### **SUPPLEMENTARY METHODS**

## Merge of the results obtained in two independent NanoString analyses

To normalize the results obtained in two Nanostring experiments (code-sets 1 and 2), first, normalization with spiked positive controls was performed [1]. A normalization factor (NF) (Vandesompele et al., 2002) was calculated for each sample and the counts were corrected:

 $\mathbf{NF}_{sample} = \frac{sum of positive controls_{sample}}{reference}$ Corrected count sample/gene = raw count sample/gene × NF sample?

where:

reference –is the sum of positive controls of the sample (the summed value is close to the average for all samples).

Then code-set calibration was implemented using four samples (2 benign and 2 PTC samples number 42 and 46 from multinodular goiters) represented in both codesets. For each gene in common ( $\times$ 21), a calibration factor (CF) was calculated:

**CF** gene = mean of <u>corrected count</u> gene sample code-set1 corrected count gene sample code-set2

The results from code-set 2 were adjusted to codeset 1 using CF as follows:

 $\begin{array}{c} \textbf{Calibrated count}_{\text{sample code-set 2/gene}} {=} \text{corrected count}_{\text{sample/gene}} \\ {\times} \text{ CF}_{\text{gene}} \end{array}$ 

#### Gene score data analysis

Predictive score for each biological sample was based on the combined expression levels of chosen five distinction genes *BMAL1*, *CHEK1*, *c-KIT*, *c-MET* and *TIMP1* exhibiting stable changes in PTCs. To ensure the reproducibility of our model, we equally divided 17 benign (Table 1) and 41 PTC (Tables 1–2) samples into training and validation sets by computer-based randomization (Supplementary Table 3; [2, 3]). The training set was used to create and optimize the gene-expression score, while the validation set allowed testing the model and evaluating its significance. Using the training set and *t*-test analysis we identified the genes that were differentially expressed in the benign and PTC samples. Five genes with high fold-change expression and high significance (*P-value* +5% *FDR* < 0.05) were chosen for the gene-expression score calculation. The group of five distinction genes includes *BMAL1*, *CHEK1*, *c-KIT*, *c-MET* and *TIMP1*. To calculate PTC prediction score, first the expression levels of distinction genes were combined to calculate a Linear Predictor Score (LPS) [4] for each sample (X) in the training set:

$$LPS(X) = \sum_{j} a_{j} X_{j}$$

Where:

X<sub>i</sub> –a gene expression value of gene j

 $a_{j}$  –a scaling factor, whose value depends on the degree, to which each gene discriminates in the subgroup. The scaling factors were chosen to be the t-statistics generated by a *t*-test for the difference in expression between PTC and benign subgroups.

To obtain the PTC prediction score comprised between 0 and 1, the following score transformation was performed: if LPS was less than 100, it was set to 100; if LPS was greater than 200, it was set to 200. The final PTC prediction score values were calculated as follows:

Score (X) = (LPS(X) - 100)/100

A similar procedure of PTC prediction score calculation was used for validation set. The score values for training and validation set samples are shown in Supplementary Table 3.

To test the performance of the score, a receiver operating characteristic (ROC) analysis with calculation of the ROC area under the curve (AUC) statistic was performed [2, 3, 5]. The ROC curve (Supplementary Figure 1) gives sensitivity and specificity of a certain test in function of the cut off value: the better the diagnostic test, the nearer the curve reaches to the top left corner, indicating 100% sensitivity and specificity.

## SUPPLEMENTARY FIGURE AND TABLES



**Supplementary Figure S1: ROC analysis.** ROC analysis with calculation of the area under the curve (AUC) statistic was performed to test in validation set (left) and all samples (right). Binormal smoothing iteration was performed to show continuous ROC curves. At the threshold 0.27 PTC diagnostic score discriminates PTC from benign cases with 90% sensitivity and 100% specificity in the validation set (left) and all sample analysis (right).

# Supplementary Table S1: NanoString probes design

Gene	Accession	<b>Target Region</b>	Gene	Accession	<b>Target Region</b>
AKT1 <sup>1</sup>	NM_005163.2	1773–1872	<i>PER2</i> <sup>1,2</sup>	<i>PER2</i> <sup>1,2</sup> NM_022817.2	
$AKT2^{1,2}$	NM_001626.2	1451-1550	<i>PER3</i> <sup>1,2</sup>	NM_016831.1	1076–1175
ALDH1 <sup>2</sup>	NM_000689.4	1161–1260	PIK3CA <sup>1</sup>	NM_006218.2	2446-2545
BCL2 <sup>1,2</sup>	NM_000657.2	6–105	PIK3CB <sup>1</sup>	NM_006219.1	2946-3045
BMAL1 <sup>1,2</sup>	NM_001030272.1	841–940	PLEKHA7 <sup>1</sup>	NM_175058.4	4137–4236
BRAF <sup>1</sup>	NM_004333.3	566–665	$PPAR\gamma^{1,2}$	<i>PPARy</i> <sup>1,2</sup> NM_015869.3	
CCND1 <sup>2</sup>	NM_053056.2	691–790	PTEN <sup>2</sup>	NM_000314.3	1676–1775
CDH1 <sup>2</sup>	NM_004360.2	536-635	<i>Rev-Erba</i> <sup>2</sup>	NM_021724.3	1081-1180
CDK1 <sup>1</sup>	NM_001786.4	179–278	$ROR\alpha^2$	NM_134261.2	1716–1815
CDKN1A <sup>2</sup>	NM_000389.2	1976–2075	SIRT1 <sup>1</sup>	NM_012238.4	841–940
CHEK1 <sup>1,2</sup>	NM_001114121.1	2226–2325	SLC5A5 <sup>2</sup>	NM_000453.2	3459–3558
CLOCK <sup>1</sup>	NM_004898.2	2351-2450	SLC26A4 <sup>2</sup>	NM_000441.1	1711–1810
CRY1 <sup>1,2</sup>	NM_004075.3	1376–1475	SOX9 <sup>2</sup>	NM_000346.2	2136–2235
<i>CRY2</i> <sup>1,2</sup>	NM_001127457.1	3326–3425	$TG^{1,2}$	NM_003235.4	6499–6598
CXCR4 <sup>1</sup>	NM_003467.2	1336–1435	TIMP1 <sup>1,2</sup>	NM_003254.2	330–429
$DBP^2$	NM_001352.2	1051-1150	$TPO^2$	NM_175719.3	297–396
DDIT3 <sup>1,2</sup>	NM_004083.4	41-140	TSHR <sup>1,2</sup>	NM_001018036.2	736–835
DIO2 <sup>2</sup>	NM_013989.3	5076-5175	<i>TP53</i> <sup>1</sup>	NM_000546.2	1331–1430
EGFR <sup>1</sup>	NM_201282.1	361-460	UCP1 <sup>2</sup>	NM_021833.4	551-650
IQGAP1 <sup>1</sup>	NM_003870.3	821–920	$VDR^1$	NM_000376.2	4386-4485
<i>c</i> - <i>KIT</i> <sup>1,2</sup>	NM_000222.1	6–105	VEGFR1 <sup>1,2</sup>	NM_002019.4	531-630
MCM2 <sup>2</sup>	NM_004526.2	2946-3045	WEE1 <sup>2</sup>	NM_003390.3	1226–1325
<i>c-MET</i> <sup>1,2</sup>	NM_000245.2	406–505	House-keeping genes		
$MYC^1$	NM_002467.3	1611–1710	ACTB <sup>1,2</sup>	NM_001101.2	1011-1110
NFIL3 <sup>2</sup>	NM_005384.2	1796–1895	$B2M^1$	NM_004048.2	26-125
NTRK1 <sup>1</sup>	NM_001012331.1	1366–1465	EEF1A1 <sup>2</sup>	NM_001402.5	791–890
PDGFRA <sup>1,2</sup>	NM_006206.4	1388–1487	GAPDH <sup>1</sup>	NM_002046.3	973–1072
PDGFRB <sup>1</sup>	NM_002609.3	809–908	HPRT1 <sup>1,2</sup>	NM_000194.1	241-340
PER1 <sup>1,2</sup>	NM_002616.2	4366-4465	RPL13A <sup>1,2</sup>	NM_012423.2	721-820

<sup>1</sup>Code-set 1, <sup>2</sup>Code-set 2

Supplementary Table S2: Thirty four transcripts assessed by NanoString analysis exhibit comparable levels in FFPE and fresh-frozen samples obtained from the same donors

	Case <sup>1</sup>	Number of significantly different transcripts <sup>2</sup> (at 5% + FC ≥ 2) <sup>3</sup>	Number of significantly different transcripts (at FDR 5% + FC ≥ 2) <sup>4</sup>		
Benign	21		0		
	22	(DDCEDD DDCEDA)			
	23	2 (PDGFKB, PDGFKA)			
	24				
PTC	1				
	5	$2(VECED \mid DDADC)$	0		
	6	2(VEGFRI, PPARG)	0		
	7				

<sup>1</sup>Donor characteristics are presented in Table 1 (subjects labeled with \*);

<sup>2</sup>Transcript list and probes design are presented in Supplementary Table 1, code-set 1;

<sup>3</sup>Number of significantly different transcripts (at 5% + FC  $\ge$  2)" corresponds to raw-*P*-value <0.05 and fold-change in gene expression  $\ge$  2;

<sup>4</sup>Number of significant genes (at FDR 5% + FC  $\geq$  2) corresponds to *P*-value with False Discovery Rate (FDR) < 0.05 and fold-change in gene expression  $\geq$  2.

Case	Histologic diagnosis	Score	Predicted diagnosis (threshold = 0.27)	Case	Histologic diagnosis	Score	Predicted diagnosis (threshold = 0.27)
18 <sup>v</sup>	benign	0.00	benign	13 <sup>II,V</sup>	PTC	0.56	РТС
19 <sup>т</sup>	benign	0.00	benign	14 <sup>II,V</sup>	PTC	0.49	РТС
20 <sup>v</sup>	benign	0.09	benign	15 <sup>II,V</sup>	PTC	0.51	РТС
21 <sup>T</sup>	benign	0.13	benign	16 <sup>II,T</sup>	PTC	0.57	РТС
22 <sup>T</sup>	benign	0.10	benign	17 <sup>II,V</sup>	PTC	0.67	РТС
23 <sup>v</sup>	benign	0.17	benign	35 <sup>I,T</sup>	PTC	0.10	benign
24 <sup>T</sup>	benign	0.09	benign	37 <sup>I,V</sup>	PTC	0.31	РТС
25 <sup>v</sup>	benign	0.25	benign	<b>39</b> <sup>I,T</sup>	PTC	0.29	РТС
26 <sup>T</sup>	benign	0.22	benign	41 <sup>I,T</sup>	PTC	0.50	РТС
27 <sup>т</sup>	benign	0.27	benign	43 <sup>I,V</sup>	PTC	0.28	РТС
28 <sup>T</sup>	benign	0.25	benign	45 <sup>II,V</sup>	PTC	0.58	PTC
29 <sup>v</sup>	benign	0.20	benign	47 <sup>II,V</sup>	PTC	0.51	PTC
30 <sup>T</sup>	benign	0.15	benign	49 <sup>II,V</sup>	PTC	0.61	PTC
31 <sup>v</sup>	benign	0.15	benign	51 <sup>II,V</sup>	PTC	0.56	PTC
32 <sup>T</sup>	benign	0.20	benign	53 <sup>II,T</sup>	PTC	0.51	PTC
33 <sup>v</sup>	benign	0.26	benign	55 <sup>II,V</sup>	PTC	0.53	РТС
34 <sup>v</sup>	benign	0.22	benign	57 <sup>II,T</sup>	PTC	0.61	РТС
1 <sup>I,V</sup>	PTC	0.61	РТС	59 <sup>I,V</sup>	PTC	0.50	РТС
2 <sup>II,V</sup>	PTC	0.51	РТС	61 <sup>I,T</sup>	PTC	0.37	РТС
3 <sup>II,T</sup>	PTC	0.95	PTC	63 <sup>I,V</sup>	PTC	0.30	PTC
4 <sup>II,T</sup>	PTC	0.63	PTC	65 <sup>I,T</sup>	PTC	0.23	benign
5 <sup>II,T</sup>	PTC	0.55	PTC	67 <sup>I,T</sup>	PTC	0.38	PTC
6 <sup>II,V</sup>	PTC	0.58	РТС	69 <sup>I,V</sup>	PTC	0.14	benign
7 <sup>II,T</sup>	PTC	0.71	PTC	71 <sup>II,T</sup>	PTC	0.36	PTC
8 <sup>I,T</sup>	PTC	0.63	PTC	73 <sup>II,V</sup>	PTC	0.65	PTC
9 <sup>II,T</sup>	PTC	0.63	PTC	75 <sup>II,T</sup>	PTC	0.49	PTC
10 <sup>II,T</sup>	PTC	0.72	РТС	77 <sup>II,V</sup>	PTC	0.19	benign
11 <sup>II,V</sup>	PTC	0.63	РТС	79 <sup>II,T</sup>	PTC	0.47	РТС
12 <sup>II,T</sup>	PTC	0.55	PTC	81 <sup>II,V</sup>	PTC	0.46	PTC

# Supplementary Table S3: Diagnostic subgroup definition using gene expression-based score

<sup>T</sup>training set, <sup>V</sup>validation set, <sup>I</sup>less aggressive PTC, <sup>II</sup>more aggressive PTC