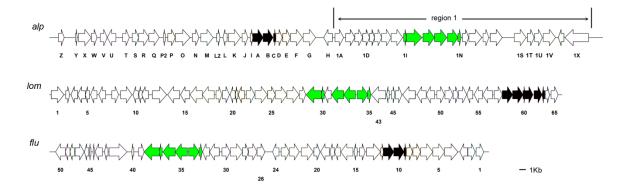
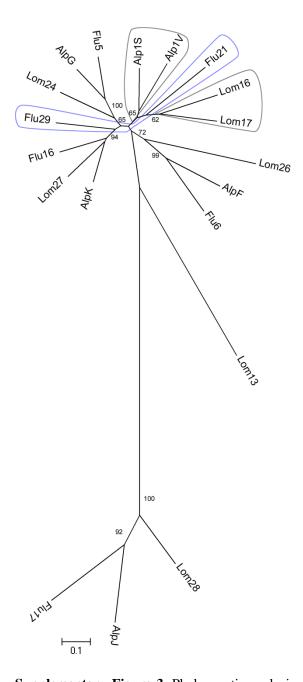
$$A \xrightarrow[OH \ 0 \]{}^{HO} \xrightarrow[N]{}^{CH_3} \xrightarrow[OH \ 0 \]{}^{HO} \xrightarrow[N]{}^{CH_3} \xrightarrow[N]{}^{CH_3}$$

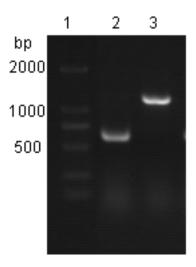
Supplementary Figure 1. The proposed biosynthetic pathways for the A-ring transformation of the kinamycin and lomaiviticin antibiotics. A, by Steven J. Gould; B, by Emily P. Balskus; C, by Bradley S. Moore.



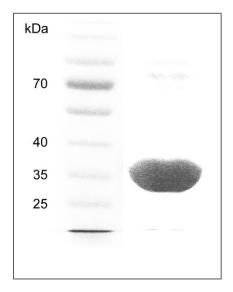
Supplementary Figure 2. Comparison of the *alp*, *lom*, and *flu* gene clusters. The three gene clusters are responsible for the biosynthesis of kinamycin, lomaiviticin and fluostatin, respectively. The DNA region labeled as "region 1" is extended in this study. The genes in black are indicated as mini-PKS, in green as the conserved six-gene subregion responsible for the diazo assembly.



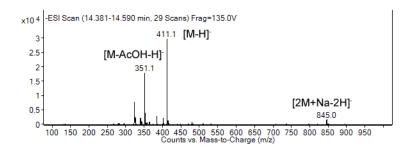
Supplementary Figure 3. Phylogenetic analysis of the oxygenases in the *alp*, *lom* and *flu* clusters. The tree was constructed by Neighbor-Joining method and the bootstrap (500 replicates) values for the main clades are shown. Scale bar represents 10% dissimilarity. The oxygenases labeled in black or blue are the candidates discussed in this study for A-ring transformation in the kinamycin, fluostatin, and lomaiviticin biosynthesis.



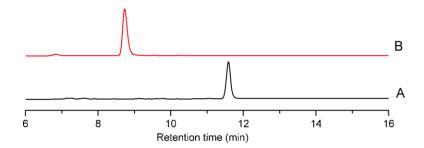
Supplementary Figure 4. PCR confirmation of alp1U double-deletion mutant. Lane 1, DNA ladder; lane 2, $\Delta\Delta alp1U$ giving a band 631 bp. lane 3, the starting strain giving a band corresponding to a fragment of 1,204 bp in length. Both detected bands were correct in size.



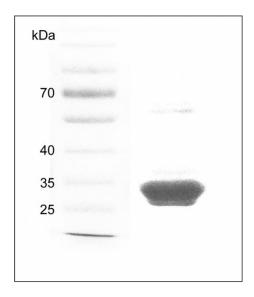
Supplementary Figure 5. SDS-PAGE analysis of the purified Alp1U. Lane 1, protein marker; lane 2, Alp1U giving a band corresponding to a protein of 37 kDa. The detected band was correct in size.



Supplementary Figure 6. LC-MS spectrum of kinamycin E (**5**) under ESI negative mode.

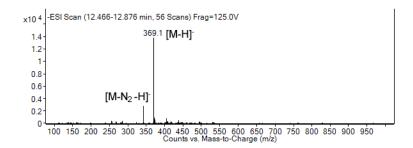


Supplementary Figure 7. HPLC analysis of kinamycin E (**5**) dissolved in (CD₃)₂SO. (A) before NMR measurement; (B) after NMR measurement.

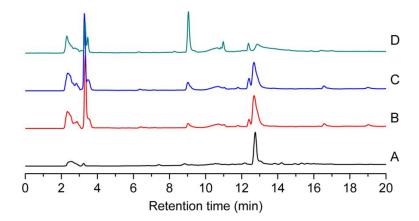


Supplementary Figure 8. SDS-PAGE analysis of the purified Lom6. Lane 1, protein marker; lane 2,

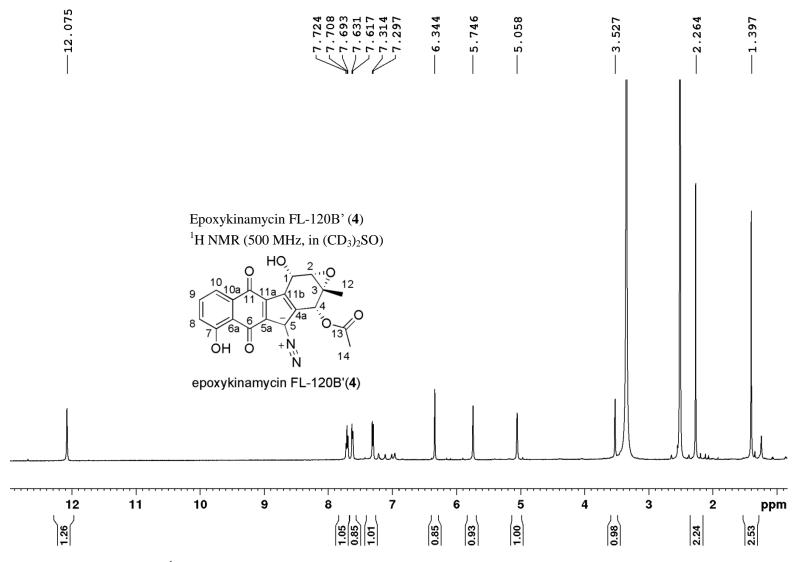
Lom6 giving a band corresponding to a protein of 30 kDa. The detected band was correct in size.



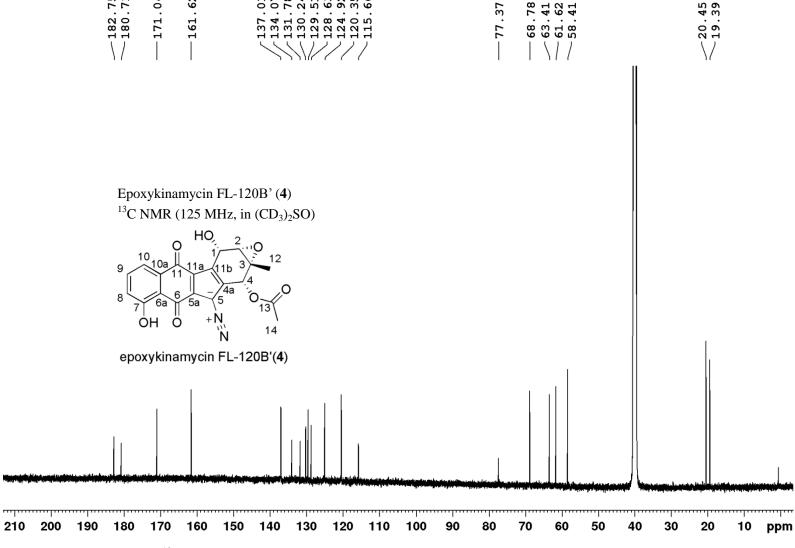
Supplementary Figure 9. LC-MS spectrum of kinamycin F (2) under ESI negative mode.



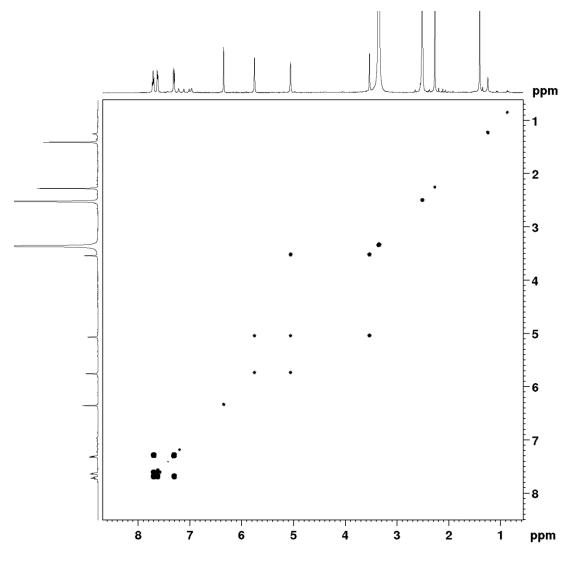
Supplementary Figure 10. HPLC profiles of Lom6 reaction with longer incubation time. (A) standard epoxykinamycin (1); (B) 10 min; (C) 20 min; (D) 50 min



Supplementary Figure 11. ¹H NMR spectrum for epoxykinamycin FL-120B' (4).



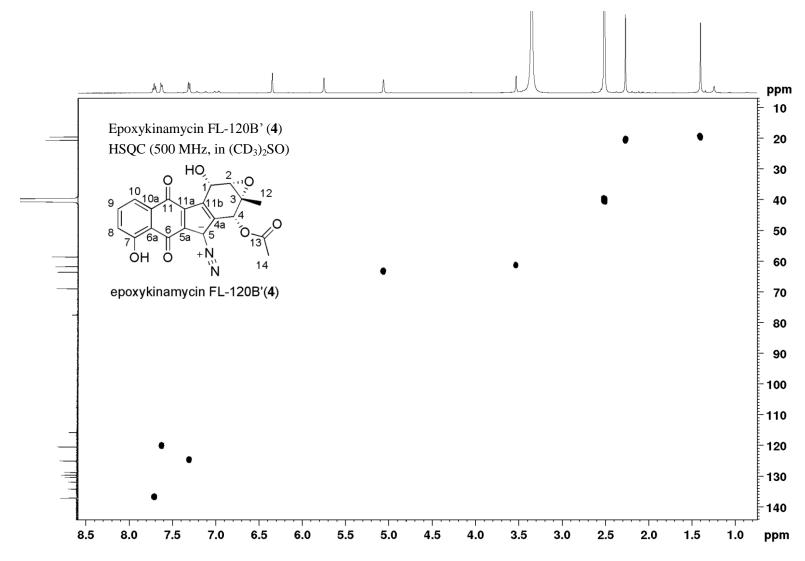
Supplementary Figure 12. ¹³C NMR spectrum for epoxykinamycin FL-120B' (4).



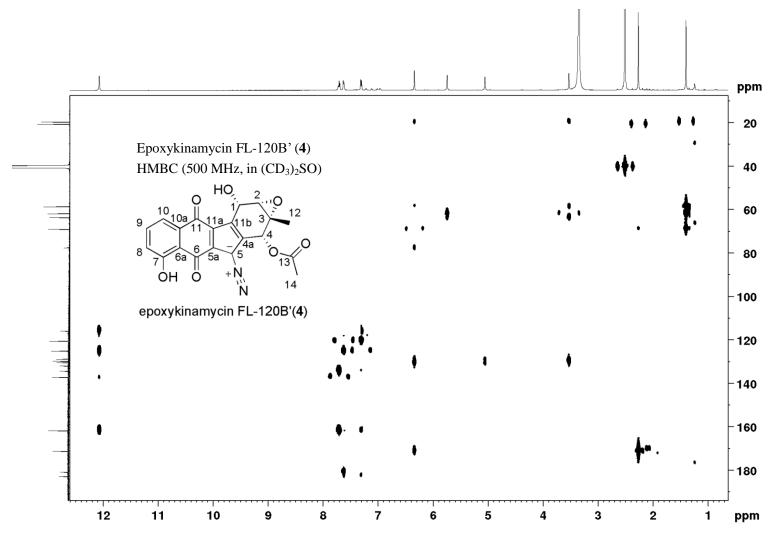
Supplementary Figure 13. COSY spectrum for epoxykinamycin FL-120B' (4).

Epoxykinamycin FL-120B' (4) COSY (500 MHz, in (CD₃)₂SO)

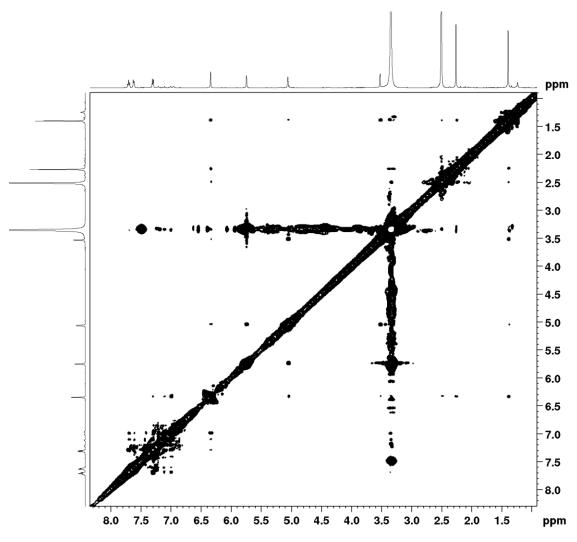
epoxykinamycin FL-120B'(4)



Supplementary Figure 14. HSQC spectrum for epoxykinamycin FL-120B' (4).



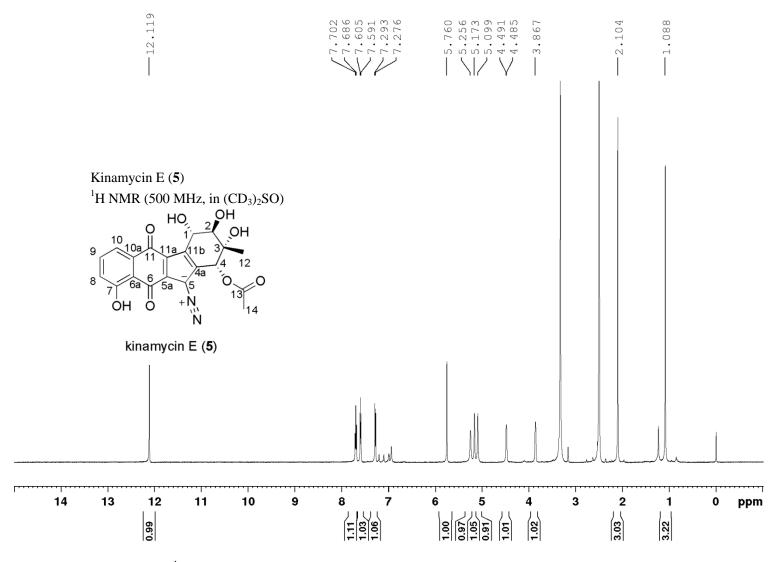
Supplementary Figure 15. HMBC spectrum for epoxykinamycin FL-120B' (4).



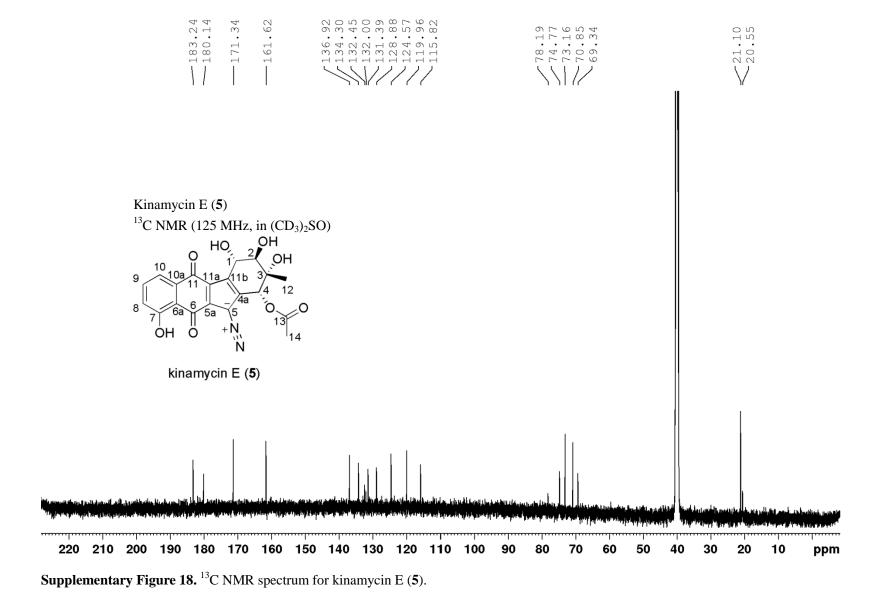
Supplementary Figure 16. NOESY spectrum for epoxykinamycin FL-120B' (4).

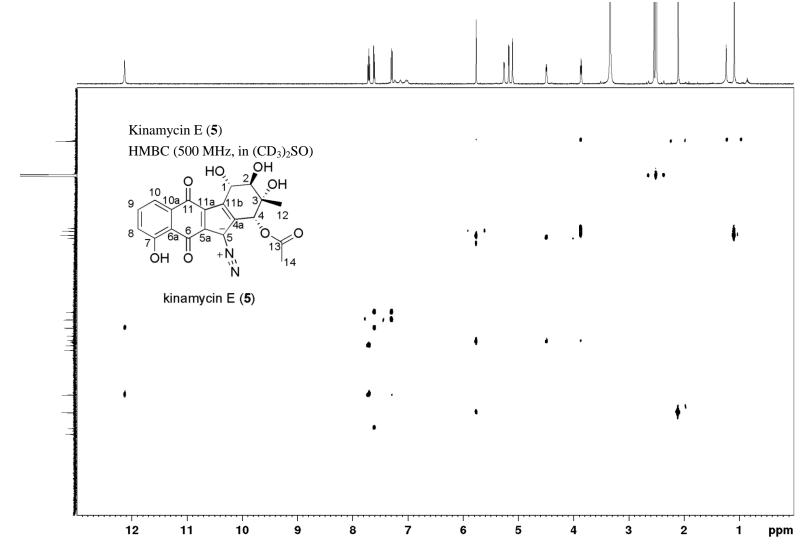
Epoxykinamycin FL-120B' (4) NOESY (500 MHz, in (CD₃)₂SO)

epoxykinamycin FL-120B'(4)

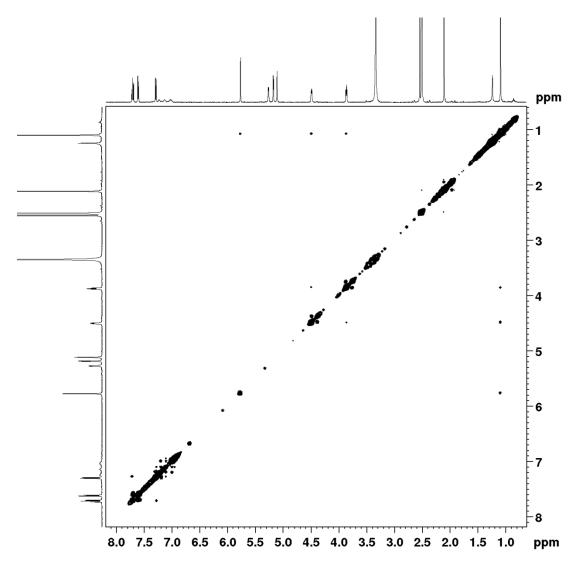


Supplementary Figure 17. ¹H NMR spectrum for kinamycin E (**5**).





Supplementary Figure 19. HMBC spectrum for kinamycin E (**5**).



Supplementary Figure 20. NOESY spectrum for kinamycin E (5).

Kinamycin E (**5**) NOESY (500 MHz, in (CD₃)₂SO)

kinamycin E (5)

Supplementary Table 1. Predicted functions of the gene products in the *alp* cluster.

gene	predicted function	note	
alpZ	gamma-butyrolactone binding protein		
alpY	hypothetical protein		
alpX	putative carboxyl transferase		
alpW	TetR-family transcriptional regulator		
alpV	SARP family pathway specific regulator		
alp U	SARP family pathway specific regulator		
alpT	SARP family pathway specific regulator		
alpS	putative thioesterase		
alpR	putative polyketide synthase		
alpQ	putative chain length factor protein hypothetical protein		
alpP2			
alpP	putative cyclase		
alpO	putative acyl-CoA dehydrogenase		
alpN	putative phosphopantetheinyl transferase	the previously proposed <i>alp</i> cluster	
alpM	peptide synthase condensation domain	the previously proposed aip cluster	
alpL2	conserved hypothetical protein		
alpL	JadX-like protein		
alpK	oxygenase		
alpJ	oxidase		
alpI	polyketide cyclase		
alpA	ketoacyl synthase		
alpB	chain length determinant		
alpC	acyl carrier protein		
alpD	polyketide ketoreductase		
alpE	cyclase/dehydratase		
alpF	oxygenase		
alpG	oxygenase		
alpH	putative O-methyltransferase		

Continued

gene	predicted function note		
alp1A	conserved hypothetical protein		
alp1B	putative myo-inositol phosphate synthase		
alp1C	putative transferase		
alp1D	putative abasic site repairing enzyme	detoxification and DNA	
alp1E	conserved hypothetical protein damage-repair putative metal-dependent hydrolase conserved hypothetical protein		
alp1F			
alp1G			
alp1H	putative type I phosphodiesterase		
alp1I	putative ferredoxin		
alp1J	hypothetical protein	diazo assembly machinery	region 1
alp1K	putative amidase		
alp1L	putative glutamine synthetase		
alp1M	putative adenylosuccinate lyase		region i
alp1N	putative acetyltransferase		
alp1O	two-component system regulator		
alp1P	putative efflux protein	export and regulation	
alp1Q	conserved hypothetical protein		
alp1R	putative antibiotic antiporter		
alp1S	putative oxygenase		
alp1T	NDP-sugar epimerase	A-ring transformation	
alp1U	epoxy hydrolase		
alp1V	putative oxygenase		
alp1W	putative glutamine amidotransferase diazo assemb		
alp1X	putative transcriptional regulator	machinery	

Supplementary Table 2. The NMR data of epoxykinamycin FL-120B' (4)

Position	¹³ C-NMR ^a	¹ H-NMR ^b	$HMBC^b$	NOE	SY
1	63.4	5.06 (s, 1H)	C-4a	H-2,	H-4,
				H_3-12	
1-OH		5.69 (s, 1H)	C-1, C-2		
2	61.6	3.52 (t, 1H, J = 6.0 Hz)	C-1, C-3, C-11b, C-12	H_3-12	
3	58.4				
4	68.8	6.34 (s, 1H)	C-3, C-5, C-11b, C-12	H_3-12	
4a	129.5				
5	77.4				
5a	128.6				
6	182.8				
6a	115.6				
7	161.6				
7-OH		12.07 (s, 1H)	C-6a, C-7, C-8		
8	124.9	7.31 (d, 1H, J = 7.5 Hz)	C-6a, C-7, C-10		
9	137.0	7.72 (t, 1H, $J = 7.5$ Hz)	C-7, C-10a		
10	120.3	7.63(d, 1H, J = 7.5 Hz)	C-6a, C-8		
10a	134.1				
11	180.7				
11a	131.8				
11b	130.2				
12	19.4	1.40 (s, 3H)	C-2, C-3, C-4		
13	171.0				
14	20.4	2.26 (s, 3H)	C-13		

^a Recorded at 125 MHz in (CD₃)₂SO. ^b Recorded at 500 MHz in (CD₃)₂SO.

Supplementary Table 3. The NMR data of kinamycin E (**5**)

Position	¹³ C-NMR ^a	¹ H-NMR ^b	$HMBC^b$	NOESY
1	69.3	4.50 (dd, 1H, J = 3.5, 6.0 Hz)	C-3, C-4a	H-2, H-12
1-OH		5.18 (d, 1H, J = 3.5 Hz)		
2	74.8	3.87 (t, 1H, J = 6.0 Hz)	C-1, C-3, C-11b, C-12	H-1
2-OH		5.26 (d, 1H, J = 6.0 Hz)		
3	73.2			
3-OH		5.10 (s, 1H)		
4	70.8	5.76 (s, 1H)	C-2, C-5, C-11b, C-12	H-12
4a	132.4			
5	78.2			
5a	128.9			
6	183.2			
6a	115.8			
7	161.6			
7-OH		12.11 (s, 1H)	C-6a, C-7, C-8	
8	124.6	7.29 (d, 1H, J = 8.5 Hz)	C-6a, C-7, C-10	
9	136.9	7.70 (t, 1H, J = 8.5 Hz)	C-7, C-10a	
10	119.9	7.61(d, 1H, J = 8.5 Hz)	C-6a, C-8	
10a	134.3			
11	180.2			
11a	131.4			
11b	132.0			
12	20.6	1.09 (s, 3H)	C-2, C-3, C-4	
13	171.3			
14	21.1	2.10 (s, 3H)	C-13	

^a Recorded at 125 MHz in (CD₃)₂SO. ^b Recorded at 500 MHz in (CD₃)₂SO.

Supplementary Table 4. List of primers used in this study.

Primers	Restriction sites	Sequences
1U-upF	HindIII	5' CCCAAGCTTGTGCAGATCCCCGACGCC 3'
1U-upR	XbaI	5' GCTCTAGAGGCCAGTTCCCGGTCCGA 3'
1U-dnF	XbaI	5' GCTCTAGATACGCCAACGCCTACAACAGC 3'
1U-dnR	EcoRI	5' GGAATTCCCCATGTCGTAGCGGATGTC 3'
CK-1U-F		5' CGAGGACATGGGTAGATGACGC 3'
CK-1U-R		5' GGCCGGCAGTTTGTCGTAGA 3'
1U-F	NdeI	5' GGAATTCCATATGCATGGGTAGATGACGCTGAAC 3'
1U-R	EcoRI	5' GGAATTCGAGGTGGCGTCAGCCGA 3'
Ealp1U-F	NdeI	5' GGAATTCCATATGCTCGCCCTGGGCACGG 3'
Ealp1U-R	HindIII	5' CCCAAGCTTGGAGGTGGCGTCAGCCGAAGAAG 3'