Synthesis of *anti*-inflammatory α-and β-linked acetamidopyranosides as inhibitors of toll-like receptor 4

Benjamin R. Eyer^a, Yukihiro Yamaguchi^b, Feng Zhang^a, Matthew D. Neal^b, Chhinder P. Sodhi^b, Misty Good^b, Maria Branca^b, Thomas Prindle Jr.^b, Peng Lu^b, Jeffrey L. Brodsky^c, David J. Hackam^b, and Peter Wipf^{a,d}*

^aDepartment of Chemistry and Center for Chemical Methodologies and Library Development, University of Pittsburgh, Pittsburgh, PA 15260, USA

^bDivision of Pediatric Surgery, Children's Hospital of Pittsburgh of University of Pittsburgh and Department of Surgery, University of Pittsburgh School of Medicine, Pittsburgh PA 15224, USA

^cDepartment of Biological Sciences, University of Pittsburgh, Pittsburgh, PA 15260, USA

^dDepartment of Bioengineering, University of Pittsburgh, Pittsburgh, PA 15260, USA

General. All glassware was dried in an oven at 140 °C for 2 h prior to use. All air and moisture-sensitive reactions were performed using syring-septum cap techniques under a dry N_2 or Ar atmosphere. 1,4-dioxane was distilled from sodium/benzophenone ketyl and degassed prior to use; aniline was distilled from KOH or CaH and stored over KOH; and toluene was purified by passage through an activated alumina filtration system. DMF was distilled and stored over 4Å molecular sieves. All other materials were obtained from commercial sources and used as received unless otherwise stated.

Reactions were monitored by thin-layer chromatography analysis using pre-coated silica gel 60 F_{254} plates (EMD, 250 µm thickness) and visualization was accomplished with a 254 nm UV light or by staining with a solution of KMnO₄ (1.5 g of KMnO₄, 10 g of K₂CO₃, and 2.5 mL of 5% aq. NaOH in 150 mL of H₂O). Flash chromatography was performed using SiO₂ (Silicycle, Silia-P Flash Silica Gel, 40-63 µm). Concentrating under reduced pressure refers to the use of a rotary evaporator connected to a membrane vacuum pump to remove solvent.

Melting points were determined using a Laboratory Devices Mel-Temp II in open capillary tubes and are uncorrected. Infrared spectra were determined as neat solids on a Smiths Detection IdentifyIR FT-IR spectrometer. Mass spectra were obtained on a Micromass Autospec double focusing instrument. ¹H and ¹³C NMR spectra were obtained on a Bruker Avance 300 MHz or 400 MHz in DMSO-d₆ unless otherwise noted. Chemical shifts (δ) were reported in parts per million with the residual solvent peak used as an internal standard δ ¹H/¹³C (Solvent); 2.50/39.52 (DMSO), 7.16/77.16 (CDCl₃); and are tabulated as follows: chemical shift, multiplicity (s = singlet, bs = broad singlet, d = doublet, t = triplet, app t = apparent triplet, q = quartet, sept = septet, m = multiplet), number of protons, and coupling constant(s). ¹³C NMR spectra were obtained at 75 MHz or 100 MHz using a proton-decoupled pulse sequence and are tabulated by observed peak. CDCl₃ was filtered through dried basic alumina prior to sample preparation.

^{*} Corresponding author. Tel.: +1-412-624-8606; fax: +1-412-624-0787; e-mail: pwipf@pitt.edu

Synthesis Information

2-Acetamido-1,3,4,6-tetra-O-acetyl-B-D-Galactopyranoside (5).¹ D-(+)-Galactosamine hydrochloride (2.00 g, 9.28 mmol) was dissolved in anhydrous pyridine (20 mL), and acetic anhydride (10.5 mL, 111.3 mmol, 12 eq) was added. The reaction mixture was stirred at room temperature until disappearance of starting material, and poured in a beaker with ice-cold water (200 mL). A white solid precipitated was collected by vacuum filtration, washed with ice-cold water and co-evaporated with toluene (3×20 mL) to remove residual water to yield 5 (2.96 g, 82%) as a powdery solid: ¹H NMR (300 MHz, $CDCl_3$ δ 5.73 (d, J = 8.8 Hz, 1 H), 5.46 (d, J = 9.5 Hz, 1 H), 5.40 (d, J = 2.8 Hz, 1 H), 5.12 (dd, J = 3.3, 11.3 Hz, 1 H), 4.52-4.43 (m, 1 H), 4.24-4.11 (m, 2 H), 4.07-4.02 (m, 1 H), 2.20 (s, 3 H), 2.16 (s, 3 H), 2.08 (s, 3 H), 2.05 (s, 3 H), 1.97 (s, 3 H). 2,3-Dihydrooxazole-3,4,6-tri-O-acetyl-a-D-galactopyranoside (6).² A solution of 5 (400 mg, 1.03 mmol) in dichloroethane (28.6 mL) was treated with TMSOTf (0.200 mL, 1.08 mmol) at room temperature, heated at 50 °C for 1 h, cooled, and treated with NEt₃ (0.440 mL, 3.08 mmol). The mixture was stirred at room temperature for 10 min, passed through a short plug of SiO_2 and washed with ethyl acetate (25 mL) and dichloromethane (30 mL). The solvent was evaporated under reduced pressure and the crude oil was purified by chromatography on SiO₂ (100% EtOAc, SiO₂ was base-washed with 1% NEt₃ prior to use) to yield **5** as a clear slightly orange oil (304 mg, 0.923 mmol, 90%): ¹H NMR (400 MHz, CDCl₃) δ 5.98 (d, J = 6.8 Hz, 1 H), 5.45 (d, J = 2.8 Hz, 1 H), 4.90 (dd, J = 3.2, 7.2Hz, 1 H), 4.26-4.16 (m, 2 H), 4.10 (dd, J = 5.6, 11.2 Hz, 1 H), 3.98 (td, J = 1.2, 7.6 Hz, 1 H), 2.11 (s, 3 H), 2.06 (s, 6 H), 2.04 (d, J = 1.2 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 170.5, 170.2, 169.8, 166.4, 101.5, 71.8, 69.5, 65.3, 63.6, 61.6, 20.8, 20.7, 20.6, 14.4; HRMS (ESI) m/z calcd for C₁₄H₂₀NO₈ [M+H]⁺ 330.1189, found 330.1187. Isopropyl 3.4.6-tri-O-acetyl-2-(acetylamino)-2-deoxy-B-D-galactopyranoside (7). A solution of 6 (0.133 g, 0.404 mmol) and anhydrous CuCl₂ in anhydrous CHCl₃ (0.76 mL) in a 2-5 mL conical sealed vessel under an atmosphere of Ar was treated with anhydrous 2-propanol (0.130 mL, 1.72 mmol). The reaction mixture was heated at 62 °C for 2 h, cooled to room temperature, diluted with acetone (15 mL) and sat. NaHCO₃ solution (7 mL) filtered through a short plug of Celite[®], and concentrated. The residue was coevaporated with toluene to remove residual water and shaken in CHCl₃ and weakly acidic ion-exchange resin (Amberlite IRC-86, ca 1.5 g). The solution was filtered, the solvent was removed under vacuum, and the residue was purified by chromatography on SiO_2 (75% EtOAc/hexanes) to give 7 (119 mg, 0.305 mmol, 75%) as a colorless solid: $[\alpha]_D$ -14.4 (c 1.0, CH₂Cl₂); Mp 188.5-189.5 °C; IR (ATR) 3264, 2980, 1735, 1648, 1568, 1380, 1256, 1232, 1215, 1124, 1072, 1053, 1023 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 5.57 (d, J = 7.8 Hz, 1 H), 5.42 (dd, J = 3.0, 11.1 Hz, 1 H), 5.35 (app d, J = 3.6 Hz, 1 H), 4.86 (d, J =8.4 Hz, 1 H), 4.16 (dd, *J* = 6.6, 11.4 Hz, 1 H), 4.10 (dd, *J* = 7.2, 11.4 Hz, 1 H), 3.95-3.90 (m, 2 H), 3.77-3.73 (m, 1 H), 2.12 (s, 3 H), 2.03 (s, 3 H), 1.99 (s, 3 H), 1.94 (s, 3 H), 1.23 (d, J = 6.6 Hz, 3 H), 1.13 (d, J = 6.6 Hz, 3 H); ¹³C NMR (150 MHz, CDCl₃) δ 170.5,

¹ (a) Traar, P.; Belaj, F.; Francesconi, K. A. *Aust. J. Chem.* **2004**, *57* (11), 1051-53. (b) Tarasiejska, Z.; Jeanloz, R. W. J. Am. Chem. Soc. **1958**, *80* (23), 6325-27. (c) Deng, S.; Gangadharmath, U.; Chang, C.-W. T. J. Org. Chem. **2006**, *71* (14), 5179-85.

² Matta, K. L.; Johnson, E. A.; Barlow, J. J. *Carbohydr. Res.* **1973**, *26* (1), 215-18.

170.3, 99.4, 72.8, 70.4, 69.6, 66.8, 61.5, 52.4, 23.5, 23.3, 22.0, 20.7 (2 C); HRMS (ESI) m/z calcd for C₁₇H₂₈NO₉ [M+H]⁺ 390.1764, found 390.1774.

Cyclohexyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-\u00b3-D-galactopyranoside (8). A solution of 6 (0.135 g, 0.411 mmol) and anhydrous CuCl₂ (55.0 mg, 0.411 mmol) in anhydrous CHCl₃ (0.68 mL) in a 2-5 mL conical sealed vessel under an atmosphere of Ar was treated with cyclohexanol (0.170 mL, 1.66 mmol). The reaction mixture was heated at 62 °C for 2 h, cooled to room temperature, diluted with ethyl acetate (15 mL), washed with 1 N HCl (2×9 mL), sat. NaHCO₃ solution (1×10 mL), and brine (1×10 mL), dried (MgSO₄), evaporated, and purified by chromatography on SiO₂ (75%) EtOAc/hexanes) to give 8 (141 mg, 0.328 mmol, 80%) as a colorless solid: $[\alpha]_D$ -14.6 (c 1.0, CH₂Cl₂); Mp 164.3-165.3 °C; IR (ATR) 3331, 2936, 2857, 1735, 1661, 1541, 1364, 1251, 1219, 1079, 1031, 984 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.48 (d, J = 8.5 Hz, 1 H), 5.41 (dd, J = 3.5, 11.5 Hz, 1 H), 5.35 (app. d, J = 3.5 Hz, 1 H), 4.89 (d, J = 8.5 Hz, 1 H), 4.17 (dd, J = 6.5, 11.5 Hz, 1 H), 4.10 (dd, J = 7.0, 11.0 Hz, 1 H), 3.91 (app t, J = 10.5Hz, 1 H), 3.80-3.74 (m, 1 H), 3.64-3.59 (m, 1 H), 2.13 (s, 3 H), 2.03 (s, 3 H), 1.99 (s, 3 H), 1.94 (s, 3 H), 1.93-1.17 (m, 10 H); ¹³C NMR (125 MHz, CDCl₃) δ 170.4, 170.3 (3 C), 99.1, 78.0, 70.4, 69.6, 66.8, 61.4, 52.5, 33.3, 31.7, 25.5, 23.9, 23.8, 23.5, 20.7 (2 C); HRMS (ESI) m/z calcd for C₂₀H₃₂NO₉ [M+H]⁺ 430.2077, found 430.2084. Geranyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-B-D-galactopyranoside (9). A solution of 6 (0.120 g, 0.364 mmol) and anhydrous CuCl₂ (49.0 mg, 0.364 mmol) in anhydrous CHCl₃ (0.84 mL) in a 2-5 mL conical sealed vessel under an atmosphere of Ar was treated with geraniol (0.270 mL, 1.47 mmol). The reaction mixture was heated at 62 °C for 2 h, cooled to room temperature, diluted with ethyl acetate (15 mL), washed with 1 N HCl $(2 \times 9 \text{ mL})$, sat. NaHCO₃ solution $(1 \times 10 \text{ mL})$, and brine $(1 \times 10 \text{ mL})$, dried (MgSO₄), evaporated, and purified by chromatography on SiO₂ (75% EtOAc/hexanes) to give 9 (149 mg, 0.308 mmol, 65%) as a colorless solid: $[\alpha]_D$ -17.0 (c 1.0, CH₂Cl₂); Mp 117.5-118.9 °C; IR (ATR) 3279, 2982, 2939, 1737, 1659, 1555, 1536, 1431, 1374, 1241, 1226, 1131, 1064, 1053, 1036, 1012, 997, 958 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 5.41 (d, J = 8.5 Hz, 1 H), 5.36-5.27 (m, 3 H), 5.09-5.06 (m, 1 H), 4.75 (d, J = 8.5 Hz, 1 H),4.29 (dd, J = 6.5, 12.0 Hz, 1 H), 4.23-4.11 (m, 3 H), 4.94-3.88 (m, 2 H), 2.13 (s, 3 H), 2.12-2.00 (m, 4 H), 2.03 (s, 3 H), 1.99 (s, 3 H), 1.94 (s, 3 H), 1.68 (s, 3 H), 1.65 (s, 3 H), 1.60 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 170.4 (2 C), 170.3, 170.2, 142.1, 131.8, 123.8, 119.3, 99.0, 70.6, 69.9, 66.9, 65.2, 61.6, 51.8, 39.6, 26.3, 25.7, 23.5, 20.7 (2 C), 17.7, 16.3; HRMS (ESI) m/z calcd for C₂₄H₃₇NO₉Na [M+H]⁺ 506.2366, found 506.2384. 2-Acetamido-2-deoxy-1,3,4,6-tetra-O-acetyl-β-D-glucopyranose (11).³ Acetic anhydride (7.76 mL, 82.3 mmol) was cooled to 0 °C and treated sequentially over 15 min with Nacetyl-D-glucosamine 10 (0.692 g, 3.13 mmol) and montmorillonite K-10 (2.40 g). The reaction mixture was stirred at room temperature for 24 h, filtered through a pad of Celite[®], and rinsed with methyl acetate (100 mL). The filtrate was concentrated and the resulting orange residue was recrystallized twice from hot methanol. filtered, and the crystals were washed with ice-cold diethyl ether $(3 \times 2 \text{ mL})$ to afford 11 as a colorless crystalline solid (350 mg, 0.900 mmol, 29%): ¹H NMR (400 MHz, CDCl₃) δ 5.69 (d, J = 8.8 Hz, 1 H), 5.41 (d, J = 9.6 Hz, 1 H), 5.17-5.09 (m, 2 H), 4.33-4.25 (m, 2 H), 4.13 (dd,

³ Knapp, S.; Huhn, R. A.; Amorelli, B. Org. Synth. 2007, 84, 68-76.

J = 2.4, 12.6 Hz, 1 H), 3.78 (ddd, *J* = 2.4, 4.4, 9.6 Hz, 1 H), 2.12 (s, 3 H), 2.09 (s, 3 H), 2.05 (s, 3 H), 2.04 (s, 3 H).

2,3-Dihydrooxazole-3,4,6-tri-O-acetyl-a-D-glucopyranoside (12).⁴ A solution of **11** (200 mg, 0.514 mmol) in 1,2-dichloroethane (14.3 mL) in a 50-mL round bottom flask was treated with TMSOTf (0.100 mL, 0.539 mmol). The reaction mixture was stirred at 50 °C for 35 min, cooled, and NEt₃ (0.220 mL, 1.54 mmol) was added. After stirring at room temperature for 10 min, the solution was passed through a short plug of SiO₂ and washed with dichloromethane (25 mL) and ethyl acetate (15 mL). The solvents were removed under reduced pressure and the crude orange oil was purified by chromatography on SiO₂ (100% EtOAc, SiO₂ was base-washed with 1% NEt₃) to give **12** (158 mg, 0.478 mmol, 93%) as a colorless oil: ¹H NMR (400 MHz, CDCl₃) δ 5.97 (d, *J* = 7.6 Hz, 1 H), 5.27 (t, *J* = 2.4 Hz, 1 H), 4.93 (app. d, *J* = 9.2 Hz, 1 H), 4.18-4.13 (m, 3 H), 3.61 (quint., *J* = 4.4 Hz, 1 H), 2.12 (s, 3 H), 2.10 (s, 3 H), 2.09 (d, *J* = 1.6 Hz, 3 H), 2.08 (s, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 170.6, 169.5, 169.2, 166.7, 99.4, 70.4, 68.4, 67.5, 65.0, 63.3, 20.9, 20.8, 20.7, 14.0.

Isopropyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-B-D-glucopyranoside (13).⁵ A solution of 12 (0.142 g, 0.424 mmol) and anhydrous CuCl₂ (58.0 mg, 0.431 mmol) in anhydrous CHCl₃ (0.82 mL) in a 2-5 mL conical sealed vessel under an atmosphere of Ar was treated with anhydrous 2-propanol (0.134 mL, 1.75 mmol). The reaction mixture was heated at 62 °C for 2 h. After cooling to room temperature, the mixture was diluted with acetone (15 mL) and sat. NaHCO₃ solution (7 mL), and filtered through a short plug of Celite[®] with acetone (20 mL). The filtrate was concentrated, co-evaporated with toluene and the crude residue was shaken with CHCl₃ and weakly acidic ion-exchange resin (Amberlite IRC-86, ca 1.5 g). The solution was filtered, the solvent was removed under vacuum, and the residue was purified by chromatography on SiO₂ (75% EtOAc/hexanes) to give **13** (146 mg, 0.375 mmol, 87%) as a colorless solid: ¹H NMR (400 MHz, CDCl₃) δ 5.51 (d, J = 8.4 Hz, 1 H), 5.41 (dd, J = 9.2, 10.4 Hz, 1 H), 5.03, (app. t, J = 10.0 Hz, 1 H), 4.84 (d, J = 8.4 Hz, 1 H), 4.24 (dd, J = 5.2, 12.0 Hz, 1 H), 4.11 (dd, J = 2.4, 12.0 Hz, 1 H), 3.93 (sept, J = 6 Hz, 1 H), 3.73-3.60 (m, 2 H), 2.07 (s, 3 H), 2.02 (s, 3 H), 2.02 (s, 3 H), 1.94 (s, 3 H), 1.22 (d, J = 6.0 Hz, 3 H), 1.13 (d, J = 6.4 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) § 170.8, 170.7, 170.2, 169.5, 99.1, 72.6, 72.1, 71.5, 68.9, 62.3, 55.5, 23.3, 23.2, 21.9, 20.8, 20.7, 20.6; HRMS (+ESI-TOF) calcd for C₁₇H₂₈NO₉ [M+H]⁺ 390.1764, found 390.1742.

Cyclohexyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-\beta-D-glucopyranoside (14).^{5,6} A solution of 12 (0.099 g, 0.301 mmol) and anhydrous CuCl₂ (41.0 mg, 0.301 mmol) in anhydrous CHCl₃ (0.68 mL) in a 2-5 mL conical sealed vessel under an atmosphere of Ar was treated with cyclohexanol (0.130 mL, 1.22 mmol). The reaction mixture was heated at 62 °C for 2.5 h, cooled to room temperature, diluted with ethyl acetate (15 mL), washed with 1 N HCl (2 × 9 mL), sat. NaHCO₃ solution (1 × 10 mL), and brine (1 × 10

⁴ (a) Nakabayashi, S.; Warren, C. D.; Jeanloz, R. W. *Carbohydr. Res.* **1986**, *150* (1), c7-c10. (b) Norberg, O.; Deng, L.; Aastrup, T.; Yan, M.; Ramström, O. *Anal. Chem.* **2010**, *83* (3), 1000-07. (c) Srivastava, V. K. *Carbohydr. Res.* **1982**, *103* (2), 286-92.

⁵ Wittmann, V.; Lennartz, D. Eur. J. Org. Chem. 2002, 2002 (8), 1363-67.

⁶ Iglesias-Guerra, F.; Romero, I.; Alcudia, F.; Vega-Pérez, J. M. *Carbohydr. Res.* **1998**, 308 (1–2), 57-62.

mL), dried (MgSO₄), evaporated, and purified by chromatography on SiO₂ (75%) EtOAc/hexanes) to give 14 (112 mg, 0.260 mmol, 86%) as a colorless solid: ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3) \delta 5.53 \text{ (d}, J = 8.4 \text{ Hz}, 1 \text{ H}), 5.40 \text{ (app. t}, J = 8.9 \text{ Hz}, 1 \text{ H}), 5.04 \text{ (t}, J = 8.9 \text{ Hz}, 1 \text{ H})$ 9.6 Hz, 1 H), 4.86 (d, J = 8.4 Hz, 1 H), 4.26 (dd, J = 4.8, 12.0 Hz, 1 H), 4.10 (dd, J = 2.4, 12.0 Hz, 1 H), 3.72-3.58 (m, 3 H), 2.07 (s, 3 H), 2.02 (s, 3 H), 2.01 (s, 3 H), 1.93 (s, 3 H), 1.92-1.18 (m, 10 H); ¹³C NMR (100 MHz, CDCl₃) δ 170.8, 170.7, 170.1, 169.5, 98.9, 77.7, 72.2, 71.6, 69.0, 62.3, 55.6, 33.3, 31.7, 25.5, 23.8, 23.7, 23.4, 20.8, 20.7 (2 C); HRMS (ESI) m/z calcd for C₂₀H₃₁NO₉Na [M+Na]⁺ 452.1897, found 452.1898. Geranyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-B-D-glucopyranoside (15). A solution of 12 (0.120 g, 0.364 mmol) and anhydrous CuCl₂ (49.0 mg, 0.364 mmol) in anhydrous CHCl₃ (0.84 mL) in a 2-5 mL conical sealed vessel under an atmosphere of Ar was treated with geraniol (0.270 mL, 1.47 mmol). The reaction mixture was heated at 62 °C for 2 h, cooled to room temperature, diluted with ethyl acetate (15 mL), washed with 1 N HCl $(2 \times 9 \text{ mL})$, sat. NaHCO₃ solution $(1 \times 10 \text{ mL})$, and brine $(1 \times 10 \text{ mL})$, dried (MgSO₄), evaporated, and purified by chromatography on SiO₂ (75% EtOAc/hexanes) to give 15 (120 mg, 0.248 mmol, 68%) as a colorless solid: $[\alpha]_D$ -20.9 (c 1.0, CH₂Cl₂); Mp 104.8-105.8 °C; IR (ATR) 3282, 2930, 1743, 1735, 1650, 1569, 1431, 1368, 1223, 1128, 1077, 1029, 975 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.40 (d, J = 8.4 Hz, 1 H), 5.30-5.26 (m, 2 H), 5.08-4.98 (m, 2 H), 4.70 (d, J = 8.4 Hz, 1 H), 4.32-4.12 (m, 4 H), 3.82 (app. q, J= 8.4 Hz, 1 H), 3.69-3.63 (m, 1 H), 2.11-2.02 (m, 4 H), 2.07 (s, 3 H), 2.02 (s, 3 H), 2.02 (s, 3 H), 1.94 (s, 3 H), 1.69 (s, 3 H), 1.66 (s, 3 H), 1.60 (s, 3 H); ¹³C NMR (125 MHz, CDCl₃) & 170.9, 170.7, 170.1, 169.4, 142.2, 131.8, 123.8, 119.2, 98.8, 72.5, 71.9, 68.7, 65.1, 62.3, 54.9, 39.6, 26.3, 25.7, 23.4, 20.7 (2 C), 20.6, 17.7, 16.3; HRMS (ESI) m/z calcd for C₂₄H₃₇NO₉Na [M+Na]⁺ 506.2366, found 506.2360. *Isopropyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-α-D-glucopyranoside* (16, C34).⁷

A solution of **11** (50.0 mg, 0.128 mmol) in a 5% HCl solution in 2-propanol (3.8 mL) was stirred at 65 °C for 1 h. The reaction mixture was evaporated to dryness, and the residue was dissolved in pyridine (0.48 mL) and treated with acetic anhydride (0.146 mL, 1.54 mmol). This mixture was stirred at room temperature for 2 d and then co-evaporated with toluene to give a brown oil which was a 3:1 mixture of α:β-isomers by NMR analysis. Purification by chromatography on SiO₂ (75% EtOAc/hexanes) gave **16** (30.4 mg, 0.0781 mmol, 61%) as a foaming colorless solid: IR (ATR) 3359, 2975, 1750, 1730, 1676, 1534, 1437, 1376, 1364, 1243, 1225, 1150, 1122, 1038, 1031, 997 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.62 (d, *J* = 9.6 Hz, 1 H), 5.19 (app t, *J* = 9.6 Hz, 1 H), 5.10 (app t, *J* = 9.6 Hz, 1 H), 4.08 (dd, *J* = 2.4, 12.4 Hz, 1 H), 4.03-3.99 (m, 1 H), 3.88 (sept, *J* = 6.0 Hz, 1 H), 2.07 (s, 3 H), 2.02 (s, 3 H), 2.01 (s, 3 H), 1.93 (s, 3 H), 1.23 (d, *J* = 6.4 Hz, 3 H), 1.14 (d, *J* = 6.4 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 171.4, 170.7, 169.8, 169.3, 95.7, 71.4, 71.0, 68.2, 67.7, 62.0, 51.8, 23.2, 23.1, 21.6, 20.7 (2 C), 20.6; HRMS (ESI) *m/z* calcd for C₁₇H₂₈NO₉ [M+H]⁺ 390.1764, found 390.1790.

Isopropyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-a-D-galactopyranoside (17).⁷ A solution of **5** (50.0 mg, 0.128 mmol) in a 5% HCl solution in 2-propanol (3.8 mL) was stirred at 65 °C for 1 h. The reaction mixture was evaporated to dryness, and the residue was dissolved in pyridine (0.50 mL) and treated with acetic anhydride (0.146 mL, 1.54

⁷ Lemieux, R. U.; James, K.; Nagabhushan, T. L.; Ito, Y. Can. J. Chem. 1973, 51, 33-41.

mmol). This mixture was stirred at room temperature for 2 d and then co-evaporated with toluene to give a brown oil that was a 3:1 mixture of α:β-isomers by NMR analysis. Purification by chromatography on SiO₂ (75% EtOAc/hexanes) gave **17** (28.6 mg, 0.0737 mmol, 57%) as a foaming colorless solid: IR (ATR) 3310, 2973, 2930, 1745, 1732, 1653, 1534, 1372, 1284, 1240, 1217, 1124, 1083, 1034, 992, 936, 878 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.54 (d, J = 9.6 Hz, 1 H), 5.35 (app. d, J = 2.8 Hz, 1 H), 5.13 (dd, J = 3.2, 11.2 Hz, 1 H), 4.95 (d, J = 4.0 Hz, 1 H), 4.56-4.50 (m, 1 H), 4.23 (app. t, J = 6.8 Hz, 1 H), 4.12-4.02 (m, 2 H), 3.88 (sept, J = 6.0 Hz, 1 H), 2.15 (s, 3 H), 2.03 (s, 3 H), 1.98 (s, 3 H), 1.95 (s, 3 H), 1.22 (d, J = 6.0 Hz, 3 H), 1.13 (d, J = 6.4 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 170.4 (2 C), 169.9, 96.3, 71.0, 68.5, 67.5, 66.7, 62.0, 47.8, 23.3, 23.1, 21.7, 20.8, 20.7, 20.6; HRMS (ESI) *m/z* calcd for C₁₇H₂₈NO₉ [M+H]⁺ 390.1764, found 390.1776.

Cyclohexyl 3,4,6-tri-O-acetyl-2-(acetylamino)-2-deoxy-a-D-glucopyranoside (18).⁸ A solution of 11 (50.0 mg, 0.128 mmol) in a 5% HCl solution in cyclohexanol (3.1 mL) was stirred at 65 °C for 1 h. The reaction mixture was evaporated to dryness, and the residue was dissolved in pyridine (0.50 mL, anhydrous) and treated with acetic anhydride (0.146 mL, 1.54 mmol). This mixture was stirred at room temperature for 31 h and co-evaporated with toluene to give a brown oil that was a 3:1 mixture of α:β-isomers by NMR analysis. Purification by chromatography on SiO₂ (75% EtOAc/hexanes) gave 18 (31.7 mg, 0.0738 mmol, 57%) as a foaming colorless solid: ¹H-NMR (400 MHz, CDCl₃) δ 5.63 (d, *J* = 9.6 Hz, 1 H), 5.19 (app. t, *J* = 10.2 Hz, 1 H), 5.08 (app. t, *J* = 10.8 Hz, 1 H), 4.96 (d, *J* = 3.6 Hz, 1 H), 4.32-4.26 (m, 1 H), 4.21 (dd, *J* = 4.8, 12.4 Hz, 1 H), 4.08 (dd, *J* = 2.4, 12.4 Hz, 1 H), 3.54 (sept, *J* = 4.0 Hz, 1 H), 2.07 (s, 3 H), 2.01 (s, 3 H), 2.00 (s, 3 H), 1.93 (s, 3 H), 1.91-1.20 (m, 10 H); ¹³C NMR (100 MHz, CDCl₃) δ 171.4, 170.7, 169.8, 169.3, 95.6, 71.4, 68.2, 67.7, 62.0, 51.9, 33.3, 31.5, 25.4, 24.1, 23.8, 23.2, 20.7 (2 C), 20.6; HRMS (ESI) *m/z* calcd for C₂₀H₃₂NO₉ ([M+H]⁺ 430.2077, found 430.2086.

Assay Information

The small intestinal cell line rat-intestinal epithelial cell (IEC)-6 and mouse-RAW 264.7 macrophages were obtained from American Type Culture Collection (ATCC, Manassas, VA) and were cultured as recommended by ATCC. Nfkb-reporter mice (BALB/c-Tg(Rela-luc)31Xen) were purchased from Taconic and used for experiments in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. Lipopolysaccharide (LPS) (Escherichia coli 0111:B4 purified by gel filtration chromatography >99% pure) was obtained from Sigma-Aldrich (St. Louis, MO). For *in vitro* experiments, both IEC6 and Raw 264.7 cells were plated overnight in 12-well plates (BD Biosciences) in growth media and pre-treated with compounds (5µg/mL, 30 min) before 6-hrs treatment with LPS (25 µg/mL for IEC6 cells, 10 ng/mg for Raw 264.7 cells). For *in vivo* experiments, Nfkb-reporter mice were pre-treated with compounds (2.5 mg/kg, 30 min) before 6-hrs treatment with LPS (2.5 mg/kg).

⁸ (a) Sauer, G.; Matsui, M.; Bloch, R.; Liang, J. S.; Fukushima, D. K. *J. Org. Chem.* **1969**, *34*, 3525-30. (b) Kadokawa, J.-i.; Nagaoka, T.; Ebana, J.; Tagaya, H.; Chiba, K. *Carbohydr. Res.* **2000**, *327*, 341-44.

Dosages of compounds and LPS were chosen based on preliminary experiments carried out to evaluate optimal time and dosages. Cells were harvested at the end of treatments for total RNA isolation using RNeasy mini kit (Qiagen) and thoroughly checked for concentration and purity by measuring the OD260 absorbance and OD260/280 ratio (BioTek Epoch Micro-Volume Spectrophotometer System) as well as by agarose gel electrophoresis. A 0.5 μg of total RNA were reverse transcribed using QuantiTect Reverse Transcription Kit (Qiagen) for cDNA synthesis and qRT-PCR assay. Gene specific amplification of transcripts was performed by quantitative PCR using IQ SYBR GRN SUPERMX and CFX96 real-time system (Bio-Rad, Hercules, CA). The primer sequences used for quantification of transcripts using delta delta CT method were: mouse/rat IL6: Fwd 5'-GGCTAAGGACCAAGACCATCCAA-3', Rev 5'-TCTGACCACAGTGAGGAATGTCCA (amplicon size =138bp); mouse IL1β: Fwd 5'-AGTGTGGATCCCAAGCAATACCCA-3', Rev 5'-TGTCCTGACCACTGTTGTTTCCCA-3' (amplicon size 175b); rat IL1β: Fwd 5'-

TAGGAAACAGCAATGGTCGGGACA-3', Rev 5'-

AGACCTGACTTGGCAGAGGACAAA-3' (amplicon size=167bp); mouse/rat TNFα: Fwd 5'- CATCTTCTCAAAATTCGAGTGACAA-3', Rev 5'-

TGGGAGTAGACAAGGTACAACCC-3' (amplicon size=175bp); mouse/rat house keeping gene RPLO: Fwd 5'-GGCGACCTGGAAGTCCAACT-3', Rev 5'-CCATCAGCACCACAGCCTTC-3' (amplicon size = 143bp).