

Table S1. Strains and plasmids used or constructed in this work

Strain or Plasmid	Genotype/relevant characteristics	Reference or source
<i>P. aeruginosa</i>		
PAO1	Reference strain fully sequenced	Laboratory collection
PAO Δ <i>ampC</i>	PAO1 Δ <i>ampC::lox</i>	(1)
PAO Δ <i>dacB</i>	PAO1 Δ <i>dacB::lox</i>	(2)
PAO Δ <i>dacC</i>	PAO1 Δ <i>dacC::lox</i>	This study
PAO Δ <i>pbpG</i>	PAO1 Δ <i>pbpG::lox</i>	This study
PAO Δ <i>dacB</i> Δ <i>dacC</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>dacC::lox</i>	This study
PAO Δ <i>dacB</i> Δ <i>pbpG</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>pbpG::lox</i>	This study
PAO Δ <i>dacC</i> Δ <i>pbpG</i>	PAO1 Δ <i>dacC::lox</i> Δ <i>pbpG::lox</i>	This study
PAO Δ <i>dacB</i> Δ <i>dacC</i> Δ <i>pbpG</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>dacC::lox</i> Δ <i>pbpG::lox</i>	This study
PAO Δ <i>dacB</i> Δ <i>ampC</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>ampC::lox</i>	This study
PAO Δ <i>dacC</i> Δ <i>ampC</i>	PAO1 Δ <i>dacC::lox</i> Δ <i>ampC::lox</i>	This study
PAO Δ <i>pbpG</i> Δ <i>ampC</i>	PAO1 Δ <i>pbpG::lox</i> Δ <i>ampC::lox</i>	This study
PAO Δ <i>dacB</i> Δ <i>dacC</i> Δ <i>ampC</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>dacC::lox</i> Δ <i>ampC::lox</i>	This study
PAO Δ <i>dacB</i> Δ <i>pbpG</i> Δ <i>ampC</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>pbpG::lox</i> Δ <i>ampC::lox</i>	This study
PAO Δ <i>dacC</i> Δ <i>pbpG</i> Δ <i>ampC</i>	PAO1 Δ <i>dacC::lox</i> Δ <i>pbpG::lox</i> Δ <i>ampC::lox</i>	This study

PAO Δ <i>dacB</i> Δ <i>pbpG</i> Δ <i>ampC</i> Δ <i>dacC</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>pbpG::lox</i> Δ <i>ampC::lox</i> Δ <i>dacC::lox</i>	This study
PAO Δ <i>dacB</i> Δ <i>dacC</i> Δ <i>pbpG</i> Δ <i>ampC</i> <i>E. coli</i>	PAO1 Δ <i>dacB::lox</i> Δ <i>dacC::lox</i> Δ <i>pbpG::lox</i> Δ <i>ampC::lox</i>	This study
XL1-Blue	F'::Tn10 <i>proA</i> + <i>proB</i> _+ <i>lacI</i> ^q Δ (<i>lacZ</i>)M15/ <i>recA1 endA1 gyrA96</i> (Nal ^r) <i>thi hsdR17</i> (r _k + m _k +) <i>mcrB1</i>	Laboratory collection ^b
S17.1 λ pyr	<i>recA pro</i> (RP4-2Tet::Mu Kan::Tn7)	Laboratory collection ^b
Plasmids		
pEX100Tlink	Ap ^r <i>sacB</i> , pUC19-based gene replacement vector with an MCS	(3)
pUCGmlox	Apr Gm ^r , pUC18-based vector containing the lox-flanked <i>aacC1</i> gene	(3)
pCM157	Tc ^r , <i>cre</i> expression vector	(3)
pEXT Δ <i>ampC</i> ::Gm	pEX100Tlink containing 5' and 3' flanking sequence of <i>ampC</i> ::Gm <i>lox</i>	(1)
pEXT Δ <i>dacB</i> ::Gm	pEX100Tlink containing 5' and 3' flanking sequence of <i>dacB</i> ::Gm <i>lox</i>	(2)
pEXT Δ <i>dacC</i> ::Gm	pEX100Tlink containing 5' and 3' flanking sequence of <i>dacC</i> ::Gm <i>lox</i>	This study
pEXT Δ <i>pbpG</i> ::Gm	pEX100Tlink containing 5' and 3' flanking sequence of <i>pbpG</i> ::Gm <i>lox</i>	This study

Table S2: Primers used for PCR amplification of both upstream (PCR1) and downstream (PCR2) regions to PAO1 genes: *pbpG*, *dacC*, *dacB* and *ampC*.

Target gene	Primer's Name	Primer's Sequence, 5'...3' ^a	PCR Product - bp	Reference
<i>pbpG</i>	pbpG-F1-ERI	-TCGAATTC <u>CACTTT</u> CAAAGCCCTACGTGC-		This study
	pbpG-RI- HDIII	-TCAAGCTTGCAGTTTCGAGTCGAGCACG-	PCR1= 375	
	pbpG-F2-HDIII	-TCAAGCTTCACTGCGATCCCGGCCGC-		
	pbpG-R2-BHI	-TCGGATCCCAGTACCGGACCCAGGAGC-	PCR2= 318	
<i>dacC</i>	dacCF-ERI	-TCGAATTCACCTTGGCCAGCCGACGC-		This study
	dacCIR-HDIII	-TCAAGCTTCTCGGCCAGGGCGACGC-	PCR1= 500	
	dacCIF-HDIII	-TCAAGCTTGAAGTGAAAGCCGGCCTCG-		
	dacCR-BHI	-TCGGATCCACGCTCGCAGGGGAATTTCG-	PCR2= 400	
<i>dacB</i>	dacB-F1-ERI	-TCGAATTC <u>CGACCA</u> TTCGGCGATATGAC-		(2)
	dacB-R1-HDIII	-TCAAGCTTGTGCGCATCAGCAGCCAG-	PCR1= 571	
	dacB-F2-HDIII	-TCAAGCTT <u>GCCAGGGC</u> AGCGTACCGC-		
	dacB-R2-BHI	-TCGGATCCC <u>CGTAAT</u> CCGAAGATCCATC-	PCR2= 693	
<i>ampC</i>	AmpC-F-ERI	-TCGAATTC <u>CGCGCGC</u> AGGGCGTTCAG-		(1)
	AmpC-I-R-HDIII	-TCAAGCTT <u>CGTCCT</u> CTTACGAGGCCAGC-	PCR1= 415	
	AmpC-I-F-HDIII	-TCAAGCTT <u>CAGGGC</u> AGCCGCTTCGAC-		
	AmpC-R-BHI	-TCGGATCCCAGGTTGGCATCGACGAAG-	PCR2= 448	

^aIn the primer's name; ERI, HDIII and BHI refer to the presence of the restriction sites (underlined sequence), EcoRI, HindIII and BamHI while F and R refer to the direction; forward and reverse respectively.

1. **Moya B, Juan C, Alberti S, Perez JL, Oliver A.** 2008. Benefit of having multiple *ampD* genes for acquiring beta-lactam resistance without losing fitness and virulence in *Pseudomonas aeruginosa*. *Antimicrobial agents and chemotherapy* **52**:3694-3700.
2. **Moya B, Dotsch A, Juan C, Blazquez J, Zamorano L, Haussler S, Oliver A.** 2009. Beta-lactam resistance response triggered by inactivation of a nonessential penicillin-binding protein. *PLoS pathogens* **5**:e1000353.
3. **Quenee L, Lamotte D, Polack B.** 2005. Combined *sacB*-based negative selection and *cre-lox* antibiotic marker recycling for efficient gene deletion in *pseudomonas aeruginosa*. *BioTechniques* **38**:63-67.

Table S3. Relative abundance and retention time of muropeptides in peptidoglycan of the mutants and PAO1 wild type.

Strain	Muropeptides (mol%) ^a								
	M3 RT= 5.2	M4G RT= 6.4	M4 RT= 9.5	M5G RT= 10.0	M5 RT= 13.0	D43 RT= 23.0	D45G RT= 24.6	D44 RT= 25.2	D45 RT= 27.0
PAO1	10.5	1.1	24.9	0.3	0.3	6.2	ND	26.3	0.4
PAO Δ <i>dacB</i>	8.5(81)	0.9(82)	22.9(94)	0.4(133)	0.5(167)	5.3(85)	ND	28.9(110)	0.6(150)
PAO Δ <i>dacC</i>	10(95)	0.8(73)	25.4(102)	0.5(167)	1.4(467)	5.9(95)	ND	24.6(94)	2.8(700)
PAO Δ <i>pbpG</i>	8.9(85)	0.9(82)	25.2(101)	ND	0.6(200)	5.1(82)	ND	29(110)	0.2(50)
PAO Δ <i>dacB</i> Δ <i>dacC</i>	7.4(71)	0.7(64)	17.9(72)	1.4(467)	7.7(2567)	4.3(69)	2.5	17.3(66)	10.1(2525)
PAO Δ <i>dacB</i> Δ <i>pbpG</i>	6.7(64)	0.7(64)	20.3(82)	0.5(167)	0.6(200)	4.9(79)	ND	31.2(119)	ND
PAO Δ <i>dacC</i> Δ <i>pbpG</i>	8.7(83)	0.8(73)	23(92)	0.6(200)	1.7(567)	5.2(84)	1	25.8(98)	4.3(1075)
PAO Δ <i>dacB</i> Δ <i>dacC</i> Δ <i>pbpG</i>	3.4(32)	0.5(46)	6.4(26)	3.6(1200)	18.6(6200)	1.8(29)	4.2	8.5(32)	24.4(6100)
Strain	Muropeptides (mol%) ^a								
	T444 RT= 33.0	T445 RT= 34.0	D44N RT= 37.5-38	D45N RT= 39.3	T444N RT= 42.5	T445N RT= 43.2			
PAO1	2.3	ND	5	0.1	1.7	0.3			
PAO Δ <i>dacB</i>	3.1(134)	ND	4.7(94)	0.1(100)	2.2(130)	0.3(100)			
PAO Δ <i>dacC</i>	2.1(91)	0.3	4.8(96)	0.5(500)	1.7(100)	0.5(167)			
PAO Δ <i>pbpG</i>	3(130)	ND	4.5(90)	0.1(100)	1.7(100)	0.4(121)			
PAO Δ <i>dacB</i> Δ <i>dacC</i>	1.8(77)	1	3.3(66)	1.6(1600)	1.4(82)	1(333)			
PAO Δ <i>dacB</i> Δ <i>pbpG</i>	4.3(187)	ND	4.5(90)	0.1(100)	2.3(135)	0.5(167)			
PAO Δ <i>dacC</i> Δ <i>pbpG</i>	2.8(122)	0.5	4.4(88)	0.6(600)	1.7(100)	0.6(200)			
PAO Δ <i>dacB</i> Δ <i>dacC</i> Δ <i>pbpG</i>	1.2(52)	3.3	1.1(22)	3.2(3200)	0.5(29)	2(667)			

^a Relative abundance in mol % of individual muropeptides. RT: retention time; ND: Not detected. Values in brackets represent percentage of the values obtained for each mutant related to wild-type PAO1.