Amniotic fluid stem cells provide considerable advantages in epidermal regeneration: B7H4 creates a moderate inflammation microenvironment to promote wound repair

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Supplementary Figure S1. Excisional wound model construction and GFP-positive hAFS cell transplantation

(a) A GFP reporter gene was delivered into hAFS cells and fibroblasts using a lentiviral vector. Immunofluorescence and flow cytometry show that almost 100% of hAFS cells and fibroblasts are GFP positive. (b) Original photographs of (Figure 4a, representative) the wounds with GFP-positive hAFS cells, fibroblasts or vehicle (sham) at days 7, 14 and 21 are shown. Over time, hAFS cell-treated wounds exhibited accelerated wound closure in BALB/c mice compared with mice treated with the fibroblast or sham groups. At 21 days, wounds in hAFS cell-treated mice achieved almost complete wound closure,

whereas no completely closed wounds were observed in the fibroblast- or sham-treated groups. The wounds are indicated by white circles.



Supplementary Figure S2. The role of the B7H4 molecule in the proliferation of T lymphocytes

(a) Analysis of the inhibitory effect of hAFS cells on human lymphocytes. The corresponding cell images of Fig. 6a. 1: human T lymphocyte; 2: human T lymphocyte + CD3; 3: human T lymphocyte + CD3 + hAFS cells; 4: human T lymphocyte + CD3 + hAFS cells+ B7H1 blocking antibody; 5: human T lymphocyte + CD3 + hAFS cells + B7H4 blocking antibody; 6: human T lymphocyte + CD3 + hAFS cells + B7H1 blocking

antibody + B7H4 blocking antibody; 7: human T lymphocyte + CD3 + hAFS cells + mIgG. (b) Analysis of the inhibitory effect of hAFS cells on mouse lymphocytes. The corresponding cell images of Fig. 6b. 1: mouse T lymphocyte; 2: mouse T lymphocyte + CD3; 3: mouse T lymphocyte + CD3 + hAFS cells; 4: mouse T lymphocyte + CD3 + hAFS cells + B7H1 blocking antibody; 5: mouse T lymphocyte + CD3 + hAFS cells + B7H4 blocking antibody; 6: mouse T lymphocyte + CD3 + hAFS cells + B7H1 blocking antibody; 7: mouse T lymphocyte + CD3 + hAFS cells + B7H1 blocking antibody; 7: mouse T lymphocyte + CD3 + hAFS cells + B7H1 blocking antibody; 7: mouse T lymphocyte + CD3 + hAFS cells + B7H1 blocking antibody; 7: mouse T lymphocyte + CD3 + hAFS cells + B7H1 blocking antibody; 7: mouse T lymphocyte + CD3 + hAFS cells + mIgG.



b







Page 6

Supplementary Figure S3. B7H4 downregulated hAFS cells nullify the advantages of hAFS on accelerated healing and moderate inflammatory reactions during wound repair

(a) Real-time PCR analysis of mRNA expression of repair-related factors (bFGF, VEGF, TGF- β 1, KGF) at days 7, 14, 21 in hAFS cell-, fibroblast- and B7H4-downregulated hAFS treatment groups. (b) Real-time PCR analysis of mRNA expression of inflammatory factors (TNF- α , Cox2, Mac3, IL-6, IL-1 β) at days 1, 4, 7, 14 and 21 in hAFS cell-treated, fibroblast and B7H4-downregulated hAFS groups. The data are presented as the mean±SD of three independent experiments: analysis was performed with GraphPad Prism; *: p<0.05.



Page 8

Supplementary Figure S4. Full-length images of RT-PCR in Figure 1a. RT-PCR assay for determining relative mRNA expression levels of several genes in keratinocytes, hAFS cells and hESCs. GAPDH was used as an internal loading control (1: keratinocyte; 2-6: three lines of hAFS cells with different passage numbers; 7: hESC. M: marker).



Supplementary Figure S5 Full-length images of RT-PCR in Figure 2b. RT-PCR assay for determining the relative mRNA expression level of several genes in hAFS cells, keratinocytes derived from hAFS cells (hAFS-K) and keratinocytes. GAPDH was used as an internal loading control (1: hAFS cells; 2-4: three lines of hAFS-K; 5: keratinocyte; M: marker).