## Diversity of gut microflora is required for the generation of B cell with regulatory properties in a skin graft model

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## Alhabbab et al. Supplemental Figure 1



**Supplemental Figure 1 Differences in total B cells and B cell subsets in the spleens and LNs of mice maintained in the CV and SPF facilities.** Spleens and LNs were harvested from B6 mice (6 weeks old) that had been maintained in SPF or CV facilities. Splenocytes and lymphocytes were analysed by flow cytometry for total B cells and B cell populations by using antibodies against CD19, CD21, CD23 and CD24. Histograms displaying mean percentages + SEM of (A) total B cells as a percentage of splenocytes, (B) total B cells as a percentage of lymphocytes in lymph node. (C) splenic B cell subsets as a percentage of total B cells, n=3. Statistics were calculated by t test, \* P<0.05.





Supplemental Figure 2 B cell subset sorting strategy. B cells were isolated from spleens of naïve B6 mice by magnetic sorting and stained with antibodies against CD21, CD24, CD23 and with DAPI. Transitional-1 (T1), transitional-2 (T2), marginal zone (MZ) and follicular (FO) B cells were purified by BD FACSAria II. Representative FACS plots of, (A) Presort gating strategy, (B) B cell subsets post-sorting.



Supplemental Figure 3: B cells isolated from mice maintained in CV facilities suppress TNF- $\alpha$  expression by CD4+ T cells. B cells were isolated from spleens of B6 mice maintained in SPF (A) or CV (B) facilities by magnetic sorting. B cell subsets were purified by FACS and co-cultured with negatively isolated CD4+ T cells and irradiated allo-DCs from mice housed in SPF facilities (25 CD4 T cells:25 B cells:1 allo-DC) for 48hrs. PMA, Ionomycin and brefeldin A were added for the last 4 hours of culture. Summary data showing TNF- $\alpha$ , IFN- $\gamma$ , CD44 and CD69 expression on CD4+ T cells. As a control no B cells were added to the co-cultures. Graph display mean±SEM, (n=3). Statistics were calculated by one-way ANOVA and Bonferroni post-tests. \*p<0.05.

Table 1: Panel of group-specific probes (16S rRNA) selected to analyze the fecal
microbiota composition by FISH-Flow

Probe	Sequence	Target	Label 5'	Working concentration	References
EUB 338	GCTGCCTCCCGTAGGAGT	Domain Bacteria	FITC	2ng/µl	1
NON 338	ACATCCTACGGGAGGC	Negative probe	Cy5	2ng/µl (control of Bac 303 and Erec 482) and 20ng/µl (control of Lab 158 and Bif 164)	2
Bac 303	CCAATGTGGGGGGACCTT	Bacteroides	Cy5	2ng/µl	3
Erec 482	GCTTCTTAGTCARGTACCG	Clostridium coccoides– Eubacterium rectale	Cy5	2ng/µl	4
Lab 158	GGTATTAGCAYCTGTTTCCA	Lactobacillus- Streptococcus group	Cy5	20ng/µl	5
Bif 164	CATCCGGCATTACCACCC	Bifidobacterium	Cy5	20ng/µl	6

**1** Amann RI, Binder BJ, Olson RJ, Chisholm SW, Devereux R, Stahl DA. Combination of 16S rRNA-targeted oligonucleotide probes with flow cytometry for analyzing mixed microbial populations. Appl Environ Microbiol. 1990 Jun;56(6):1919-25. PubMed PMID: 2200342. Epub 1990/06/01. eng.

**2** Wallner G, Amann R, Beisker W. Optimizing fluorescent in situ hybridization with rRNAtargeted oligonucleotide probes for flow cytometric identification of microorganisms. Cytometry. 1993;14(2):136-43. PubMed PMID: 7679962. Epub 1993/01/01. eng.

**3** Manz W, Amann R, Ludwig W, Vancanneyt M, Schleifer KH. Application of a suite of 16S rRNA-specific oligonucleotide probes designed to investigate bacteria of the phylum cytophaga-flavobacter-bacteroides in the natural environment. Microbiology. 1996 May;142 (Pt 5):1097-106. PubMed PMID: 8704951. Epub 1996/05/01. eng.

**4**. Franks AH, Harmsen HJ, Raangs GC, Jansen GJ, Schut F, Welling GW. Variations of bacterial populations in human feces measured by fluorescent in situ hybridization with group-specific 16S rRNA-targeted oligonucleotide probes. Appl Environ Microbiol. 1998 Sep;64(9):3336-45. PubMed PMID: 9726880. Epub 1998/09/03. eng.

**5** Harmsen HJ, Kengen HM, Akkermans AD, Stams AJ, de Vos WM. Detection and localization of syntrophic propionate-oxidizing bacteria in granular sludge by in situ hybridization using 16S rRNA-based oligonucleotide probes. Appl Environ Microbiol. 1996 May;62(5):1656-63. PubMed PMID: 8633864. Epub 1996/05/01. eng.

**6** Langendijk PS, Schut F, Jansen GJ, Raangs GC, Kamphuis GR, Wilkinson MH, et al. Quantitative fluorescence in situ hybridization of Bifidobacterium spp. with genus-specific 16S rRNA-targeted probes and its application in fecal samples. Appl Environ Microbiol. 1995 Aug;61(8):3069-75. PubMed PMID: 7487040. Epub 1995/08/01. eng.