

Physiology, genomics, and pathway engineering of an ethanol-tolerant strain of  
*Clostridium phytofermentans*

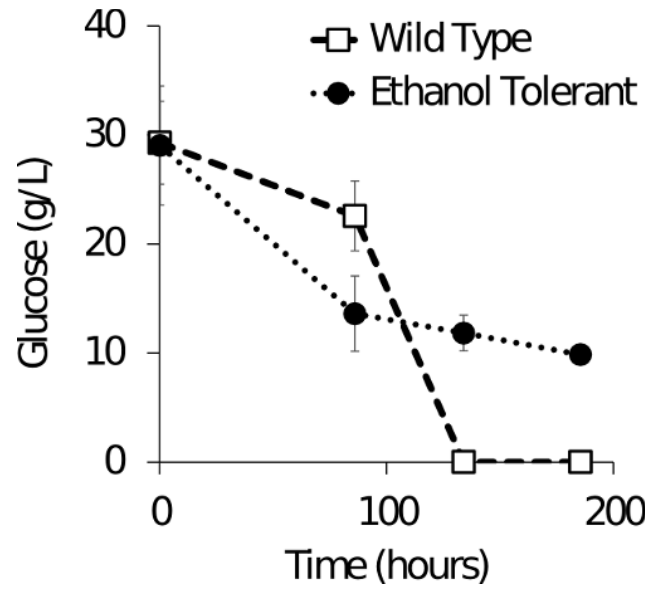
**Supplementary information**

**Page 2: Fig S1** Glucose consumption by *C. phytofermentans* WT and ET strains.

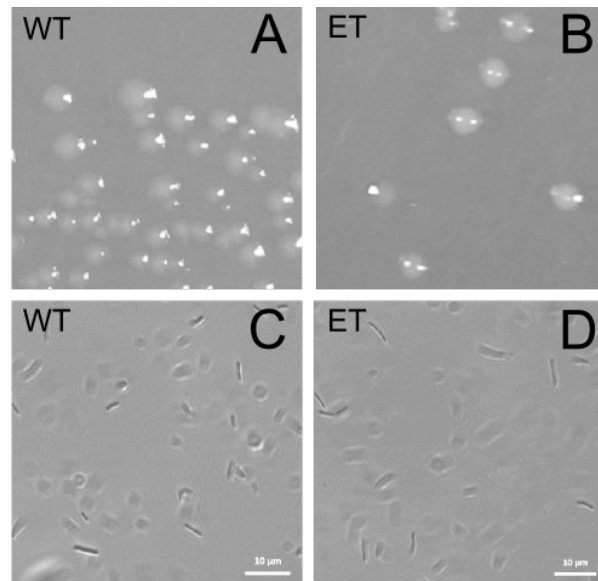
**Page 3: Fig S2** *C. phytofermentans* WT and ET strain morphologies.

**Page 4: Table S1** Generation times (hours) and maximum cell densities (OD<sub>600</sub>) of *C. phytofermentans* wild-type (WT) and ethanol-adapted (ET) strains grown in GS2 medium supplemented with different ethanol concentrations.

**Page 5-6: Supplementary Text** Analysis of the genome sequence of the *C. phytofermentans* ethanol tolerant (ET) strain



**Fig S1** *C. phytofermentans* ET strain shows reduced glucose consumption relative to wild-type when growing in GS2 medium initially containing 30 g/l glucose. Data points are averages of 4 independent cultures and error bars are 1 standard deviation.



**Fig S2** *C. phytofermentans* WT and ET strains have similar morphologies for **A-B** colonies growing on GS2 agar plates and **C-D** cells visualized by bright field microscopy using a Zeiss AxioScope-A1 with a Zeiss EC Plan-173 NEOFLUAR 100x/1.3 oil immersion objective. Scale bar is 10 micrometers.

**Table S1** Generation times (hours) and maximum cell densities (OD<sub>600</sub>) of *C. phytofermentans* wild-type (WT) and ethanol-adapted (ET) strains grown in microtiter plates at 30°C in GS2 medium supplemented with 0, 2, 4, 6, 7% (v/v) ethanol.

| Strain | Ethanol added (v/v) | Generation time (h) | Maximum cell density (OD600) |                    |
|--------|---------------------|---------------------|------------------------------|--------------------|
|        |                     |                     | Mean                         | Standard Deviation |
| WT     | 0                   | 2.47                | 0.87                         | 0.02               |
| WT     | 2                   | 2.51                | 0.82                         | 0.06               |
| WT     | 4                   | ND                  | 0.19                         | 0.05               |
| WT     | 6                   | ND                  | 0.08                         | 0.05               |
| WT     | 7                   | ND                  | 0.07                         | 0.04               |
| ET     | 0                   | 4.15                | 0.62                         | 0.04               |
| ET     | 2                   | 4.22                | 0.66                         | 0.05               |
| ET     | 4                   | 6.42                | 0.46                         | 0.04               |
| ET     | 6                   | 5.76                | 0.30                         | 0.06               |
| ET     | 7                   | ND                  | 0.15                         | 0.02               |

**Supplementary text:** Analysis of the genome sequence of the *C. phytofermentans* ethanol tolerant (ET) strain

FASTQ-formatted DNA sequencing files for the *C. phytofermentans* ET genome were submitted to the European Nucleotide Archive under Primary Accession PRJEB7255.

DNA sequencing reads were quality-filtered using Picard tools (<https://github.com/broadinstitute/picard>). Specifically, reads in fastq format were merged into bam files and reverted with RevertSam to restore original qualities from the OQ field to the QUAL field and to remove duplicate read flags and all alignment information. 'MarkIlluminaAdapters' was run to clear any existing adapter-trimming tags for PAIRED\_END adapter sequences. The reads were aligned to the NCBI reference genome NC\_010001.1 ([http://www.ncbi.nlm.nih.gov/nucleotide/NC\\_010001.1](http://www.ncbi.nlm.nih.gov/nucleotide/NC_010001.1)) with bwaMem version 0.7.7 using the -M flag to mark shorter split hits as secondary. After alignment, duplicate records and optical duplicates were flagged using MarkDuplicates.

DNA variants in ET genome were identified using the Broad Institute GATK (<https://www.broadinstitute.org/gatk/>). SNP and indel calling was performed using the GATK version 3.1 UnifiedGenotyper with the following parameters:

```
java -jar GenomeAnalysisTK-3.1-144-g00f68a3.jar -R [fasta file of Clostridium phytofermentans reference genome] -I [bam file of Clostridium phytofermentans ET genome sequence] -T UnifiedGenotyper -ploidy 1 -stand_call_conf 30 -stand_emit_conf 10
```

Raw output produced by the above GATK analysis:

```
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT Cphy_Ethanol_Tolerant
Cphy 144943 . C T 8841 .
AC=1;AF=1.00;AN=1;BaseQRankSum=-0.584;DP=247;DeIs=0.00;FS=2.508;HaplotypeScore=11.9565;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ0=0;MQRankSum=-0.277;QD=28.17;ReadPosRankSum=1.745 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL
1:12,234:247:99:1:1.00:8871,0
Cphy 263809 . T A 5569 .
AC=1;AF=1.00;AN=1;BaseQRankSum=-0.337;DP=249;DeIs=0.00;FS=1.221;HaplotypeScore=12.5372;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ0=0;MQRankSum=0.187;QD=22.37;ReadPosRankSum=-0.005 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL
1:55,192:249:99:1:1.00:5599,0
Cphy 289172 . C T 9246 .
AC=1;AF=1.00;AN=1;BaseQRankSum=-0.107;DP=250;DeIs=0.00;FS=2.586;HaplotypeScore=7.1707;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ0=0;MQRankSum=-0.060;QD=32.72;ReadPosRankSum=1.224 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL
1:11,239:250:99:1:1.00:9276,0
Cphy 333974 . G A 7470 .
AC=1;AF=1.00;AN=1;BaseQRankSum=0.656;DP=209;DeIs=0.00;FS=0.000;HaplotypeScore=7.1964;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ0=0;MQRankSum=-1.343;QD=27.32;ReadPosRankSum=-0.272 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL
1:13,196:209:99:1:1.00:7500,0
Cphy 697063 . C A 8661 .
AC=1;AF=1.00;AN=1;BaseQRankSum=-1.428;DP=248;DeIs=0.00;FS=4.733;HaplotypeScore=12.7853;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ0=0;MQRankSum=-0.991;QD=34.92;ReadPosRankSum=1.131 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL
1:18,230:248:99:1:1.00:8691,0
```

Cphy 1644823 . G T 52 .  
AC=1;AF=1.00;AN=1;DP=18;DeIs=0.00;FS=0.000;HaplotypeScore=0.0000;MLEAC=1;MLEAF=1.00  
;MQ=20.00;MQ0=16;QD=2.89 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL 1:16,2:18:82:1:1.00:82,0

Cphy 3712793 . T C 9011 .  
AC=1;AF=1.00;AN=1;DP=224;DeIs=0.00;FS=0.000;HaplotypeScore=13.5533;MLEAC=1;MLEAF=1.00  
;MQ=60.00;MQ0=0;QD=31.94 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL  
1:0,223:224:99:1:1.00:9041,0

Cphy 3717529 . C T 8214 .  
AC=1;AF=1.00;AN=1;BaseQRankSum=0.774;DP=249;DeIs=0.00;FS=0.000;HaplotypeScore=10.20  
34;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ0=0;MQRankSum=0.596;QD=32.99;ReadPosRankSum=0.780  
GT:AD:DP:GQ:MLPSAC:MLPSAF:PL 1:22,226:249:99:1:1.00:8244,0

Cphy 3960216 . G GTTGCATGAGAAAACCTGCAT 15937.97 .  
AC=1;AF=1.00;AN=1;BaseQRankSum=1.517;DP=235;FS=0.547;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ  
0=0;MQRankSum=-  
0.769;QD=10.17;RPA=1,2;RU=TTGCATGAGAAAACCTGCAT;ReadPosRankSum=5.925;STR  
GT:AD:DP:GQ:MLPSAC:MLPSAF:PL 1:76,112:235:99:1:1.00:15977,0

Cphy 4533687 . C A 8824 .  
AC=1;AF=1.00;AN=1;BaseQRankSum=1.103;DP=249;DeIs=0.00;FS=0.000;HaplotypeScore=5.461  
4;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ0=0;MQRankSum=1.764;QD=29.71;ReadPosRankSum=-0.791  
GT:AD:DP:GQ:MLPSAC:MLPSAF:PL 1:16,233:249:99:1:1.00:8854,0

Cphy 4631034 . TA T 11686.97 .  
AC=1;AF=1.00;AN=1;BaseQRankSum=0.155;DP=249;FS=1.645;MLEAC=1;MLEAF=1.00;MQ=60.00;MQ  
0=0;MQRankSum=0.870;QD=26.93;RPA=7,6;RU=A;ReadPosRankSum=-0.184;STR  
GT:AD:DP:GQ:MLPSAC:MLPSAF:PL 1:6,238:248:99:1:1.00:11726,0

Cphy 4823500 . G A 8929 .  
AC=1;AF=1.00;AN=1;BaseQRankSum=-  
2.003;DP=246;DeIs=0.00;FS=3.092;HaplotypeScore=5.1933;MLEAC=1;MLEAF=1.00;MQ=60.00;M  
Q0=0;MQRankSum=0.412;QD=31.80;ReadPosRankSum=-2.821 GT:AD:DP:GQ:MLPSAC:MLPSAF:PL  
1:8,238:246:99:1:1.00:8959,0