Figure S1

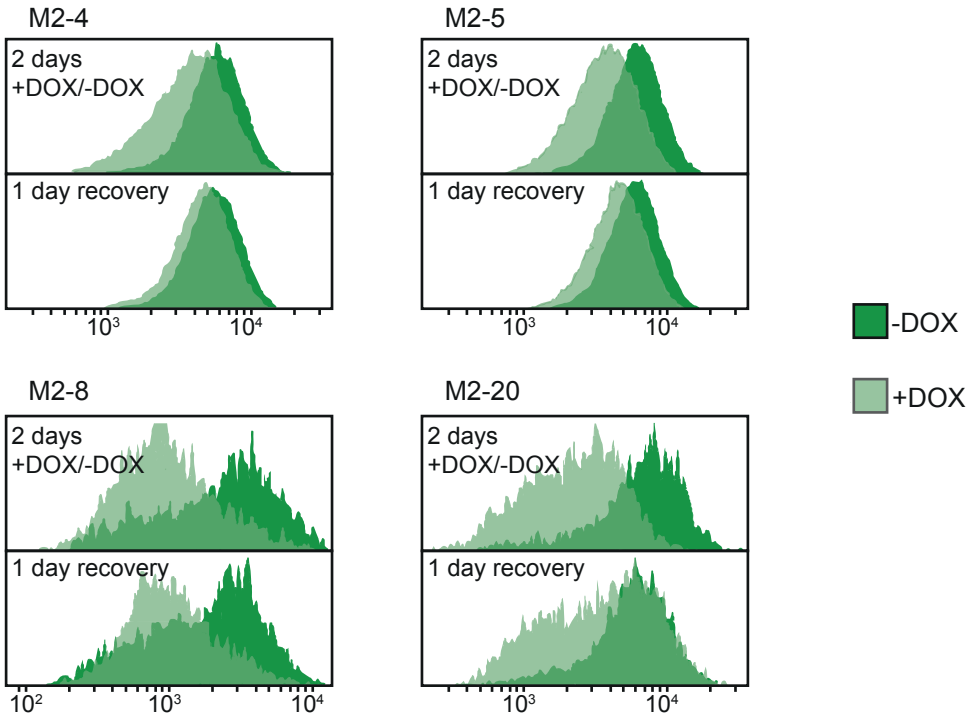

FIG S1 Representative histograms of EGFP fluorescence intensity (FI) distribution in clones M2-4, M2-5, M2-8 and M2-20 as determined by FACS analysis. Dark green represents uninduced cells, and light green the cells treated with doxycycline. Upper panels shows repression after two days of doxycycline treatment as compared to untreated control, and lower panels shows the respective situation one day later, after doxycycline has been washed out.

Figure S2

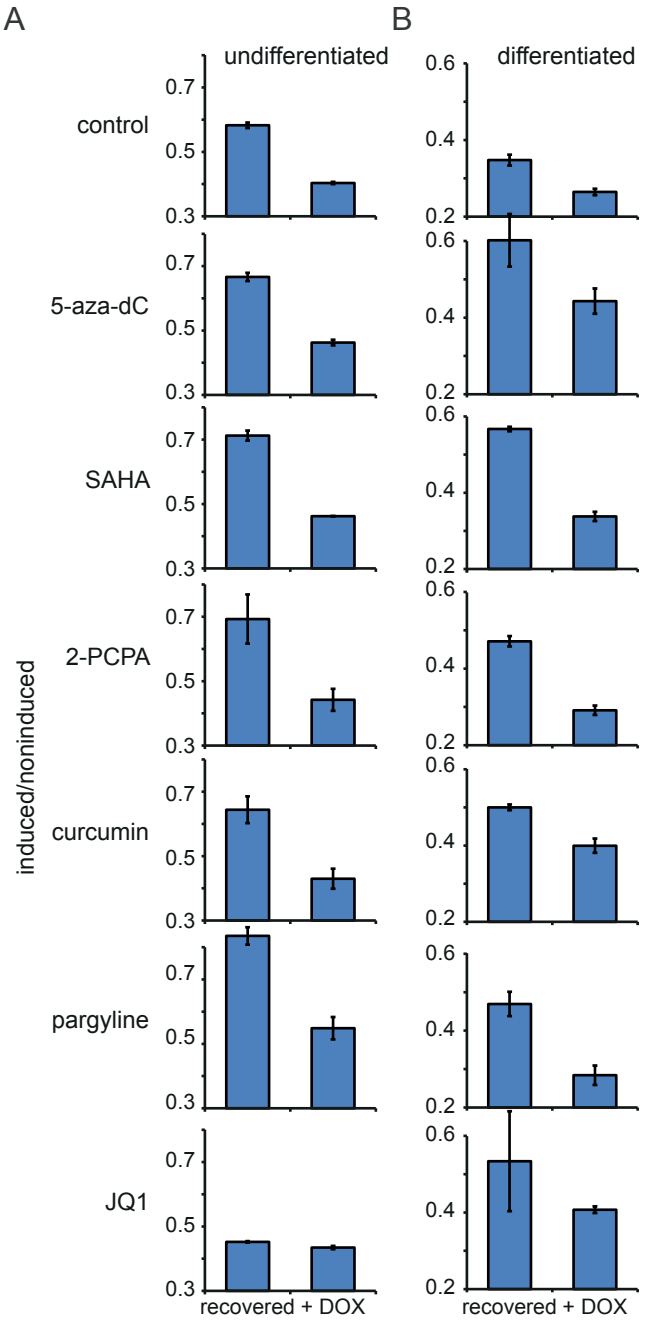

FIG S2 Interference of small molecule inhibitors with chromatin modifiers. (A) Mean EGFP FI of induced/noninduced M2-3 cells after two days of doxycycline treatment (+DOX) and after two days of doxycycline treatment followed by one day of recovery (recovered). Small molecule inhibitors used are indicated. Error bars represent SD of three independent experiments. (B) Same as (A) but in differentiating M2-3 cells. Time schedule as in Figure 5A but all experimental steps taken one day earlier and cells analyzed at day 2 of differentiation. Error bars represent SD of three independent experiments.

Table S1

name	abbreviation	targets	inhibition
5-aza-2'-deoxycytidine	5-aza-dC	DNMTs	DNA methylation
suberanilohydroxamic acid	SAHA	HDACs 1-9	histone deacetylation
2,4-pyridinedicarboxylic acid	2,4-PDCA	JARID1/JMJD2 family	H3K4me3/H3K36me3 demethylation
tranylcypromine	2-PCPA	LSD1	H3K4me1/2 demethylation
curcumin	CUR	HATs	histone acetylation
pargyline	PAR	LSD1	H3K9me1/2 demethylation
thieno-triazolo-1,4-diazepine	JQ1	BET family/BRD4	BRD4/transcriptional elongation

TABLE S1 Small molecule inhibitors and their effects on histone modifying enzymes. First column gives name, second abbreviation, third enzymes targeted and fourth biological process inhibited.

#	SEQUENCE	DESCRIPTION
228	ACACGCAGCTCATTGTAG	First strand primer for strand-specific *Actb* expression
226	GATATCGCTGCGCTGGTCGT	FOR primer *Actb* expression
227	AGATCTTCTCCATGTCGTCC	REV primer *Actb* expression
217	CTTCTCGTTGGGGTCTTTGC	First strand primer for strand-specific EGFP expression
106	AGGGCATCGACTTCAAGGAG	FOR primer EGFP expression
107	CACCTTGATGCCGTTCTTCTG	REV primer EGFP expression
104	CAAGATCCGCCACAACATCG	First strand primer for strand-specific ptet proximal expr.
222	TTTCACTGCATTCTAGTTGTGGT	FOR primer ptet proximal expression
223	GGTACCCGGGGATCCTCTA	REV primer ptet proximal expression
220	TGGTAATCGTGCGAGAGGG	First strand primer for strand-specific ptet distal expr.
231	TCCCCTTCTCCCTCTCCAG	FOR primer ptet distal expression
232	CTGCAGAATTCTAGAGCCGC	REV primer ptet distal expression
195	CTCTGACTGACCGCGTTACT	FOR primer ChIP for CAG promoter
196	TTTCACGCAGCCACAGAAAA	REV primer ChIP for CAG promoter
323	CACATTTGTAGAGGTTTTACTTGCT	FOR primer ChIP for GFP-ptet
324	AGCTGCAATAAACAAGTTAACAACA	REV primer ChIP for GFP-ptet
317	GAGCTCGAATTCTCCAGGCG	FOR primer ChIP for ptet promoter
318	GTATGTCGAGGTAGGCGTG	REV primer ChIP for ptet promoter
343	TCTCCCAGCATCCTCTACACA	FOR primer ChIP for H3K36me3 control in *Mcm2*
344	CTATGGTATGTGTGGTGGGCA	REV primer ChIP for H3K36me3 control in *Mcm2*
331	GCTAGGTTAGGAGAGCCCAGA	FOR primer ChIP for H3K4me3 control in *Mrps14*
332	AGGTCTCAATCATCCGACTCTC	REV primer ChIP for H3K4me3 control in *Mrps14*
204	GGATTTTTTTTGTTTTAAATTTGTG	FOR primer for bisulfite sequencing amplicon
207	AAATAAACTTCAAAATCAACTTACC	REV primer for bisulfite sequencing amplicon
209	GTAAAACGACGGCCAG	FOR primer for amplification from bacteria (M13 -20 FOR)
208	CAGGAAACAGCTATGAC	REV primer for amplification from bacteria (M13 REV)
302	ATTTAGGTGACACTATAG	Sequencing primer for bisulfite sequencing (Sp6)

TABLE S2 Primers used in this study.