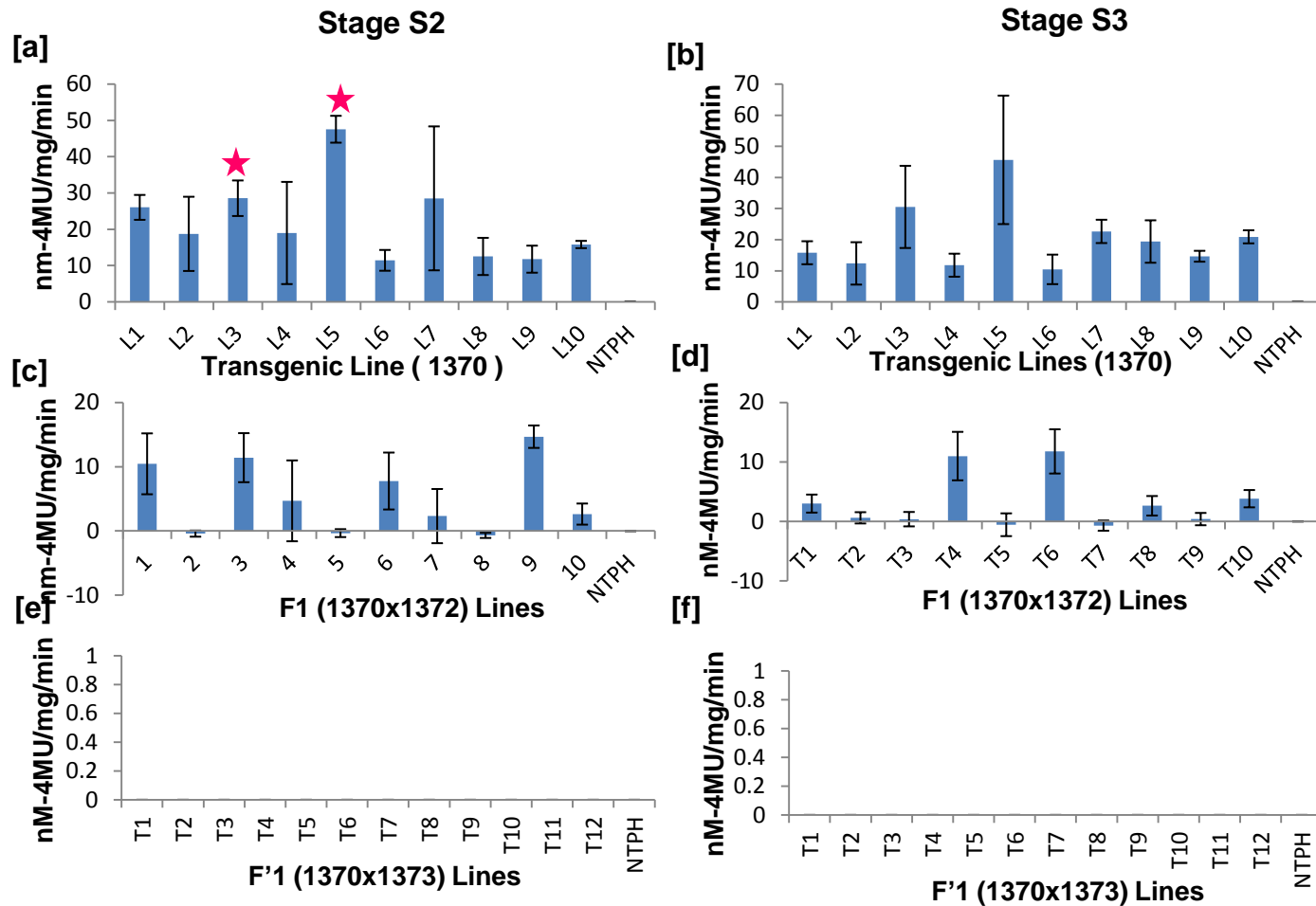


Fig. S1 Schematic representation of expression cassettes and analysis of pollen fertility.

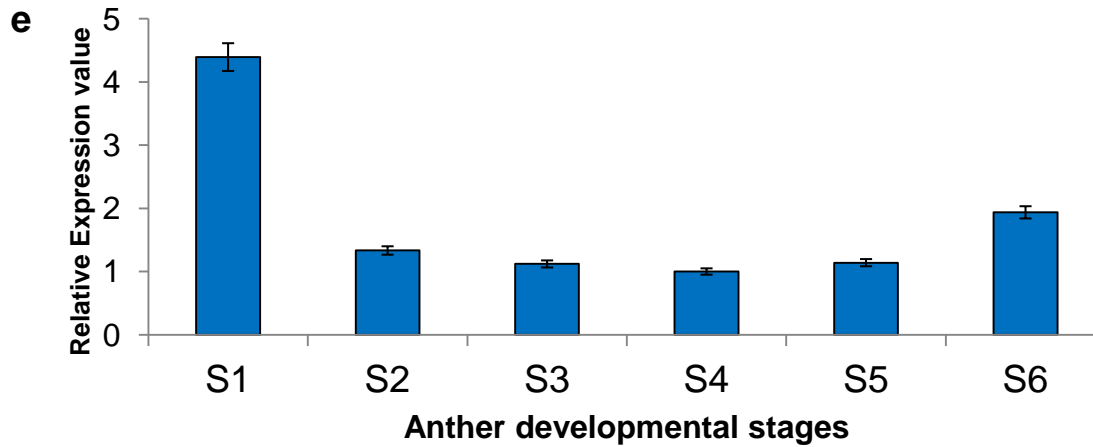
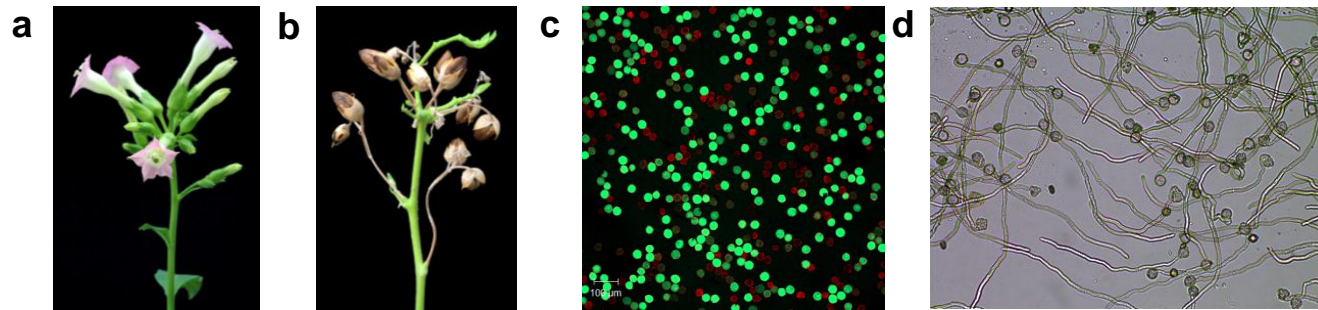
[a] Construct 1370 with two transcriptional units; expression component (*TA*_{29(TGTA)}:*gusA*:Tnos) which is regulated by regulatory component (Pcec:*HFR1^{NT131}*-*TBPm3*:Tnos), cloned in LB and RB of binary vector pBI101 [I]. Construct 1371 where *gusA* gene was replaced by Arabidopsis *BECLIN1/ATG6* in 1370 making two-component ES (*TA*_{29 (TGTA)}:*AtBECLIN1*: Tnos:: Pcec:*HFR1^{NT131}*-*TBPm3*:Tnos) cloned in LB and RB of binary vector pBI101 [II]. Construct 1372 expressing *COP1* using A9 promoter, cloned in LB and RB of modified pCambia1300 binary vector [III]. Construct 1373 expressing *COP1^{L105A}* using A9 promoter, cloned in LB and RB of modified pCambia1300 binary vector [IV]. [*COP1^{L105A}*= *COP1* with mutation in the CLS to increase nuclear abundance, modified pCambia1300= modified by replacing CaMV35S promoter by pNOS promoter to avoid leaky expression of *COP1* due to bidirectional expression possibility by CaMV35S promoter]

Pollen viability, in-vitro pollen germination [b], and seed setting per pod (in mg) [c] in control, 1370, 1372, 1373 transgenic lines, F1 (1370x1372) and F'1 (1370x1373). Error bar represents SD of 10 independent replicates.



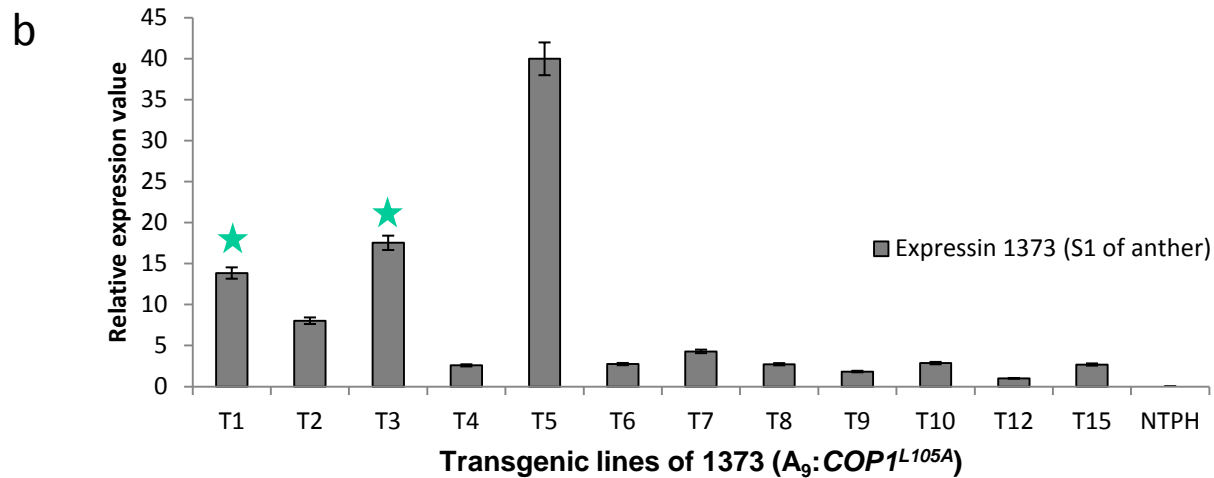
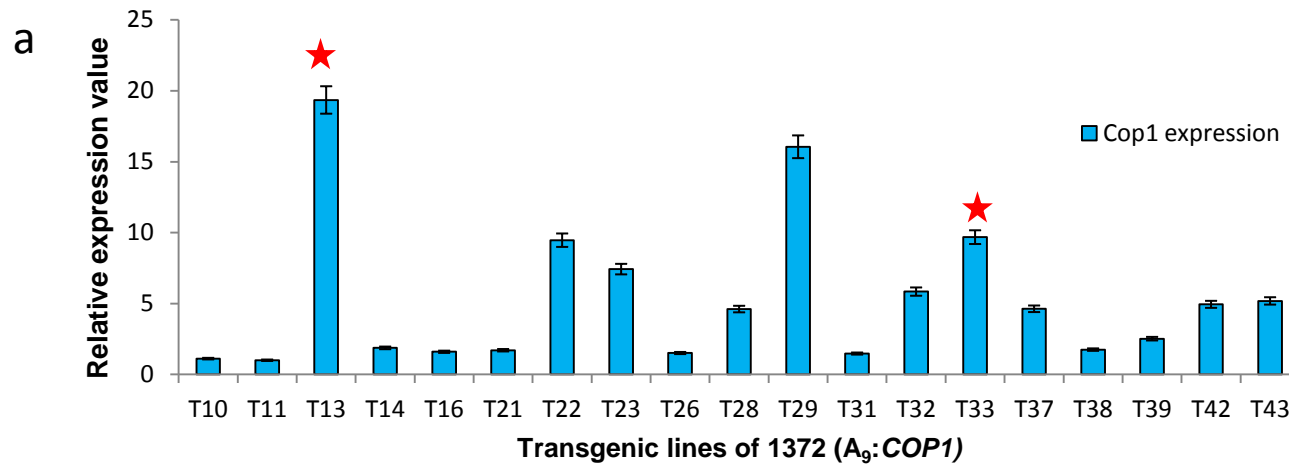
Supplementary Figure S2. Fluor-metric GUS expression analysis.

[a-b] GUS expression analysis in 1370 ($TA_{29(m)}:GUS:Pcec:HFR1^{NT131}-TBPm3$) transgenic lines of anther developmental stage 2 [a] and 3 [b]. **[c-d]** GUS expression analysis in the F1 anther stage 2 [c] and 3 [d]. The F1 generated by crossing with 1370 (♀) line 3 (L3) and line 5 (L5) with 1372 ($A9:COP1$) lines (♂) T13 and T33. **[e-f]** GUS expression analysis in the F'1 anther stage 2 [e] and 3 [f]. The F'1 generated by crossing with 1370 (♀) line 3 (L3) and line 5 (L5) with 1373 ($A_9:COP1^{L105A}$) lines (♂) T1 and T3. (The expression values are normalized with control (NTPH) values on their respective stages 2 and 3. star indicates the lines of 1370 selected for crossing as female parent. Error bar represent three replicate, each with n=3)



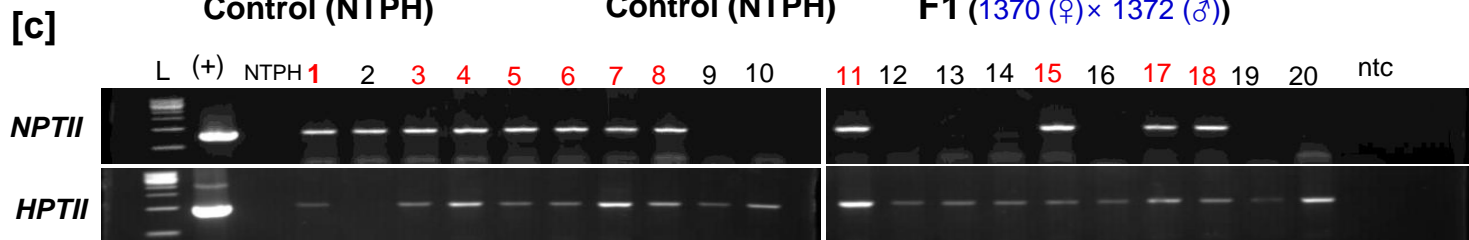
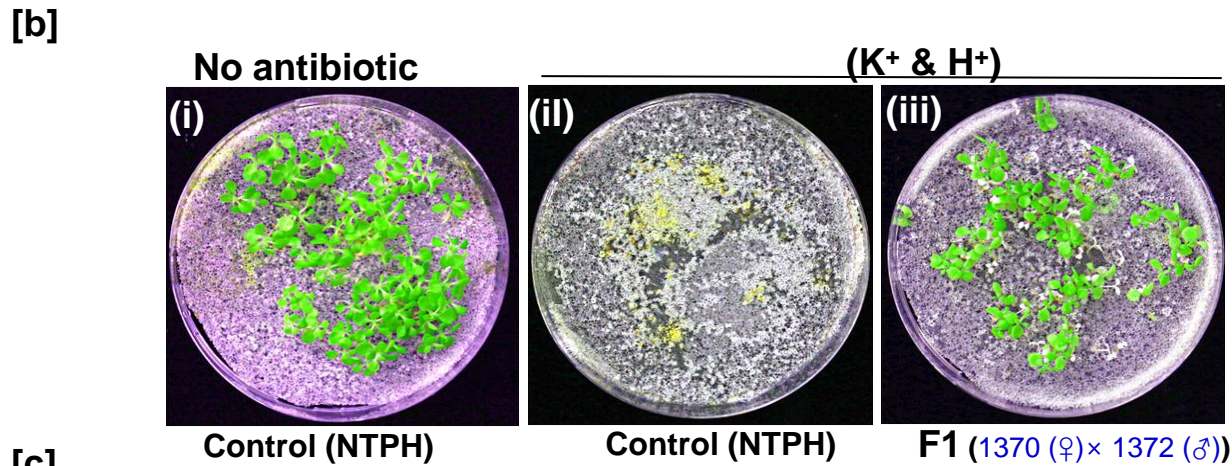
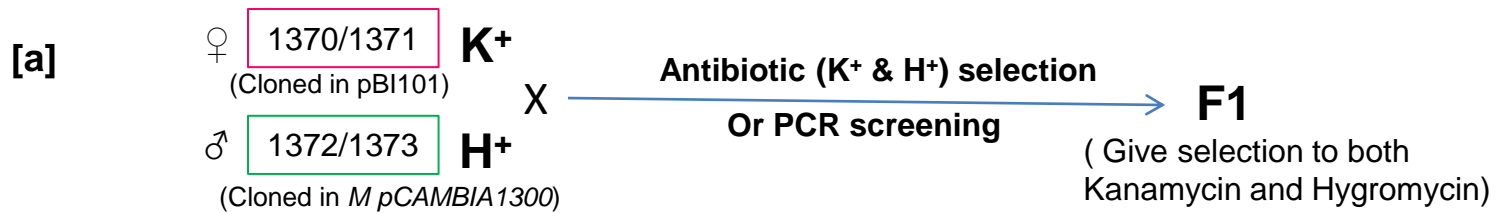
Supplementary Figure S3. Analysis of 1372 (A9: *COP1*) transgenic lines.

[a-d] Transgenic line of 1372 with normal flowering [a] and seed setting [b]. Pollens fertility as evaluated by pollen viability [c] and in-vitro pollen germination [d]. [e] qRT-PCR analysis for relative expression of *COP1* in the anther developmental stage 1 of 1372 transgenic lines. *COP1* expression was found to be specific to the stage 1 of the anther. Error bar represent SE of three independent Experiments. *UBIQ10* used as internal control.



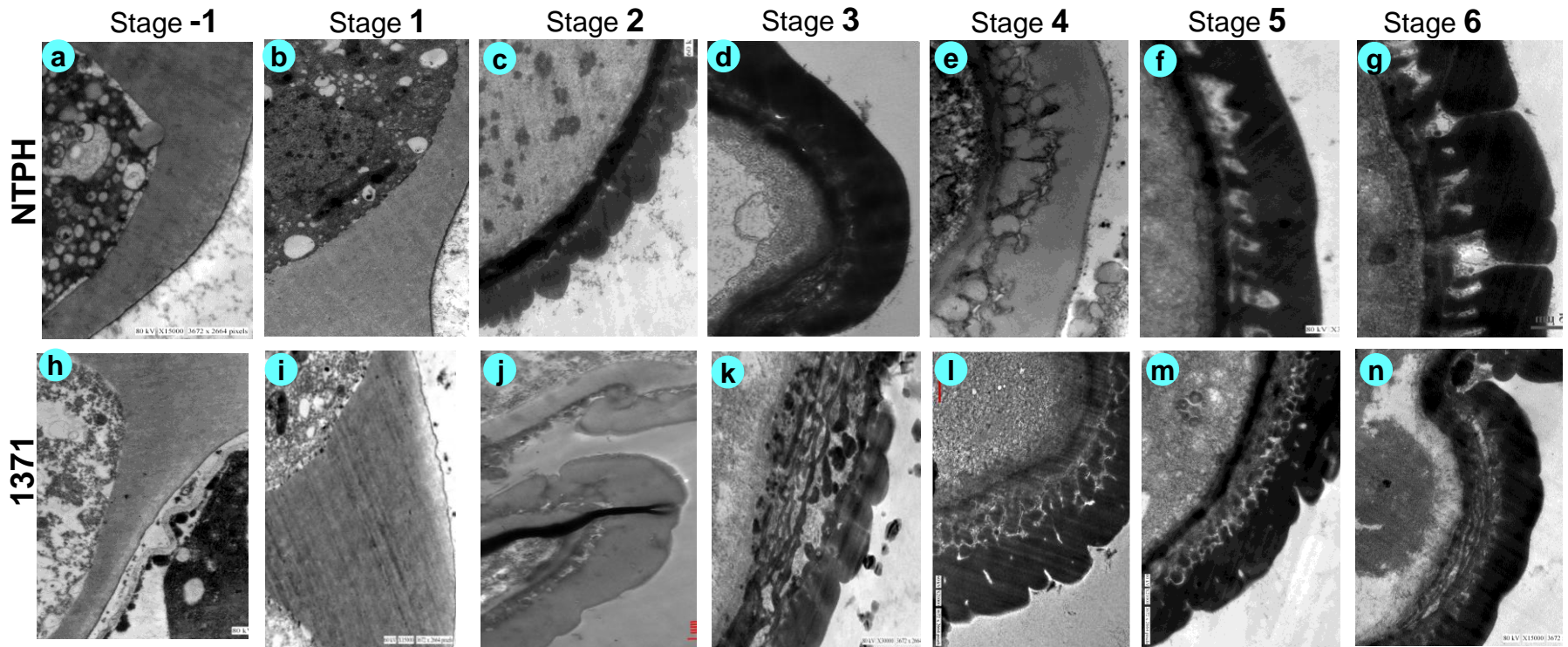
Supplementary Figure S4. Screening of best expressing transgenic lines.

[a] qRT-PCR analysis of stage 1 anther of different transgenic lines of 1372 ($A_9:COP1$), transgenic line 13 (T13) and 33 (T33) were selected for crossing (as male parent). **[b]** qRT-PCR analysis of stage 1 anther of different transgenic lines of 1373 ($A_9:COP1^{L105A}$), transgenic line 1(T1) and 3 (T3) were selected for crossing (as male parent). Error bar represents SE of three independent experiment (n=3) Tobacco *UBIQ10* taken as internal control.



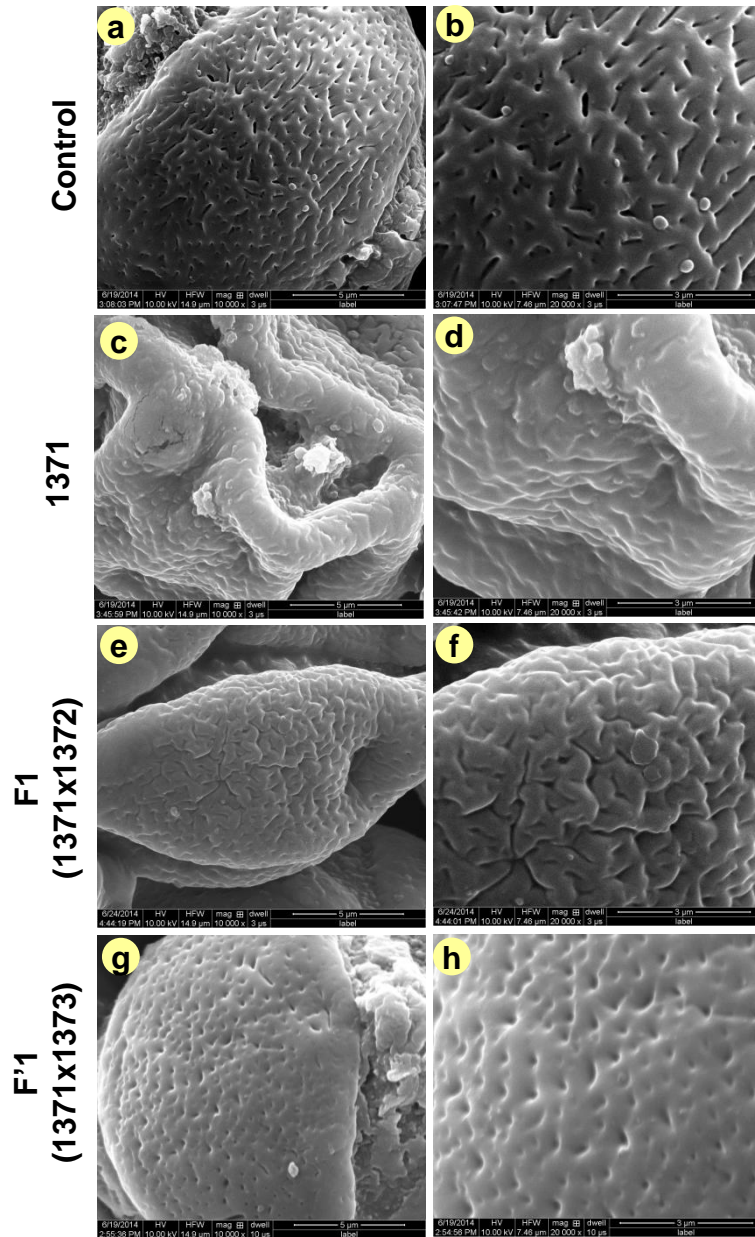
Supplementary Figure S5. Screening of F1 lines.

[a] Strategy of the F1 screening. **[b]** Screening of F1 seeds on the double antibiotic selection ; Control (NTPH) seeds were sown without antibiotic selection (i) and with double selection of K⁺ and H⁺ (ii), while F1 seeds germinated on the double selection (iii). **[c]** PCR screening of F1s; F1 seeds obtained were sown directly in the soil and genomic DNA was isolated from their seedlings, slandered PCR screening was performed using *nptII* and *hptII* specific primer . F1 lines giving *nptII* and *HptII* positive amplification are the F1s. Positive control taken were vector construct and *nph* as negative control while non-templated control (ntc) as null control.



Supplementary Figure S6. Pollen wall architecture in control (NTPH) and Beclin1 transgenic (1371).

Stage -1 pollens covered with callose deposition in both control [a] and 1371 transgenic [b] which is consistent in Stage 1 the ([b] and [i]). Stage 2 pollen showing exine and intine development in control [c] while undecomposed callose was remain deposited over the 1371 [j]. Exine sculpturing was progressively developed in control in stage 3 [d], stage 4 [e], stage 5 [f] and stage 6 [g] resulting in the well developed wall architecture, whereas in 1371 transgenic defective pollen wall development started in stage 3 [k], followed by stage 4 [l], 5 [m] and complete collapse in stage 6 [n].



Supplementary Figure S7. Pollen surface morphology in control (NTPH), Beclin1 transgenic (1371), F1 and F'1.

Pollen surface of control showing reticulated exine architecture (**[a]** and **[b]**). In 1371 transgenic deformed, shrunken pollen wall without pore was found (**[c]** and **[d]**) whereas some degree of pollen deformation was limited in F1 (**[e]** and **[f]**). F'1 showing surface architecture (**[g]** and **[h]**) similar to control. Bar= 5 μ m in [a], [c], [e], and [g] and 3 μ m in [b], [d], [f], and [h].

Supplementary Tables

Table_S1 : List of primers used in this study				
S.No.	Gene/target	Primers	restriction site	Used for
1	TA29	Forward: 5'-CGCGGATCCAGATCTTCCAACACCATTTACTCCAAGGG-3'	<i>Bam</i> Hl	promoter amplification
		Reverse: 5'-CTAGTCTAGATTTAGCTAATTT CTTTAAGTAAAACT-3'	<i>Xba</i> l	
		Forward: 5' TGTATGTCTTGTGCTCTGTATATGCCCTTGTGGTG 3'		Site directed mutagenesis
		Reverse: 5' CACCACAAGGGCA TATACAGAGCACAAGACATACA 3'		
2	<i>Beclin1/ATG6</i>	Forward: 5'-CTAGTCTAGAATGAGGAAAGAGGAGATTCCAGA-3'	<i>Xba</i> l	Gene amplification
		Reverse: 5'-CGTCGAGCTCCTAAGTTT TTTTACATGAAGGCTTA-3'	<i>Sac</i> l	
		Forward: 5'AGGGCATTCTCCACGTC3'		qRT-PCR
		Reverse: 5' AAGAGACA GATTGTGAGAACCACCA 3'		
3	<i>HFR1</i>	Forward: 5'CCATCGATATGTCGAATAATCAAGCTTTCATGG 3'	<i>Cl</i> aI	Gene amplification
		Reverse: 5'CCATCGATTCTTGTAAGTCTCCGATTCATC3'	<i>Cl</i> aI	
4	COP1	Forward: 5'CCGCTCGAGATGGAAGAGATTTTCGACGGATCC3'	<i>Xho</i> l	Gene amplification
		Reverse: 5'CGAGCTCTCACGCAGCGAGTACCAGAACTT 3'	<i>Sac</i> l	
		Forward: 5'GCTTTACCCTAATTTTCGCGGCCCGATAAGCTATTGAAGAAACTTC 3'		Site directed mutagenesis
		Reverse: 5'GTTTTCTTCAATAGCTTATCGGCCGCGAAATTAGGGTAAAGCTG 3'		
		Forward: 5' TAATTTCTTGCTCGATAAAGGCAGCGAAGAAACTTCAGCTCGGC 3'		qRT-PCR
		Reverse: 5' CGAGCTGAAGTTTTCTTCGCTGCCTTATCGAGCAAGAAATTAGG 3'		
		Forward: 5'AAGCGGCGGTTCTGAGATT3'		qRT-PCR (COP1m)
		Reverse: 5'ACCACAAGCCGTGAGGAAAG3'		
Forward: 5'-AAGCGGCGGTTCTGAGATT-3'				
Reverse: 5'-ACCACAAGCCGTGAGGAAAG-3'				
5	A9	Forward: 5'-ACGCGTCTAGACATAACGGTGAGAGTTAA-3'	<i>Sal</i> l	promoter amplification
		Reverse: 5'-TCCCATCGATTCTAATTAGATACTATATTGTTTGTAC-3'	<i>Cl</i> aI	
6	<i>UbiQ10</i>	Forward: 5'CCACGGAGACGGAGGACAA3'		qRT-PCR
		Reverse: 5'GAAGCA GCTCGAGGATGGAA3'		