nature neuroscience

Corresponding Author:	Dr. Scott Russo	# Main Figures:	3
Manuscript Number:	NN-BC45686B	# Supplementary Figures:	10
Manuscript Type:	Brief Communication	# Supplementary Tables:	0
		# Supplementary Videos:	0

Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read Reporting Life Sciences Research.

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).
- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.
- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.
- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.
- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

		TEST USED		JSED n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
	FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
example	1a	one-way ANOVA	Fig. legend	9, 9, 10, 15	mice from at least 3 litters/group	Methods para 8	error bars are mean +/- SEM	Fig. legend	p = 0.044	Fig. legend	F(3, 36) = 2.97	Fig. legend
example	results, para 6	unpaired t- test	Results para 6	15	slices from 10 mice	Results para 6	error bars are mean +/- SEM	Results para 6	p = 0.0006	Results para 6	t(28) = 2.808	Results para 6
+ -	1d	one-way ANOVA	Fig. legend	11,8,9	cells per experimental group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0141	Fig. legend	F(2,25) = 1.152	Fig. legend

		TEST USED		n		DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE		
	FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
+	1h	one-way ANOVA	Fig. legend	8,8,7	cells per experimental group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.5246	Fig. legend	F(2,20) = 0.268	Fig. legend
+	2b	unpaired t- test	Fig. legend	5,6	cells per experimental group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0042	Fig. legend	t=3.796 df=9	Fig. legend
+	2d left	unpaired t- test	Fig. legend	5,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0039	Fig. legend	t=4.017 df=8	Fig. legend
+	2d right	two-way ANOVA	Fig. legend	15,14	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0194 (interaction)	Fig. legend	F (1, 54) = 5.809 (interaction)	Fig. legend
+	2d right	unpaired t- test	Fig. legend	15,14	mice/group	Fig. legend	scatterplot with mean	Fig. legend	p = 0.0413	Fig. legend	t=2.143 df=27	Fig. legend
+	2f left	unpaired t- test	Fig. legend	4,4	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0141	Fig. legend	t=3.421 df=6	Fig. legend
+	2f right	two-way ANOVA	Fig. legend	7,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.034 (interaction)	Fig. legend	F (1, 20) = 5.170 (interaction)	Fig. legend
+	2f right	unpaired t- test	Fig. legend	7,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0131	Fig. legend	t=3.013 df=11	Fig. legend
+	3b	two-way ANOVA	Fig. legend	16,17	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0023 (interaction)	Fig. legend	F (1,62) = 10.15 (interaction)	Fig. legend
+	3b	unpaired t- test	Fig. legend	16,17	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0005	Fig. legend	t = 3.894 df =31	Fig. legend
+	3c	two-way ANOVA	Fig. legend	11,11	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.9894 (interaction)	Fig. legend	F (1, 40) = 0.0001775 (interaction)	Fig. legend
+	3c	unpaired t- test	Fig. legend	11,11	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.3601	Fig. legend	t = 1.050 df =20	Fig. legend
+	3e	two-way ANOVA	Fig. legend	14,18	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0278	Fig. legend	F (1,60) = 5.087	Fig. legend
+	3e	unpaired t- test	Fig. legend	14,18	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0088	Fig. legend	t = 2.802 df = 30	Fig. legend
+	3f	two-way ANOVA	Fig. legend	5,6	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.9533 (interaction)	Fig. legend	F (1, 18) = 0.003529 (interaction)	Fig. legend
+	3f	unpaired t- test	Fig. legend	5,6	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.8314	Fig. legend	t=0.2191 df=9	Fig. legend
+	Supp 1b	one-way ANOVA	Fig. legend	4,4,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p =0.0203	Fig. legend	F(2,10) = 5.826	Fig. legend
+	Supp 1c	linear regression	Fig. legend	13	all mice in experiment	Fig. legend	best-fit values with 95% confidence intervals	Fig. legend	p=0.0055	Fig. legend	R square = 0.5190	Fig. legend
+ -	Supp 1e	one-way ANOVA	Fig. legend	4,3,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p=0.5330	Fig. legend	F(2,9) = 0.7205	Fig. legend
+	Supp 1f	linear regression	Fig. legend	12	all mice in experiment	Fig. legend	best-fit values with 95% confidence intervals	Fig. legend	p = 0.3036	Fig. legend	R square = 0.1052	Fig. legend
+	Supp 2	one-way ANOVA	Fig. legend	10,6,8	cells per experimental group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.3708	Fig. legend	F(2,21) = 0.3615	Fig. legend

+ -	Supp 4a	linear regression	Fig. legend	10	all mice in experiment	Fig. legend	best-fit values with 95% confidence intervals	Fig. legend	p = 0.0243		R square = 0.4896	Fig. legend
+ -	Supp 4b	two-way ANOVA	Fig. legend	15,14	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0454 (interaction)	Fig. legend	F (1, 54) = 4.195 (interaction)	Fig. legend
+ -	Supp 4c	two-way ANOVA	Fig. legend	15,14	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.3155 (interaction)	Fig. legend	F (1, 54) = 1.027 (interaction)	Fig. legend
+	Supp 5a	linear regression	Fig. legend	8	all mice in experiment	Fig. legend	best-fit values with 95% confidence intervals	Fig. legend	p = 0.8626	Fig. legend	R square = 0.005410	Fig. legend
+ -	Supp 5b	two-way ANOVA	Fig. legend	7,6	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0717 (interaction)	Fig. legend	F (1, 22) = 4.392 (interaction)	Fig. legend
+ -	Supp 5c	two-way ANOVA	Fig. legend	7,6	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.7053 (interaction)	Fig. legend	F (1, 22) = 0.1468 (interaction)	Fig. legend
+ -	Supp 6c	unpaired t- test	Fig. legend	5,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p =0.1120	Fig. legend	t=1.785 df=8	Fig. legend
+ -	Supp 6d	unpaired t- test	Fig. legend	5,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.7964	Fig. legend	t=0.2658 df=8	Fig. legend
+ -	Supp 6e	unpaired t- test	Fig. legend	5,5	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.1575	Fig. legend	t=1.542 df=8	Fig. legend
+ -	Supp 6f	unpaired t- test	Fig. legend	4,4	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.5513	Fig. legend	t=0.6498 df=6	Fig. legend
+ -	Supp 6g	unpaired t- test	Fig. legend	4,4	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.4935	Fig. legend	t=0.7528 df=6	Fig. legend
+ -	Supp 6h	unpaired t- test	Fig. legend	4,4	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.6656	Fig. legend	t=0.4658 df=6	Fig. legend
+ -	Supp 7a	paired t-test	Fig. legend	16,17	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0745	Fig. legend	t=1.908 df=16	Fig. legend
+ -	Supp 7b	two-way ANOVA	Fig. legend	16,17	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p= 0.1874 (interaction)	Fig. legend	F (1, 62) = 1.777 (interaction)	Fig. legend
+ -	Supp 7c	unpaired t- test	Fig. legend	11,11	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0485	Fig. legend	t=2.102 df=20	Fig. legend
+ -	Supp 7d	two-way ANOVA	Fig. legend	11,11	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.4024 (interaction)	Fig. legend	F (1,40) = 0.7162 (interaction)	Fig. legend
+ -	Supp 8c	two-way ANOVA	Fig. legend	14,18	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0458 (interaction)	Fig. legend	F (1, 60) = 6.893 (interaction)	Fig. legend
+ -	Supp 8d	two-way ANOVA	Fig. legend	6,6	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.5183 (interaction)	Fig. legend	F (1, 20) = 0.4324 (interaction)	Fig. legend
+ -	Supp 8e	two-way ANOVA	Fig. legend	14,18	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.3258 (interaction)	Fig. legend	F (1, 60) = 0.9815 (interaction)	Fig. legend
+ -	Supp 8f	two-way ANOVA	Fig. legend	6,6	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.4716 (interaction)	Fig. legend	F (1, 20) = 0.5385 (interaction)	Fig. legend
+ -	Supp 9a	unpaired t- test	Fig. legend	16,14	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0045	Fig. legend	t=3.087 df=27	Fig. legend
+ -	Supp 9a	unpaired t- test	Fig. legend	16,14	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.1060	Fig. legend	t=1.672 df=27	Fig. legend
+ -	Supp 9b	unpaired t- test	Fig. legend	16,14	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0325 (interaction)	Fig. legend	t=2.372 df=14 (interaction)	Fig. legend
+ -	Supp 9c	unpaired t- test	Fig. legend	16,14	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.3617	Fig. legend	F (1, 34) = 0.8549 (interaction)	Fig. legend
+ -	Supp 10a	two-way ANOVA	Fig. legend	9,10	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0459 (interaction)	Fig. legend	F (1, 34) = 4.294 (interaction)	Fig. legend
+ -	Supp 10a	unpaired t- test	Fig. legend	9,10	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0408	Fig. legend	t=2.214 df=17	Fig. legend
+ -	Supp 10b	two-way ANOVA	Fig. legend	9,10	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.6404 (interaction)	Fig. legend	F (1, 34) = 0.2221 (interaction)	Fig. legend
+ -	Supp 10c	two-way ANOVA	Fig. legend	9,10	mice/group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.8132 (interaction)	Fig. legend	F (1, 34) = 0.05669 (interaction)	Fig. legend

Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

If so, what figure(s)?

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

If so, where is this reported (section, paragraph #)?

Yes, IHC and electrophysiology traces Figures 1b,c,f,g; 2a,c,e; 3a,d Supp Figures 1a,d,g,h,i,j; 3a,b; 6a,b; 9a,b

Yes, all experiments were done w mutliple mice per group. All data collected was collapsed to an average per animal or experimental condition. N per group is listed in the figure legends

For spine density experiments, ~ 10 dendrites/animal~ 1,000 spines per group were analyzed. This is listed in the methods under Imaging and Analysis on pg 20

Statistics and general methods

1. Is there a justification of the sample size?

If so, how was it justified?

Where (section, paragraph #)?

Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

2. Are statistical tests justified as appropriate for every figure?

Where (section, paragraph #)?

- a. If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?
- b. Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?

Where is this described (section, paragraph #)?

c. Is there any estimate of variance within each group of data?

Is the variance similar between groups that are being statistically compared?

Where is this described (section, paragraph #)?

d. Are tests specified as one- or two-sided?

e. Are there adjustments for multiple comparisons?

No justification of sample size is included in the manuscript. However, sample size was based on previously published work (Krishnan et al., Cell, 2007; Christoffel et al., J. Neuro., 2011; Christoffel et al., Neuropsychopharm., 2012) and animal protocol (approved by IACUC). This is stated in the methods section. A sample size of 5-15 is appropriate to see a significant difference both in behavior tests, 5-10 cells for electrophysiology, 4-6 for dendritic spine density and VGLUT puncta, and 3 for c-Fos mapping experiments without unnecessary sacrifice.

Yes. in methods under statistics

Yes

Yes in methods under statistics

Yes, F tests for variance were carried out for all analysis, where appropriate a Bartlett's test for equal variances was performed for the one-way ANOVAs.

It is stated in the methods statistics section that the data has equal variance.

Yes in methods statistics section

N/A

Are criteria for excluding data points reported?
 Was this criterion established prior to data collection?

Where is this described (section, paragraph #)?

 Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data.

If no randomization was used, state so.

Where does this appear (section, paragraph #)?

5. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included?

If no blinding was done, state so.

Where (section, paragraph #)?

6. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included?

Where (section, paragraph #)?

7. Is the species of the animals used reported?

Where (section, paragraph #)?

8. Is the strain of the animals (including background strains of KO/ transgenic animals used) reported?

Where (section, paragraph #)?

9. Is the sex of the animals/subjects used reported?

Where (section, paragraph #)?

10. Is the age of the animals/subjects reported?

Where (section, paragraph #)?

- For animals housed in a vivarium, is the light/dark cycle reported?
 Where (section, paragraph #)?
- 12. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported?

Where (section, paragraph #)?

13. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)?

Where (section, paragraph #)?

No, we did not have data points to exclude.

Animals were randomized by cages before surgeries. For example, if there were 30 animals in an experiment, with 5 animals per cage, animals were randomly assigned to be control or manipulation groups. Additionally, the order of the animals were randomized prior to behavioral tests.

This is mentioned in the text under animals section in methods

This is mentioned in the imaging analysis and in vitro electrophysiology section of the methods. All behavioral data collected was automated through the use of video tracking software.

Yes, methods animals

Yes, methods animals

Yes, methods animals

Yes , methods animals

Yes , methods animals

Yes , pg 13

No, animals were housed 5 per cage until performance of surgery. Animals were then singly housed till the end of the experiment

Yes , methods animals

14. Is the pr administ	revious history of the animals/subjects (e.g. prior drug tration, surgery, behavioral testing) reported?	This did not occur
Where (section, paragraph #)?	
a.	If multiple behavioral tests were conducted in the same group of animals, is this reported?	
	Where (section, paragraph #)?	
15. If any ar Where (nimals/subjects were excluded from analysis, is this reported? section, paragraph #)?	Animals were excluded for the following reasons.1. missed viral injection2. missed cannula placement3. no/low viral expression
		This is listed under perfusion and tissue processing, of the methods
a.	How were the criteria for exclusion defined?	See above
	Where is this described (section, paragraph #)?	
b.	Specify reasons for any discrepancy between the number of animals at the beginning and end of the study.	For spine morphology analysis we processed tissue from only a subset of the total animals run through social defeat behavior. This was due to the labor intensive nature of this analysis and our
	Where is this described (section, paragraph #)?	previous results which suggested that we only need between 4-7 animals for appropriate power (Christoffel et al., J Neurosci, 2011; Golden et al., Nat Med., 2013).

▶ Reagents

- 1. Have antibodies been validated for use in the system under study (assay and species)?
 - a. Is antibody catalog number given?

Where does this appear (section, paragraph #)?

b. Where were the validation data reported (citation, supplementary information, Antibodypedia)?

Where does this appear (section, paragraph #)?

2. If cell lines were used to reflect the properties of a particular tissue or disease state, is their source identified?

Where (section, paragraph #)?

a. Were they recently authenticated?

Where is this information reported (section, paragraph #)?

Yes.

yes under immunohistochemistry in methods

Antibodies used for protein quantification were mentioned on page the immunohistochemistry section in the methods. These were chosen based on previous published work although references were not give in the methods.

NA

Data deposition

Data deposition in a public repository is mandatory for:

- a. Protein, DNA and RNA sequences
- b. Macromolecular structures
- c. Crystallographic data for small molecules
- d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available here. We encourage the provision of other source data in supplementary information or in unstructured repositories such as Figshare and Dryad.

We encourage publication of Data Descriptors (see Scientific Data) to maximize data reuse.

1. Are accession codes for deposit dates provided?

NA

NA

NA

Where (section, paragraph #)?

Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

- 1. Identify all custom software or scripts that were required to conduct the study and where in the procedures each was used.
- If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "Code availability" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

Human subjects

1. Which IRB approved the protocol?

Where is this stated (section, paragraph #)?

- Is demographic information on all subjects provided? Where (section, paragraph #)?
- Is the number of human subjects, their age and sex clearly defined?
 Where (section, paragraph #)?
- Are the inclusion and exclusion criteria (if any) clearly specified? Where (section, paragraph #)?

NA

INA

NA

NA

5. How well were the groups matched?

Where is this information described (section, paragraph #)?

6. Is a statement included confirming that informed consent was obtained from all subjects?

Where (section, paragraph #)?

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?

Where (section, paragraph #)?

fMRI studies

For papers reporting functional imaging (fMRI) results please ensure that these minimal reporting guidelines are met and that all this information is clearly provided in the methods:

NA

1.	Were an data was	y subjects scanned but then rejected for the analysis after the collected?	NA
	a.	If yes, is the number rejected and reasons for rejection described?	
		Where (section, paragraph #)?	
2.	Is the nu or subjec	mber of blocks, trials or experimental units per session and/ ts specified?	NA
	Where (s	section, paragraph #)?	
3.	Is the ler	ngth of each trial and interval between trials specified?	NA
4.	Is a block please sp design w	ked, event-related, or mixed design being used? If applicable, becify the block length or how the event-related or mixed as optimized.	NA
5.	Is the tas	k design clearly described?	NA
	Where (s	section, paragraph #)?	
6.	How was	behavioral performance measured?	NA
7.	Is an ANG	DVA or factorial design being used?	ΝΑ
8.	For data	acquisition, is a whole brain scan used?	NA
	If not, sta	ate area of acquisition.	
	a.	How was this region determined?	NA

NA

9. Is the field strength (in Tesla) of the MRI system stated?

a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated?

- b. Are the field-of-view, matrix size, slice thickness, and TE/TR/ flip angle clearly stated?
- Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated?
- 11. Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)?
- 12. If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)?
- 13. How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.?
- 14. Were any additional regressors (behavioral covariates, motion etc) used?
- 15. Is the contrast construction clearly defined?
- 16. Is a mixed/random effects or fixed inference used?
 - a. If fixed effects inference used, is this justified?
- 17. Were repeated measures used (multiple measurements per subject)? NA
 - a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated?
- 18. If the threshold used for inference and visualization in figures varies, is NA this clearly stated?
- 19. Are statistical inferences corrected for multiple comparisons?
 - a. If not, is this labeled as uncorrected?

NA

NA

NA

NA

NA

NA

NA

NA

NA

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- 20. Are the results based on an ROI (region of interest) analysis?
 - a. If so, is the rationale clearly described?
 - b. How were the ROI's defined (functional vs anatomical localization)?
- 21. Is there correction for multiple comparisons within each voxel?
- 22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined?

Additional comments

Additional Comments

NA			
NA			
NA			