

**Table E1: Comparison of observational controls and active arm before PNOIT**

	Geometric Mean (95% CI) Observational controls	Geometric Mean (95% CI) Baseline in active arm	p-value
Number of multimer-positive memory cells per million B cells	3.1 (1.6,6)	4 (2.2,7.2)	0.37
Number of multimer-positive naive cells per million B cells	4.5 (2.4,8.5)	6.8 (3.9,12)	0.19
Number of multimer-positive plasmablasts per million B cells	1.5 (1,2.2)	1.3 (0.9,2)	0.60
Number of multimer-positive memory cells per million memory B cells	8.2 (4.1,16.5)	11.1 (6.1,20.5)	0.32
Number of multimer-positive naive cells per million naive B cells	8.2 (4.9,13.9)	13.1 (7.7,22.1)	0.14
Number of multimer-positive plasmablasts per million plasmablasts	11.8 (6.8,20.6)	9.7 (5.5,16.8)	0.47

**Table E2: Categorization of time in longitudinal model**

Group	N	Median (weeks)	Mean (weeks)	Minimum (weeks)	Maximum (weeks)
1	16	0	0	0	0
2	14	2	3	2	6
3	16	7	8	6	11
4	16	14	13	11	15
5	15	18	18	15	20
6	14	25	36	20	50
7	16	59	58	51	65
8	15	71	76	66	100

**Table E3: NGS sequences per sample**

<b>Patient</b>	<b>Time</b>	<b>IgA</b>	<b>IgD</b>	<b>IgE</b>	<b>IgG</b>	<b>IgM</b>	<b>Total</b>
1	2 weeks	2513	1243	666	3579	18842	26843
2	2 weeks	65241	26235	73	161465	361123	614137
3	2 weeks	93312	41716	771	133762	384236	653797
1	14 weeks	28388	14876	582	51793	250996	346635
2	14 weeks	94994	35160	809	224421	497492	852876
3	14 weeks	116127	31213	479	181498	388318	717635

**Table E4: Multimer selected sequence similarity to NGS repertoires**

		NGS repertoire multimer-similar sequences												
		Subject 1				Subject 2				Subject 3				
Subject of multimer sequence	Isotype of multimer sequences	IgA	IgD	IgG	IgM	IgA	IgD	IgE	IgG	IgM	IgA	IgD	IgG	IgM
1	IgM	1	0	1	14	4	6	0	17	49	5	2	24	52
1	IgA	15	0	4	0	0	0	0	0	0	15	0	4	0
1	IgM	0	0	0	0	0	0	0	0	0	0	2	0	0
1	IgM	0	0	0	0	0	0	0	0	5	0	0	0	0
1	IgG	0	0	0	0	1	0	0	27	5	0	0	54	16
2	IgA	1	0	0	20	18	2	0	4	25	4	0	3	22
2	IgM	1	0	0	1	0	0	0	1	18	1	0	0	3
2	IgM	0	0	0	0	0	0	0	0	4	0	0	0	0
2	IgM	0	0	0	0	0	0	0	2	8	0	0	0	1
2	IgA	0	0	0	0	0	3	0	0	2	0	0	0	0
2	IgG	0	0	0	0	0	0	1	4	0	0	0	0	0
2	IgG	0	0	0	0	0	0	0	1	0	0	0	0	0
3	IgM	0	0	0	0	0	0	0	0	0	0	0	0	0
3	IgG	0	0	0	0	0	0	0	0	0	1	0	41	2
3	IgA	0	0	0	0	2	0	0	0	0	0	0	0	0
3	IgG	0	0	0	0	0	0	0	1	0	0	0	6	1
3	IgA	0	0	0	0	21	0	0	21	2	3	0	47	1
3	IgG	0	0	0	0	0	0	0	0	1	0	0	0	0
3	IgG	0	0	0	0	0	0	0	1	0	0	0	1	0
3	IgA	0	0	0	0	0	0	0	31	1	0	0	5	0
3	IgA	0	0	0	0	0	0	0	0	0	0	0	1	0
3	IgG	0	0	0	0	16	3	0	1	1	1	0	17	1
3	IgM	0	0	0	0	0	0	0	0	7	0	0	0	0
3	IgM	0	1	0	1	0	0	0	0	1	0	1	0	1
3	IgG	0	0	0	0	2	1	0	51	2	0	0	0	0
3	IgG	0	0	0	0	0	0	0	16	3	0	0	32	0
3	IgG	0	0	0	0	6	0	0	2	0	0	0	0	0
3	IgG	0	0	0	0	0	0	0	0	0	0	0	3	0
3	IgG	0	0	0	0	0	0	0	0	0	1	0	35	0
3	IgG	0	0	0	0	0	0	0	0	0	1	0	0	0
3	IgM	0	0	0	0	0	0	0	0	1	0	0	0	0
3	IgG	0	0	0	0	0	0	0	0	0	1	0	13	0

		NGS repertoire multimer-similar sequences												
		Subject 1				Subject 1					Subject 1			
Subject of multimer sequence	Isotype of multimer sequences	IgA	IgD	IgG	IgM	IgA	IgD	IgE	IgG	IgM	IgA	IgD	IgG	IgM
3	IgG	0	0	0	0	0	1	0	6	0	0	0	0	0
3	IgM	1	0	0	17	0	1	0	0	4	1	0	0	19
3	IgA	0	0	0	0	1	0	0	19	3	3	1	17	2
3	IgA	0	0	0	0	9	0	0	49	0	18	0	25	0
3	IgG	0	0	0	0	0	0	0	7	0	0	0	1	0
3	IgA	0	0	0	0	3	0	0	2	0	0	0	11	2
3	IgG	0	0	0	0	0	0	0	3	0	0	0	1	0
3	IgG	1	0	1	5	0	0	0	0	2	1	1	1	8
3	IgA	0	0	0	0	0	1	0	9	0	3	0	12	0
3	IgG	0	0	0	0	1	0	0	0	0	0	0	0	0
3	IgM	1	4	0	10	0	1	0	0	19	1	6	0	13
3	IgM	0	0	0	1	0	2	0	6	15	5	2	0	4
3	IgM	0	0	0	0	0	0	0	0	7	0	1	1	9
3	IgA	0	0	0	0	0	0	0	15	0	0	0	0	0

1 **Table E1: Comparison of observational controls and active arm before**  
2 **PNOIT**

3 The frequency of multimer-positive B cells both within each B cell subset and  
4 within the entire B cell population were compared between subjects in the  
5 observational arm and the active arm before onset of PNOIT. No significant  
6 differences are found.

7

8 **Table E2: Categorization of time in longitudinal model**

9 Time period groups used in the longitudinal analysis are noted here with the  
10 number of observations (N) as well as the median, mean, minimum, and  
11 maximum number of weeks characterizing each time period.

12

13 **Table E3: NGS sequences per sample**

14 Within each of the six samples, with 2 time points during PNOIT of each of the  
15 three patients, the number of IgH sequences within each isotype are shown.

16

17 **Table E4: Multimer selected sequence similarity to NGS repertoires**

18 Each individual multimer-positive sequence with similar sequences in the NGS  
19 repertoire are listed, with both their originating subject and isotype, in each row.

20 Columns reflect the NGS repertoires from different subjects by isotype

21 compartment.

22 **Figure E1: Frequency of multimer+ B cell subsets in allergic and non-**  
23 **allergic subjects**

24 The frequency of multimer-positive cells was normalized to the subset (naïve,  
25 memory, or plasmablast) population and described as the number of cells per  
26 million subset cells or per million CD19<sup>+</sup> B cells in either peanut allergic patients  
27 from the PNOIT trial before the start of PNOIT compared to non-allergic control  
28 patients. Basophil staining of the non-allergic control patients is shown in  
29 Figure 1c.

30

31 **Figure E2: Single cell immunoglobulin amplification**

32 Immunoglobulin amplification of heavy and light chains was performed using  
33 single cell, nested RT-PCR. In this example, gel electrophoresis demonstrates  
34 recovery of 17 out of 24 paired heavy and light chains.

35

36 **Figure E3: Antibody cloning vectors**

37 Recombinant antibodies were produced after insertion of the multimer-positive  
38 single cell amplified sequences into heavy or light (either kappa or lambda,  
39 respectively) vectors, as previously published in Tiller, et.al.

40

41 **Figure E4: NGS analysis pipeline**

42 The next generation sequencing (NGS) analysis pipeline for processing of raw  
43 Illumina MiSeq 2x250 reads is used to generate the heavy chain immunoglobulin  
44 sequences used for downstream analysis.

45

46 **Figure E5: Representative results from NGS data analysis**

47 The numbers of reads retained at each step of the analysis pipeline for one of the  
48 subjects' samples is shown.

49 **Figure E6: Ara h 2 specific heavy chain clonal groups**

50 The distribution of clonal groups (same V-J gene usage and similar CDR3)  
51 demonstrates that a subset (20%) have more than one sequence.

52

53 **Figure E7: Inference of antigen selection of Ara h 2-positive B cell IgH**

54 Heavy chain immunoglobulin sequences of Ara h 2 affinity selected B cells have  
55 increased frequency of CDR replacement mutations ( $R_{\text{CDR}}$ ) compared to the total  
56 number of mutations in the V region ( $M_{\text{V}}$ ), suggestive of antigen selection. The  
57 gray area represents the 90-95% confidence interval for the probability of random  
58 mutations.

59

60 **Figure E8: Recombinant antibody affinity for Ara h 2**

61 Ara h 2 affinity of the recombinant IgG1 antibodies produced from multimer-  
62 positive single cells was characterized using biolayer interferometry (Octet). The  
63 colors of the bars reflect the original multimer-positive single cell isotype from  
64 which the recombinant antibody was produced.



Figure E1

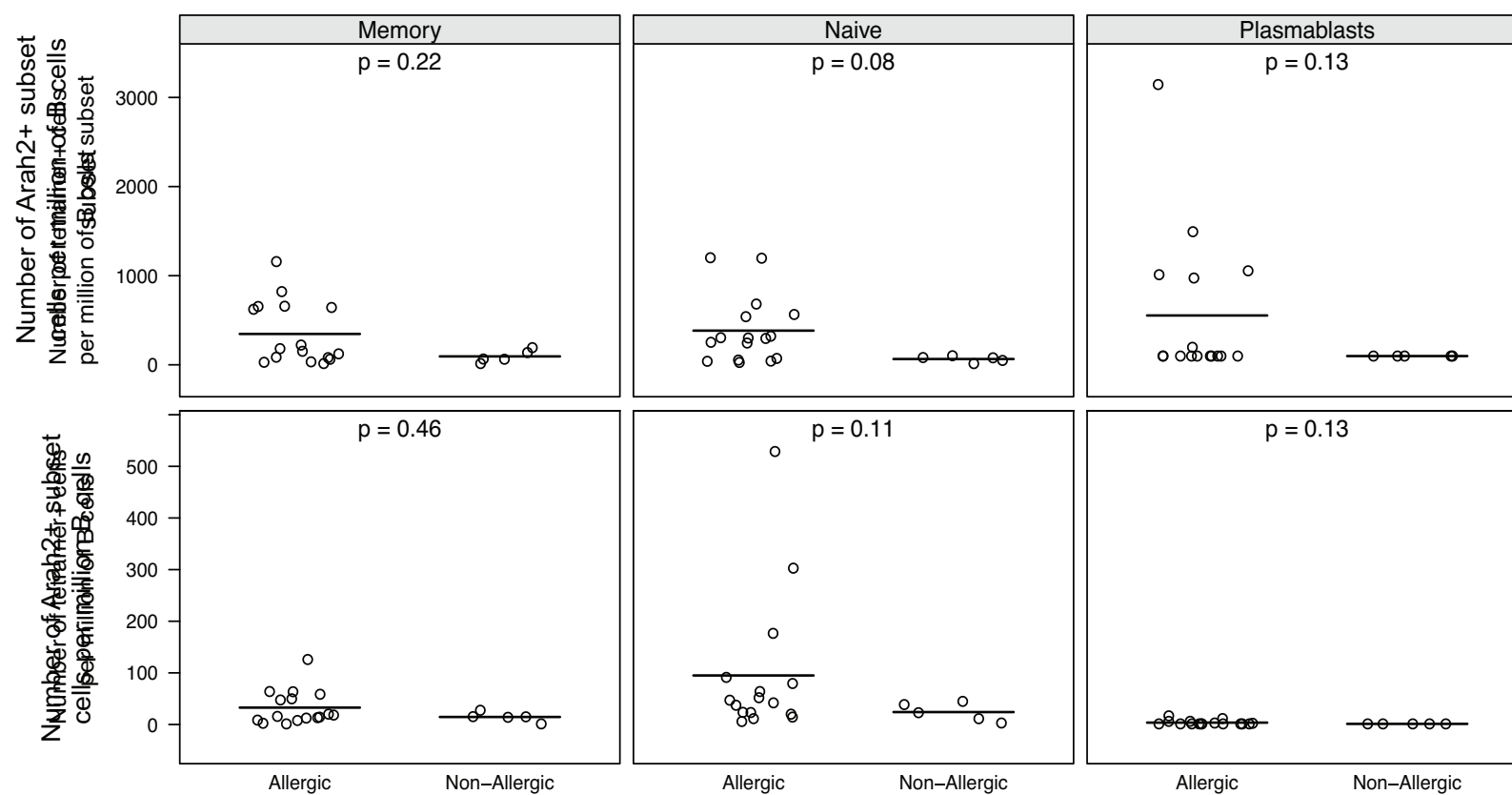


Figure E2

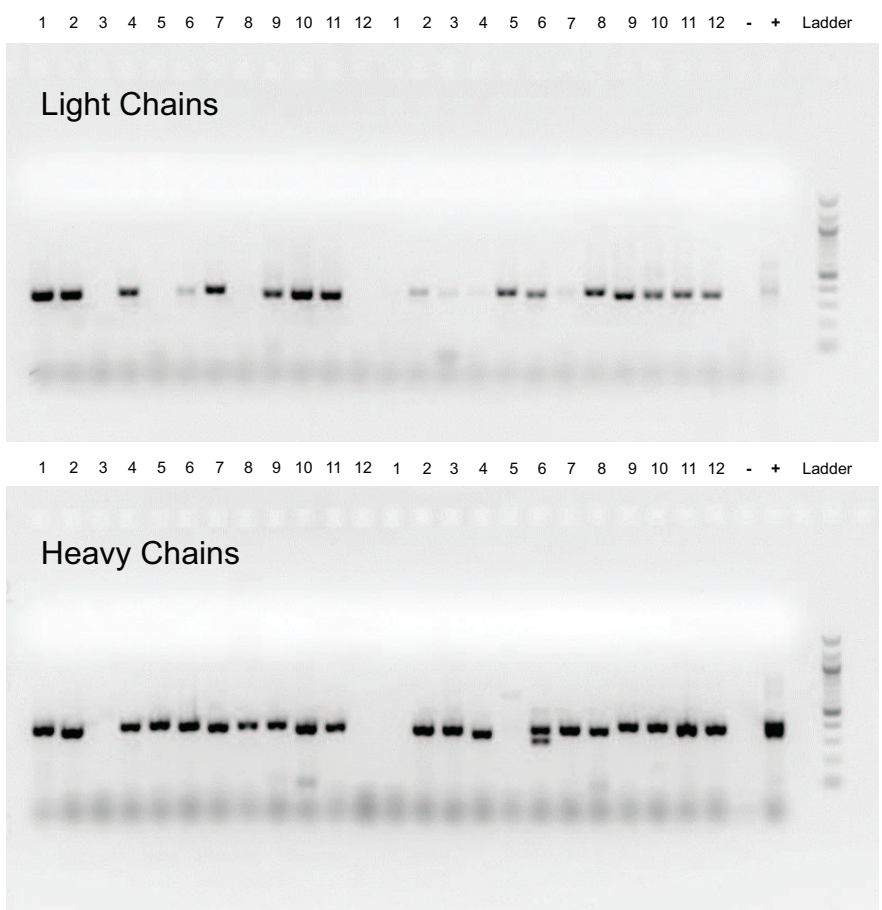


Figure E3

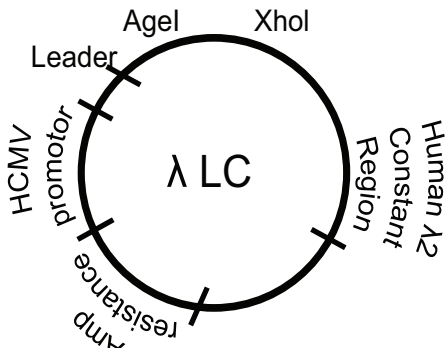
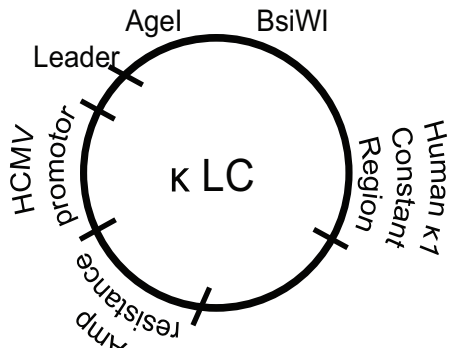
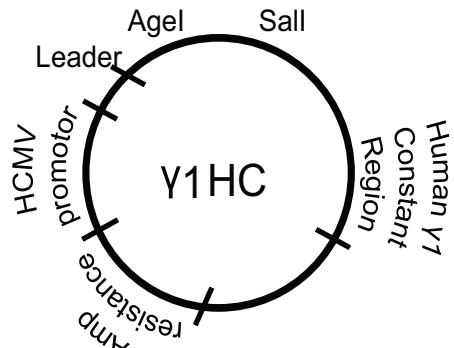


Figure E4

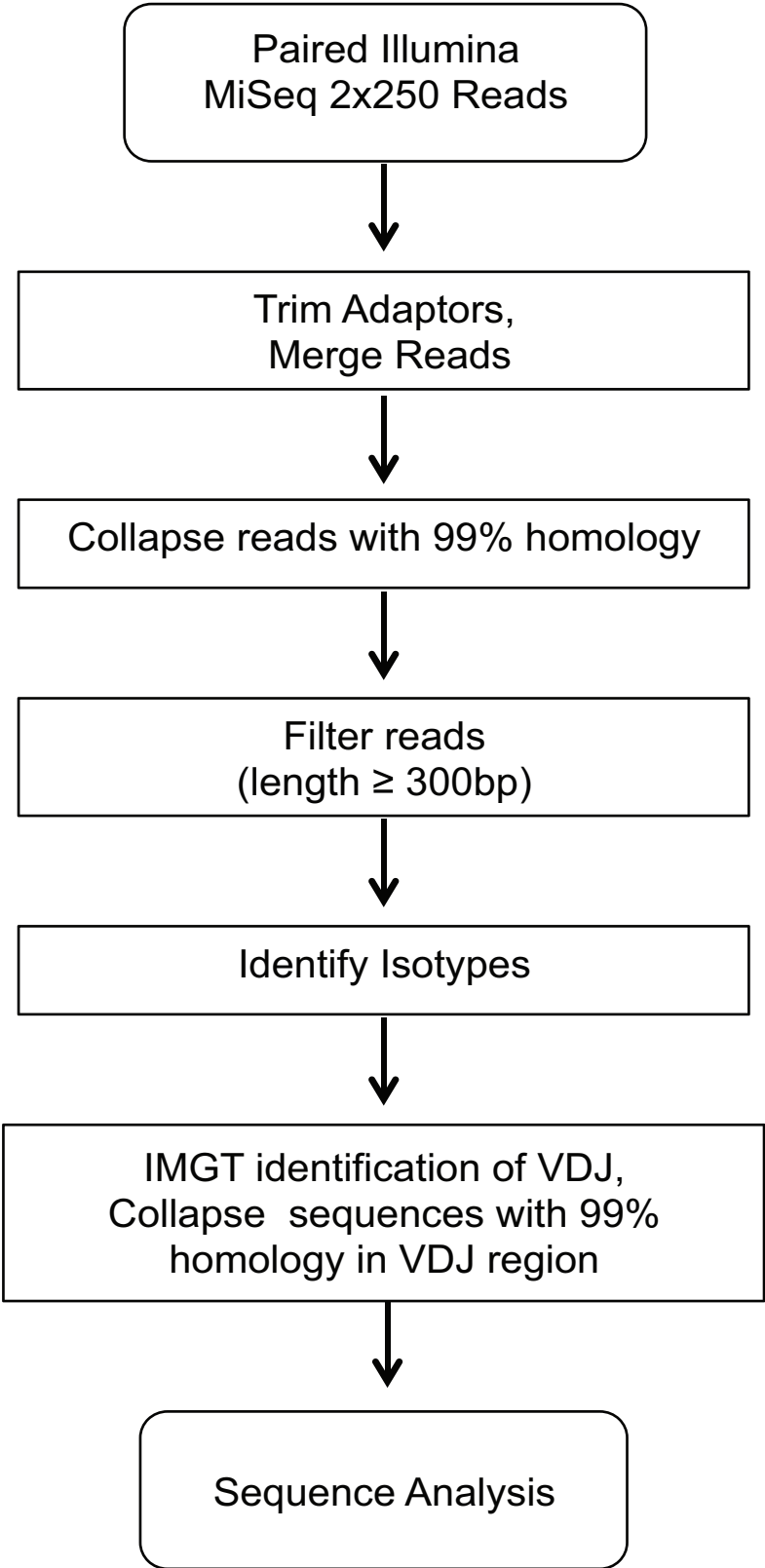


Figure E5

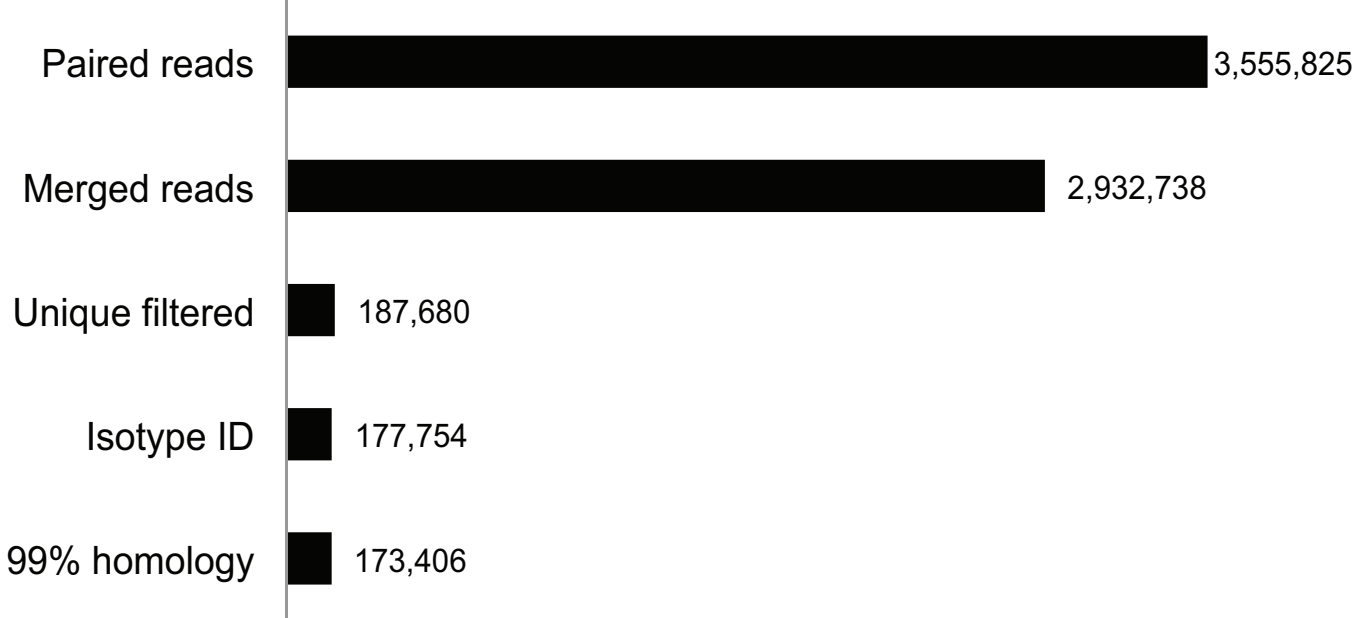


Figure E6

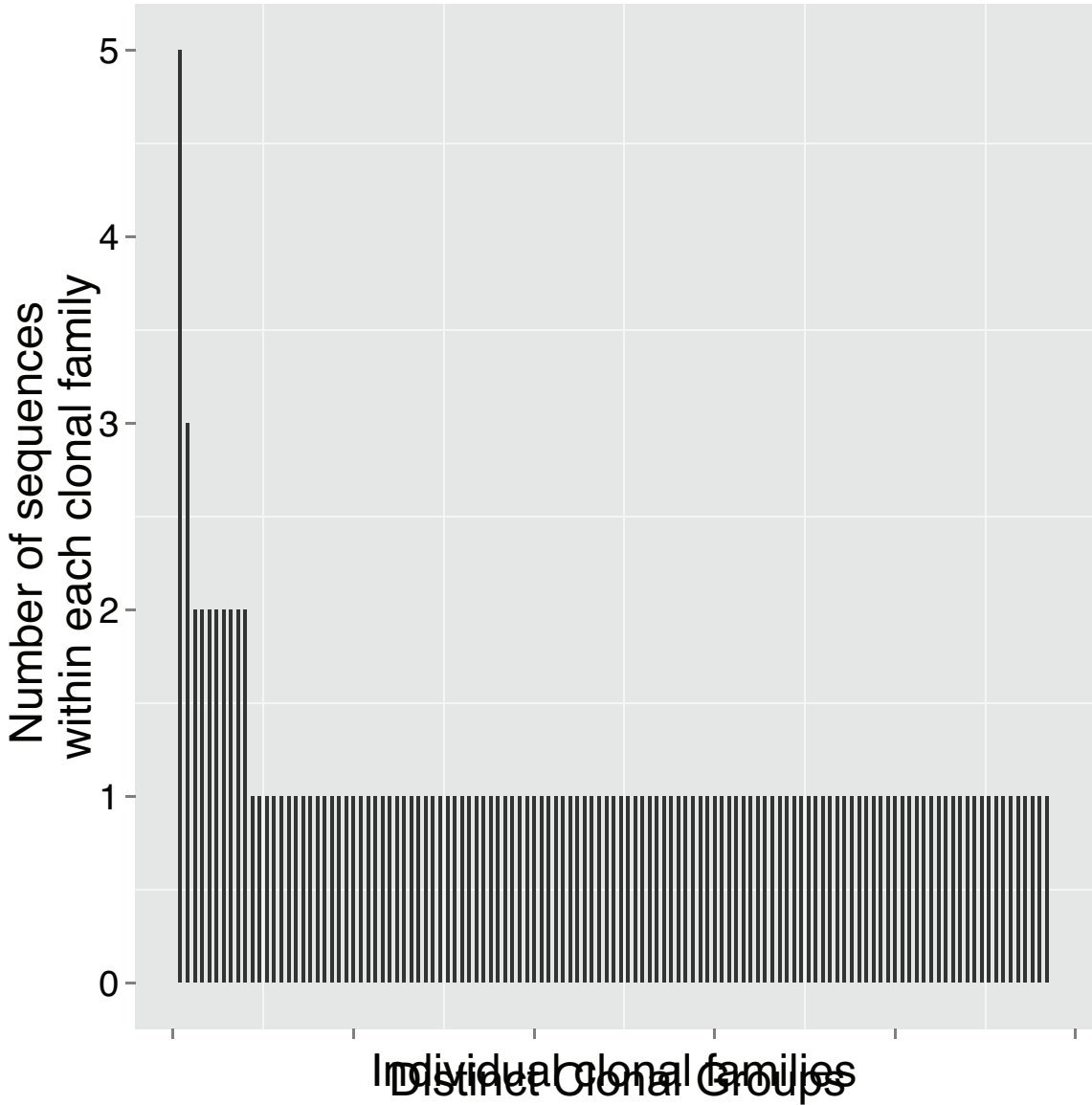


Figure E7

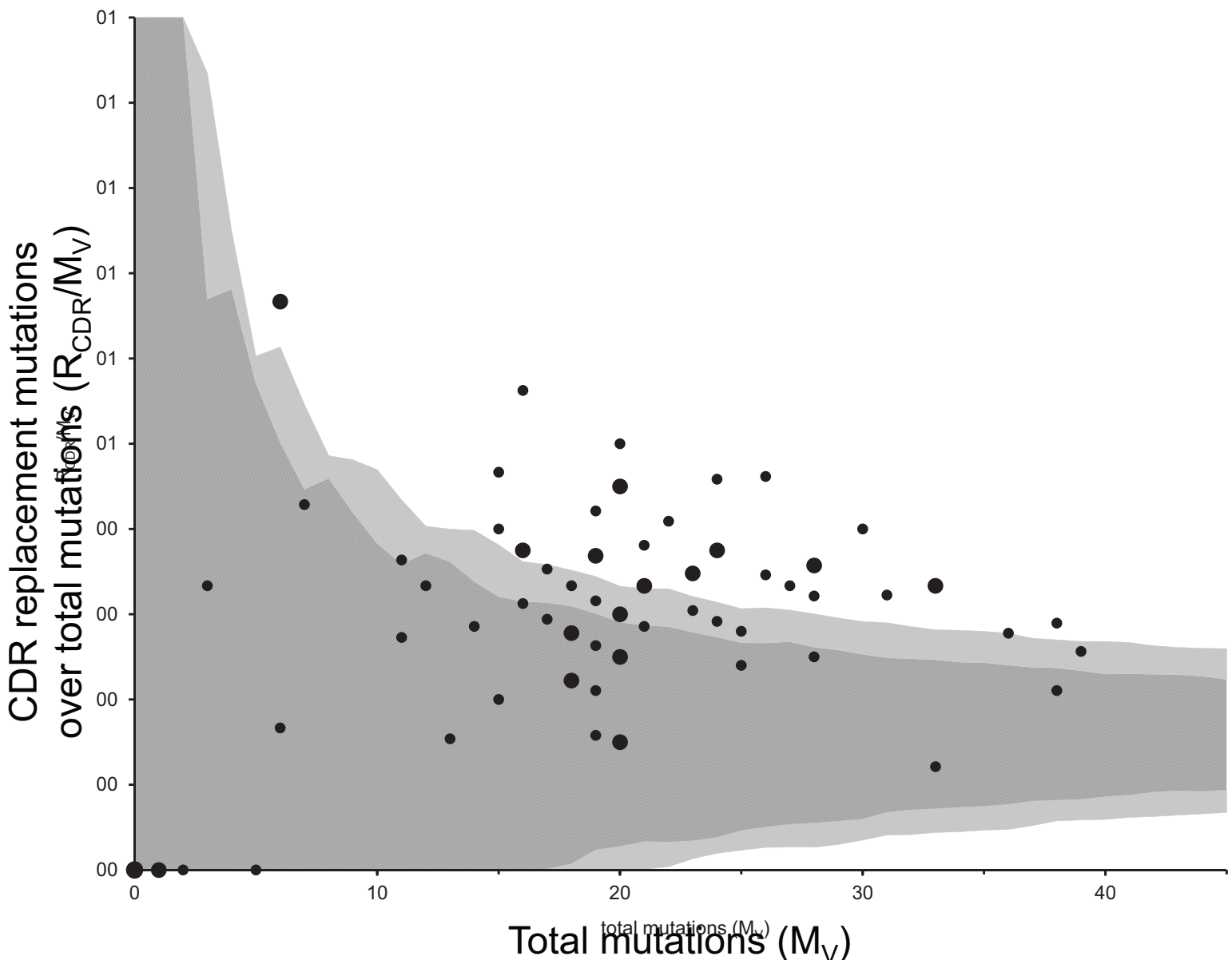


Figure E8

