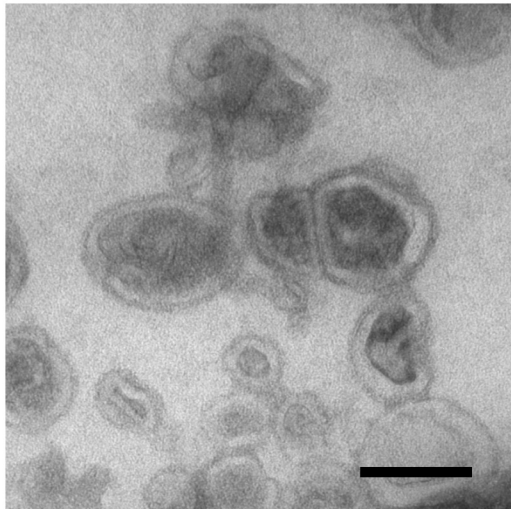


Supplementary Figure 1. Western blots of Fig. 1c, d. (a, b) Western blot analysis of pelleted proteins from fractions collected following sucrose gradient centrifugation of PalmGFP- (a) and Palmtomato-labeled EVs (b). Cells lysate of 293-PalmGFP or -Palmtomato were used as a positive control. Immunoblotting with anti-tdTomato antibody revealed a band at 25 kDa predominantly in the cell lysate but not in the EV-containing fractions, indicating it may be cleaved monomers of tdTomato.

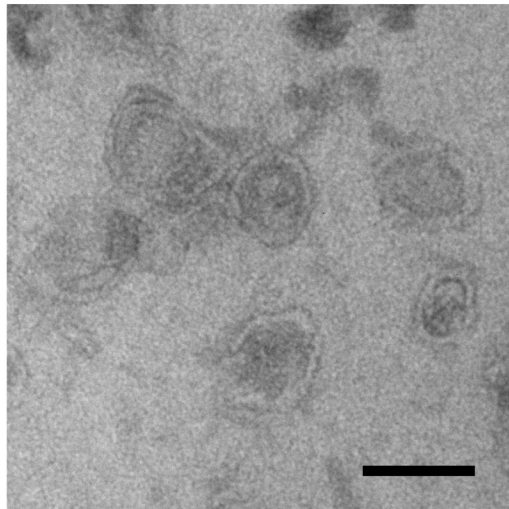
anti-rabbit
protein A gold (5 nm)

PalmGFP-EV



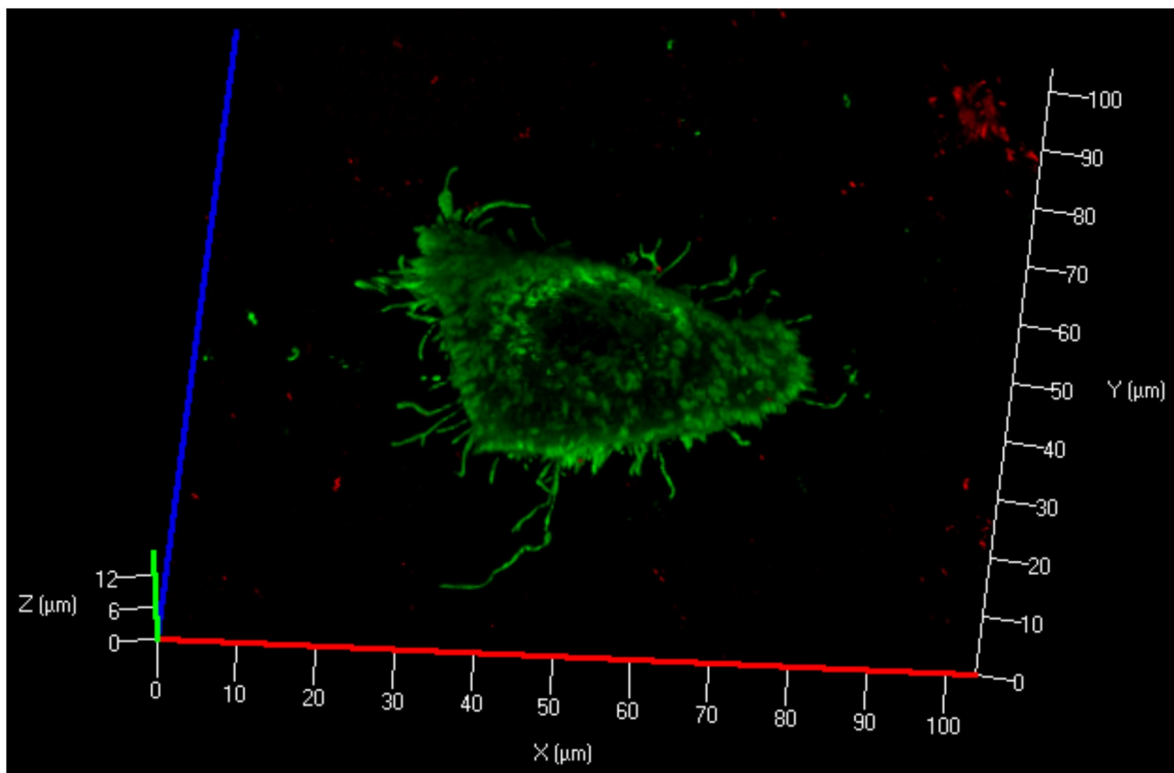
anti-mouse
protein A gold (10 nm)

PalmtdTomato-EV

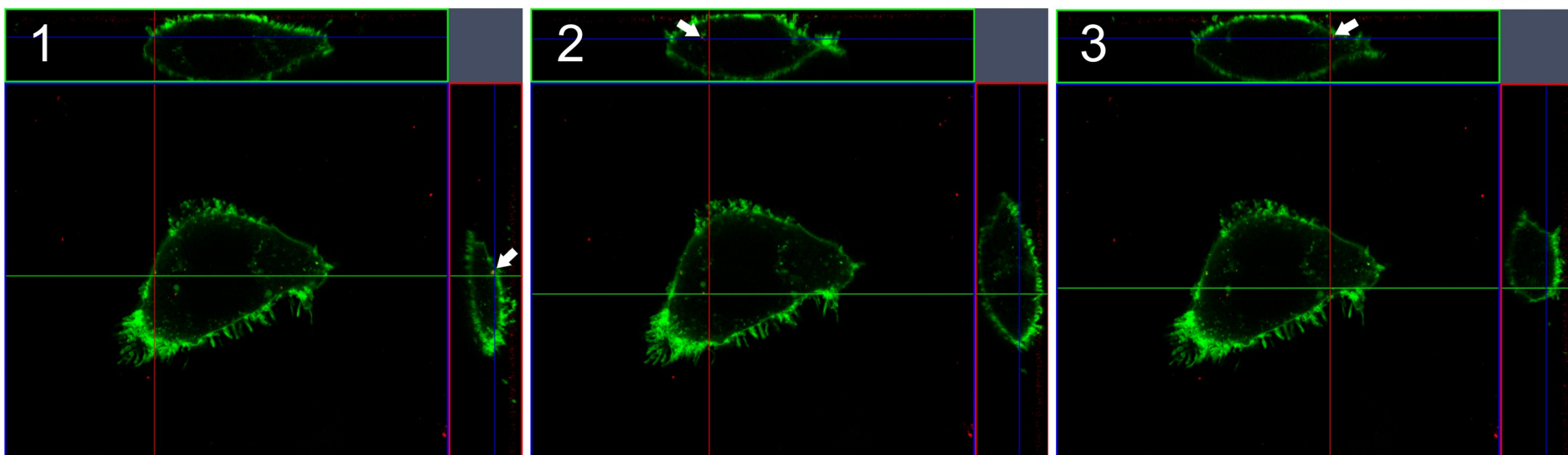
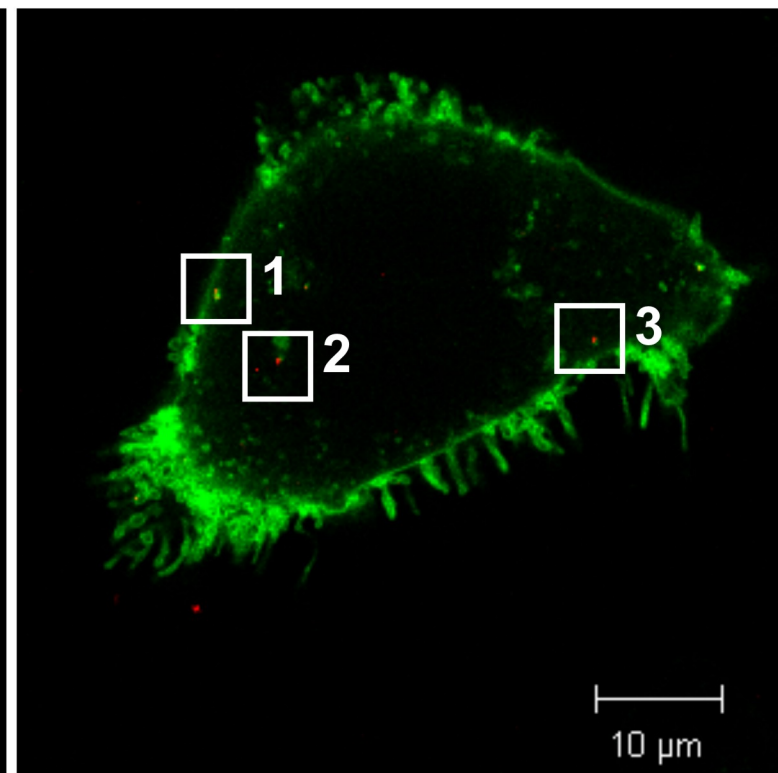


Supplementary Figure 2. Secondary antibody control immunolabeling of PalmGFP- and PalmtdTomato-EVs. PalmGFP- and PalmtdTomato-EVs were immunolabeled with anti-mouse protein A gold (5 nm) and anti-rabbit protein A gold (10 nm) antibodies, respectively. TEM shows no non-specific binding of the secondary antibodies to PalmGFP (left) and PalmtdTomato-EVs (right). Bar, 100 nm.

a

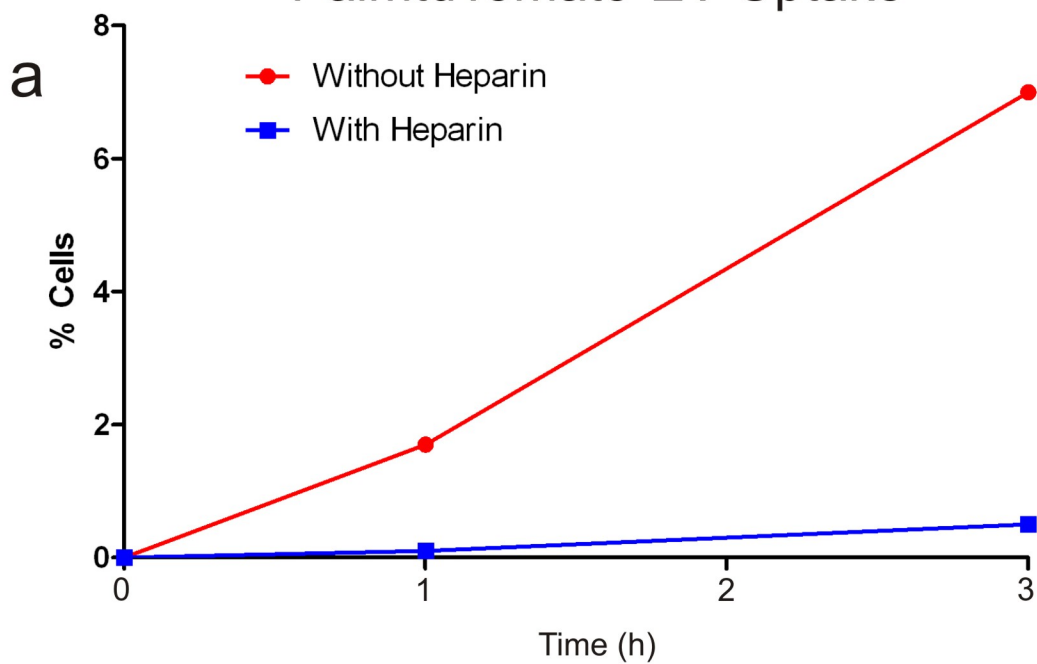


b

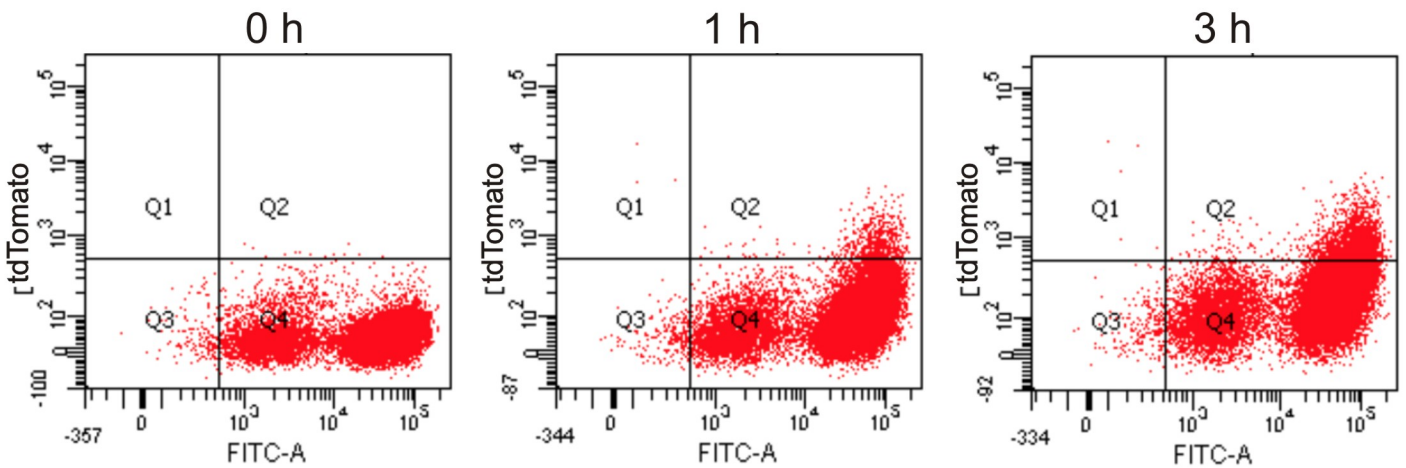


Supplementary Figure 3. EV docking and internalization by recipient cells. (a) 3D reconstruction of Z-stack images showing 293T-PalmGFP cells exposed to 293T-PalmtmTomato⁺ EVs for 1.75 h. (b) Single Z-plane confocal image and orthogonal projections from Z-stack images (subpanel 1-3) demonstrating EV docking on the plasma membrane (arrow; subpanel 1 and 3) and uptake by the recipient cell (arrow, subpanel 3). Bar, 10 μ m.

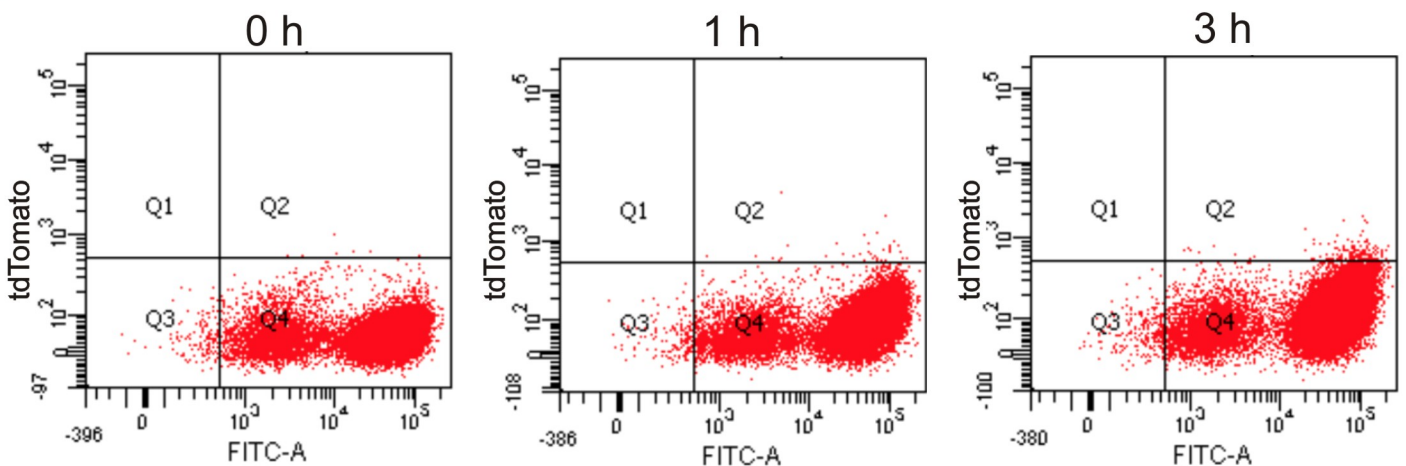
PalmtTomato-EV Uptake



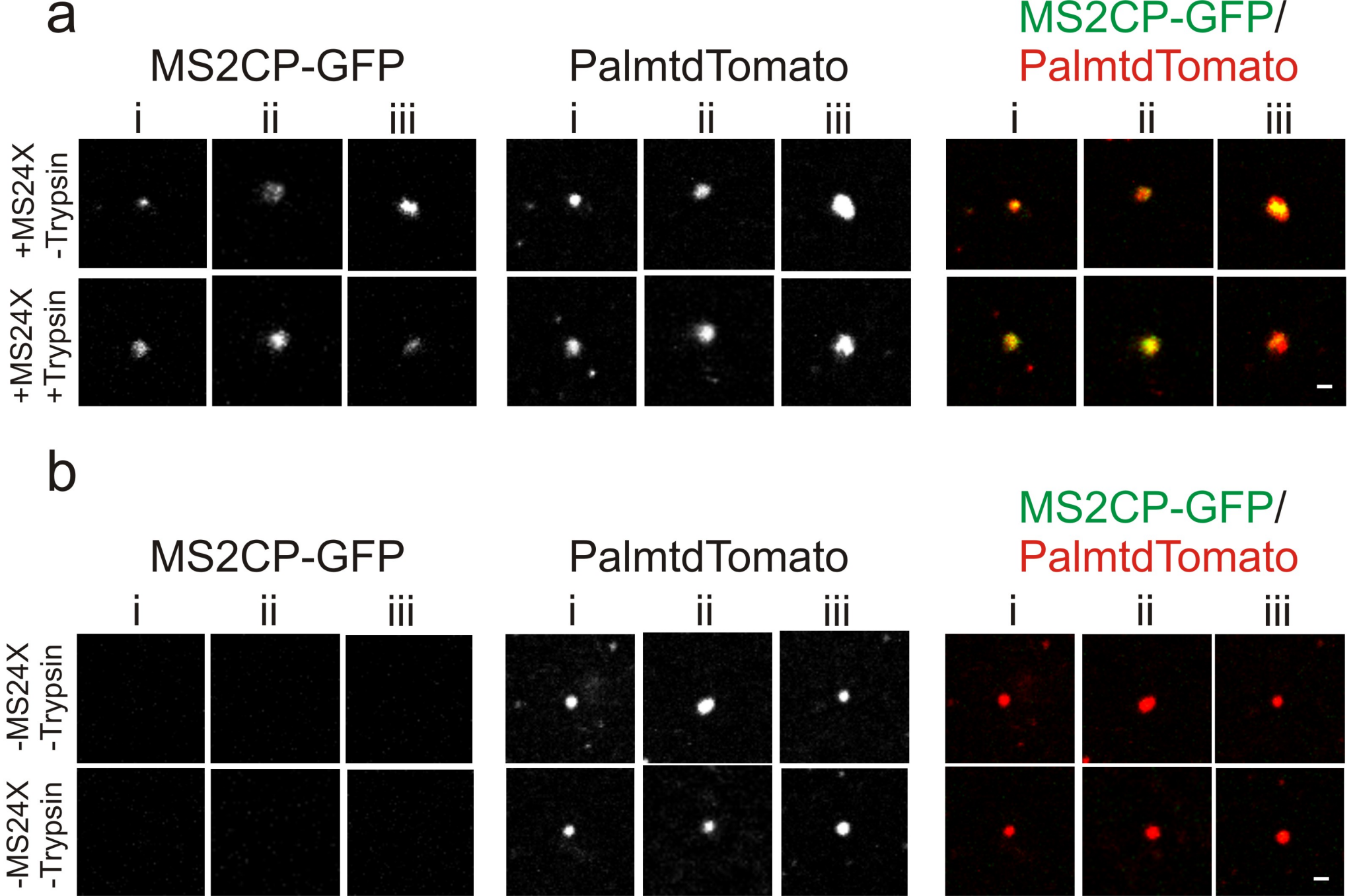
b



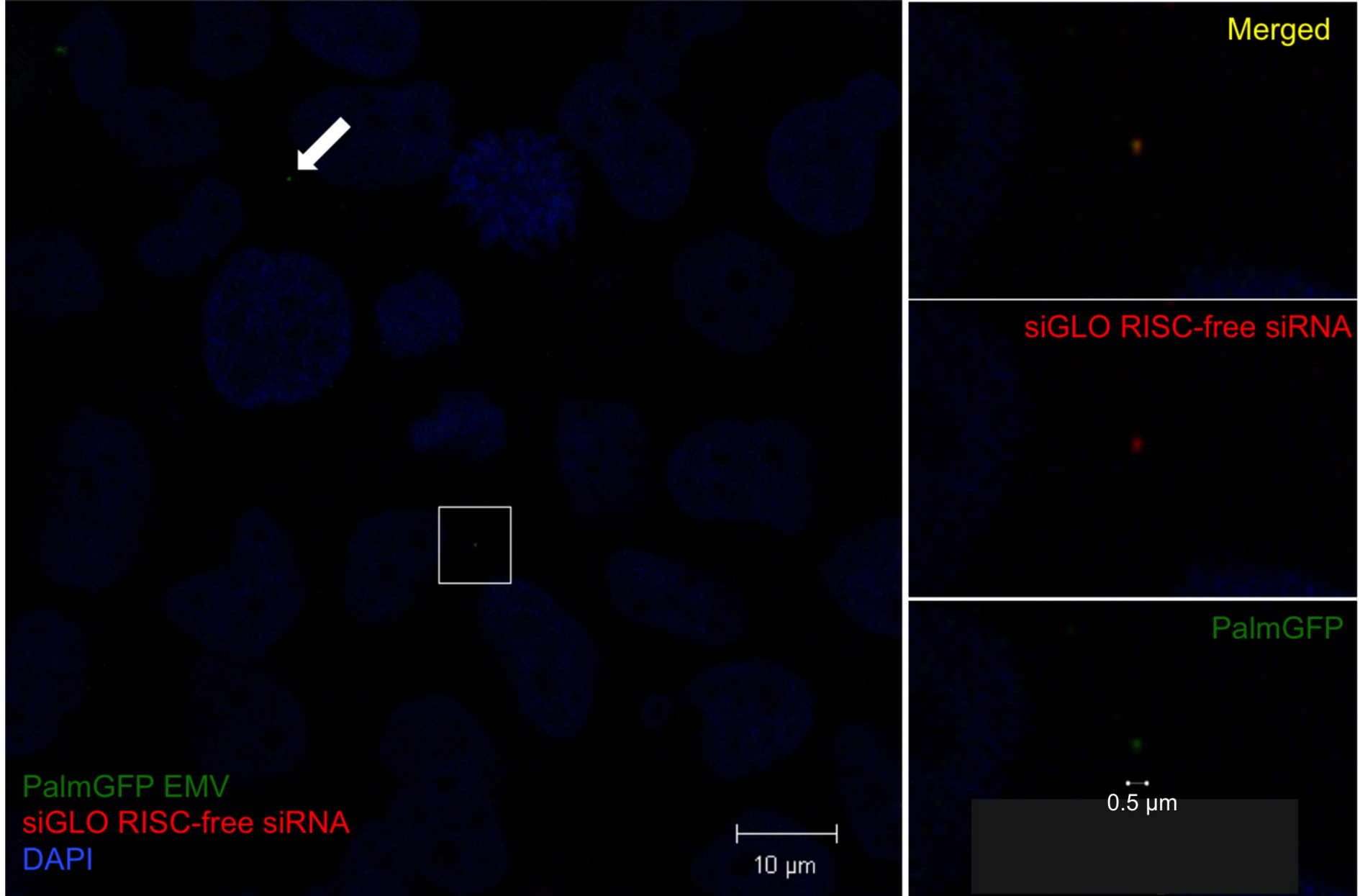
c



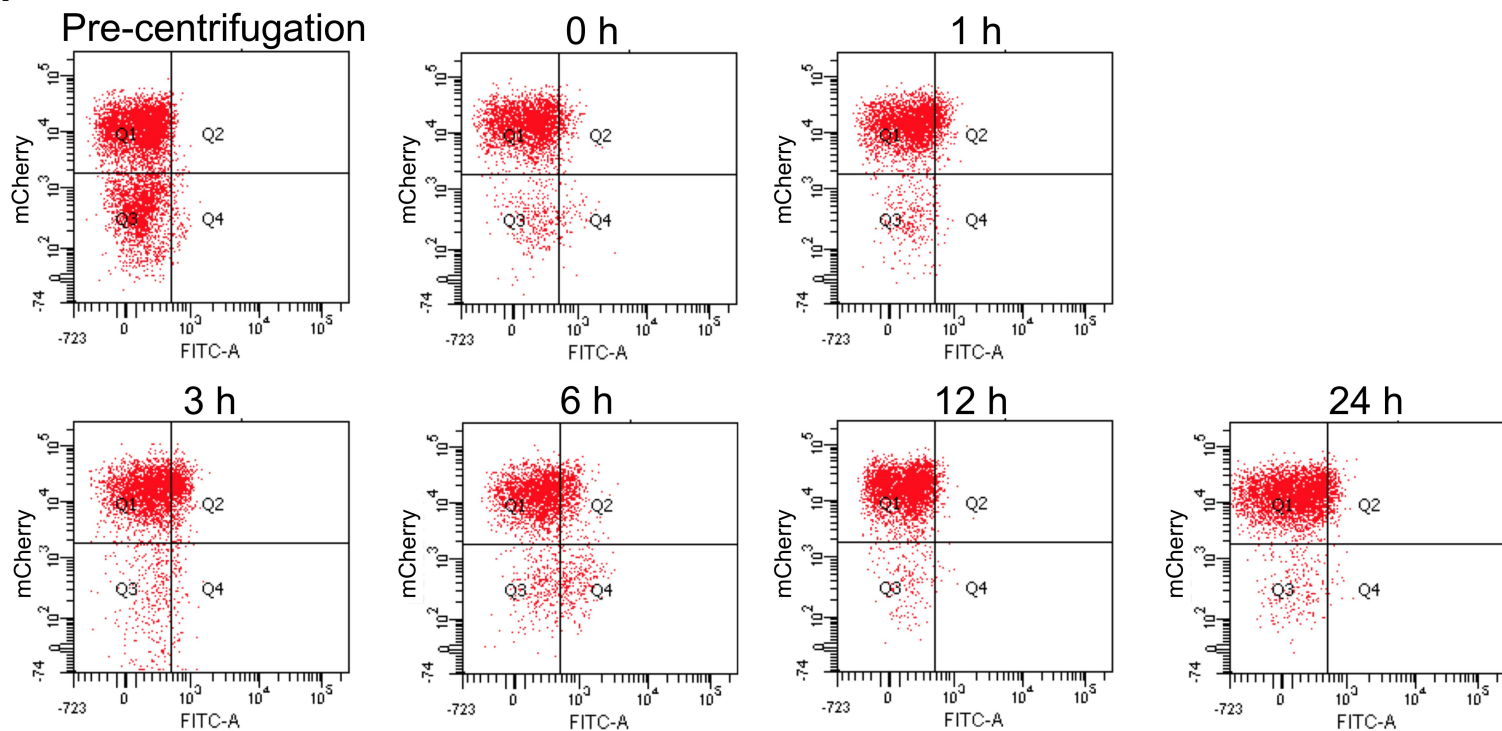
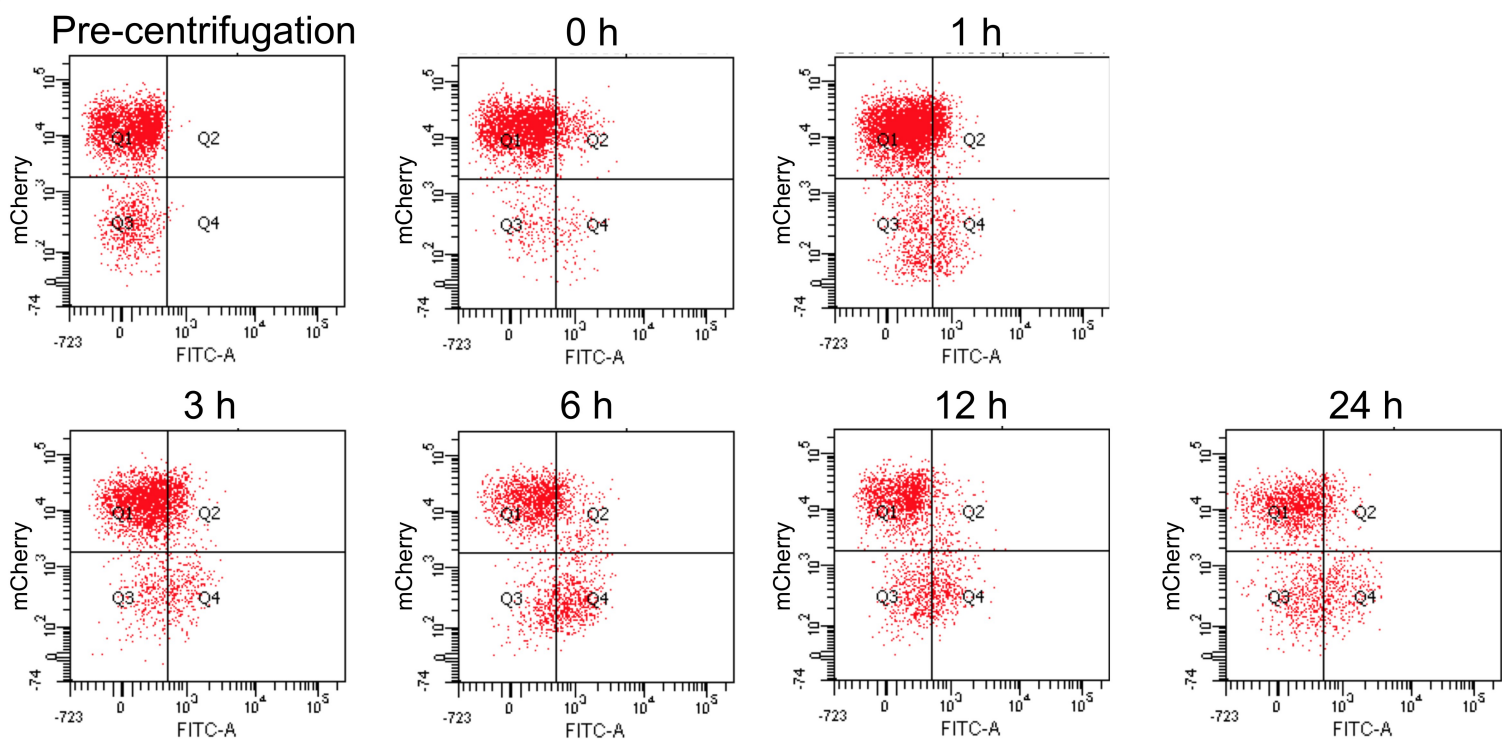
Supplementary Figure 4. Heparin-mediated inhibition of 293T-PalmtTomato⁺ EV uptake by Gli36 cells. (a) Gli36-PalmGFP cells were treated with 293T-PalmtTomato⁺ EVs and centrifuged at 4 °C for 1.5 h followed by a wash with PBS and replaced with EV-depleted media in the presence or absence of heparin (200 µg/mL). Cells were then collected at 0, 1 and 3 h to assess EV uptake by FACS analysis. (b, c) FACS analyses of Gli36-PalmGFP cells in the absence (b) or presence (c) of heparin.



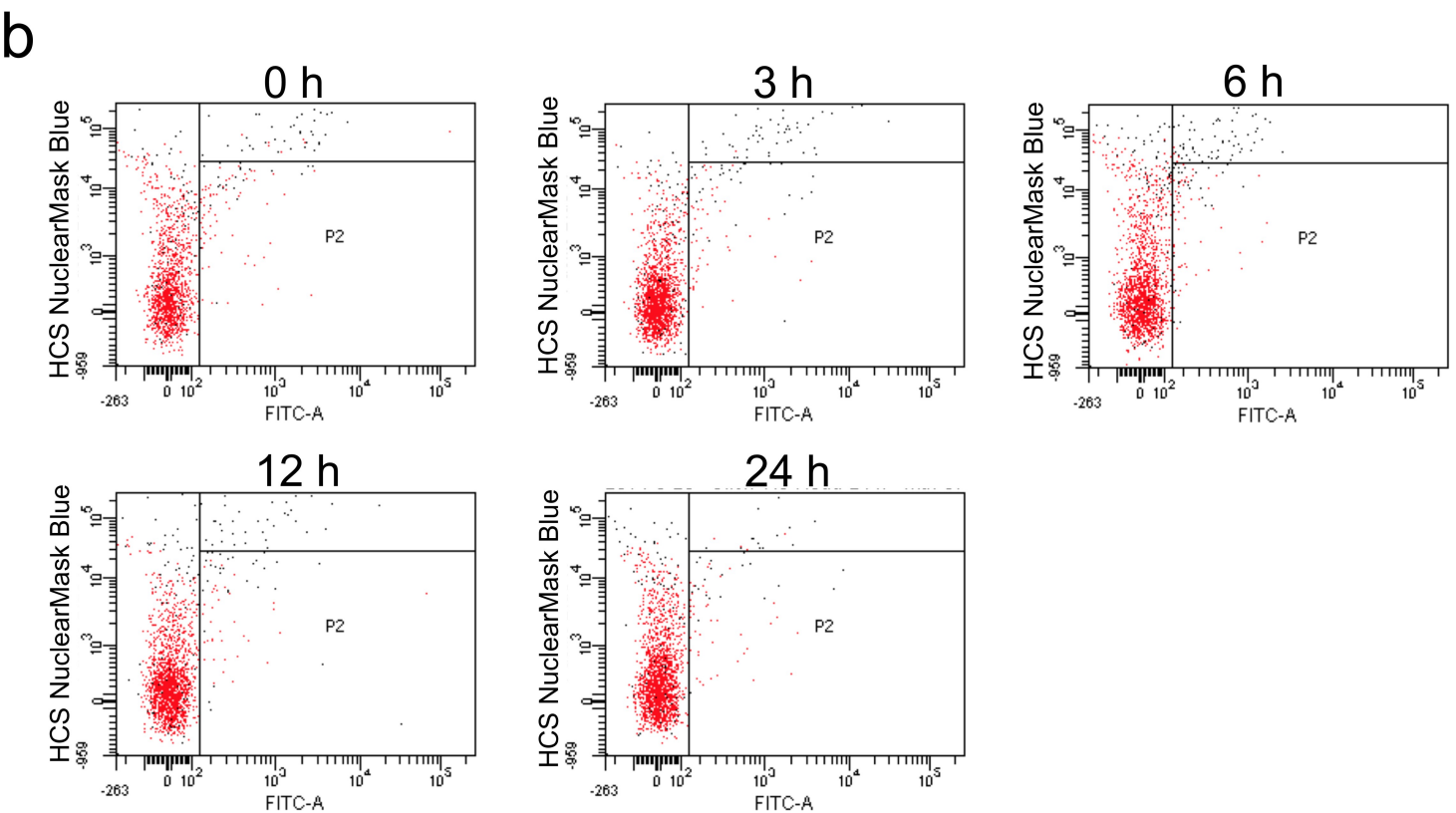
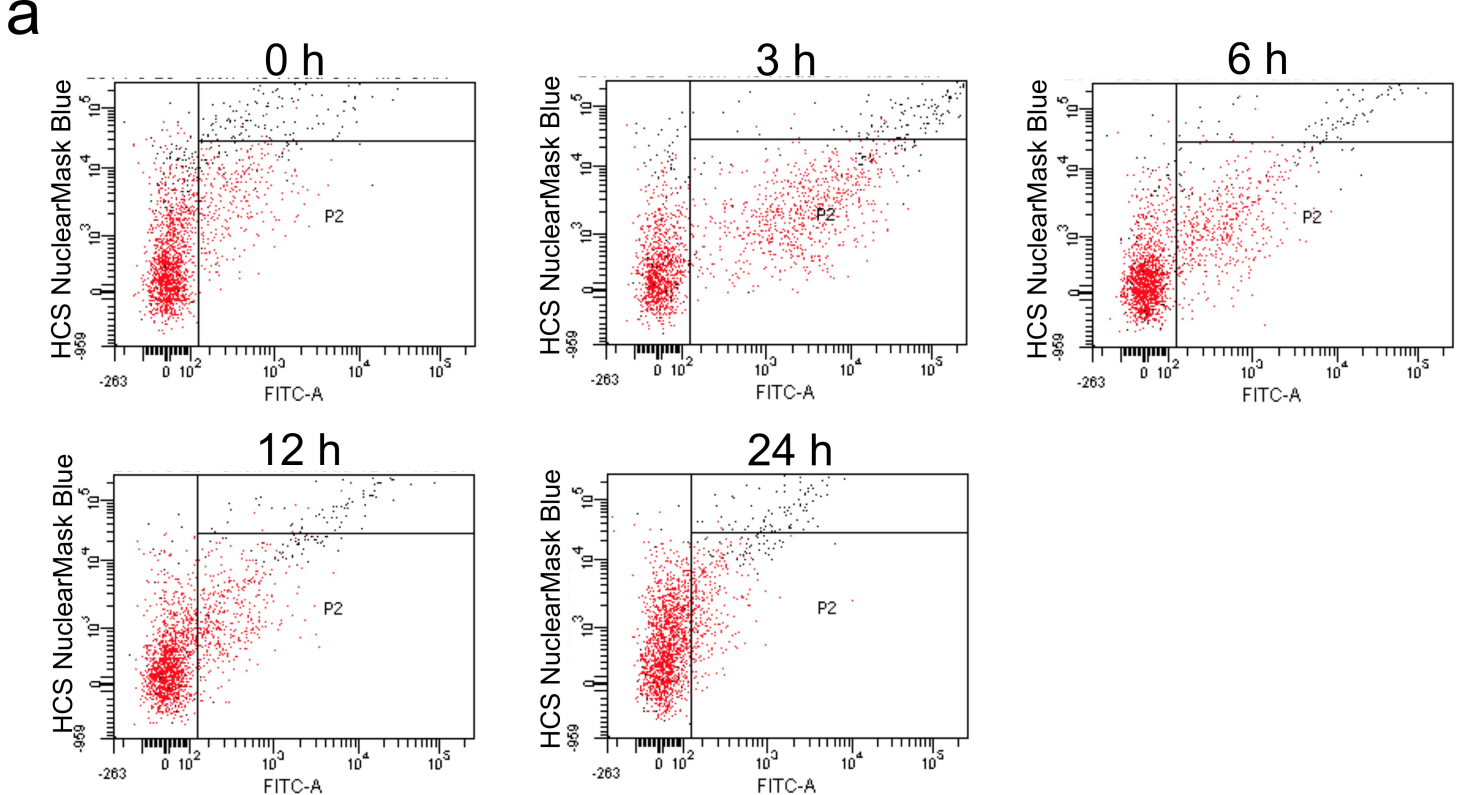
Supplementary Figure 5. Trypsin does not remove MS2CP-GFP from EVs. (a, b) EVs (subpanel i-iii) isolated from Gli36 cells co-expressing MS2CP-GFP and Palmtomato-MS24X (a) or Palmtomato (b) were imaged in the presence or absence of trypsin. Bar, 1 μ m. MS2CP-GFP signals were detected with Palmtomato⁺ EV in the presence and absence of trypsin when the MS24X tag is present. No appreciable MS2CP-GFP signal was detected with Palmtomato⁺ when the MS24X tag is absent in the EV donor cells.



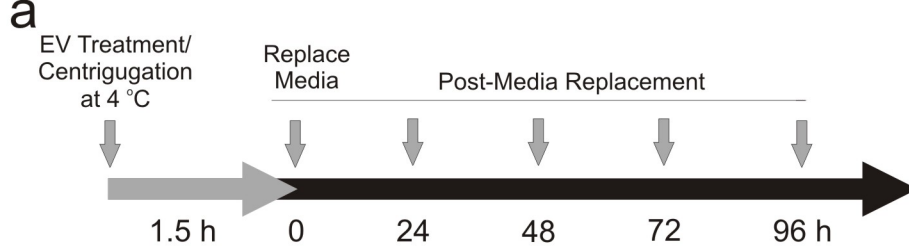
Supplementary Figure 6. Visualizing and tracking EV-delivered siRNAs. Gli36 glioma cells were seeded in the bottom dish of a transwell setup to serve as EV recipient cells. As EV donors, 293T-PalmGFP cells were transfected with siGLO RISC-free siRNA conjugated with DY-547, seeded into the upper transwell chamber (connected to the lower chamber through 1 µm pores) and allowed to incubate for 24 h. Gli36 cells were then fixed, stained with DAPI to reveal nuclei, and imaged with confocal microscopy. *Square box*, co-localization between PalmGFP⁺ EVs and siGLO RISC-free siRNA in recipient cells. *Arrow*, imaging negative control for DY-547 illustrating only PalmGFP⁺ EV signals. *Right panels*, magnified images showing PalmGFP⁺ EV associated siRNAs.

a**b**

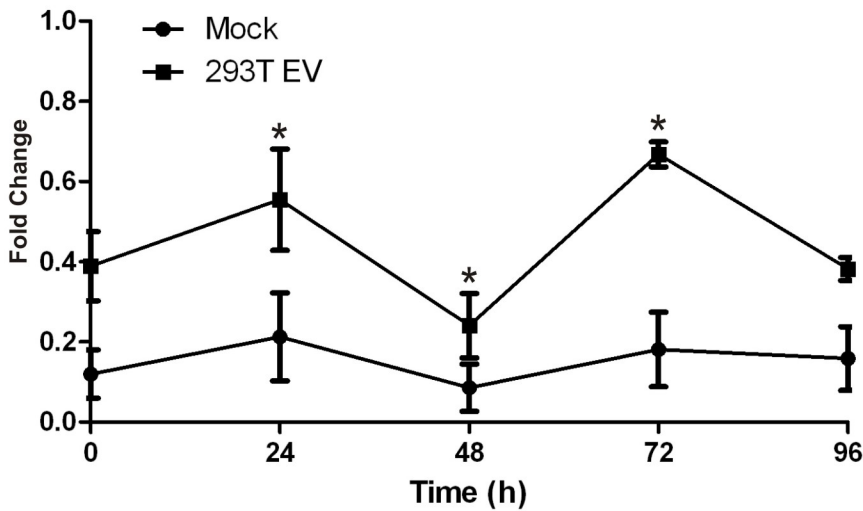
Supplementary Figure 7. FACS Analysis of PalmGFP⁺ EV uptake by Gli36-mCherry GBM cells. Gli36 cells stably transduced with mCherry were treated with PalmGFP+GlucB⁺ EVs (as described in legend to Fig. 6) in the presence (a) or absence (b) of CHX (20 μ g/ml). Cell samples were analyzed by FACS at different time points from pre-centrifugation (4°C) to 24 h post-centrifugation (37°C).



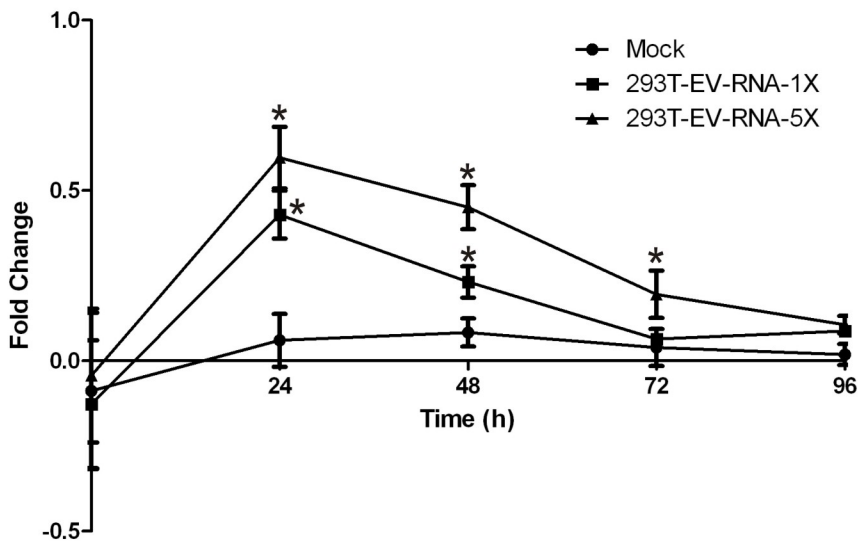
Supplementary Figure 8. FACS Analysis of nascent protein synthesis. Gli36 glioma cells stably transduced with mCherry exposed to PalmGFP+GlucB⁺ EVs (as described in legend to Fig. 6) were treated with (a) or without (b) CHX (20 μ g/ml) at 37°C. At each time point, cells were incubated with Click-iT[®] HPG for 30 min, collected for reaction with Alexa Fluor[®] 488 azide, and nascent protein synthesis was measured by FACS analysis.



b



c



Supplementary Figure 9. EV and EV-RNA increase NFkB activity in EV recipient cells. (a) Schematic of EV treatment and sample collection time to assess NFkB-Gluc activity. PBS (control) or EVs isolated from wildtype 293T cells were added to 293T cells stably expressing NFkB-Gluc followed by centrifugation at 4 °C for 1.5 h. The EV-containing media was then removed, washed with PBS and replaced with EV-depleted media. Conditioned media was collected at indicated time points to assess NFkB activity (luciferase activity) of EV-treated cells. (b) NFkB-Gluc assay of EV-treated 293T cells showed a significant increase in NFkB activity starting at 48 h and up to 72 h post-EV treatment. * $p < 0.05$ by paired t test. (c) Transfection of RNA isolated from 293T-EVs into 293T-NFkB-Gluc cells induced a robust increase in NFkB activity starting at 24 h and up to 72 h post-transfection. 1X represents the amount of RNA (54 ng) isolated from the volume of EVs used to treat 293T-NFkB-Gluc in (b). 5X indicates 5-fold more RNA (270 ng) than the 1X used for the transfection. Mock transfected samples were used as a control for Lipofectamine® 2000 transfection-induced NFkB activity while untransfected cells were used as baseline NFkB-Gluc activity for normalization. * $p < 0.05$ by one way analysis of variance with Tukey's *post hoc* test when compared to the control (mock).

Inserts	Sense/Antisense
PalmGFP	5' – ACCGGTCCACCATGCTGTGCTGTA – 3' / 5' - AATTCTCGAGGGGCCCTTACTTGTACAGCTCGTC – 3'
PalmtdTomato	5'– ACCGGTCCACCATGCTGTGCTGTATGAGAAGAACCAAACAGG TTGAAAAGAATGATGAGGACCAAAGATCATGGTGAGCAAG GGCGAG – 3'/ 5' – AATTCTCGAGGGGCCCTTACTTGTACAGCTCGTC – 3'

Supplementary Table 1. List of Primers