

Supplementary Information

Dominant and novel clades of *Candidatus Accumulibacter phosphatis* in 18 globally distributed full-scale wastewater treatment plants

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database. Sequences belonging to *Accumulibacter* clades IIIH and II-I are from the clone libraries. The node labels refer to bootstrap support values.

Figure S4. Agarose gel electrophoresis images of the qPCR products targeting the *ppk1* genes in clades IIB and IIC respectively. Plasmids containing specific copy numbers of *ppk1* genes were used for drawing the standard curves. Some *ppk1* genes amplified from the activated sludge samples by using the *ppk1* gene primer sets were visualized. Sterile water was used for the negative control.

Table S1. Optimal primers information and qPCR conditions for this study.

Target	Primer	Sequence (5' – 3')	T ^a (°C)	Primer concentration (nM)	Standard curve correlation efficient (R ²)	PCR efficiency (%)
Acc-IA <i>ppk1</i>	Acc-ppk1-763f	GACGAAGAAGCGGTCAAG	61	500	0.99 ± 0.01	105 ± 2
	Acc-ppk1-1170r	AACGGTCATCTTGATGGC				
Acc-IIA <i>ppk1</i>	Acc-ppk1-893f	AGTTCAATCTCACCGAGAGC	61	400 ^a	0.98 ± 0.01	92 ± 5
	Acc-ppk1-997r	GGAACTTCAGGTCGTTGC				
Acc-IIB <i>ppk1</i>	Acc-ppk1-870f	GATGACCCAGTTCCTGCTCG	61	400	0.98 ± 0.01	88 ± 2
	Acc-ppk1-1002r	CGGCACGAACTTCAGATCG				
Acc-IIC <i>ppk1</i> excluding OTU NS D3 ^b	Acc-ppk1-1123f	GAACAGTCCGCCAACGACC	63	500	0.97 ± 0.01	97 ± 5
	Acc-ppk1-1376r	ACGATCATCAGCATCTTGCC				
Acc-IID <i>ppk1</i>	Acc-ppk1-375f	GGGTATCCGTTTCCTCAAGCG	63	400	0.97 ± 0.01	97 ± 4
	Acc-ppk1-522r	GAGGCTCTTGTTGAGTACACGC				
Acc 16S rRNA genes	518f	CCAGCAGCCGCGGTAAT	65	400	0.99 ± 0.01	106 ± 1
	846r	GTTAGCTACGGCACTAAAAGG				
Bacterial 16S rRNA genes	341f	CCTACGGGAGGCAGCAG	60	400	0.98 ± 0.01	87 ± 1
	534r	ATTACCGCGGCTGCTGG				

^a Modified qPCR condition from previously published paper ¹.

^b The relative abundance of Accumulibacter in Clade IIC was estimated by the qPCR assay using primer sets targeting Accumulibacter IIC excluding OTU NS D3 ¹.

Table S2. Relative distribution (%) of indicated clade of *Accumulibacter* in the bacterial communities according to the qPCR analysis using *ppk1* gene as biomarker.

Sample	I	IIA	IIB	IIC	IID
UK-WL-OW	1.64	0.11	0.25	0.08	0.60
UK-NW-NW	0.10	0.00	0.20	0.06	1.45
UK-SB-SB	0.03	0.01	0.00	0.06	0.00
UK-LB-LB	0.79	0.24	0.26	0.05	0.24
US-CO-CO	0.02	0.01	0.22	0.01	0.06
US-GR-PC	0.01	0.22	0.11	0.22	0.00
CA-GP-GP	0.00	0.00	0.00	0.00	0.00
JP-A2O-TK	0.03	0.00	0.47	0.60	0.06
JP-STD-TK	0.00	0.32	0.06	0.14	0.00
SG-SG-UP	3.68	0.19	0.48	0.60	2.07
CN-BJ-BX	0.04	0.01	0.05	0.07	0.10
CN-NJ-SJ	0.00	0.00	0.00	0.01	0.91
CN-SH-MH	0.00	0.31	0.09	0.38	0.07
CN-GZ-DT	0.00	0.35	0.29	0.40	0.28
CN-WH-LW	0.00	2.02	1.36	0.50	0.00
CN-QD-TD	0.00	0.00	0.05	0.07	0.31
CN-HK-ST	0.00	0.00	0.00	0.00	0.00
CN-HK-SH	0.00	0.00	0.14	0.33	0.01

Table S3. Relative distribution (%) of indicated clade within the Accumulibacter lineage by estimating the total abundance of the Accumulibacter lineage based on the qPCR assays targeting the 16S rRNA genes.

Sample	I	IIA	IIB	IIC	IID	Other clades
UK-WL-OW	49.2	3.4	7.5	2.3	17.9	19.7
UK-NW-NW	5.2	0.0	10.6	3.4	78.3	2.5
UK-SB-SB	26.6	7.0	3.5	62.6	0.0	0.0
UK-LB-LB	39.7	12.1	12.9	2.5	12.0	20.8
US-CO-CO	3.7	1.0	32.7	1.3	8.7	52.6
US-GR-PC	1.0	14.6	7.2	14.9	0.0	62.4
CA-GP-GP	0.0	0.0	0.0	0.0	0.0	0.0
JP-A2O-TK	1.8	0.0	30.9	39.6	3.7	24.0
JP-STD-TK	0.0	39.9	7.6	17.0	0.0	35.5
SG-SG-UP	26.0	1.4	3.4	4.3	14.6	50.5
CN-BJ-BX	3.3	0.5	4.3	6.3	9.3	76.3
CN-NJ-SJ	0.0	0.0	0.0	0.9	99.5	0.0
CN-SH-MH	0.0	12.7	3.8	15.5	2.8	65.3
CN-GZ-DT	0.0	7.5	6.3	8.7	6.0	71.6
CN-WH-LW	0.0	25.6	17.2	6.3	0.0	50.8
CN-QD-TD	0.0	0.0	0.5	0.7	3.1	95.6
CN-HK-ST	0.1	0.0	0.0	0.2	0.5	99.3
CN-HK-SH	0.0	0.0	4.4	10.6	0.3	84.7

Table S4. Summary statistics for redundancy analysis based on the relative abundances of different *Accumulibacter* clades in the bacterial communities.

	Explain	P-value (focus scaling on inter-clades correlation)	P-value (focus scaling on inter-sample distance)
TP	0.17	0.08	0.08
Latitude	0.11	0.16	0.16
COD	0.04	0.42	0.42
Sampling temperature	0.04	0.63	0.63
TN	0.02	0.56	0.56

Abbreviations: TP, total phosphorus; COD, chemical oxygen demand; TN, total nitrogen.

Table S5. Summary statistics for redundancy analysis based on the relative abundances of different *Accumulibacter* clades in the *Accumulibacter* lineage.

	Explain	P-value (focus scaling on inter-clades correlation)	P-value (focus scaling on inter-sample distance)
TP	0.15	0.04	0.04
COD	0.14	0.05	0.06
TN	0.12	0.06	0.07
Sampling temperature	0.02	0.85	0.86
Latitude	0.01	0.98	0.98

Abbreviations: TP, total phosphorus; COD, chemical oxygen demand; TN, total nitrogen.

Table S6. Reference 16S rRNA gene sequences of Accumulibacter lineage.

PAO	Clade	Accession NO.	Ref.
Accumulibacter	I	AF204244	1
		AF255641	
		AF502224	
		AF502225	
		AJ224937	
		AY064178	
		AY064179	
		EF565148	
	IIA	AF204247	
		AF502227	
		AF502229	
		AF502230	
		AF502231	
		EF565147	
		EF565153	
		EF565155	
	IIB	EF565156	
		EF565149	
		EF565150	
		EF565152	
	IIC & IID	EF565157	
AY062125			
EF565151			
EF565158			
EF565159			
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EF565161			

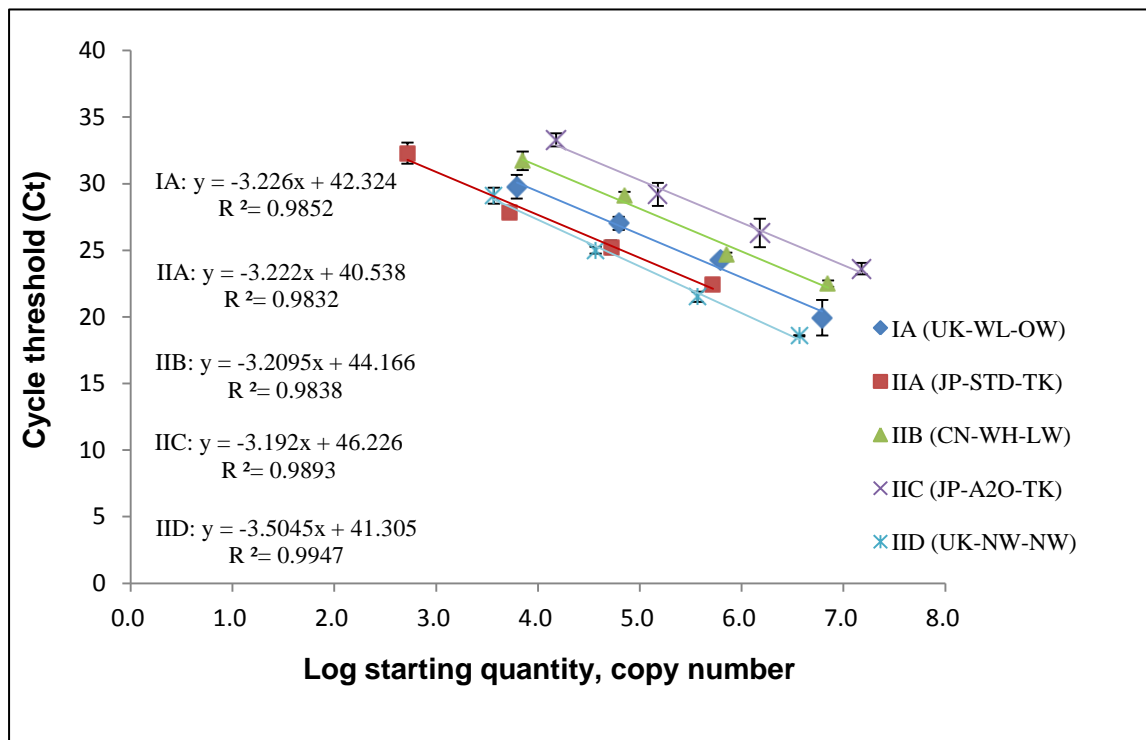
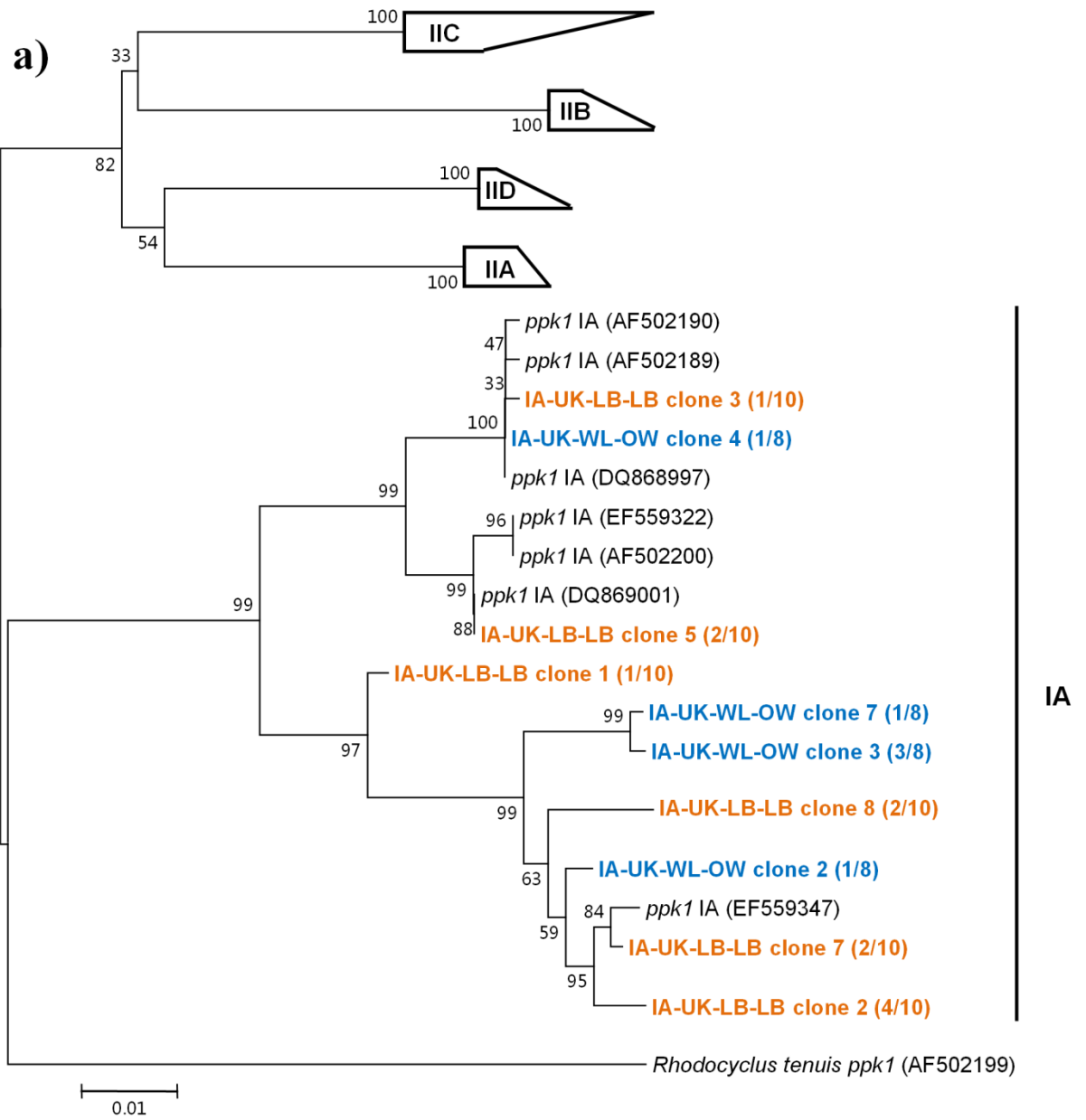
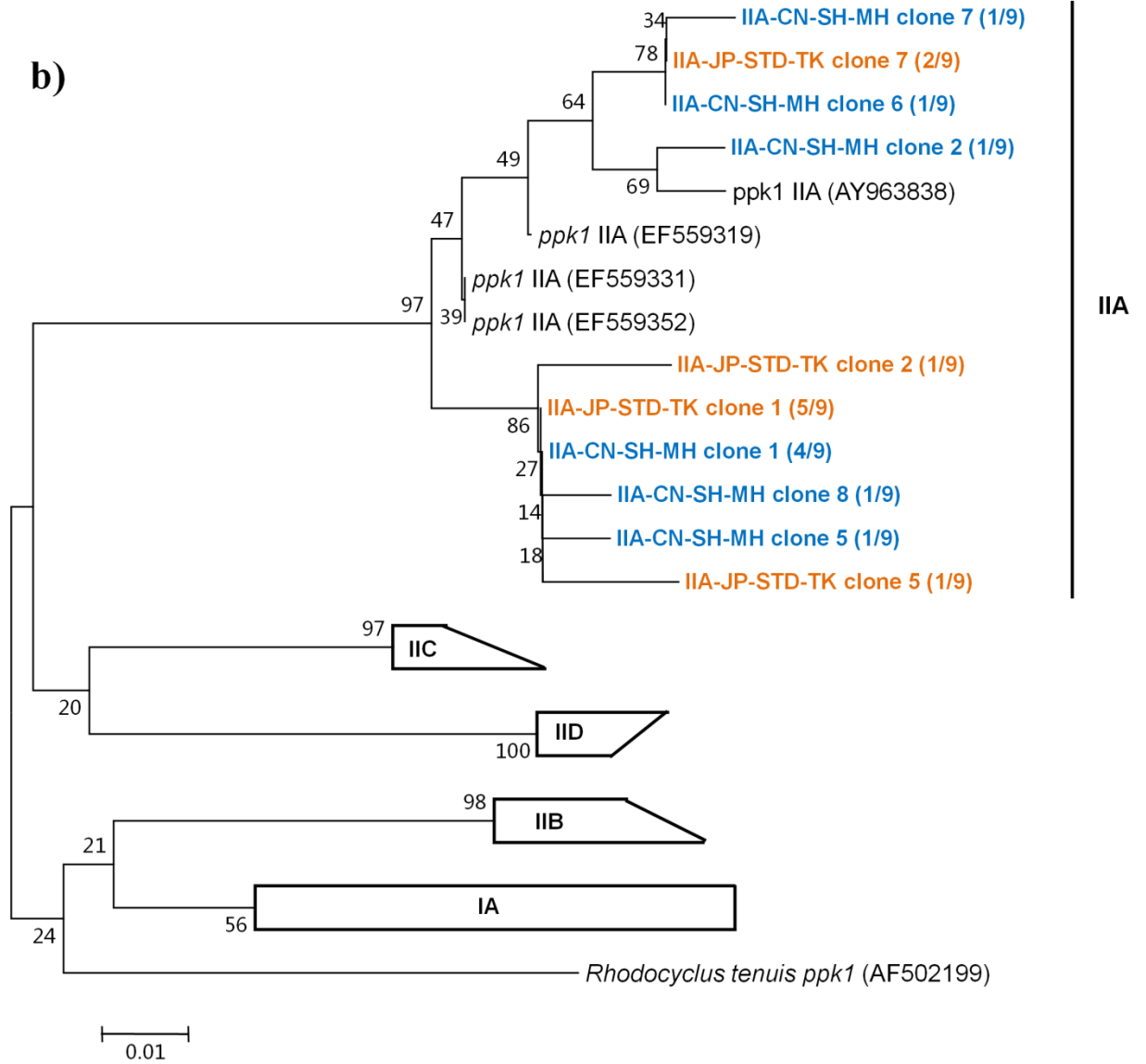
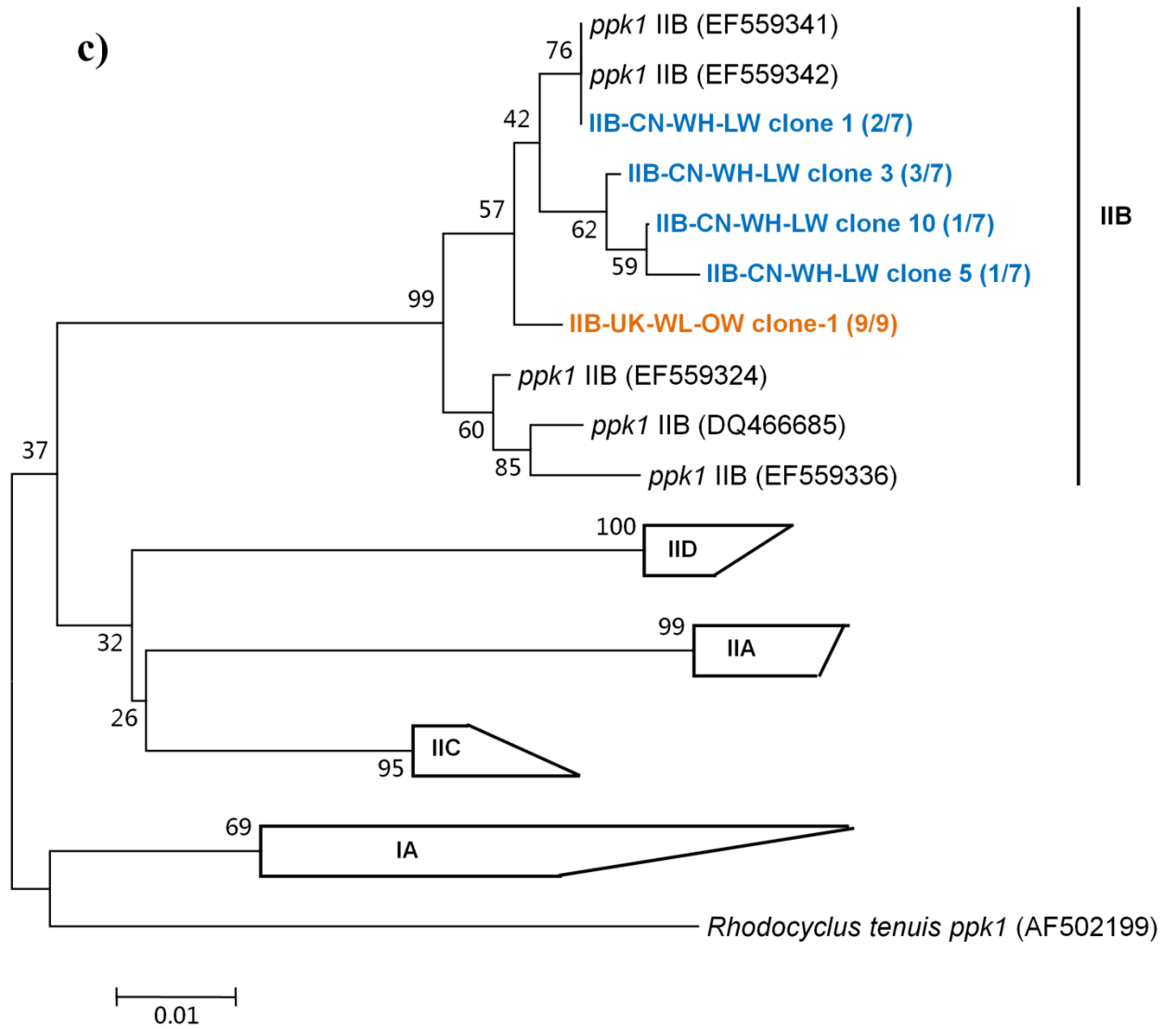


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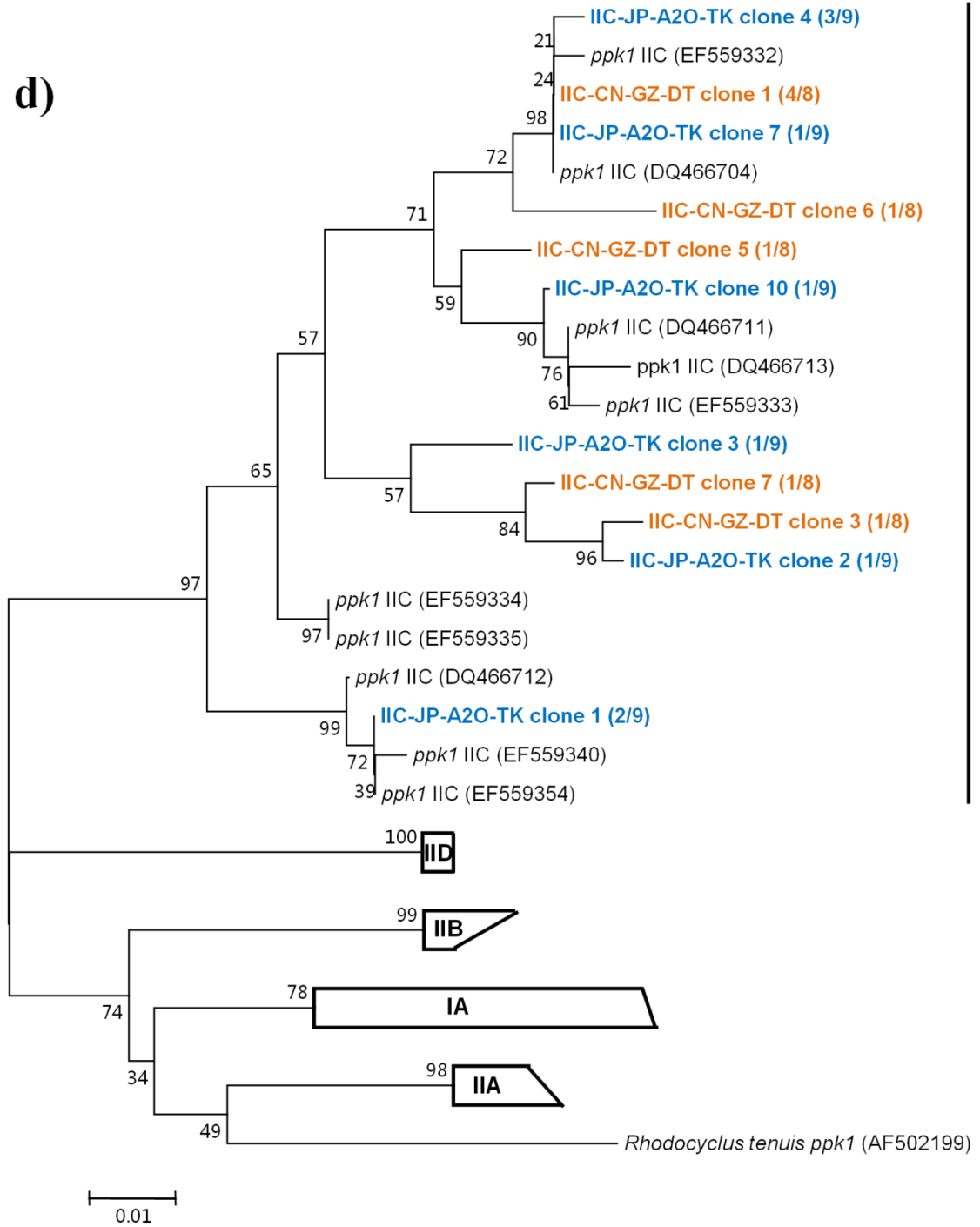


b)





d)



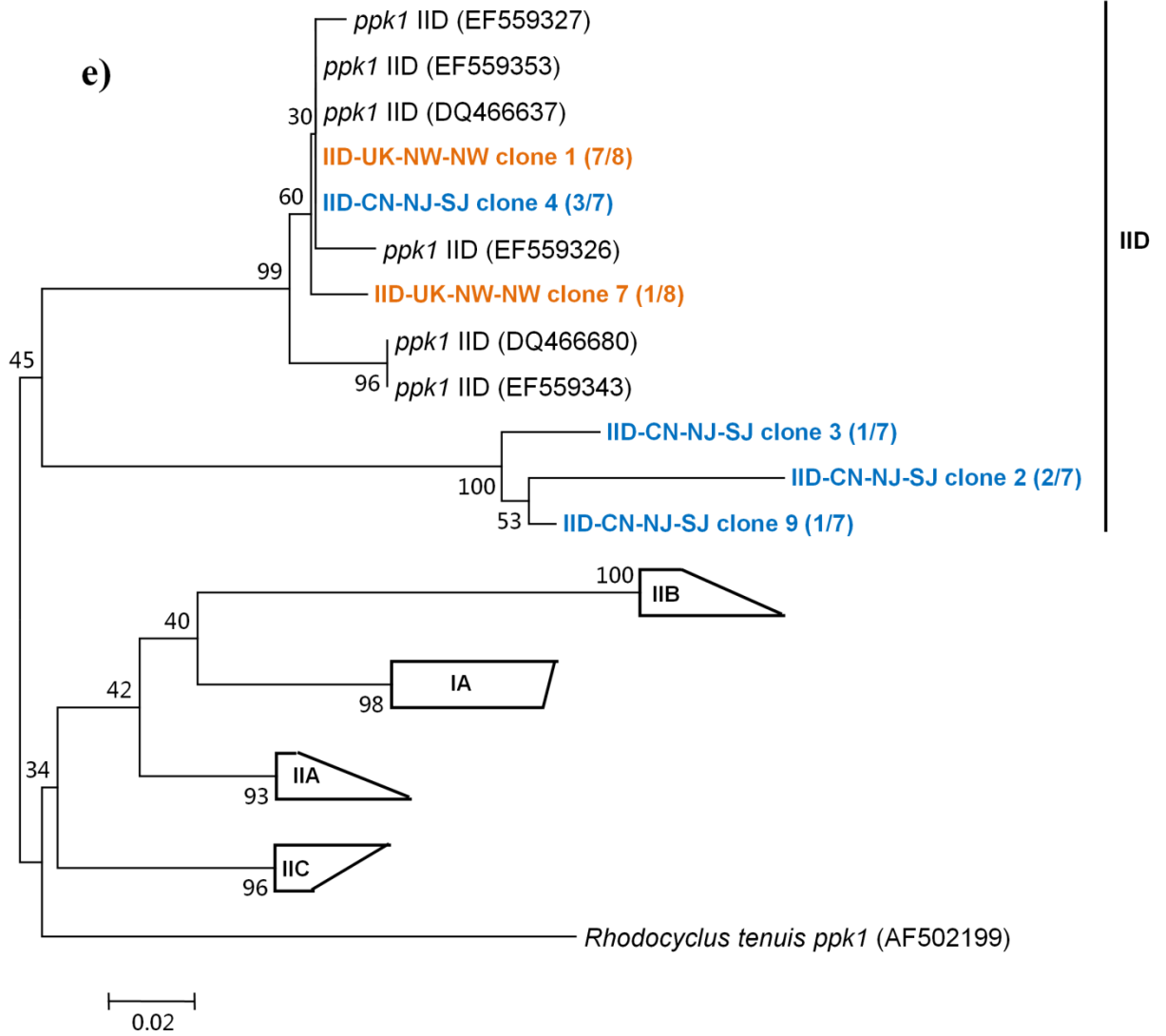


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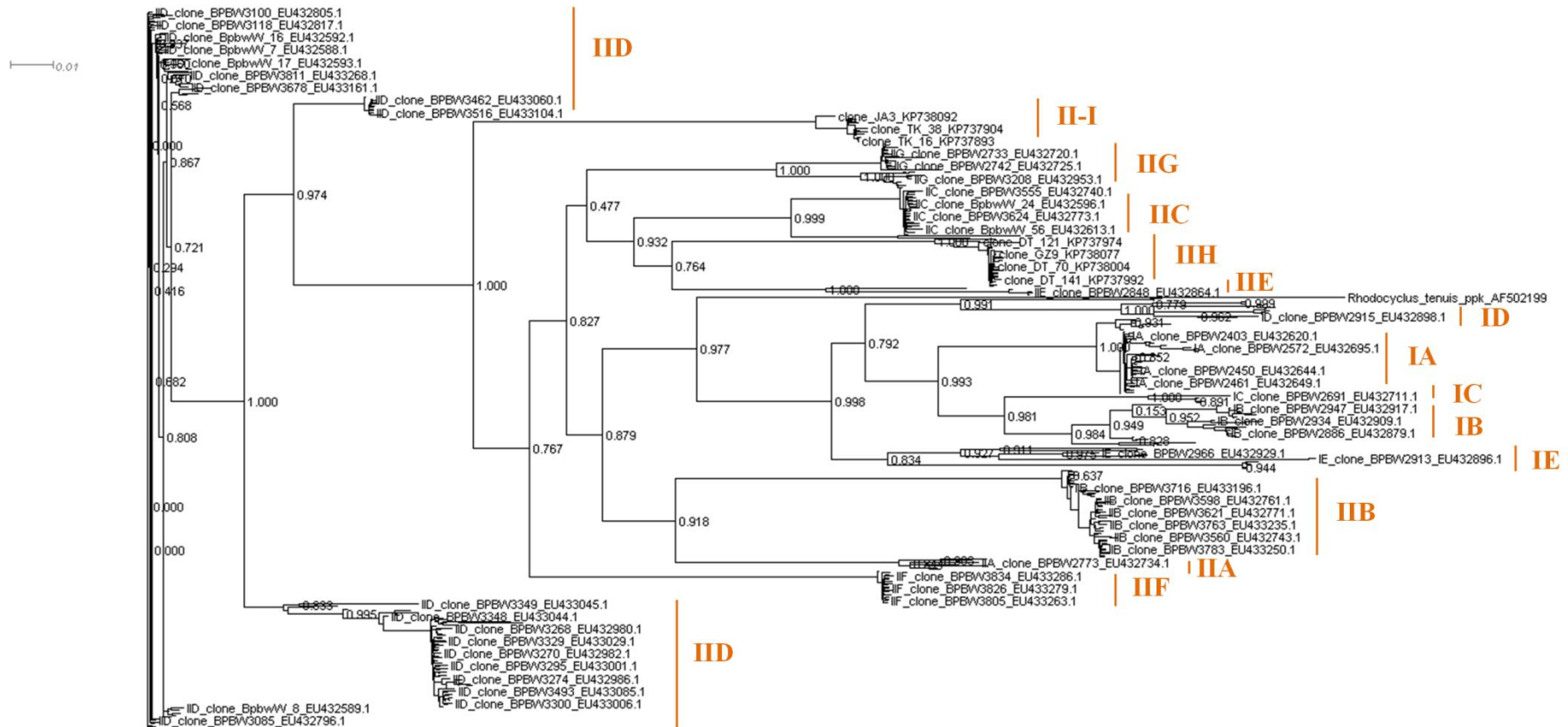
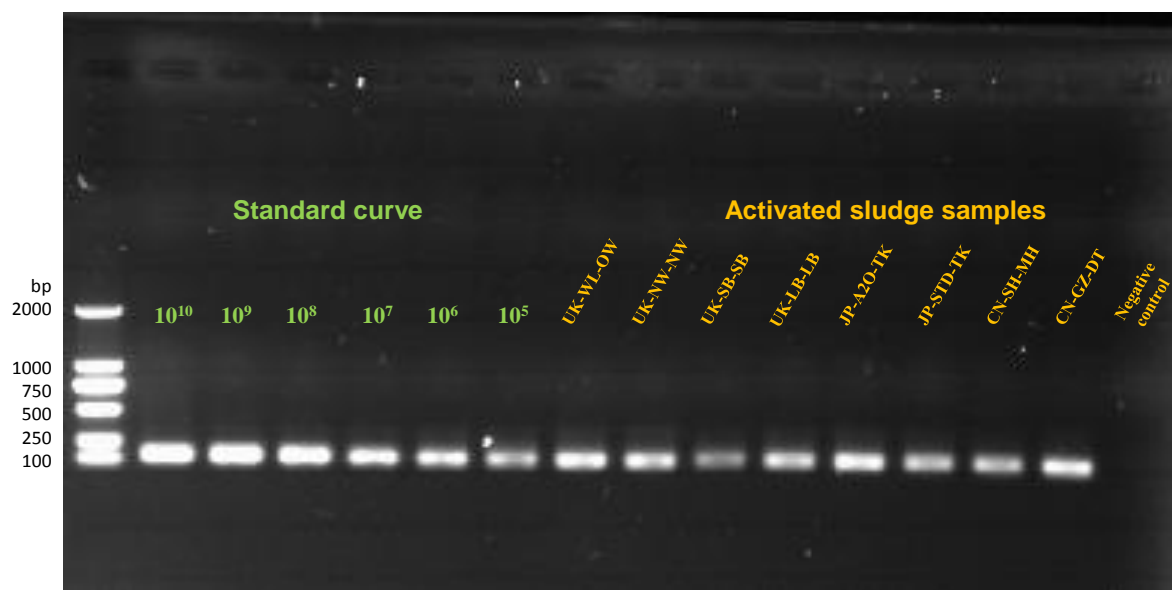


Figure S3. Maximum likelihood phylogenetic tree of *Accumulibacter ppk1* gene sequences. Over 360 of published *ppk1* nucleotide reference sequences² were downloaded from NCBI database. Sequences belonging to *Accumulibacter* clades IIH and II-I are from the clone libraries. The node labels refer to bootstrap support values.

qPCR results targeting *ppk1* genes in clade IIB



qPCR results targeting *ppk1* genes in clade IIC

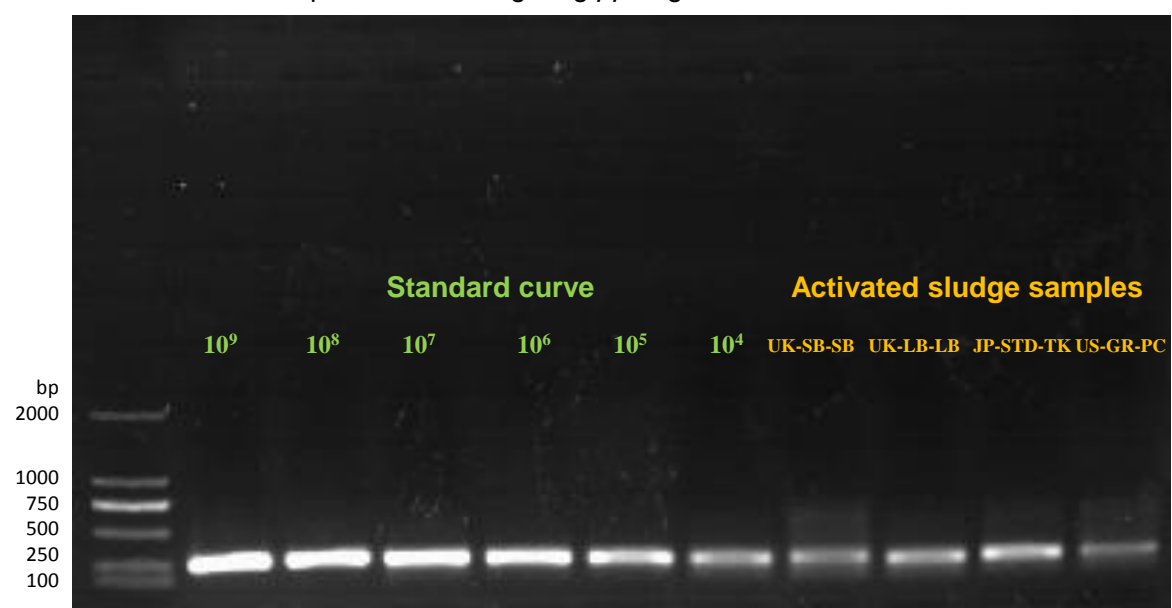


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1. He, S., Gall, D. L. & McMahon, K. D. "*Candidatus Accumulibacter*" population structure in enhanced biological phosphorus removal sludges as revealed by polyphosphate kinase genes. *Appl. Environ. Microbiol.* **73**, 5865-5874, doi:10.1128/aem.01207-07 (2007).
2. Peterson, S. B., Warnecke, F., Madejska, J., McMahon, K. D. & Hugenholtz, P. Environmental distribution and population biology of *Candidatus Accumulibacter*, a primary agent of biological phosphorus removal. *Environ. Microbiol.* **10**, 2692-2703, doi:10.1111/j.1462-2920.2008.01690.x (2008).