

**Supplementary Figure 1. Gating strategy for flow cytometry analysis of mouse aorta.** Cell suspensions from mouse aorta digested with enzyme cocktail were stained with propidium iodide (PI), anti-CD45 (FITC), anti-CD11b (PerCy5.5), anti-CD19 (APC.Cy7), and anti-CD11c (PE), anti-CD3 (APC), and anti-NK1.1(PE.Cy7) antibodies; numbers in the gate in the top two panels represent the percentage of PI<sup>-</sup> and CD45<sup>+</sup> cells of all analyzed cells. The numbers in the bottom five panels represent the percentages of CD11b<sup>+</sup>, CD19<sup>+</sup>, CD11c<sup>+</sup>, CD3<sup>+</sup> and NK1.1<sup>+</sup>cells among all PI<sup>-</sup> CD45<sup>+</sup> cells.









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**Supplementary Figure 2. Macrophage area, necrotic core area and fibrous cap thickness.** (a) Cross sections from the aortic roots of ApoE<sup>-/-</sup> and ApoE<sup>-/-</sup>,  $\beta$ -D849V mice after 8 weeks of WD were stained with MOMA2 antibody(green). (b) Macrophage staining was quantified from ApoE<sup>-/-</sup> and ApoE<sup>-/-</sup>,  $\beta$ -D849V mice in (a) and expressed as percent of total plaque area (n=6). (c) Representative images of aortic root cross sections from ApoE<sup>-/-</sup> and ApoE<sup>-/-</sup>,  $\beta$ -D849V mice after 16 weeks of WD stained with H&E. Blue lines show the boundary of the necrotic core. (d) Quantification of necrotic core size in aortic roots from mice in (c) (n=7). (e) Representative sections of aortic roots from ApoE<sup>-/-</sup>,  $\beta$ -D849V mice after 16 weeks of WD stained with H&E. Bracketed regions show a representative measurement of the fibrous cap thickness.(f) Quantitative analysis of fibrous cap thickness in (e) (n=7), All data were assessed using Student's *t*-test and are present as mean ± s.e.m. \**P*<0.01. Scale bar, 100µm.



Supplementary Figure 3. Expression of M1 and M2 macrophage markers. (a) RNA was extracted from aortic arches of ApoE<sup>-/-</sup> and ApoE<sup>-/-</sup>,  $\beta$ -D849V mice after 16 weeks of WD. mRNA levels of a common macrophage marker CD68 were measured and normalized to GAPDH by qRT-PCR(n=3). (b) Expression of M1 macrophage markers (MCP-1, IL-6) and M2 macrophage markers (CD163, CD204) in aortic arches of ApoE<sup>-/-</sup> and ApoE<sup>-/-</sup>,  $\beta$ -D849V mice after 16 weeks of WD was quantified and normalized to CD68 expression by qRT-PCR(n=3). \**P*<0.05 by Student's *t*-test. All data represent mean ± s.e.m.



Supplementary Figure 4. F4/80 and MOMA2 staining of macrophages in the descending aorta. Consecutive cross sections from ApoE<sup>-/-</sup>,  $\beta$ -D849V mice after 8 weeks of WD were stained with F4/80 antibody (red) and MOMA2 antibody (green). Scale bar, 100µm.



Supplementary Figure 5. Survival curve for ApoE<sup>-/-</sup> and ApoE<sup>-/-</sup>,  $\beta$ -D849V mice on WD. ApoE<sup>-/-</sup> and ApoE<sup>-/-</sup>,  $\beta$ -D849V mice were fed WD at 4 weeks of age for 24 weeks. ApoE<sup>-/-</sup>,  $\beta$ -D849V mice show significantly decreased survival on WD.





















Supplementary Figure 6. Full scans of Western blots in the main text.

| Name   | Forward                  | Reverse                  |
|--------|--------------------------|--------------------------|
| IL-1β  | ATGGGCAACCACTTACCTATTT   | GTTCTAGAGAGTGCTGCCTAATG  |
| TNF-α  | CTGAGTTCTGCAAAGGGAGAG    | CCTCAGGGAAGAATCTGGAAAG   |
| IFN- α | GATGCCCAGCAGATCAAGAA     | CATGCAGCAGATGAGTCCTT     |
| IFN-β  | ATCCAAGAGATGCTCCAGAATG   | CCAGGAGACGTACAACAATAGTC  |
| IFN-γ  | CTCTTCCTCATGGCTGTTTCT    | TTCTTCCACATCTATGCCACTT   |
| CCL2   | AGCACCAGCCAACTCTCACT     | CGTTAACTGCATCTGGCTGA     |
| CCL3   | ATACAAGCAGCAGCGAGTACCAGT | ACAGAGAAGAACAGCAAGGGCAGT |
| CCL5   | TCGTGCCCACGTCAAGGAGTATTT | TCTTCTCTGGGTTGGCACACACTT |
| CCL6   | AAATACCAGGGCAGGCCAGAGAAT | ACCCAAGGACAGCCACAAGGATAA |
| CCL7   | ACCAACCTAGGAGCCAAGAAGCAA | AAGACCATTCCTTAGGCGTGACCA |
| CCL9   | GCCGGGCATCATCTTTATCAGCAA | TGGCAGTTCACACCCTTCTCTCA  |
| CCL12  | ACCATCAGTCCTCAGGTATTGGCT | ACTGGCTGCTTGTGATTCTCCTGT |
| CXCL9  | ATCTTCCTGGAGCAGTGTGGAGTT | AGGGATTTGTAGTGGATCGTGCCT |
| CXCL10 | ATATCGATGACGGGCCAGTGAGAA | AATGATCTCAACACGTGGGCAGGA |
| CXCL11 | AACGGCTGCGACAAAGTTGAAGTG | TATGAGGCGAGCTTGCTTGGATCT |
| Col1a2 | TCTCCTGGAAATGTTGGCCCATCT | AATCCGATGTTGCCAGCTTCACCT |
| Col3a1 | AGGTGGACCAGGCAATGATGGAAA | TTCCTTTAGGACCAGGGAAACCCA |
| Tnc    | TATCGCAACTGGAAGGCCTATGCT | TTGCTCAGGTTATCCAGTCCAAGC |
| Vcan   | TGTGGATCATCTGGATGGCGATGT | CAAAGCCATTTCTCCAAGCTGCCT |
| Bgn    | AGCTCCTCCAGGTTGTCTAT     | CAAGAGGGCCTACTATAATGGC   |
| MCP-1  | CTCACCTGCTGCTACTCATTC    | ACTACAGCTTCTTTGGGACAC    |
| IL-6   | CCAGAGTCCTTCAGAGAGATACA  | CCTTCTGTGACTCCAGCTTATC   |
| CD163  | TGACGACAACTTCAGCAAAGA    | CCAGAACCAGCTCCCAATTTA    |
| CD206  | CCAAATGATGAGCTGTGGATTG   | GCTCTCCAGGAAGCCATTTA     |
| GAPDH  | CTGGAGAAACCTGCCAAGTA     | AAGAGTGGGAGTTGCTGTTG     |

## Supplementary Table1. Primers for Real-Time PCR