

The global variability of diatomaceous earth toxicity: a  
physicochemical and *in vitro* investigation

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**SUPPLEMENTARY MATERIAL**

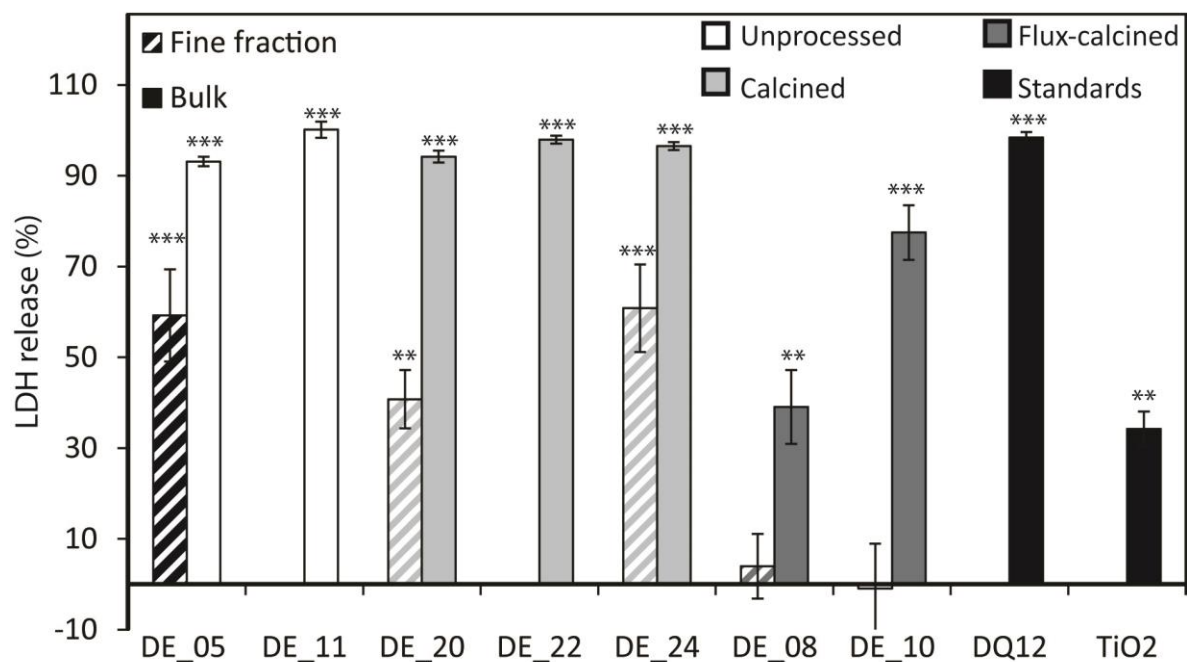
## The toxicity of soluble components of DE

To determine if soluble components of DE were haemolytic, two highly haemolytic DE powders (DE\_05, DE\_11) were leached in saline for one hour at the top treatment concentration (1000  $\mu\text{g}/\text{ml}$ ) by shaking gently, the suspension was then centrifuged at 5000 rcf for 15 minutes and filtered through a 0.2  $\mu\text{m}$  Nalgene syringe filter, and the collected leachates were used in the haemolysis assay. All samples used in the cytotoxicity assays were also leached in complete medium for 24 hours at 63  $\mu\text{g}/\text{ml}$  (in the absence of cells), centrifuged at 5000 rcf for 10 minutes and the collected leachate used to treat the cells, and their toxicity assessed using the alamarBlue<sup>®</sup> assay.

The leachates were not haemolytic, nor cytotoxic in the alamarBlue<sup>®</sup> assay and, therefore, soluble components of the DE were not the cause of the observed haemolytic and cytotoxic responses, which must be particle dependent.

## Results

### Lactate Dehydrogenase release

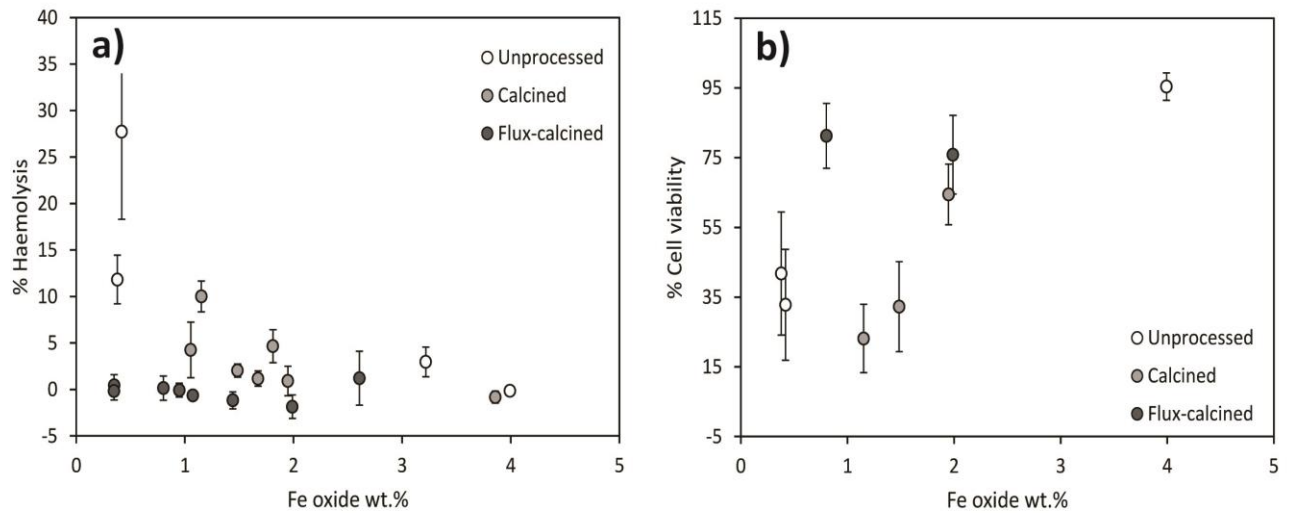


**Figure S1:** Lactate dehydrogenase release from J774 macrophages treated with 500  $\mu\text{g}/\text{ml}$  of the fine fraction (hashed) and bulk (solid) samples of unprocessed (white), calcined (light grey), and flux-calcined (dark grey) DE for 24 h (n=4). Fine fractions were not analysed for DE\_11 or DE\_22. Error bars represent standard error (n=4), \*\*\* p = <0.001 difference from untreated control.

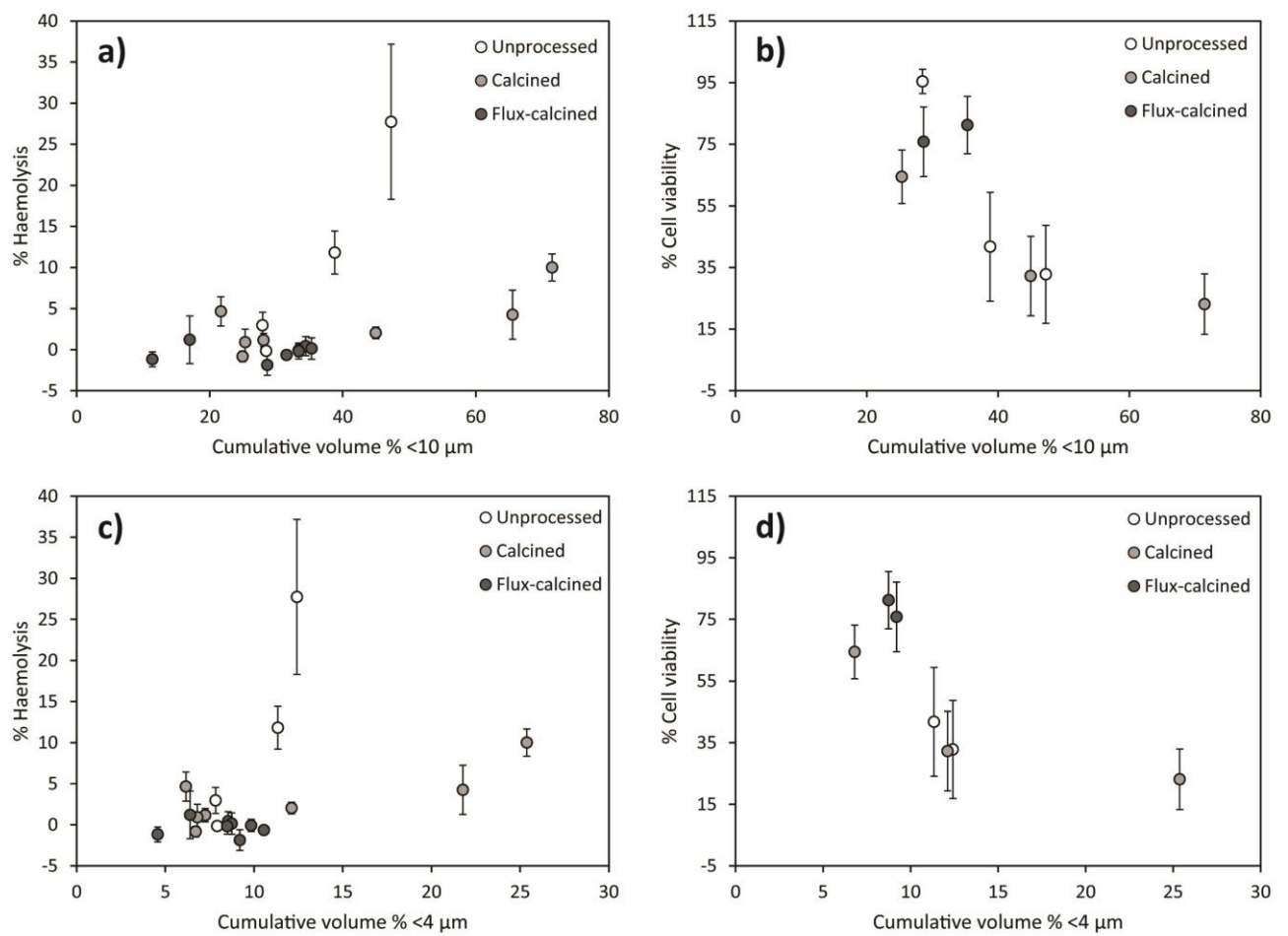
*Cytokine production*

**Table S1:** Cytokine release from J774 macrophages exposed to the separated fine fractions of DE for 24h. None of these cytokines were produced in concentrations significantly greater than the negative control.

Sample	Concentration ( $\mu\text{g/ml}$ )	Cytokine release (pg/ml)		
		KC	IL-1 $\beta$	IL_10
<b>DE_05</b>				
	<b>125</b>	0.28	1.33	0.41
	<b>63</b>	0.22	2.69	0.00
	<b>31</b>	0.21	0.20	0.00
	<b>8</b>	0.40	0.18	0.04
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<b>DE_20</b>				
	<b>125</b>	0.88	0.47	0.00
	<b>63</b>	0.22	7.54	0.64
	<b>31</b>	0.21	9.55	0.00
	<b>8</b>	1.08	7.82	3.79
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<b>DE_24</b>				
	<b>125</b>	0.26	4.96	0.21
	<b>63</b>	0.22	20.17	0.00
	<b>31</b>	0.25	40.87	0.00
	<b>8</b>	0.42	25.30	0.00
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<b>DE_08</b>				
	<b>125</b>	0.42	2.58	0.00
	<b>63</b>			
	<b>31</b>			
	<b>8</b>			
<hr/>				
<b>DE_10</b>				
	<b>125</b>	0.75	16.56	1.90
	<b>63</b>			
	<b>31</b>			
	<b>8</b>			
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<b>DQ12</b>				
	<b>125</b>	0.32	2.97	0.00
	<b>63</b>	0.50	13.68	0.00
	<b>31</b>	0.43	1.49	0.00
	<b>8</b>			
<hr/>				
<b>Untreated control</b>				
		0.27	13.65	0.07



**Figure S2:** Correlation between bulk iron oxide content and **a)** haemolysis post-exposure to 1 mg/ml bulk DE, and **b)** cell viability post-exposure to 500  $\mu$ g/ml bulk DE.



**Figure S3:** Correlation between cumulative volume % particle size <10  $\mu$ m and **a)** haemolysis post-exposure to 1 mg/ml bulk DE, and **b)** cell viability post-exposure to 500  $\mu$ g/ml bulk DE, and between cumulative volume % <4  $\mu$ m **c)** haemolysis, and **d)** cell viability.