#### SUPPLEMENTARY INFORMATION

# Preliminary Characterization and In Vivo Studies of Structurally Identical <sup>18</sup>F- and <sup>125</sup>I-Labeled Benzyloxybenzenes for PET/SPECT Imaging of $\beta$ -Amyloid Plaques

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### **Chemical Synthesis**

#### 1-(Benzyloxy)-4-(2-fluoroethoxy)benzene (5a)

A mixture of 4-(benzyloxy)phenol (2.00 g, 10.0 mmol) and KOH (0.56 g, 10.0 mmol) in dry EtOH (30 mL) was stirred under reflux for 30 min. 1-bromo-2-fluoroethane (1.52 g, 12.0mmol) was then added dropwise, and the mixture was further stirred for 1 h, and evaporated under a vacuum. A white precipitate was formed by adding 50 mL of 1M NaOH, which was then filtered, washed with 50 mL water and recrystallized from methanol to obtain a white solid of **5a** (2.23 g, 90.4%). mp: 69.1-69.6 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.46 – 7.28 (m, 5H), 6.91 (d, *J* = 9.2 Hz, 2H), 6.86 (d, *J* = 9.3 Hz, 2H), 5.02 (s, 2H), 4.81 – 4.64 (m, 2H), 4.22 – 4.10 (m, 2H); MS (EI): m/z calcd for C<sub>15</sub>H<sub>15</sub>FO<sub>2</sub> 246; found 246 M<sup>+</sup>.

#### 2-(4-(Benzyloxy)phenoxy)ethanol (5b)

The procedure described above for the preparation of **5a** was employed to obtain a white solid of **5b** from 4-(benzyloxy)phenol and 2-chloroethanol (2.35 g, 48.1%). mp: 105.8-106.4 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.46 – 7.30 (m, 5H), 6.91 (d, *J* = 9.1 Hz, 2H), 6.85 (d, *J* = 9.0 Hz, 2H), 5.02 (s, 2H), 4.08 – 3.99 (m, 2H), 3.97 – 3.87 (m, 2H), 1.95 (s, 1H); MS (EI): m/z calcd for C<sub>15</sub>H<sub>16</sub>O<sub>3</sub> 244; found 244 M<sup>+</sup>.

#### 4-(2-Fluoroethoxy)phenol (6a)

To a solution of **5a** (2.08 g, 8.44 mmol) in anhydrous MeOH (10 mL) was added 10% Pd/C (89.4 mg, 0.84 mmol). The mixture was stirred for 4 h at 50 °C under 1 atm of hydrogen atmosphere. The catalyst was filtered while hot and washed with MeOH, and the filtrate was concentrated under reduced pressure to give a white solid of **6a** (1.32 g, 57.4%). mp: 95.7-96.5 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.83 (d, *J* = 8.9 Hz, 2H), 6.77 (d, *J* = 9.0 Hz,

2H), 4.80 - 4.64 (m, 2H), 4.42 (s, 1H), 4.23 - 4.09 (m, 2H); MS (EI): m/z calcd for C<sub>8</sub>H<sub>9</sub>FO<sub>2</sub> 156; found 156 M<sup>+</sup>.

#### 4-(2-Hydroxyethoxy)phenol (6b)

The procedure described above for the preparation of **6a** was employed to obtain a white solid of **6b** from **5b** (1.35 g, 100%). mp: 92.1-92.8 °C; <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ )  $\delta$  8.86 (s, 1H), 6.74 (d, J = 8.6 Hz, 2H), 6.66 (d, J = 8.8 Hz, 2H), 4.78 (t, J = 5.2 Hz, 1H), 3.90 – 3.80 (m, 2H), 3.70 – 3.61 (m, 2H); MS (EI): m/z calcd for C<sub>8</sub>H<sub>10</sub>O<sub>3</sub> 154; found 154 M<sup>+</sup>.

#### 1-(2-Fluoroethoxy)-4-((4-iodobenzyl)oxy)benzene (7a)

To a solution of **6a** (468.5 mg, 3.0 mmol) and 1-(bromomethyl)-4-iodobenzene (890.8 mg, 3.0 mmol) in anhydrous DMF (5 mL), K<sub>2</sub>CO<sub>3</sub> was added (414.6 mg, 3.0 mmol). The resulting mixture was stirred at 90 °C for 30 min. After cooling to room temperature, a white precipitate was formed by adding 50 mL of water, which was then filtered, washed with 50 mL of water and recrystallized from methanol to obtain a white solid of **7a** (986.8 mg, 88.4%). mp: 108.3-108.8 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.70 (d, *J* = 8.3 Hz, 2H), 7.16 (d, *J* = 8.2 Hz, 2H), 6.90 – 6.84 (m, 4H), 4.96 (s, 2H), 4.79 – 4.65 (m, 2H), 4.21 – 4.11 (m, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  153.11 (C<sub>q</sub>), 152.93 (C<sub>q</sub>), 137.62 (2 × CH), 136.94 (C<sub>q</sub>), 129.24 (2 × CH), 115.89 (2 × CH), 115.79 (2 × CH), 93.35 (C<sub>q</sub>), 82.01 (d, *J* = 170.6 Hz, CH<sub>2</sub>), 69.96 (CH<sub>2</sub>), 67.87 (d, *J* = 20.5 Hz, CH<sub>2</sub>); HRMS (EI): m/z calcd for C<sub>15</sub>H<sub>14</sub>FIO<sub>2</sub> 372.0023; found 372.0029 M<sup>+</sup>.

#### 2-(4-((4-Iodobenzyl)oxy)phenoxy)ethanol (7b)

The procedure described above for the preparation of **7a** was employed to obtain a white solid of **7b** from **6b** and 1-(bromomethyl)-4-iodobenzene (1.38 g, 72.9%). mp: 129.3-129.9

°C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.70 (d, *J* = 8.3 Hz, 2H), 7.17 (d, *J* = 8.4 Hz, 2H), 6.89 – 6.83 (m, 4H), 4.96 (s, 2H), 4.06 – 4.00 (m, 2H), 3.96 – 3.91 (m, 2H); MS (EI): m/z calcd for C<sub>15</sub>H<sub>15</sub>IO<sub>3</sub> 370; found 370 M<sup>+</sup>.

#### 1-Bromo-4-((4-(2-fluoroethoxy)phenoxy)methyl)benzene (7c)

The procedure described above for the preparation of **7a** was employed to obtain a white solid of **7c** from **6a** and 1-bromo-4-(bromomethyl)benzene (1.55 g, 95.2%). mp: 115.6-116.4  $^{\circ}$ C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.50 (d, *J* = 8.4 Hz, 2H), 7.29 (d, *J* = 8.4 Hz, 2H), 6.90 – 6.84 (m, 4H), 4.97 (s, 2H), 4.80 – 4.65 (m, 2H), 4.21 – 4.11 (m, 2H); MS (EI): m/z calcd for C<sub>15</sub>H<sub>14</sub>BrFO<sub>2</sub> 324; found 324 M<sup>+</sup>.

#### 2-(4-((4-Iodobenzyl)oxy)phenoxy)ethyl 4-methylbenzenesulfonate (8a)

A mixture of **7b** (740.4 mg, 2.0 mmol) and Et<sub>3</sub>N (10 mL) was stirred in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (10 mL) in an ice bath, and tosyl chloride (571.9 mg, 3.0 mmol) was slowly added. The reaction mixture was stirred for 4 h at room temperature, and the solvent was evaporated under reduced pressure. Water was added (50 mL), and the mixture was extracted by CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). Combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under a vacuum. The crude mixture was purified by silica gel chromatography (petroleum ether/AcOEt = 4/1, v/v) to obtain a white solid of **7a** (425.7 mg, 40.6%). mp: 120.8-121.5 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.81 (d, *J* = 8.3 Hz, 2H), 7.70 (d, *J* = 8.3 Hz, 2H), 7.33 (d, *J* = 8.1 Hz, 2H), 7.15 (d, *J* = 8.3 Hz, 2H), 6.83 (d, *J* = 9.1 Hz, 2H), 6.72 (d, *J* = 9.1 Hz, 2H), 4.94 (s, 2H), 4.36 – 4.31 (m, 2H), 4.12 – 4.07 (m, 2H), 2.44 (s, 3H); MS (EI): m/z calcd for C<sub>22</sub>H<sub>21</sub>IO<sub>5</sub>S 524; found 524 M<sup>+</sup>.

#### Tributyl(4-((4-(2-fluoroethoxy)phenoxy)methyl)phenyl)stannane (8b)

A mixture of **7c** (650.3 mg, 2.0 mmol), (Bu<sub>3</sub>Sn)<sub>2</sub> (2.32 g, 4.0 mmol), (Ph<sub>3</sub>P)<sub>4</sub>Pd (231.7 mg, 0.2 mmol) and Et<sub>3</sub>N (1 mL) in toluene (10 mL) was stirred under reflux overnight. The mixture was concentrated under reduced pressure and purified by silica gel chromatography (petroleum ether/AcOEt = 15/1, v/v) to give a colorless oil of **8b** (325.6 mg, 30.4%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.54 – 7.41 (m, 2H), 7.37 (d, *J* = 7.8 Hz, 2H), 6.92 (d, *J* = 9.3 Hz, 2H), 6.87 (d, *J* = 9.3 Hz, 2H), 4.99 (s, 2H), 4.80 – 4.65 (m, 2H), 4.22 – 4.11 (m, 2H), 1.58 – 1.50 (m, 6H), 1.39 – 1.27 (m, 6H), 1.14 – 0.96 (m, 6H), 0.88 (t, *J* = 7.3 Hz, 9H); MS (EI): m/z calcd for C<sub>27</sub>H<sub>41</sub>FO<sub>2</sub>Sn 536; found 536 M<sup>+</sup>.

#### 4-(2-Fluoroethoxy)benzaldehyde (9a)

To a solution of 4-hydroxybenzaldehyde (2.44 g, 20 mmol) and 1-bromo-2-fluoroethane (2.54 g, 20 mmol) in anhydrous DMF (5 mL), K<sub>2</sub>CO<sub>3</sub> (5.53 g, 40 mmol) was added. The resulting mixture was stirred at 90 °C for 2h, and the solvent was evaporated under reduced pressure. 50 mL of water was added, and the mixture was extracted by CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). Combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under a vacuum. The crude mixture was purified by silica gel chromatography (petroleum ether/AcOEt = 4/1, v/v) to give a white solid of **9a** (2.95 g, 87.8%). mp: 53.3-54.5 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.90 (s, 1H), 7.86 (d, *J* = 8.7 Hz, 2H), 7.04 (d, *J* = 8.6 Hz, 2H), 4.88 – 4.71 (m, 2H), 4.37 – 4.24 (m, 2H); MS (EI): m/z calcd for C<sub>9</sub>H<sub>9</sub>FO<sub>2</sub> 168; found 168 M<sup>+</sup>.

#### 4-(2-Bromoethoxy)benzaldehyde (9b)

The procedure described above for the preparation of **9a** was employed to obtain a white solid of **9b** from 4-hydroxybenzaldehyde and 1,2-dibromoethane (1.32 g, 28.9%). mp:

51.2-51.7 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.90 (s, 1H), 7.85 (d, *J* = 8.8 Hz, 2H), 7.02 (d, *J* = 8.7 Hz, 2H), 4.38 (t, *J* = 6.2 Hz, 2H), 3.67 (t, *J* = 6.2 Hz, 2H).

#### (4-(2-Fluoroethoxy)phenyl)methanol (10a)

To a stirring mixture of **9a** (2.95 g, 17.6 mmol) in anhydrous MeOH (10 mL) in ice bath, NaBH<sub>4</sub> (1.33 g, 35.2 mmol) was slowly added. The reaction mixture was stirred for 30 min at 0 °C and 10 mL of water was added to quench the reaction. MeOH was evaporated under reduced pressure and the mixture was neutralized with 1 M HCl and then extracted by CH<sub>2</sub>Cl<sub>2</sub> (3 × 10 mL). Combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure to give a yellow oil of **6a** (2.73 g, 91.1%). mp: 50.6-51.5 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.31 (d, *J* = 8.5 Hz, 2H), 6.93 (d, *J* = 8.5 Hz, 2H), 4.84 – 4.68 (m, 2H), 4.63 (s, 2H), 4.27 – 4.17 (m, 2H); MS (EI): m/z calcd for C<sub>9</sub>H<sub>11</sub>FO<sub>2</sub> 170; found 170 M<sup>+</sup>.

#### (4-(2-Bromoethoxy)phenyl)methanol (10b)

The procedure described above for the preparation of **10a** was employed to obtain a white solid of **10b** from **9b** (1.14 g, 95.8%). mp: 88.7-89.6 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.30 (d, *J* = 8.5 Hz, 2H), 6.91 (d, *J* = 8.6 Hz, 2H), 4.63 (s, 2H), 4.30 (t, *J* = 6.3 Hz, 2H), 3.64 (t, *J* = 6.3 Hz, 2H); MS (EI): m/z calcd for C<sub>9</sub>H<sub>11</sub>BrO<sub>2</sub> 230; found 230 M<sup>+</sup>.

#### 1-(Bromomethyl)-4-(2-fluoroethoxy)benzene (11a)

To a stirring solution of **10a** (2.73 g, 16.0 mmol) in anhydrous  $CH_2Cl_2$  (25 mL) at 0 °C, phosphorus tribromide (4.33 g, 16.0 mmol) was added dropwise. The resulting mixture was stirred at room temperature for 30 min and the reaction was quenched by addition of 20 mL of water. The mixture was neutralized with NaHCO<sub>3</sub> and then extracted by  $CH_2Cl_2$  (3 × 10 mL). Combined organic layers were dried over anhydrous MgSO<sub>4</sub>, filtered and concentrated under reduced pressure to give a colorless oil of **11a** (3.54 g, 95.0%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33 (d, *J* = 8.6 Hz, 2H), 6.89 (d, *J* = 8.6 Hz, 2H), 4.84 – 4.68 (m, 2H), 4.50 (s, 2H), 4.27 – 4.16 (m, 2H); MS (EI): m/z calcd for C<sub>9</sub>H<sub>11</sub>BrFO 232; found 232 M<sup>+</sup>.

#### 1-(2-Bromoethoxy)-4-(bromomethyl)benzene (11b)

The procedure described above for the preparation of **11a** was employed to obtain a white solid of **11b** from **10b** (1.29 g, 99.0%). mp: 53.9-54.7 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33 (d, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.7 Hz, 2H), 4.49 (s, 2H), 4.29 (t, *J* = 6.3 Hz, 2H), 3.63 (t, *J* = 6.3 Hz, 2H); MS (EI): m/z calcd for C<sub>9</sub>H<sub>10</sub>Br<sub>2</sub>O 292; found 292 M<sup>+</sup>.

#### 1-(2-Fluoroethoxy)-4-((4-iodophenoxy)methyl)benzene (12a)

The procedure described above for the preparation of **7a** was employed to obtain a white solid of **12a** from 4-iodophenol and **11a** (623.4 mg, 94.1%). mp: 127.6-128.2 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.55 (d, *J* = 8.9 Hz, 2H), 7.34 (d, *J* = 8.6 Hz, 2H), 6.94 (d, *J* = 8.6 Hz, 2H), 6.74 (d, *J* = 8.9 Hz, 2H), 4.95 (s, 2H), 4.83 – 4.68 (m, 2H), 4.27 – 4.16 (m, 2H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)  $\delta$  158.65 (C<sub>q</sub>), 158.39 (C<sub>q</sub>), 138.23 (2 × CH), 129.24 (C<sub>q</sub>), 129.21 (2 × CH), 117.34 (2 × CH), 114.81 (2 × CH), 83.00 (C<sub>q</sub>), 81.87 (d, *J* = 170.8 Hz, CH<sub>2</sub>), 69.80 (CH<sub>2</sub>), 67.20 (d, *J* = 20.6 Hz, CH<sub>2</sub>); HRMS (EI): m/z calcd for C<sub>15</sub>H<sub>14</sub>FIO<sub>2</sub> 372.0023; found 372.0028 M<sup>+</sup>.

#### 1-(2-Bromoethoxy)-4-((4-iodophenoxy)methyl)benzene (12b)

The procedure described above for the preparation of **7a** was employed to obtain a white solid of **12b** from 4-iodophenol and **11b** (493.7 mg, 92.7%). mp: 118.6-119.7 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.55 (d, J = 8.9 Hz, 2H), 7.34 (d, J = 8.6 Hz, 2H), 6.92 (d, J = 8.6 Hz,

2H), 6.74 (d, *J* = 8.9 Hz, 2H), 4.96 (s, 2H), 4.30 (t, *J* = 6.3 Hz, 2H), 3.64 (t, *J* = 6.3 Hz, 2H); MS (EI): m/z calcd for C<sub>15</sub>H<sub>14</sub>BrIO<sub>2</sub>432; found 432 M<sup>+</sup>.

#### 1-Bromo-4-((4-(2-fluoroethoxy)benzyl)oxy)benzene (12c)

The procedure described above for the preparation of **7a** was employed to obtain a white solid of **12c** from 4-bromophenol and **11a** (1.03 g, 76.1%). mp: 116.7-117.3 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.39 – 7.31 (m, 4H), 6.94 (d, *J* = 8.0 Hz, 2H), 6.84 (d, *J* = 8.2 Hz, 2H), 4.96 (s, 2H), 4.84 – 4.68 (m, 2H), 4.28 – 4.17 (m, 2H); MS (EI): m/z calcd for C<sub>15</sub>H<sub>14</sub>BrFO<sub>2</sub> 324; found 324 M<sup>+</sup>.

#### 2-(4-((4-Iodophenoxy)methyl)phenoxy)ethyl 4-methylbenzenesulfonate (13a)

A mixture of **12b** (402.5 mg, 0.93 mmol) and silver *p*-toluenesulfonate (519.1 mg, 1.86 mmol) in acetonitrile (20 mL) was stirred for 12 h at 90 °C. The mixture was concentrated under reduced pressure and purified by silica gel chromatography (petroleum ether/AcOEt = 4/1) to give a white solid of **13a** (277.6 mg, 57.0%). m.p. 141.7-142.5 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.82 (d, *J* = 8.3 Hz, 2H), 7.55 (d, *J* = 8.9 Hz, 2H), 7.34 (d, *J* = 8.0 Hz, 2H), 7.29 (d, *J* = 8.6 Hz, 2H), 6.79 (d, *J* = 8.6 Hz, 2H), 6.73 (d, *J* = 8.9 Hz, 2H), 4.94 (s, 2H), 4.39 – 4.35 (m, 2H), 4.17 – 4.14 (m, 2H), 2.45 (s, 3H); MS (EI): m/z calcd for C<sub>22</sub>H<sub>21</sub>IO<sub>5</sub>S 524; found 524 M<sup>+</sup>.

#### Tributyl(4-((4-(2-fluoroethoxy)benzyl)oxy)phenyl)stannane (13b)

The procedure described above for the preparation of **8b** was used to obtain a colorless oil of **13b** from **12c** (134.6 mg, 25.1%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.37 (d, *J* = 8.7 Hz, 2H), 7.32 – 7.26 (m, 2H), 6.99 – 6.93 (m, 4H), 5.00 (s, 2H), 4.83 – 4.68 (m, 2H), 4.28 – 4.17 (m,

2H), 1.58 - 1.50 (m, 6H), 1.39 - 1.27 (m, 6H), 1.14 - 0.96 (m, 6H), 0.88 (t, J = 7.3 Hz, 9H); MS (EI): m/z calcd for C<sub>27</sub>H<sub>41</sub>FO<sub>2</sub>Sn 536; found 536 M<sup>+</sup>.

# **Supplementary Tables**

	12a
Data collection	
Formula sum	$C_{15}H_{14}FIO_2$
Formula weight (g/mol)	372.16
Crystal system	monoclinic
Space group	P2(1)/c
Cell dimensions	
<i>a</i> , <i>b</i> , <i>c</i> (Å)	17.65, 10.49, 7.66
α, β, γ (°)	90, 96.4, 90
Cell volume (Å <sup>3</sup> )	1408.94
Z	4
F(000)	728
Crystal size (mm)	0.27×0.20×0.11
Calc. density (g/cm <sup>3</sup> )	1.754
Completeness (%)	99.9
Refinement	
$\mathbf{R}[\mathbf{F}^2 > 2\sigma(\mathbf{F}^2)]$	0.0180
$wR(F^2)$	0.0409
GoF	1.036
Reflections	2550
Parameters	172
Restraints	0

 Table S1 | Crystal data and structure refinements for compound 12a (CCDC 1063679)

Table S2 | Purity of key target compounds

Compounda	Flow rate	Mobile phase	Column	Retention time	D
Compounds	(mL/min)	(CH <sub>3</sub> CN %)	(Venusil MP C18)	(RT, min)	Purity (%)
7a	1	80	$4.6 \times 250 \text{ mm}$	8.81	99.4
[ <sup>125</sup> I] <b>7a</b>	1	80	$4.6 \times 250 \text{ mm}$	9.28	99.2
7a	1	80	$4.6 \times 250 \text{ mm}$	8.12	99.2
[ <sup>18</sup> F] <b>7a</b>	1	80	$4.6 \times 250 \text{ mm}$	8.53	99.7
12a	1	80	$4.6 \times 250 \text{ mm}$	8.21	99.7
[ <sup>125</sup> I] <b>12a</b>	1	80	$4.6 \times 250 \text{ mm}$	8.63	99.0
12a	1	80	$4.6 \times 250 \text{ mm}$	8.96	99.2
[ <sup>18</sup> F] <b>12a</b>	1	80	$4.6 \times 250 \text{ mm}$	9.61	99.9

Organ	2 min	10 min	30 min	60 min
	[ <sup>125</sup> I] <b>7a</b> (log	$g D = 3.96 \pm 0.22$ , SA $\approx$	≈ 81 GBq/ $\mu$ mol)	
blood	$5.03 \pm 0.20$	$3.03 \pm 0.48$	$2.17 \pm 0.31$	$1.96 \pm 0.31$
brain	$7.04 \pm 0.89$	$2.73 \pm 0.25$	$1.05 \pm 0.22$	$0.55\ \pm 0.11$
heart	$9.70 \pm 2.25$	$2.01\ \pm 0.14$	$1.65 \pm 0.43$	$1.00\ \pm 0.18$
liver	$22.45 \pm 3.88$	$14.43 \pm 2.37$	$12.35 \pm 2.83$	$5.15\ \pm 1.09$
spleen	$3.86 \pm 0.21$	$1.32 \pm 0.13$	$1.26\ \pm 0.20$	$1.14\ \pm 0.56$
lung	$10.78 \pm 2.33$	$7.44 \pm 1.59$	$4.25 \pm 0.41$	$4.04 \pm 1.63$
kidney	$12.03 \pm 1.20$	$9.53 \pm 1.58$	$6.71 \pm 0.78$	$6.56 \pm 1.10$
pancreas	$8.15 \pm 1.09$	$2.07 \pm 0.30$	$2.61 \pm 0.59$	$1.97 \pm 0.48$
muscle	$3.72 \pm 0.51$	$1.28 \pm 0.11$	$1.29 \pm 0.31$	$1.36 \pm 0.25$
thyroid <sup>b</sup>	$0.12 \pm 0.02$	$0.09 \pm 0.02$	$0.12 \pm 0.01$	$0.16 \pm 0.03$
stomach <sup>b</sup>	$1.32 \pm 0.21$	$1.08 \pm 0.24$	$4.38 \pm 1.00$	$2.44 \pm 0.97$
intestine <sup>b</sup>	$6.46 \pm 0.79$	$17.21 \pm 2.33$	$19.88 \pm 4.22$	$20.21 \pm 7.78$
	[ <sup>125</sup> I] <b>7a</b> (log	$g D = 3.96 \pm 0.22$ , SA	≈ 60 GBq/ $\mu$ mol)	
blood	$3.30 \pm 0.43$	$2.31 \pm 0.22$	$1.39 \pm 0.15$	$0.83 \pm 0.06$
brain	$5.39 \pm 0.36$	$3.51 \pm 0.36$	$0.97 \pm 0.11$	$0.37 \pm 0.05$
heart	$7.34 \pm 0.69$	$2.32 \pm 0.15$	$0.96 \pm 0.13$	$0.55 \pm 0.10$
liver	$12.12 \pm 2.01$	$11.30 \pm 0.58$	$6.29 \pm 0.22$	$3.31 \pm 0.37$
spleen	$3.63 \pm 0.83$	$1.53 \pm 0.23$	$0.62 \pm 0.03$	$0.36 \pm 0.03$
lung	$7.77 \pm 0.18$	$6.50 \pm 0.69$	$4.30 \pm 0.58$	$1.91 \pm 0.29$
kidney	$9.80 \pm 1.05$	$10.01 \pm 1.43$	$6.25 \pm 1.23$	$3.40 \pm 0.48$
pancreas	6.79 ±1.53	$2.79 \pm 0.35$	$1.06 \pm 0.16$	$0.57 \pm 0.05$
muscle	$3.37 \pm 0.50$	$1.42 \pm 0.20$	$0.95\ \pm 0.08$	$0.72 \pm 0.20$
thyroid <sup>b</sup>	$0.12 \pm 0.03$	$0.11 \pm 0.01$	$0.12 \pm 0.04$	$0.22 \pm 0.05$
stomach <sup>b</sup>	$1.15 \pm 0.21$	$0.74 \pm 0.09$	$1.67 \pm 0.33$	$1.04 \pm 0.30$
intestine <sup>b</sup>	$4.57 \pm 0.96$	$8.51 \pm 0.48$	$18.87 \pm 2.27$	$25.30 \pm 3.94$
	[ <sup>18</sup> F] <b>7a</b> (log	$g D = 3.88 \pm 0.17$ , SA $\approx$	≈ 60 GBq/µmol)	
blood	5.99 ±0.16	4.29 ±0.14	$4.36 \pm 0.20$	$4.97 \pm 0.12$
brain	$6.14 \pm 0.52$	$4.78 \pm 0.11$	$3.85 \pm 0.24$	$3.48 \pm 0.12$
heart	$9.32 \pm 0.27$	$4.03 \pm 0.34$	$4.20 \pm 0.52$	$4.26 \pm 0.13$
liver	$17.83 \pm 1.41$	$6.07 \pm 0.27$	$4.31 \pm 0.32$	$3.84 \pm 0.21$
spleen	$5.92 \pm 1.30$	$3.33 \pm 0.20$	$3.38 \pm 0.23$	$3.28 \pm 0.29$
lung	$9.34 \pm 0.45$	$4.63 \pm 0.35$	$4.21 \pm 0.18$	$4.32 \pm 0.17$
kidney	$12.70 \pm 0.69$	$7.69 \pm 1.15$	$6.12 \pm 0.79$	$4.77 \pm 0.52$
pancreas	$8.18 \pm 0.79$	$3.96 \pm 0.15$	$3.62 \pm 0.30$	$2.89 \pm 0.37$
muscle	$5.03 \pm 0.63$	$3.46 \pm 0.19$	$3.82 \pm 0.40$	$4.31 \pm 0.38$
bone	$2.53 \pm 0.43$	$1.55 \pm 0.50$	$2.70 \pm 0.48$	4.16 ±0.59
stomach <sup>b</sup>	$1.55 \pm 0.11$	$1.24 \pm 0.13$	$1.82 \pm 0.48$	$1.58 \pm 0.18$
intestine <sup>b</sup>	6.38 ±1.04	$5.38 \pm 0.33$	$5.95 \pm 0.89$	8.13 ±1.01
-		$g D = 3.62 \pm 0.15$ , SA		
blood	$4.96 \pm 0.35$	3.88 ±0.79	$3.89 \pm 0.53$	$3.01 \pm 1.07$ 1

**Table S3** | Biodistribution of radioactivity after intravenous injection of  $[^{125}I]$ **7a**,  $[^{18}F]$ **7a**,  $[^{125}I]$ **12a** and  $[^{18}F]$ **12a** in normal ICR mice <sup>*a*</sup>

brain	$5.27 \pm 0.98$	$2.28 \pm 0.27$	$0.81\ \pm 0.06$	$0.37 \pm 0.06$
heart	$6.61 \pm 1.42$	$2.12\ \pm 0.16$	$1.67\ \pm 0.09$	$1.17\ \pm 0.45$
liver	$18.12 \pm 2.34$	$12.07 \pm 1.06$	$8.68 \pm 1.02$	$4.64 \pm 0.77$
spleen	$2.85\ \pm 0.30$	$1.29\ \pm 0.06$	$1.03\ \pm 0.10$	$0.84 \pm 0.31$
lung	$10.30 \pm 2.16$	$4.16 \pm 0.26$	$3.56 \pm 0.38$	$3.11 \pm 1.02$
kidney	$9.16 \pm 0.59$	$7.61 \pm 1.23$	$7.44 \pm 0.90$	$6.09 \pm 2.17$
pancreas	$6.45 \pm 0.56$	$2.44\ \pm 0.28$	$1.75\ \pm 0.10$	$1.37 \pm 0.41$
muscle	$3.47 \pm 0.45$	$1.31 \pm 0.12$	$0.99 \pm 0.17$	$1.20 \pm 0.26$
thyroid <sup>b</sup>	$0.13 \pm 0.02$	$0.13 \pm 0.01$	$0.19 \pm 0.04$	$0.34 \pm 0.10$
stomach <sup>b</sup>	$1.30 \pm 0.14$	$1.34 \pm 0.16$	$2.02 \pm 0.49$	$2.27 \pm 0.40$
intestine <sup>b</sup>	$4.81 \pm 0.97$	$10.68 \pm 2.60$	$16.61 \pm 3.88$	$18.84 \pm 8.75$
	[ <sup>18</sup> F] <b>12a</b> (lo	$g D = 3.84 \pm 0.07$ , SA	$\approx 60 \text{ GBq}/\mu \text{mol})$	
blood	$7.53 \pm 0.80$	$4.55 \pm 0.36$	$5.91 \pm 0.33$	$5.87 \pm 0.19$
brain	$6.76 \pm 0.41$	$5.73 \pm 0.44$	$5.02 \pm 0.32$	$4.26 \pm 0.18$
heart	$9.11 \pm 1.07$	$4.40 \pm 0.36$	$5.41 \pm 0.36$	$4.76 \pm 0.40$
liver	$13.20 \pm 1.45$	$4.74 \pm 0.36$	$4.84 \pm 0.28$	$4.64 \pm 0.21$
spleen	$4.98 \pm 0.32$	$3.55 \pm 0.26$	$4.29 \pm 0.36$	$3.87 \pm 0.34$
lung	$9.17 \pm 1.10$	$4.91 \pm 0.60$	$5.34 \pm 0.31$	$5.19 \pm 0.36$
kidney	$11.63 \pm 0.86$	$6.19 \pm 0.46$	$6.19 \pm 0.37$	$5.48 \pm 0.41$
pancreas	$8.75 \pm 0.33$	$3.96 \pm 0.27$	$4.55 \pm 0.36$	$3.95 \pm 0.25$
muscle	$5.38 \pm 0.47$	$3.92 \pm 0.41$	$4.71 \pm 0.50$	$4.51 \pm 0.49$
bone	$2.55 \pm 0.29$	$2.13 \pm 0.27$	$3.31 \pm 0.21$	$3.51 \pm 0.78$
stomach <sup>b</sup>	$1.70\ \pm 0.08$	$1.54 \pm 0.32$	$2.09 \pm 0.32$	$1.70 \pm 0.29$
intestine <sup>b</sup>	$7.72 \pm 0.54$	$6.94 \pm 0.53$	$8.09 \pm 0.43$	8.50 ±1.23

<sup>*a*</sup> Expressed as % injected dose per gram. Each value represents the mean  $\pm$  SD for 4-5 mice at each interval.

<sup>b</sup> Expressed as % injected dose per organ.

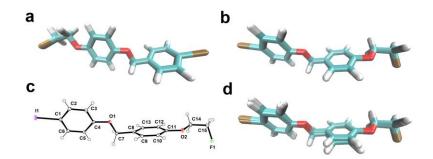
		-		[ <sup>18</sup> F]7a			
Organ	Post-injection time (min)	Metabolite [ <sup>125</sup> I]7a-1	Metabolite [ <sup>125</sup> I]7a-2	Parent tracer [ <sup>125</sup> I]7a	Metabolite [ <sup>18</sup> F]7a-1 and 2	Parent tracer [ <sup>18</sup> F]7a	
	2	4.1%	0.0%	95.9%	15.3%	84.7%	
D	10	5.1%	8.9%	86.0%	30.2%	69.8%	
Brain	30	10.0%	10.6%	79.4%	31.1%	68.9%	
	60	10.3%	17.6%	72.1%	66.0%	34.0%	
	2	14.8%	0.0%	85.2%	56.9%	43.1%	
Plasma	10	58.7%	0.0%	41.3%	78.4%	21.6%	
	30	77.9%	0.0%	22.1%	78.2%	21.8%	
	60	87.7%	0.0%	12.3%	92.8%	7.2%	
	2	60.5%	4.3%	35.2%	20.5%	79.5%	
Liver	10	72.3%	14.2%	13.5%	37.4%	62.6%	
Liver	30	86.3%	4.9%	8.8%	51.3%	48.7%	
	60	93.0%	3.7%	3.3%	76.6%	23.4%	
	2	0.0%	0.0%	0.0%	100%	0.0%	
Urine	10	97.4%	0.0%	2.6%	100%	0.0%	
Urme	30	100%	0.0%	0.0%	100%	0.0%	
	60	100%	0.0%	0.0%	100%	0.0%	
	2	77.1%	0.0%	22.9%	46.4%	53.6%	
Б	10	41.2%	0.0%	58.8%	81.2%	18.8%	
Feces	30	98.0%	0.0%	2.0%	89.5%	10.5%	
	60	91.7%	8.3%	0.0%	92.7%	7.3%	

**Table S4** | Percentages of metabolites extracted from the brain, plasma, liver, urine and feces of ICR mice after intravenous injection of  $[^{125}I]$ **7a** and  $[^{18}F]$ **7a** 

			[ <sup>125</sup> I]12a	[ <sup>18</sup> F]12a		
Organ	Post-injection time (min)	Metabo lite [ <sup>125</sup> I]12a-1	Metabolite [ <sup>125</sup> I]12a-2	Parent tracer [ <sup>125</sup> I]12a	Metabolite [ <sup>18</sup> F]12a-1 and 2	Parent tracer [ <sup>18</sup> F]12a
	2	7.9%	0.0%	92.1%	12.1%	87.9%
Brain	10	8.2%	9.7%	82.1%	34.3%	65.7%
Drain	30	9.6%	12.1%	78.3%	50.8%	49.2%
	60	12.7%	18.7%	68.6%	70.2%	29.8%
	2	11.6%	0.0%	88.4%	26.3%	73.7%
Plasma	10	70.5%	0.0%	29.6%	59.6%	40.4%
r iasilia	30	73.7%	0.0%	26.3%	65.9%	34.1%
	60	95.3%	0.0%	4.68%	86.1%	13.9%
	2	48.7%	2.2%	49.1%	8.9%	91.1%
Liver	10	76.7%	1.8%	21.5%	14.7%	85.3%
LIVEI	30	90.5%	3.2%	6.3%	17.8%	82.2%
	60	94.7%	1.5%	3.8%	78.3%	21.7%
	2	0.0%	0.0%	0.0%	100%	0.0%
Urine	10	100%	0.0%	0.0%	100%	0.0%
OTINE	30	100%	0.0%	0.0%	100%	0.0%
	60	98.9%	1.1%	0.0%	100%	0.0%
	2	0.0%	0.0%	0.0%	53.3%	46.7%
Feces	10	0.0%	0.0%	0.0%	79.1%	20.9%
reces	30	0.0%	0.0%	0.0%	83.2%	16.8%
	60	0.0%	0.0%	0.0%	82.6%	17.4%

**Table S5** | Percentages of metabolites extracted from the brain, plasma, liver, urine and fecesof ICR mice after intravenous injection of  $[^{125}I]$ **12a** and  $[^{18}F]$ **12a** 

# **Supplementary Figures**



**Figure S1** | **Chemical structures of 7a and 12a.** Optimized structures of benzyloxybenzene derivatives **7a** (a) and **12a** (b). (c) X-ray crystal structure of **12a** (CCDC 1063679). Superposition of **12a** optimized and its X-ray crystal structure was show in d. RMSD = 0.190 Å.

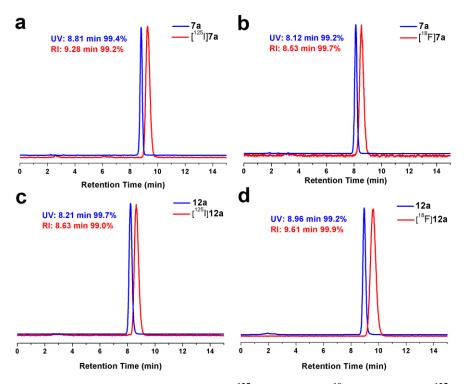


Figure S2 | Co-injection HPLC profiles of 7a and [<sup>125</sup>I]7a; 7a and [<sup>18</sup>F]7a; 12a and [<sup>125</sup>I]12a; 12a and [<sup>18</sup>F]12a. HPLC conditions: Venusil MP C18 column (Agela Technologies, 5  $\mu$ m, 4.6 × 250 mm), CH<sub>3</sub>CN/H<sub>2</sub>O = 80%/20%, 1 mL/min, UV, 254 nm.

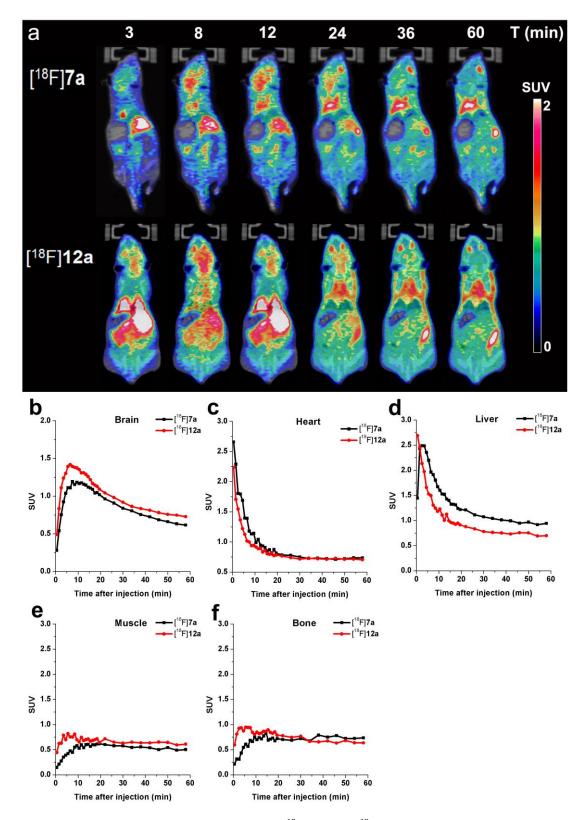


Figure S3 | Dynamic microPET/CT imaging of  $[^{18}F]$ 7a and  $[^{18}F]$ 12a in normal ICR mice. (a) Whole body time-radioactivity biodistribution by dynamic microPET/CT imaging. PET image color intensities are expressed as standardized uptake value (SUV). (b-f) Time-activity curves (TACs) of  $[^{18}F]$ 7a and  $[^{18}F]$ 12a in brain, heart, liver, muscle and bone for the entire 60 min PET scan.

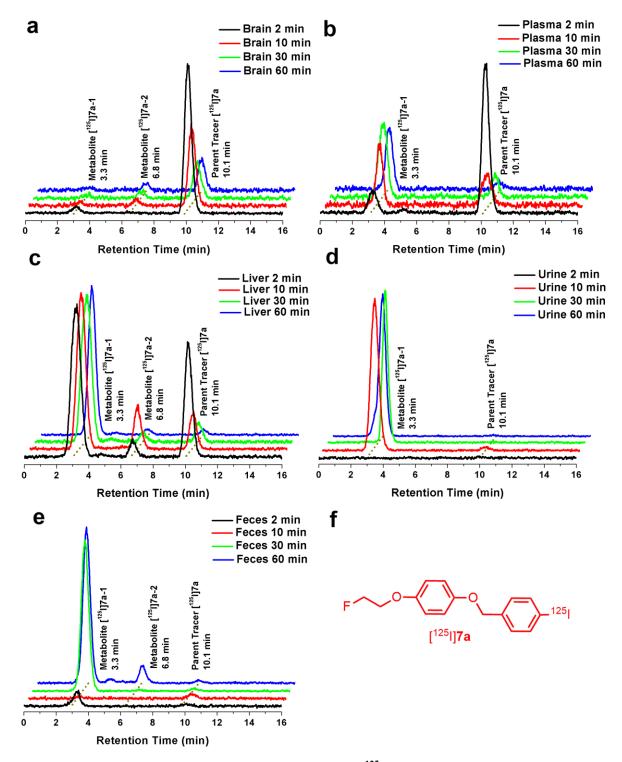


Figure S4 | HPLC profiles for radioactive metabolites of [<sup>125</sup>I]7a in ICR mice brain (a), plasma (b), liver (c), urine (d) and feces (e) at 2, 10, 30 and 60 min post-injection time points. Reversed-phase HPLC performed on a Venusil MP C18 reverse phase column (Agela Technologies, 5  $\mu$ m, 4.6 mm × 250 mm) using a binary gradient system (acetonitrile/water : 80%/20%) at a 1.0 mL/min flow rate. (f) Chemical structure of [<sup>125</sup>I]7a.

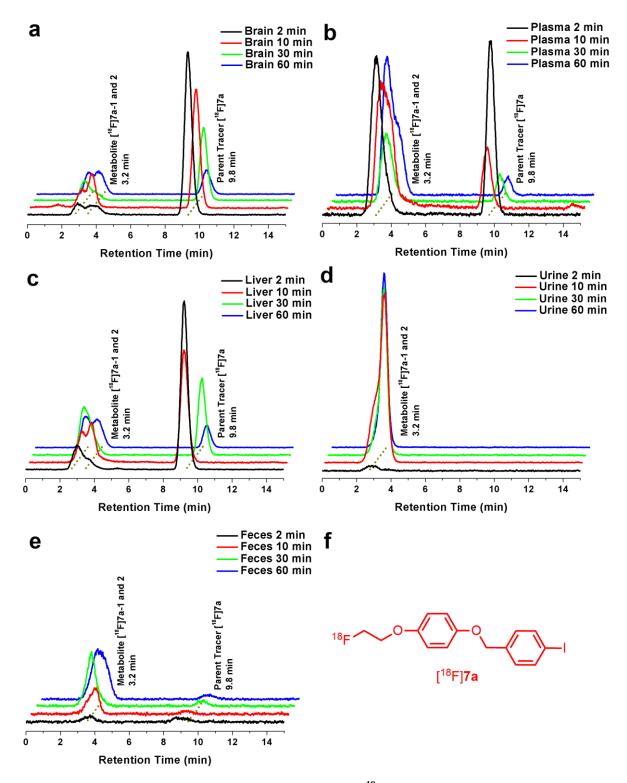


Figure S5 | HPLC profiles for radioactive metabolites of [<sup>18</sup>F]7a in ICR mice brain (a), plasma (b), liver (c), urine (d) and feces (e) at 2, 10, 30 and 60 min post-injection time points. Reversed-phase HPLC performed on a Venusil MP C18 reverse phase column (Agela Technologies, 5  $\mu$ m, 4.6 mm × 250 mm) using a binary gradient system (acetonitrile/water : 80%/20%) at a 1.0 mL/min flow rate. (f) Chemical structure of [<sup>18</sup>F]7a.

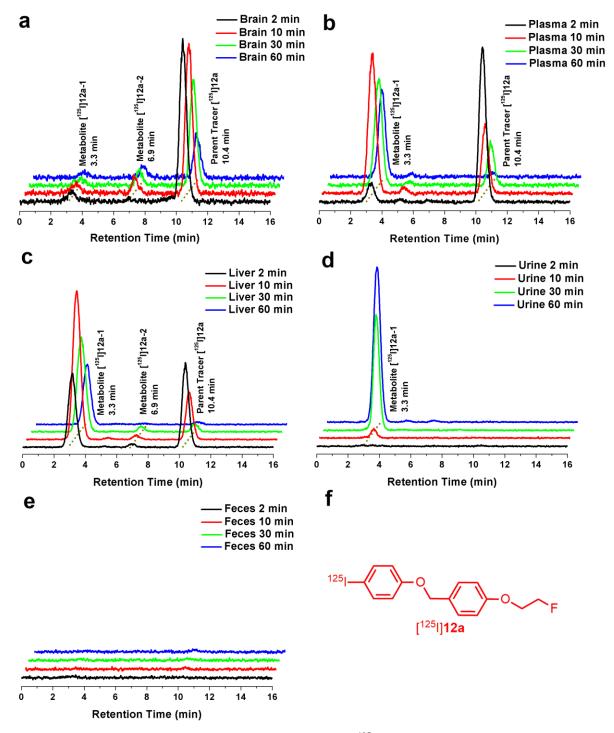


Figure S6 | HPLC profiles for radioactive metabolites of [<sup>125</sup>I]12a in ICR mice brain (a), plasma (b), liver (c), urine (d) and feces (e) at 2, 10, 30 and 60 min post-injection time points. Reversed-phase HPLC performed on a Venusil MP C18 reverse phase column (Agela Technologies, 5  $\mu$ m, 4.6 mm × 250 mm) using a binary gradient system (acetonitrile/water : 80%/20%) at a 1.0 mL/min flow rate. (f) Chemical structure of [<sup>125</sup>I]12a.

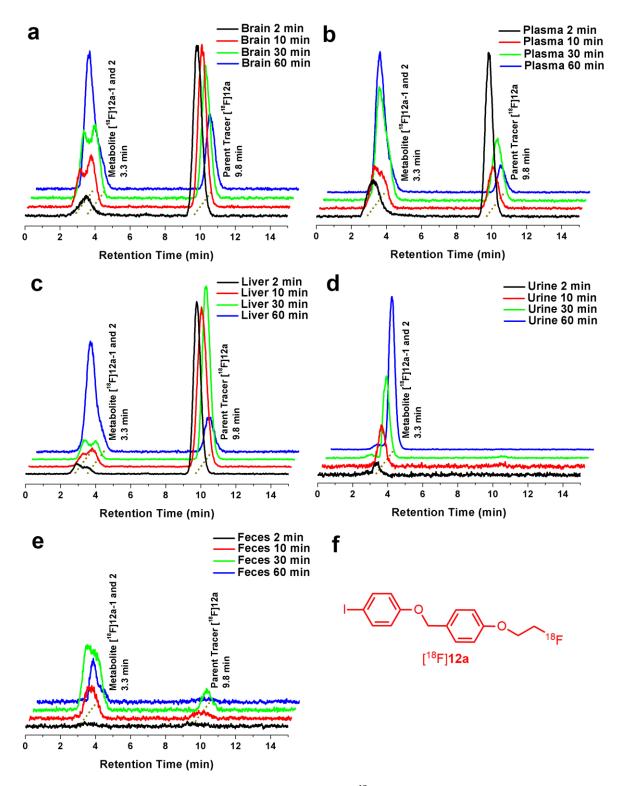
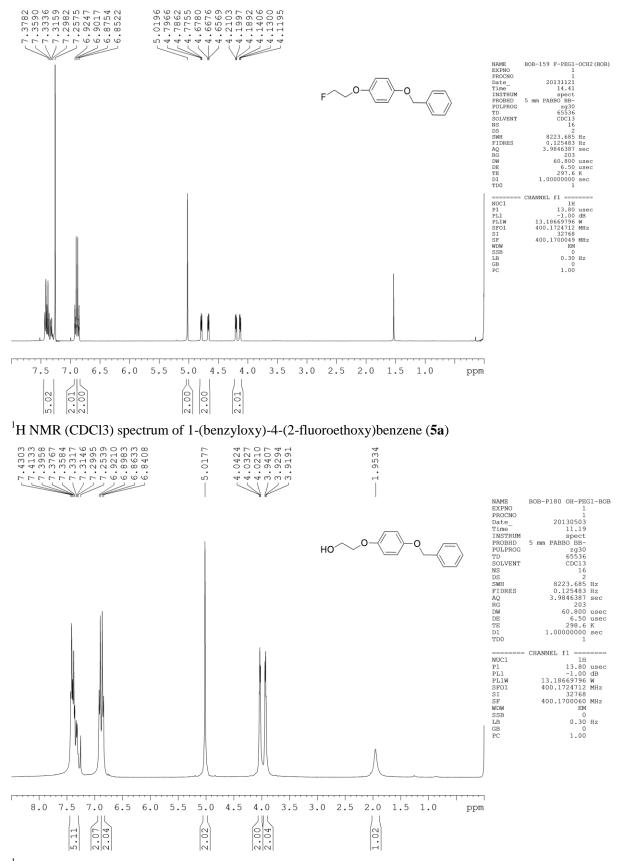
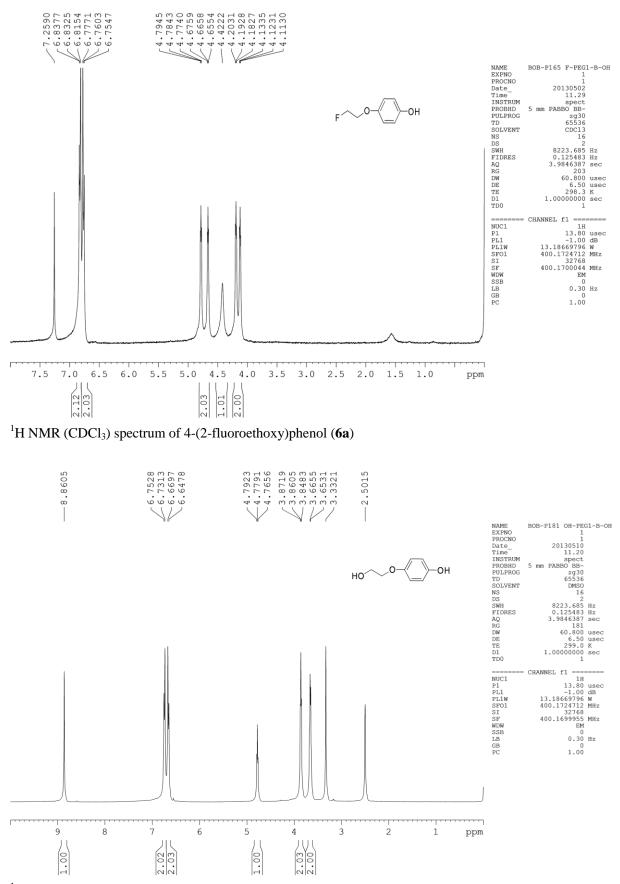


Figure S7 | HPLC profiles for radioactive metabolites of [<sup>18</sup>F]12a in ICR mice brain (a), plasma (b), liver (c), urine (d) and feces (e) at 2, 10, 30 and 60 min post-injection time points. Reversed-phase HPLC performed on a Venusil MP C18 reverse phase column (Agela Technologies, 5  $\mu$ m, 4.6 mm × 250 mm) using a binary gradient system (acetonitrile/water : 80%/20%) at a 1.0 mL/min flow rate. (f) Chemical structure of [<sup>18</sup>F]12a.

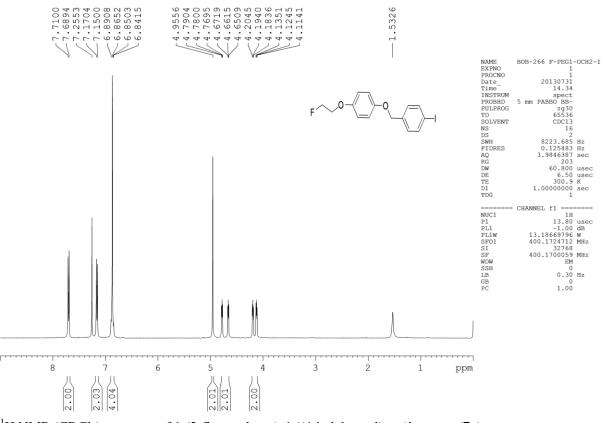




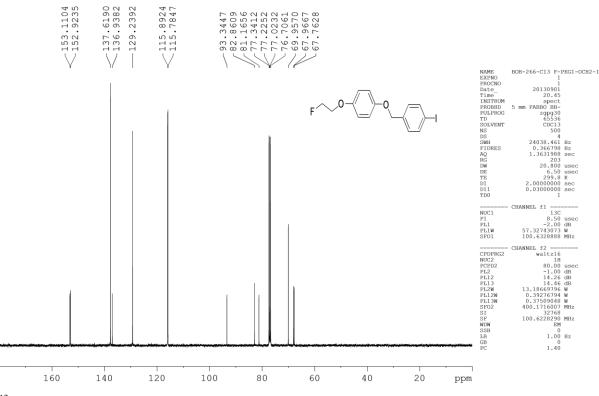
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of 2-(4-(benzyloxy)phenoxy)ethanol (**5b**)



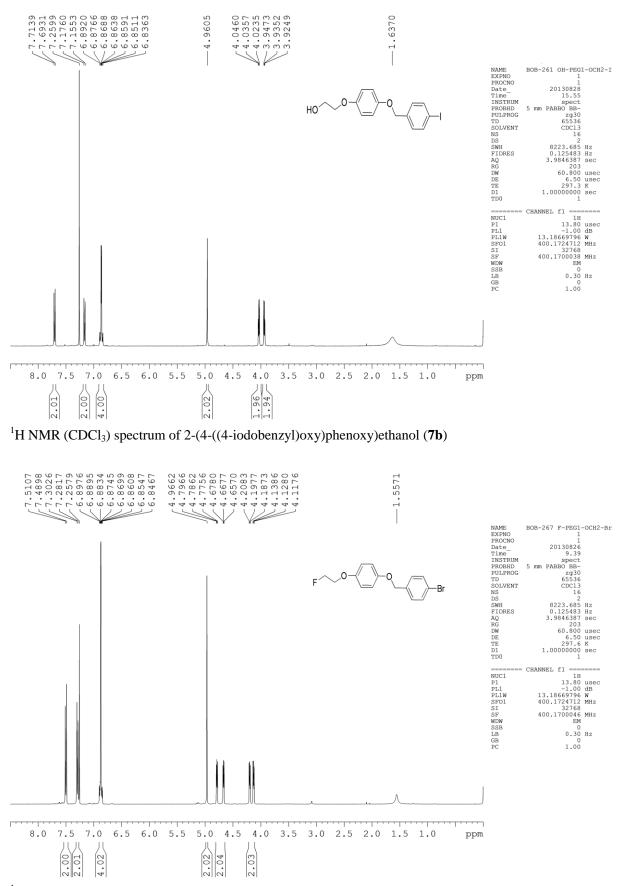
<sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>) spectrum of 4-(2-hydroxyethoxy)phenol (**6b**)



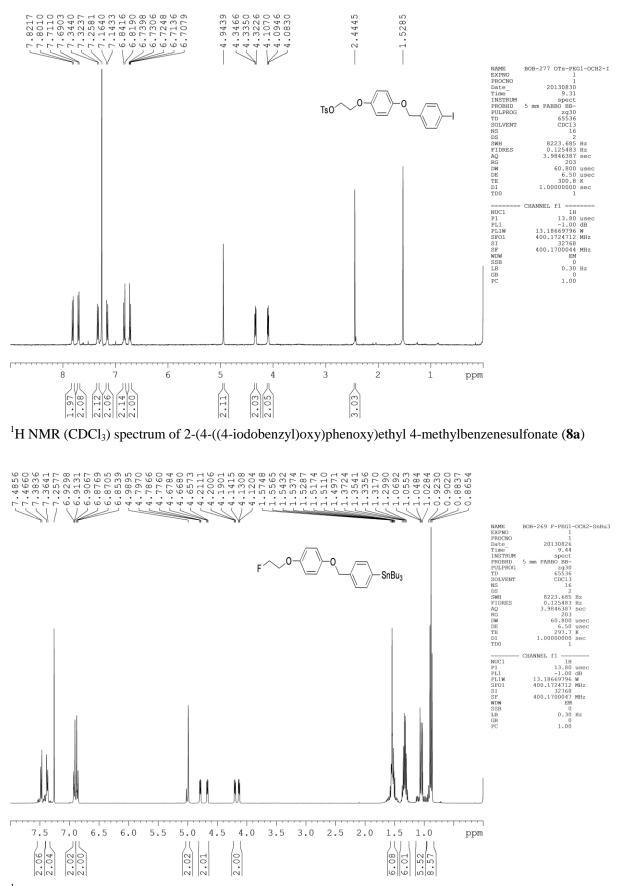
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of 1-(2-fluoroethoxy)-4-((4-iodobenzyl)oxy)benzene (7**a**)



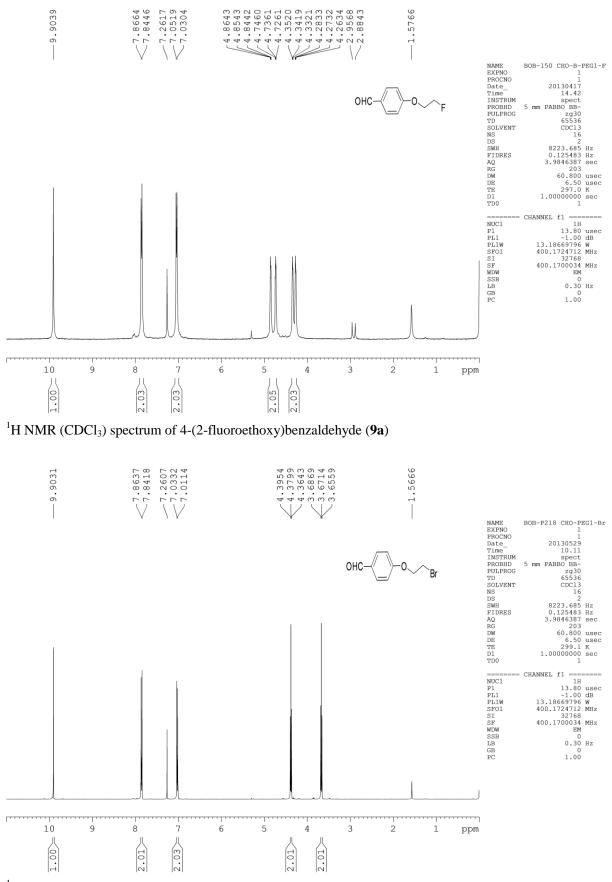
 $^{13}C\ NMR\ (CDCl_3)\ spectrum\ of\ 1-(2-fluoroethoxy)-4-((4-iodobenzyl)oxy) benzene\ (\textbf{7a})$ 



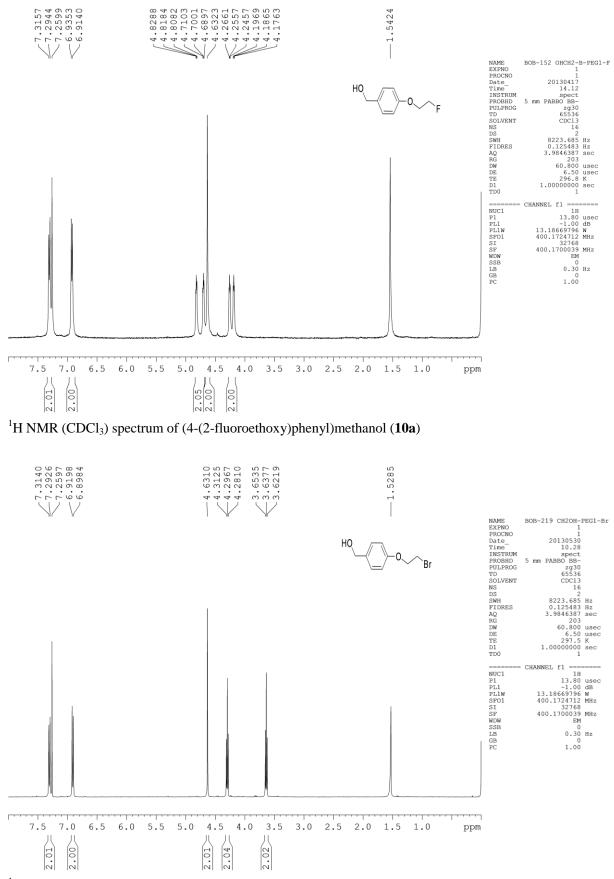
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of 1-bromo-4-((4-(2-fluoroethoxy)phenoxy)methyl)benzene (7c)



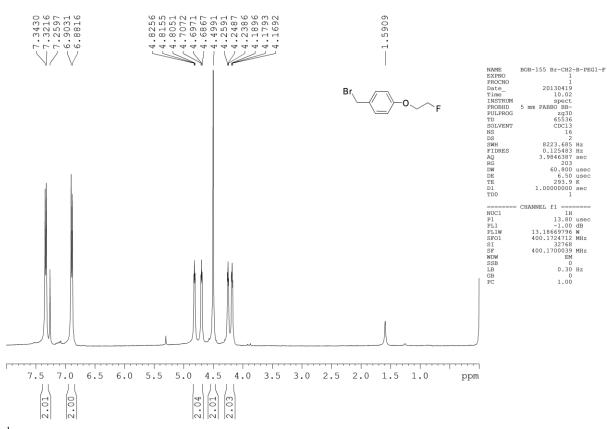
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of tributyl(4-((4-(2-fluoroethoxy)phenoxy)methyl)phenyl)stannane (**8b**)



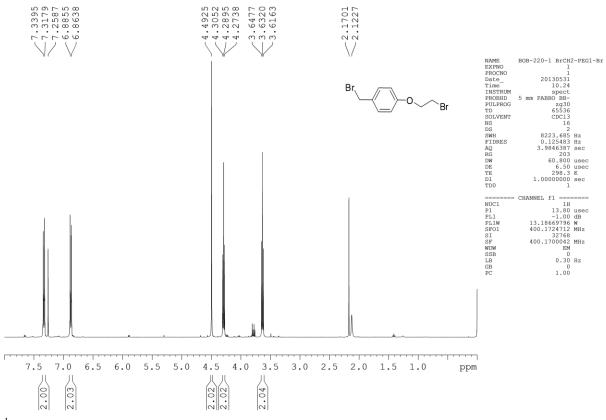
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of 4-(2-bromoethoxy)benzaldehyde (**9b**)



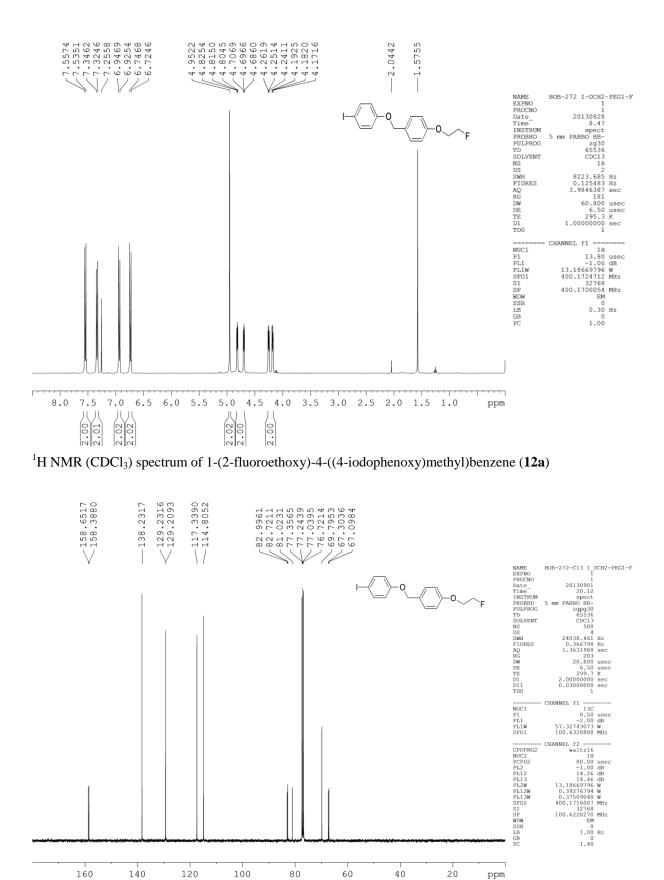
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of (4-(2-bromoethoxy)phenyl)methanol (**10b**)



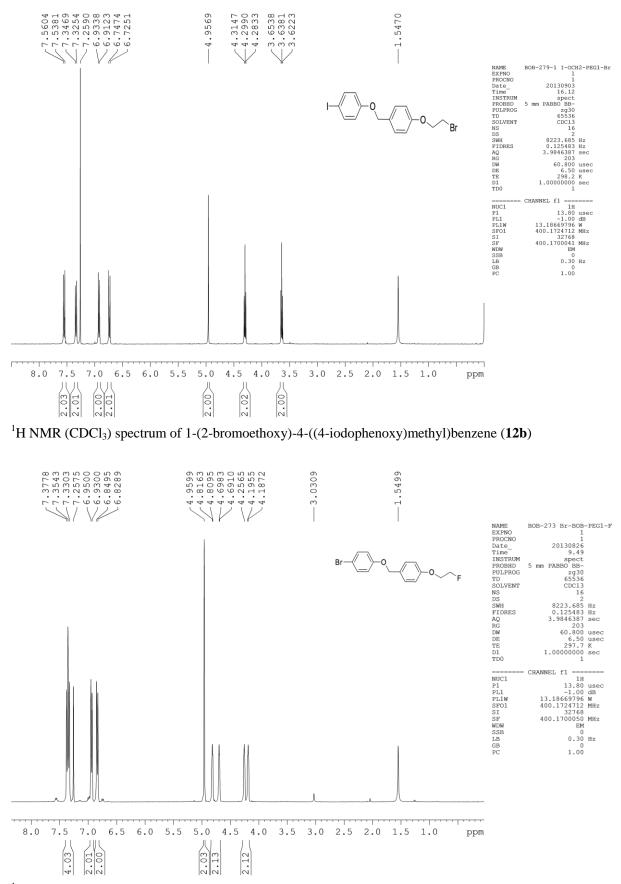
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of 1-(bromomethyl)-4-(2-fluoroethoxy)benzene (**11a**)



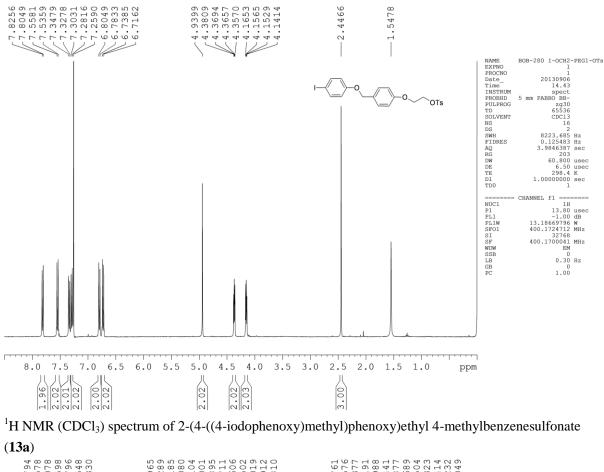
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of 1-(2-bromoethoxy)-4-(bromomethyl)benzene (**11b**)

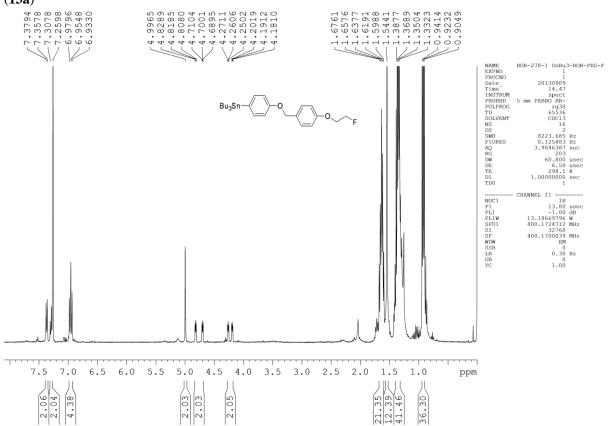


<sup>13</sup>C NMR (CDCl<sub>3</sub>) spectrum of 1-(2-fluoroethoxy)-4-((4-iodophenoxy)methyl)benzene (**12a**)

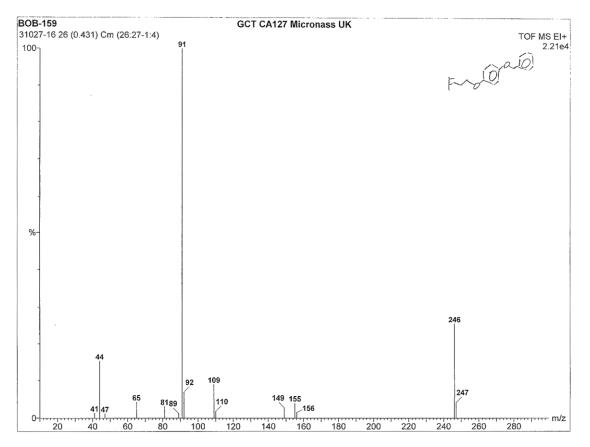


<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of 1-bromo-4-((4-(2-fluoroethoxy)benzyl)oxy)benzene (12c)

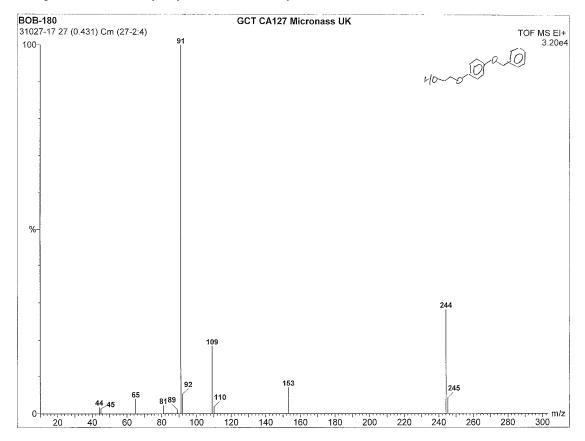




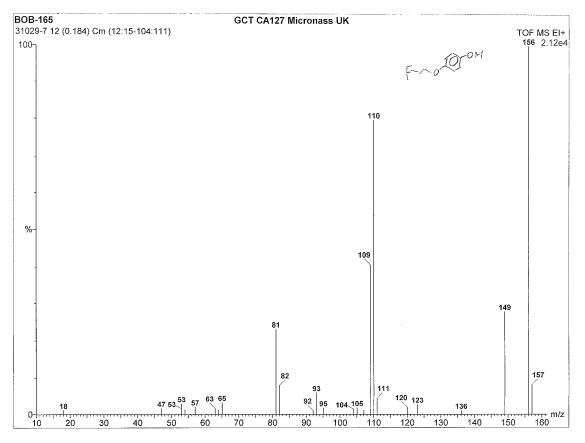
<sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum of tributyl(4-((4-(2-fluoroethoxy)benzyl)oxy)phenyl)stannane (**13b**)



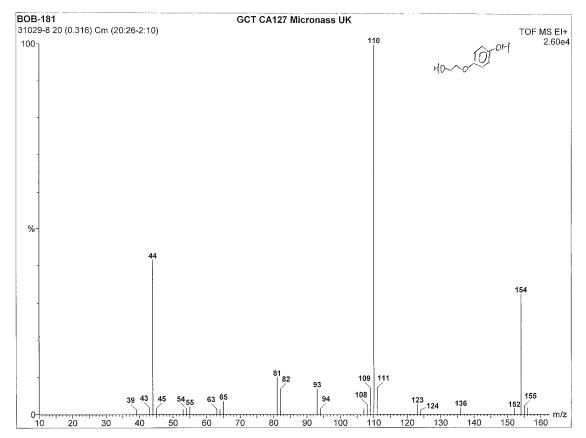
MS spectrum of 1-(benzyloxy)-4-(2-fluoroethoxy)benzene (5a)



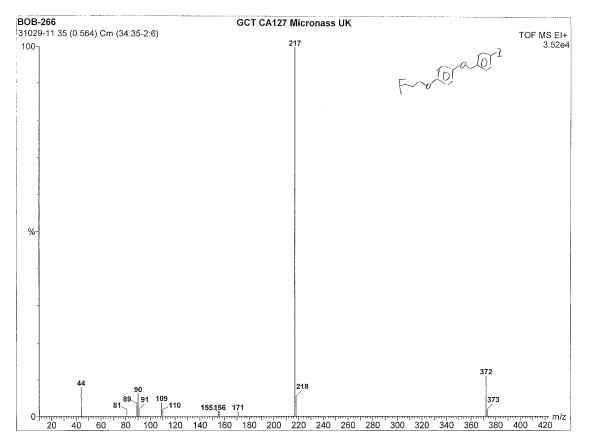
MS spectrum of 2-(4-(benzyloxy)phenoxy)ethanol (5b)



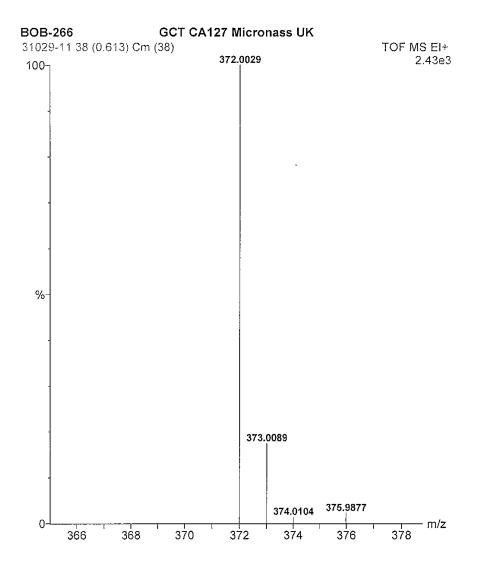
MS spectrum of 4-(2-fluoroethoxy)phenol (6a)



MS spectrum of 4-(2-hydroxyethoxy)phenol (6b)



MS spectrum of 1-(2-fluoroethoxy)-4-((4-iodobenzyl)oxy)benzene (7a)



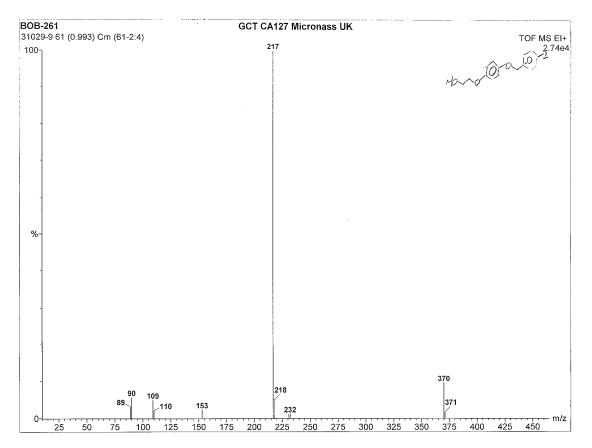
Elemental Composition Report

Tolerance = 10.0 PPM / DBE: min = -1.5, max = 50.0 Isotope cluster parameters: Separation = 1.0 Abundance = 1.0%

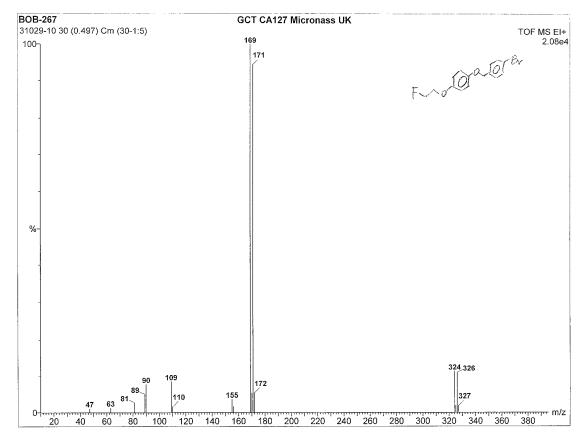
Monoisotopic Mass, Odd and Even Electron Ions 87 formula(e) evaluated with 3 results within limits (up to 50 closest results for each mass)

Minimum: Maximum: Mass 372.0029 I	80.00 100.00 RA Calc. Mass 100.00 372.0023	200.0 10.0 mDa PPM 0.6 1.7	-1.5 50.0 DBE Score 8.0 1	Formula C15 H14 O2 F
				$\sim \sim \sim$

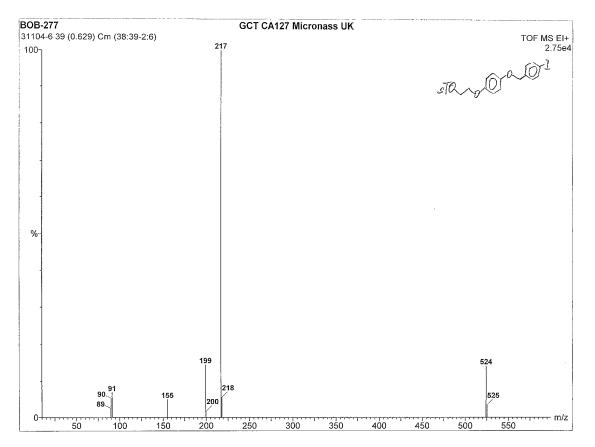
HRMS spectrum of 1-(2-fluoroethoxy)-4-((4-iodobenzyl)oxy)benzene (7a)



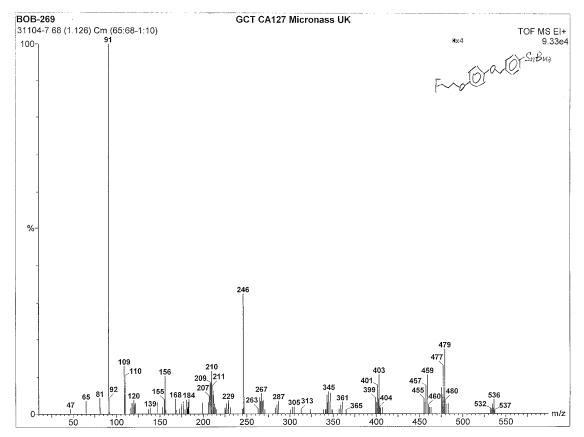
MS spectrum of 2-(4-((4-iodobenzyl)oxy)phenoxy)ethanol (7b)



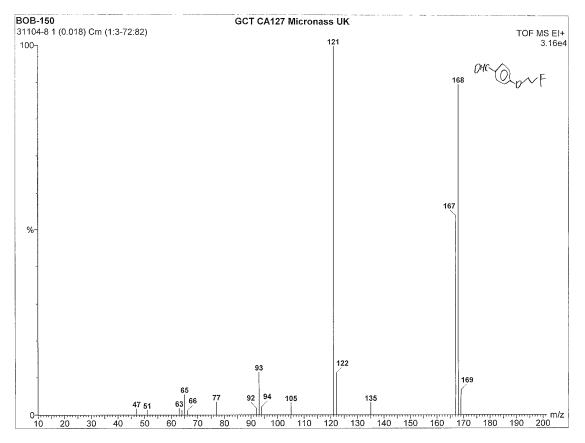
MS spectrum of 1-bromo-4-((4-(2-fluoroethoxy)phenoxy)methyl)benzene (7c)



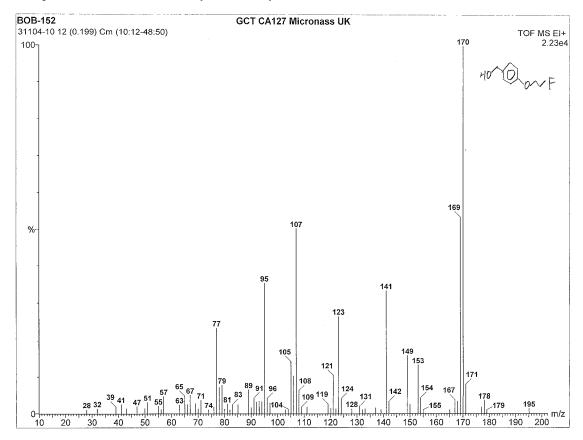
MS spectrum of 2-(4-((4-iodobenzyl)oxy)phenoxy)ethyl 4-methylbenzenesulfonate (8a)



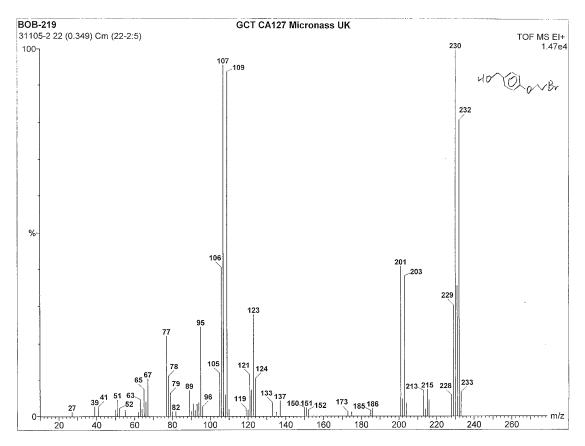
MS spectrum of tributyl(4-((4-(2-fluoroethoxy)phenoxy)methyl)phenyl)stannane (8b)



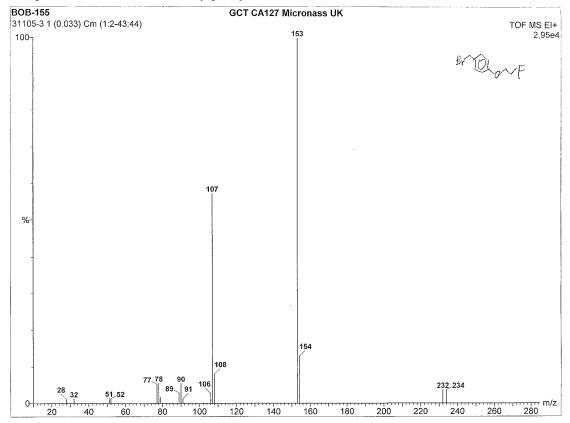
MS spectrum of 4-(2-fluoroethoxy)benzaldehyde (9a)



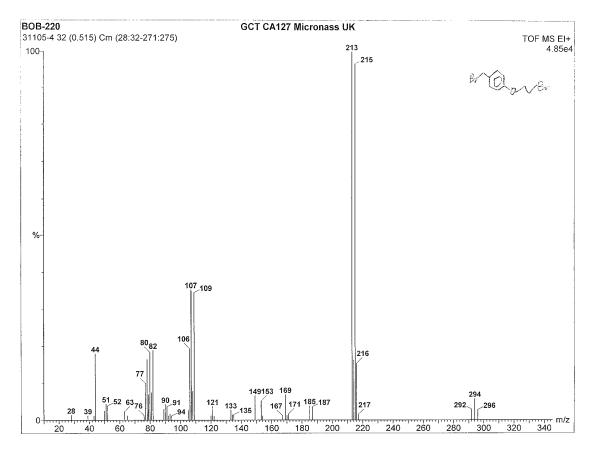
MS spectrum of (4-(2-fluoroethoxy)phenyl)methanol (10a)



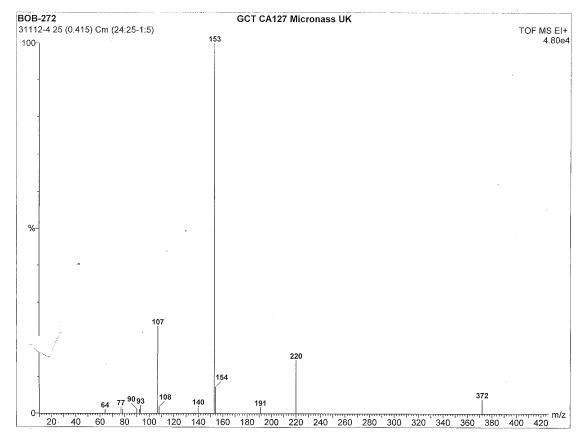




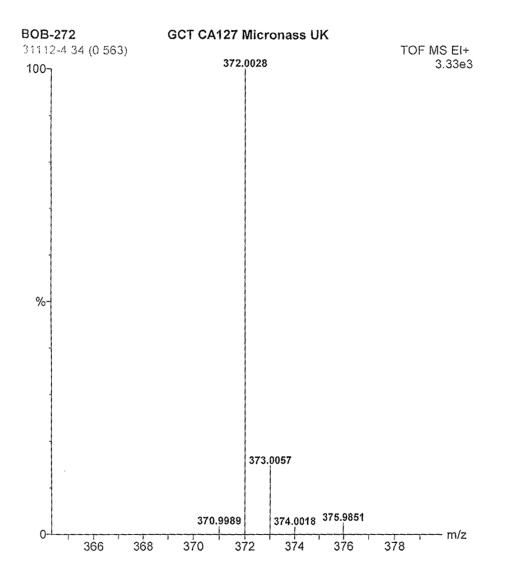
MS spectrum of 1-(bromomethyl)-4-(2-fluoroethoxy)benzene (11a)



MS spectrum of 1-(2-bromoethoxy)-4-(bromomethyl)benzene (11b)



MS spectrum of 1-(2-fluoroethoxy)-4-((4-iodophenoxy)methyl)benzene (12a)



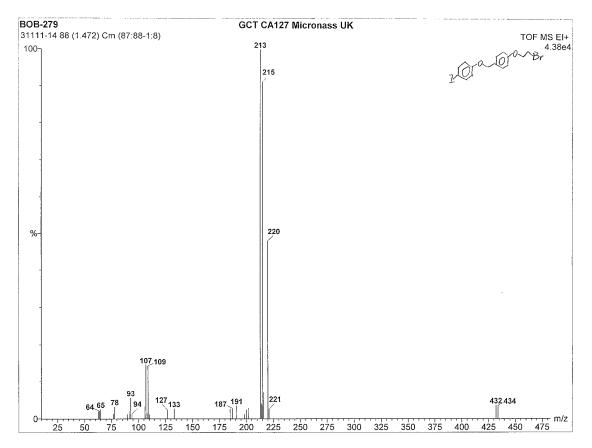
Elemental Composition Report

Tolerance = 10.0 PPM / DBE: min = -1.5, max = 50.0 Isotope cluster parameters: Separation = 1.0 Abundance = 1.0%

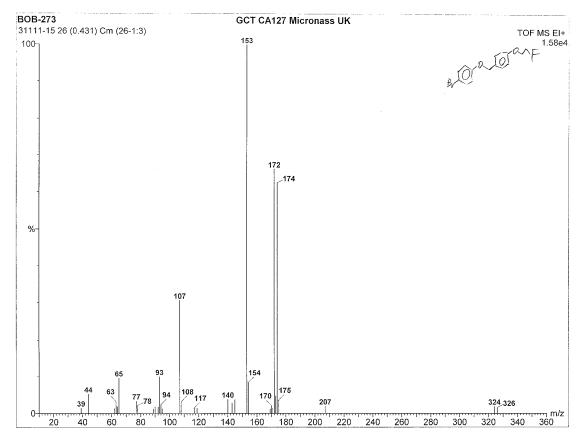
Monoisotopic Mass, Odd and Even Electron lons 82 formula(e) evaluated with 3 results within limits (up to 50 closest results for each mass)

Minimum: Maximum: Mass 372.0028	80.00 100.00 RA 100.00	Calc. Mass 372.0023	200.0 mDa 0.5	10.0 PPM 1.5	-1.5 50.0 DBE 8.0	Score 2	Formula C15 H14 O2 F I
						ľ	Draldran F

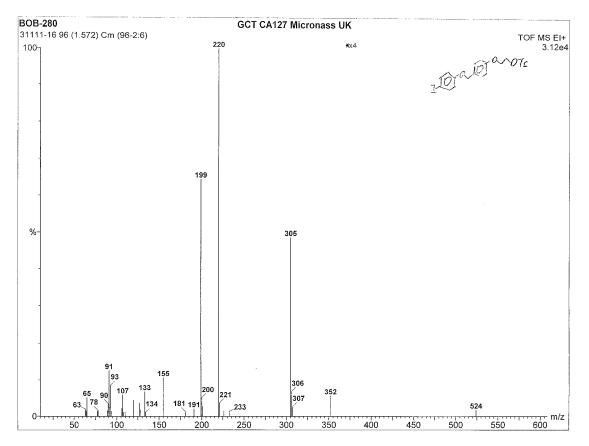
HRMS spectrum of 1-(2-fluoroethoxy)-4-((4-iodophenoxy)methyl)benzene (12a)



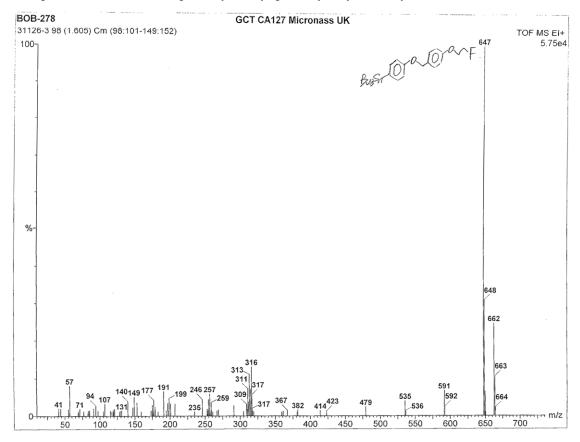
MS spectrum of 1-(2-bromoethoxy)-4-((4-iodophenoxy)methyl)benzene (12b)



MS spectrum of 1-bromo-4-((4-(2-fluoroethoxy)benzyl)oxy)benzene (12c)



MS spectrum of 2-(4-((4-iodophenoxy)methyl)phenoxy)ethyl 4-methylbenzenesulfonate (13a)



MS spectrum of tributyl(4-((4-(2-fluoroethoxy)benzyl)oxy)phenyl)stannane (13b)