Carboxyl-terminal domain of MUC16 imparts tumorigenic and metastatic functions through nuclear translocation of JAK2 to pancreatic cancer cells



Supplementary Material

Supplemental Figure 1. Requirement of N-terminal signal peptide for appropriate membrane targeting of carboxyl-terminal portion of MUC16. HeLa cells transiently transfected with CMV9-F114HA (left panel, **B** and **D**) and CMV10-F114HA (right panel, **F** and **H**) constructs along with their respective controls (CMV9: left panel, **A** and **C**; CMV10: right panel, **E** and **G**) were analyzed for the surface and intracellular localization of both FLAG and HA-tagged portions of MUC16-Cter under non- permeabilizing (Non-Perm) and permeabilizing (Perm) conditions using immunofluorescence microscopy. Cells were analyzed for the surface an intracellular localization of both FLAG (green) and HA-tagged (red) portions of MUC16-Cter. DAPI was used to stain the nucleus (DNA). Scale bars, 5µm.



Supplemental Figure 2. MUC16-Cter does not up regulate *JAK2* at the mRNA level and inhibition of JAK2 in MiaPaCa-2 cells leads to up regulation of basal level of *LMO2* and *NANOG* independent of MUC16-Cter. (A) Quantitative real-time PCR analysis of *JAK2* expression from control and MUC16-Cter expressing cells. Data represent mean \pm s.e.m, n=3. (B and C) MiaPaCa-2 cells stably transfected with either control (CMV9) or MUC16-Cter (F114HA) were treated with either DMSO or TG101209 (3 μ M) for 3 h. Expression of *LMO2* and *NANOG* was analyzed using real-time PCR. (D) K562 cells were treated with either DMSO or TG101209 for 3 h. Expression of *LMO2* and *NANOG* was analyzed using real-time PCR.



Supplemental Figure 3. MUC16-Cter imparts metastatic phenotypes PC cells. (A) Representative photographs of mice receiving MUC16-Cter expressing MiaPaCa-2 cells showing extensive peritoneal metastasis (yellow arrows), which was not observed in the mice receiving control cells. (B) *Ex vivo* GFP fluorescence images of representative metastatic sites (peritoneum and intestine) demonstrate the enhanced metastatic capability of MUC16-Cter-transfected MiaPaCa-2 and T3M4 cells over the control cells. Scale bars, 10 mm. (C) Sections of primary tumors from mice injected with control and MUC16-Cter-expressing cells were stained with haematoxylin and eosin (H&E, top panel) or stained with anti-HA antibodies (α -HA, bottom panel). Scale bars, 500 µm. (D) Sections of metastatic tumor from mice injected with MUC16-Cter-expressing T3m4 cells were stained with haematoxylin and eosin (H&E, left) or stained with anti-HA antibody (α -HA, right). Scale bars, 500 µm.

Supplemental Table-1 Primers used to generate the constructs (Restriction Sites are underlined)

F114HA (in p3X-FLAG-CMV9 and p3X-FLAG-CMV10 vector)

Forward Primer: 5'-CTGAAGATCTAACTTCTCGCCACTGGCTCGGAGAG-3'

Reverse Primer: 5'-TACG<u>GGATCC</u>TCAAGCGTAATCTGGAACATCGTATGGGTAACCACCTTGCAGAT CCTCCAGGTCTAGGTG-3'

HA114Myc (in pSecTag2C vector)

Forward Primer: 5'-CTTA<u>AAGCTT</u>TATACCCATACGATGTTCCAGATTACGCTTTCATCAATGGCTATG CAC-3'

Reverse Primer: 5'-TACG<u>CTCGAG</u>TTGCAGATCCTCCAGGTCTAGGTGTGACTGGTAGTAGCCTGGG CAC-3'

Supplemental Table-2. Primers used in the study (for Real-Time PCR)

LMO2 Forward primer: 5'- CGGCGCCTCTACTACAAACT-3'

LMO2 Reverse primer: 5'-GAATCCGCTTGTCACAGGAT-3'

NANOG Forward primer: 5'-GAAACAGAAGACCAGAACTGTG-3'

NANOG Reverse primer: 5'- GGTCTTCACCTGTTTGTAGC-3'

JAK2 Forward primer: 5'-GTCCACCAGCGGAATTTATGCG-3'

JAK2 Reverse primer: 5'-CATATAGATCTCATCTGGGCATCC-3'

β-ACTIN Forward primer: 5'-TGGACATCCGCAAAGACCTG-3'

β-ACTIN Reverse primer: 5'-TGGACATCCGCAAAGACCTG-3'

Lmo2 Forward Primer: 5'-CAGCAGAACATAGGGGACCG-3'

Lmo2 Reverse Primer: 5'-TCCTGACCAAAAAGCCTGAGAT-3'

Nanog Forward Primer: 5'-TACCTCAGCCTCCAGCAGAT-3'

Nanog Reverse Primer: 5'-GGTGCTGAGCCCTTCTGAAT-3'