

Supplemental Figure Legends

Fig. S1. FXR binding peaks at the *Lsd1* gene. (A) FXR binding peaks (red bars) detected within the *Lsd1* gene from mouse liver ChIP-seq analysis (Lee et al., Hepatology, 2012). (B) Sequences for 16 IR1 motifs that are potential FXR response elements detected within 50 Kb the *Lsd1* gene are presented.

Fig. S2. Partially purified FXR and RXR α used in in the gel mobility shift assays. Flag-RXR and 3flag-FXR were expressed in Cos-1 cells and purified by M2 agarose. Eluted proteins were visualized by Coomassie blue staining. Approximate concentrations of proteins were estimated according to bovine serum albumin standards.

Fig. S3. Gel mobility shift assays using mouse liver nuclear extracts. γ -³²P-labeled oligonucleotide probe containing the IR motif #6 was incubated with 10 μ g of mouse liver nuclear extracts, 2 μ g poly-dIdC and 0.25 μ g of sonicated salmon sperm DNA, and 2.5 μ g of BSA. For antibody (Ab) supershift experiments (lanes 2-5) and competitor experiments (lanes, 6-11), Ab (0.4 μ g) or double-stranded unlabeled oligonucleotides containing the IR motif #6 (WT) or mutated (Mut) IR1 motif (1, 10, 30 ng) were preincubated with nuclear extracts for 10 min before adding the probe.

Fig. S4. Hepatic protein levels of LSD1 detected by IHC. (A, B) WT mice and FXR-KO mice were treated with GW4064 for 3 h (A) or fed 0.5 % CA-supplemented chow for 6 h (B) and hepatic LSD1 protein levels were detected by IHC.

Fig. S5. CDCA treatment increases LSD1/SHP co-localization. Immunofluorescent detection of LSD1 (green) and SHP (red) images showing co-localization (Merge) in Hepa1c1c7 cells treated with CDCA for 3 h.

Fig. S6. Cell-based luciferase reporter assays: effects of overexpression or downregulation of LSD1 on *CYP7A1*-luc activity. HepG2 cells were transfected with plasmids as indicated and treated with 50 μ M CDCA overnight and luciferase activity was measured. The values for firefly luciferase activities were normalized by dividing by the β -galactosidase activities. The mean and SEM (n=3) are plotted.

Fig. S7. Effects of downregulation of SHP on *CYP7A1* and *CYP8B1* mRNA levels in HepG2 cells. HepG2 cells were infected with Ad-shSHP or control virus and 60 h later, cells were treated with vehicle or 50 μ M CDCA for 8 h. The mRNA levels of indicated genes were measured by q-RT-PCR (SEM, n=3).

Fig. S8. Effects of downregulation of *LSD1* on occupancy of SHP and histone H3K4-me3 levels at *CYP7A1* and *CYP8B1* genes. HepG2 cells were infected with Ad-shLSD1 or control virus and 60 h later, cells were treated with 50 μ M CDCA or vehicle for 8 h and ChIP assays were performed. Three independent assays were performed (SEM, n=3).

Fig. S9. Hepatic protein levels of LSD1 detected by IHC. Mice were infected with Ad-shLSD1 or control virus and 6 days later, mice were fed a 0.5% CA-supplemented diet for 6 h. Hepatic LSD1 protein levels were detected by IHC. The scale bar is 100 μ m.

Fig. S10. Effects of downregulation of *Lsd1* on mRNA levels of *SHP* and *LSD1* after 6 h of CA feeding. Mice were infected with Ad-shSHP or control virus and 6 days later, mice were fed a 0.5 % CA-supplemented diet for 6 h. *Shp* and *Lsd1* mRNA levels were measured by q-RT-PCR (SEM, n=3 mice).

Fig. S11. Effects of downregulation of *Lsd1* on mRNA levels of *SHP* and *LSD1* after 6 days of CA feeding. Mice were infected with Ad-shSHP or control virus and 6 days later, mice were fed 0.5 % CA-supplemented diet for 6 days. *Shp* and *Lsd1* mRNA levels were measured by q-RT-PCR (SEM, n=3 mice).

Fig. S12. Effects of downregulation of *Lsd1* on mRNA levels of BA transporters. Mice were infected with Ad-shLSD1 or control virus for 6 days and then were fed CA chow for 6 days. The mRNA levels of the indicated genes involved in BA transport were measured by q-RT-PCR. The mean and SEM (n=3) are plotted and statistical significance was measured using Student's t-test. *p<0.05.

Fig. S13. Effects of downregulation of *Lsd1* on activation of ERK and JNK kinases in mouse liver. Mice were tail vein injected with Ad-shLSD1 or control Ad-Empty virus and 6 days later, mice were fed normal chow (-CA) or 0.5% CA-supplemented chow (+CA) for 6 days and livers were pooled from 3 mice. Liver extracts were prepared, and protein levels of activated phosphorylated ERK (p-ERK) or JNK (p-JNK) and total levels of ERK (t-ERK) and JNK (t-JNK) were detected by IB.

Fig. S14. Effects of downregulation of SHP or Prox1 on occupancy of LSD1 at *Cyp7a1* gene. (A) Primary mouse hepatocytes were infected with Ad-shSHP or control virus and 60 h later, cells were treated with CDCA or vehicle and ChIP assays were performed. (B) Endogenous Prox1 in hepatocytes was downregulated using siRNA and then, cells were treated with CDCA or vehicle and ChIP assays were performed. Reduced SHP and Prox1 protein levels are shown below. Three independent combined shRNA or siRNA/ChIP experiments were performed and the mean and SEM (n=3) are plotted.

Fig. S4

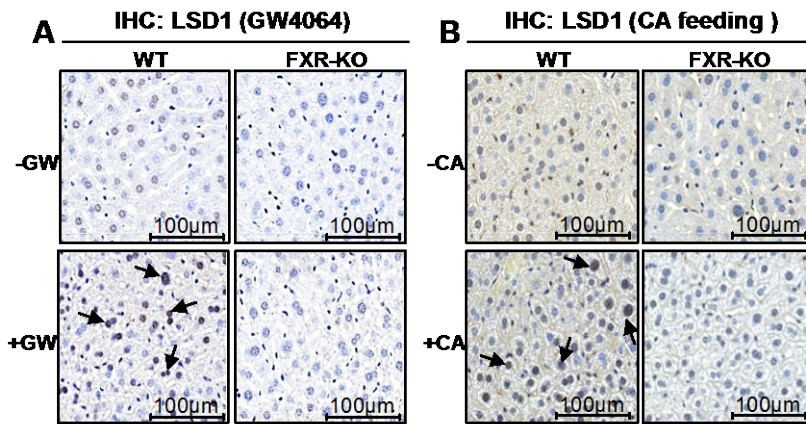


Fig. S5

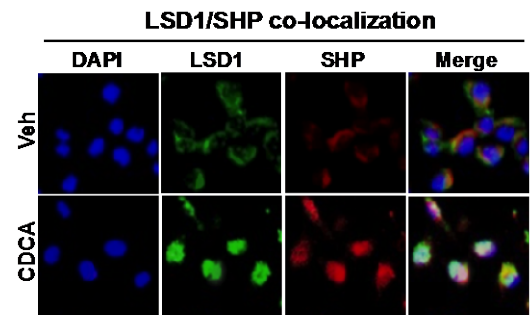


Fig. S6

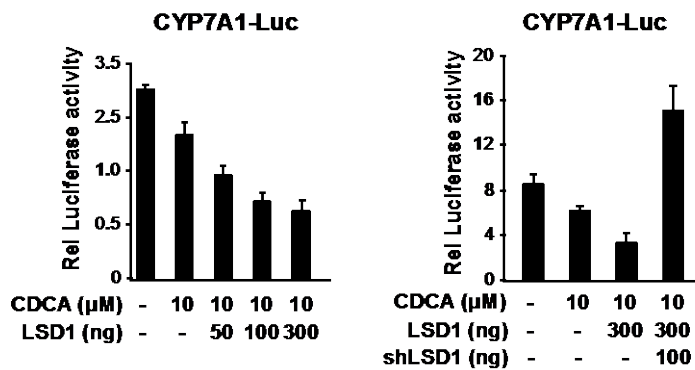


Fig. S7

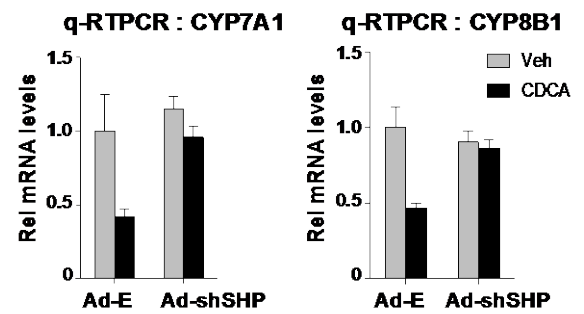


Fig. S8

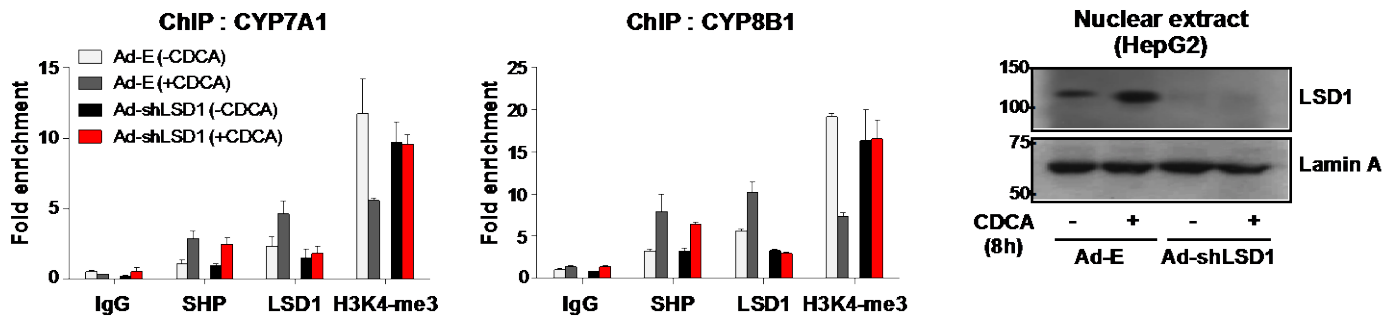


Fig. S9

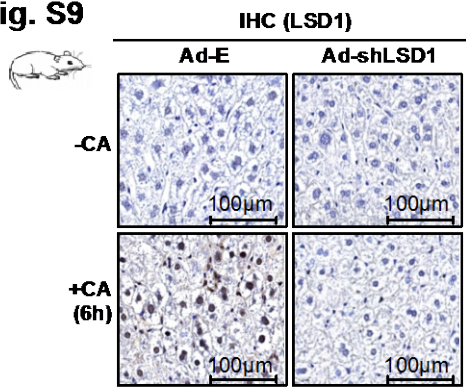


Fig. S10

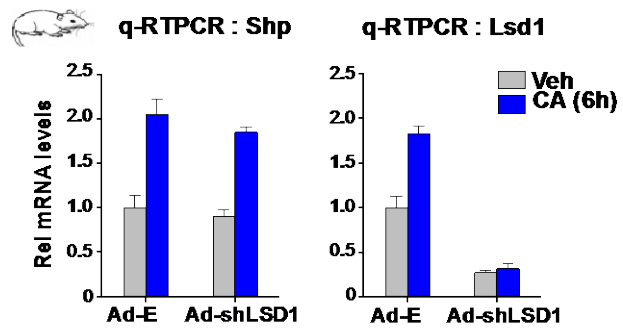


Fig. S11

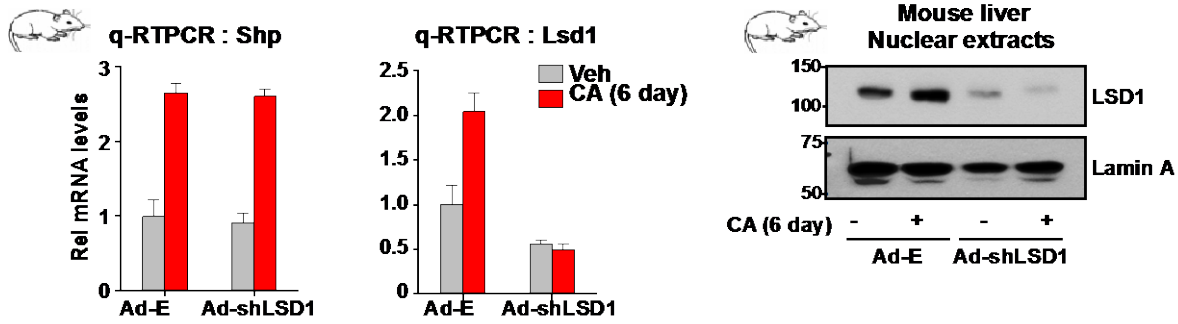


Fig. S12

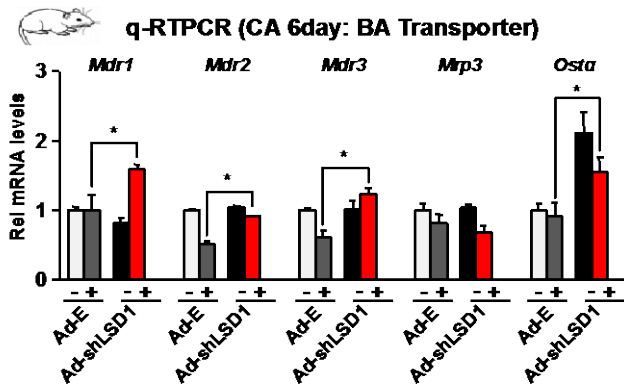


Fig. S13

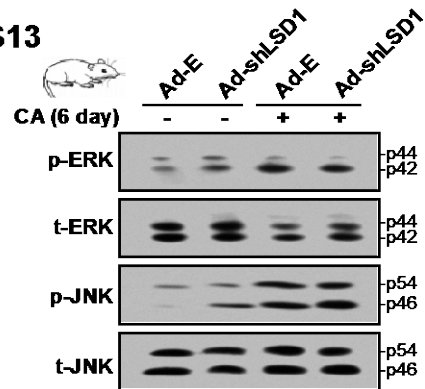


Fig. S14

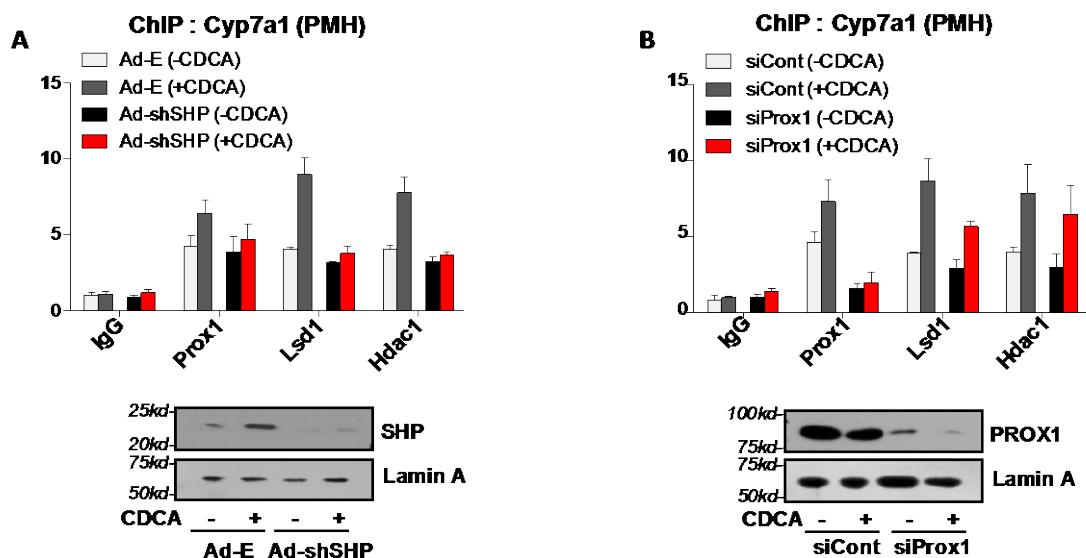


Fig. S15

Primers used in this study (ChIP and q-RT-PCR)

	Purpose	Definition	Sense Primer	Anti-sense Primer
human	ChIP	CYP7A1	GCCCATCTTAAACAGGTT	TCCACAGGTATCAGAAGT
human	ChIP	CYP8B1	CAGCCAGCCTCAGGAGAAATG	CCCCGACCCAGCGACCAGCCA
human	mRNA	36B4	TGCTGAACATGCTCAAC	GTCGAACACCTGCTGGATGAC
human	mRNA	CYP7A1	TGGGCATCGCAAGCAA	CTTTCATTGCTTCTGGGTTCTTA
human	mRNA	CYP8B1	TTCGCTTCTGCTATTACATCTT	TCCTGCTCCTTGTCCTTC
mouse	ChIP	Cyp7a1	ATATGCACAGGACCATGATC	CTTTGGTAGGTGAGCTCTTC
mouse	ChIP	Cyp8b1	AAGCATGGGGATGTGTTTAC	CAAACCTGCGGAACCTCATG
mouse	ChIP	Ntcp	TACCTCCTCCCTGATGCCTTTC	TGCGTCTGCAGCTTGGATTTA
mouse	ChIP	Shp	ACAGCAGCGATAAGCCACTT	GCCTGGGACTGCTGTTTCTA
mouse	mRNA	36B4	CCCTGAAGTGCTCGACATCA	TGCGGACACCCTCCAGAA
mouse	mRNA	Bsep	CAATGTTCAGTTCCTCCGTTCA	TTTGGTGTGTCCCCSTSTCTG
mouse	mRNA	Cyp27a1	GACAACCTCCTTTGGGACTTAC	GTGGTCTCTTATTGGTACTTGC
mouse	mRNA	Cyp2b10	TCCTGACCAGTTCCTGGATG	CTGGAGGATGGACGTGAAGAA
mouse	mRNA	Cyp3a11	CGCCTCTCCTTGCTGTCACA	CTTTGCCTTCTGCCTCAAGT
mouse	mRNA	Cyp7a1	AACGGGTTGATTCCATACCTGG	GTGGACATATTTCCCATCAGTT
mouse	mRNA	Cyp7b1	GACGATCCTGAAATAGGAGCAC	AATGGTGTGTTGCTAGAGAGGCC
mouse	mRNA	Cyp8b1	GAATCTAACAGGCCATGCT	AGGAGCTGGCACCTAGACT
mouse	mRNA	IL1b	GGACCCATATGAGCTGAAAGCT	TGTCGTTGCTTGGTTCTCCTT
mouse	mRNA	Lsd1	AGCAGCCTGTTTCCCAGACA	TGCAATGTGCGATTCTGAT
mouse	mRNA	Mdr1	CTGGTGTGCTCATAGTTG	CCTAATCTTGTGATCTGTCTT
mouse	mRNA	Mrp2	TATCCCCGGGAAATCTGTTT	TAACCAACATTCTCCGCGC
mouse	mRNA	Mrp3	AAGAGGAGATAGCAGAGA	ATACAGGAGGCAGATAGA
mouse	mRNA	Mrp4	CACCATTGCTCACAGACT	CAAGACATACGGCTCATCA
mouse	mRNA	Oatp1	CCTGGAGCAGCAATATGGAAA	CCAAGGCATACTGGAGGCAA
mouse	mRNA	Osta	TGGACCCTGGAAGACATA	TAACCACTGATAAGGCTGAG
mouse	mRNA	Ostβ	ATCTTGATGACTCCATAATG	GTCTTTCTCTTCAACTCA
mouse	mRNA	Shp	TCTGCAGGTCGTCGACTAT	CAGGCAGTGGCTGTGAGAT
mouse	mRNA	Sult2a1	AGGACCACGACTCATAACCTCCCA	CCGAGTGACCCTGGATTCTTACA
mouse	mRNA	Ugt1a1	TCTGAGCCCTGCATCTATCTG	CCCCAGAGGCGTTGACATA
mouse	Pre-mRNA	LSD1 (In1-Ex2)	AAGTCTGAAAGCAATTAGTT	ATTCATCTTCTGAGAGGTT

