

SUPPLEMENTAL FIGURES

Characterization of *Trypanosoma cruzi* sirtuins as possible drug targets for Chagas Disease

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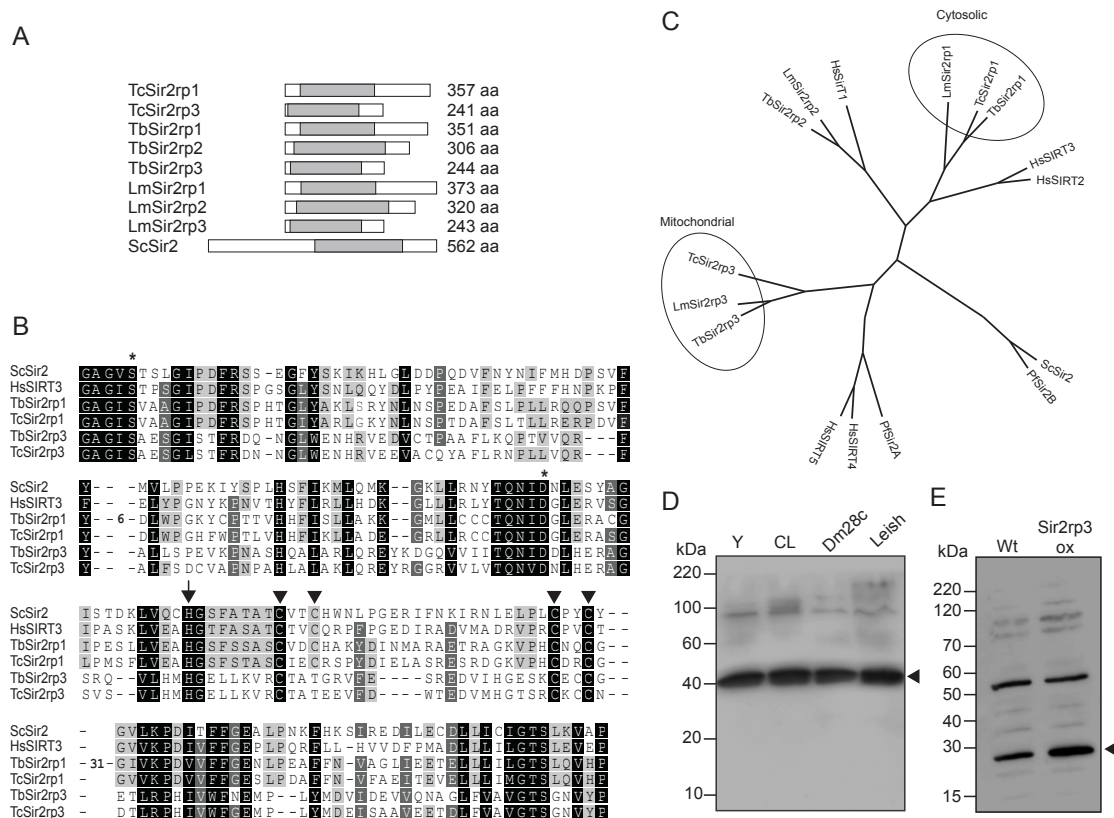


FIG S1. *T. cruzi* sirtuins amino acid sequence analysis. (A) Schematic representation of *T. cruzi* sirtuin domain (showed in grey) compared with *L. major* (LmSir2rp1-3), *T. brucei* (TbSir2rp1-3) and *S. cerevisiae* (ScSir2). (B) ClustalW amino acid sequence alignment between the core deacetylase domain of *T. cruzi* sirtuins and sirtuins of *S. cerevisiae* (ScSir2, NP_010242), *H. sapiens* (HsSIRT3, NP_036371.1), and *T. brucei* TbSir2rp1, AAX70528.1 and TbSir2rp3, AAX79070.1). Arrowheads indicate a putative Zn²⁺-binding motif, arrow indicates critical catalytic residues, and asterisks correspond to NAD⁺-binding sites. (C) Unrooted tree derived from phylogenetic analysis of the above sequences based in the Neighbor-Joining (NJ) method using the Clustal 1.8 and TreeView. *S. cerevisiae* (ScSir2, NP_010242), *H. sapiens* (HsSIRT1, AAH12499.1; HsSIRT2, AAK51133.1; HsSIRT3, NP_036371.1; HsSIRT4, NP_036372.1 and HsSIRT5, Q9NXA8.2); *T. brucei* TbSir2rp1, AAX70528.1; TbSir2rp2, AAZ13079 and TbSir2rp3, AAX79070.1); *L. major* (LmSir2rp1, CAJ040546; LmSir2rp2, CAJ04804 and LmSir2rp3, CAJ07927); *P. falciparum* (PfSir2A, Q8IE47.1; PfSir2B, Q8IKW2.1). (D) Western blot of total extracts of 1 x 10⁷ epimastigotes of Y, CL-Brener (CL), Dm28c and promastigotes forms of *Leishmania major* (Leish) probed with anti-Sir2rp1. (E) Western blot of total extracts of 1 x 10⁷ epimastigotes of wild type Y strain, or the Y strain overexpressing TcSir2rp3 probed with anti-Tc-Sir2rp3. Size markers are shown on the left side of each panel.

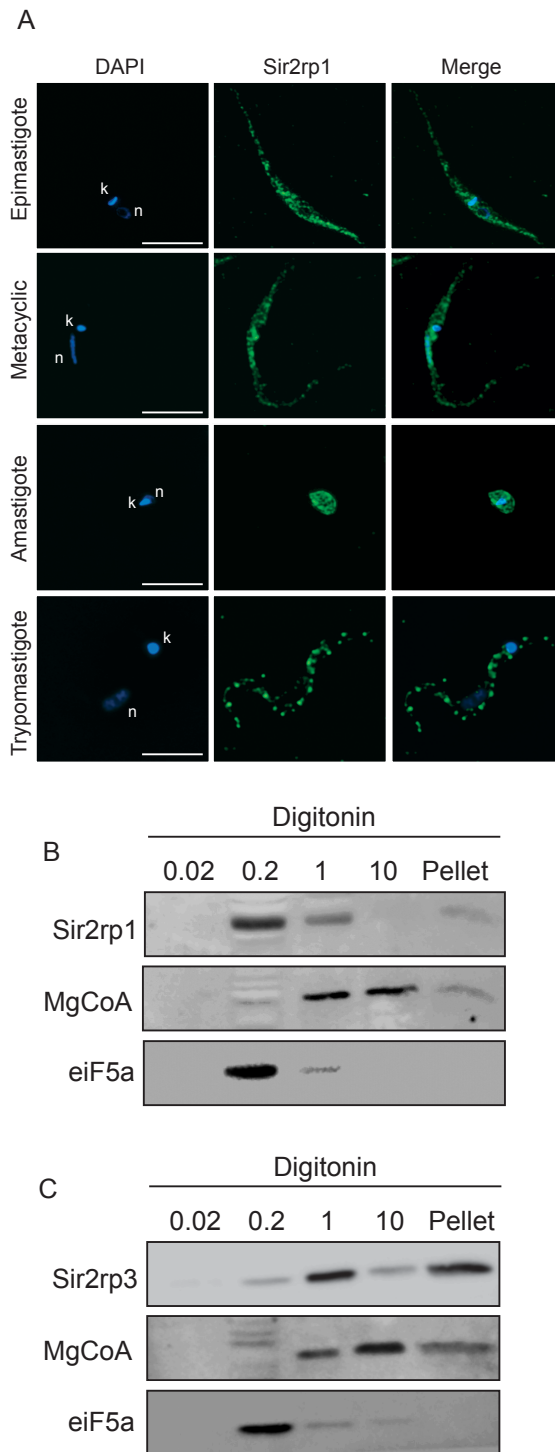


FIG S2. Subcellular localization of TcSir2rp1 and TcSir2rp3. (A) Immunofluorescence of epimastigotes, metacyclic-trypomastigotes, intracellular amastigotes and trypomastigotes derived from mammalian cells, probed with anti-Sir2rp1. The TcSir2rp1 labeling is shown in green and the DAPI in blue. Bar = 5 μ m. Western blot of solubilized proteins extracted with digitonin at the indicated concentrations in mM of TcSir2rp1-ox (B) and TcSir2rp3-ox (C) parasites probed with anti-HA antibodies for TcSir2rp1-ox and the indicated antibodies for the other blots.

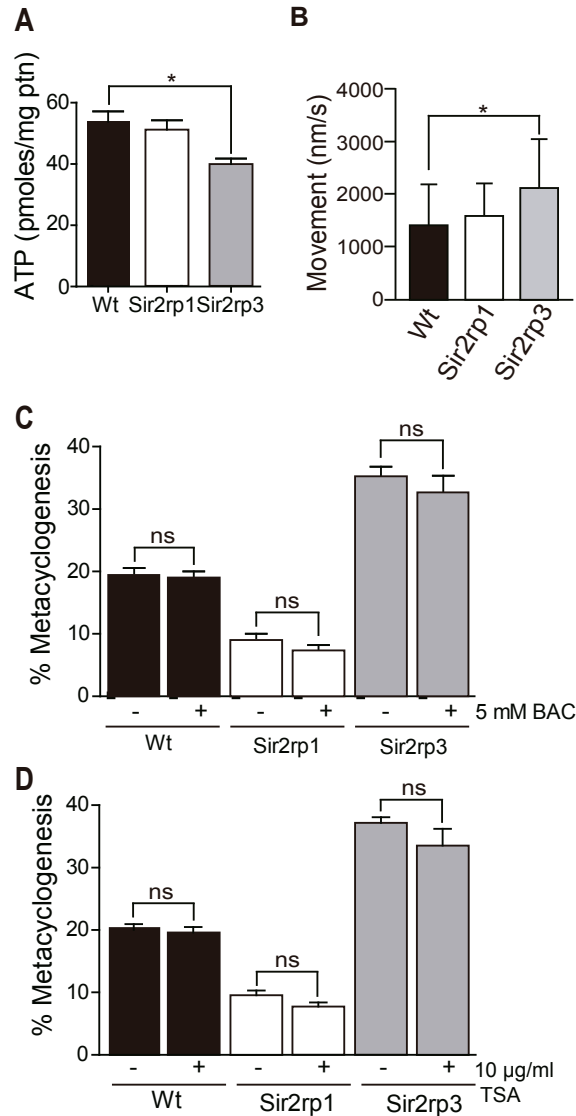


FIG S3. Phenotypic analysis of the sirtuin overexpressors. Panel (A) panel (B) shows respectively the intracellular ATP levels and the movement of wild type (dark bars), TcSir2rp1-ox (empty bars), and TcSir2rp3-ox (grey bars) epimastigotes. (C) and (D) show respectively the metacyclogenesis in the absence (-) and presence (+) of 5 mM butyric acid (BAC) or 10 µg/ml trichostatin A using wild type (dark bars); TcSir2rp1-ox (empty bars) and TcSir2rp3-ox (grey bars) epimastigotes. The values are mean \pm standard deviation (n = 3) * (p<0.01).

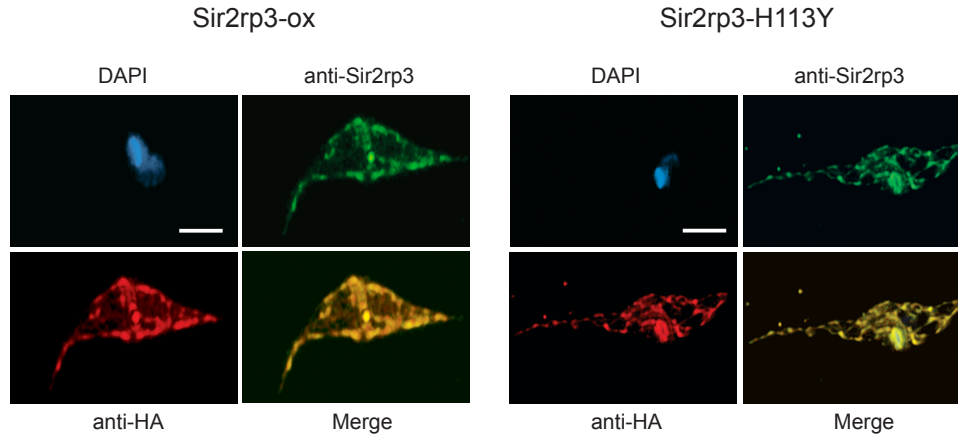


FIG S4. Wild type and overexpressor TcSir2rp3 have similar localizations. Indirect immunofluorescences of TcSir2rp3-ox and TcSir2rp3-His113Y epimastigotes using anti-Sir2rp3 (green) and anti-HA (red). The figures also show the DAPI labeling and the merged fluorescence. Bars = 2.5 μ m.

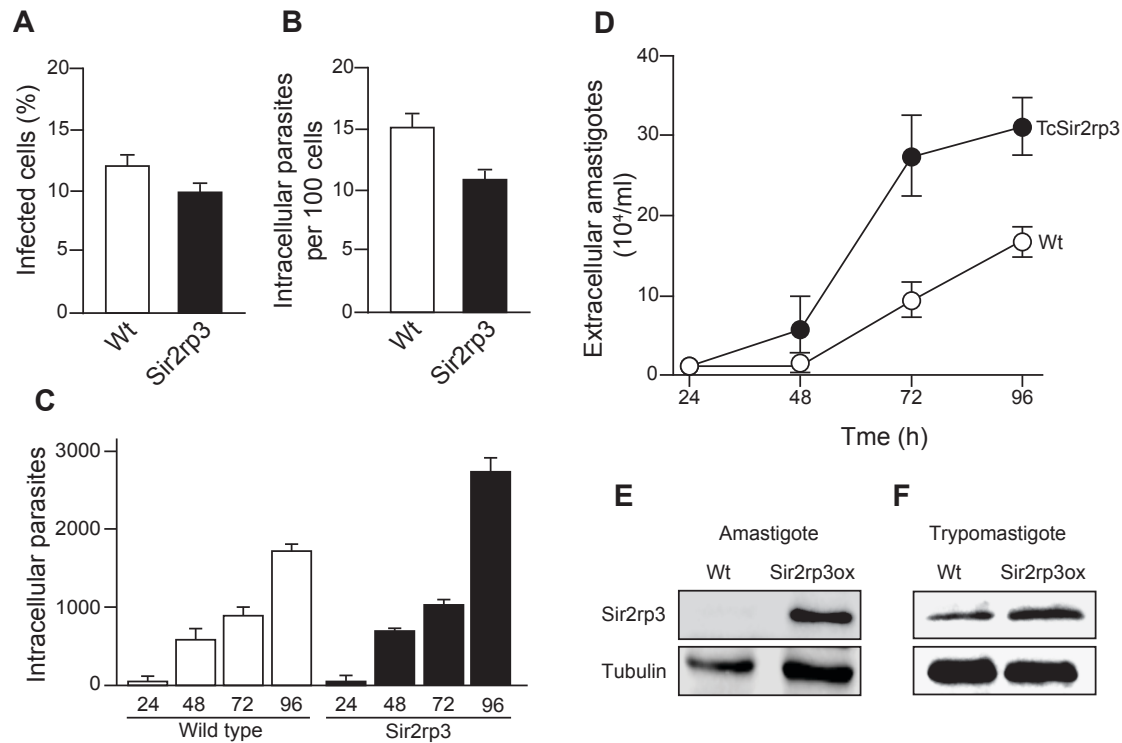


FIG S5. Trypomastigote invasion is decreased while intracellular multiplication is enhanced in TcSir2rp3 overexpressors resulting in the release of amastigotes. Wild type and TcSir2rp3-ox trypomastigotes were incubated for 1h with LLC-MK₂ cells and the number of infected cells (**A**) and intracellular parasites (**B**) measured after infection. Panel (**C**) shows the number of intracellular parasites following invasion of wild type and TcSir2rp3-ox parasites. The numbers are mean \pm standard deviation of triplicates. (**D**) Shows the number of extracellular amastigotes released from infected mammalian cells during the progression of infection with trypomastigotes from wild type (empty circles) and TcSir2rp3-ox (dark circles) parasites. (**E**) and (**F**) Western blots of wild type and TcSir2rp3ox intracellular amastigotes and trypomastigotes revealed with anti-TcSir2rp3 and anti- α -tubulin antibodies.

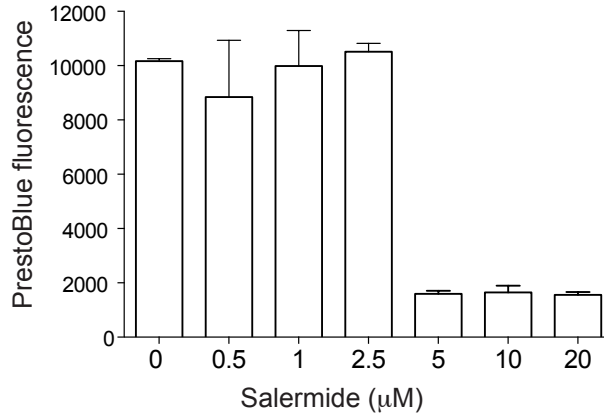


FIG S6 Cytotoxicity of salermide to LLCMK₂ cells. Cell viability was determined for the different concentrations of concentrations of salermide incubated after 48 h using PrestoBlue Viability reagent. The numbers are mean ± standard deviation (n = 3), and the experiment was repeated three times.