Table S1. Primers used in this work.

Primer	Nucleotide sequence	Application
16s nono left	AGCTTGTTGGTGGGGTAGTG	qRT-PCR control
16s nono r	TCACCGCTACACCAGGAATT	
hrdB rev	CTGGCGGATGCGCTCACGCGTGAC	qRT-PCR control
hrdB for	CTCGCTGGCCAAGCGCTACACCGG	
dbv1 rt rev	GCCTTCGACCGGGTGTTCCTG	qRT-PCR-*dbv1*
dbv1 rt for	TTGCGGATGTCACGTGGCCTG	
dbv3 rt rev	GCCGGCTGGACGTGAAGGTGA	qRT-PCR-*dbv3*
dbv3 rt for	CGACACTCCGCCCAGCAGCAG	
dbv4 rt rev	GCTCCAACCGGCCTCTCACATC	qRT-PCR -*dbv4*
dbv4 rt for	GGGGTGAAACAACAATCTCGGTGA	
dbv6 rt rev	AGCCTGGGCGCCGACGACTAT	qRT-PCR -*dbv6*
dbv6 rt for	GCGCGGGACGGTTTCGATCA	
dbv7 rt rev	GGTGGGGTAGCCGCCGTGGT	qRT-PCR –*dbv7*
dbv7 rt for	GGGTGCTCGCCGTGCTCCTG	
dbv8 rt rev	ATGGGCGACTACAAGGTGAA	qRT-PCR –*dbv8*
dbv8 rt for	AATAGATGTCGGGGATCAGC	
dbv14 rt rev	AGCATGTCGTCGCCGGGATC	qRT-PCR - *dbv14*
dbv14 rt for	GGTGCTGGGCTCGGACAAGTTC	
dbv15 rt rev	GATGAGACTCTTCGGCCGGATGT	qRT-PCR - *dbv15*
dbv15 rt for	CGGCTCCTTCCTCGTGCTCGT	
dbv17 rt rev	TCGGCAGCGCAAATCAGGTGA	qRT-PCR - *dbv17*
dbv17 rt for	CCTCGACACCCCGCAGCTCC	
dbv19 rt rev	GGCAAGTTCACCGCGATCATGG	qRT-PCR - *dbv19*
dbv19 rt for	CGTCGGCGAACAGCAGATCC	
dbv20 rt rev	GCCGCTGATCGAGGAACGCC	qRT-PCR - *dbv20*
dbv20 rt for	CGCCCGAATCGTGTCATGGAA	
dbv22 rt rev	GCAGCAGCAGGCGGCCATTC	qRT-PCR - *dbv22*
dbv22 rt for	CCAGCGTCAGCAGCGATTCCAG	
dbv23 rt rev	CAGGCTCACGTTCGCGATGCT	qRT-PCR - *dbv23*
dbv23 rt for	CGACCGCCCACTTGTTGATCTTCT	
dbv25 rt rev	CGCCTCCCGATCCACCTTCC	qRT-PCR - *dbv25*
dbv25 rt for	GCGCGGCTATCTGGGTCGTG	
dbv28 rt rev	TCCGCTACTGGGGGCACGC	qRT-PCR - *dbv28*
dbv28 rt for	CGGCGGATATGGCGGTAGAGAA	
*dbv29*rt rev	CCTGATCGAGCCGGGCAACAC	qRT-PCR - *dbv29*
dbv29 rt for	CCCGCTGTTCCGCTCGTACCA	
dbv32 rt rev	GCCGGAGTGCACCAGCCAATG	qRT-PCR - *dbv32*
dbv32 rt for	ATGCAACGCGGGGCTGAACG	
dbv34 rt for	CGGCCAGGACCTCAAGGAACG	qRT-PCR - *dbv34*
dbv34 rt for	GGCCACGCACCGATCCAGCT	
dbv36 rt rev	CGGGCAGGACGTGGCAGGAG	qRT-PCR - *dbv36* gene
dbv36 rt for	CGGGCACAGTATGGGCAGTTTG	
dbv37 rt rev	ACCGCCTCCCCACGCTGAAG	qRT-PCR - *dbv37* gene
dbv37 rt rev	CGGGTTGAGCAGGCTGATCGAC	
dbv3-apra for	GCCGGCCGCGGTGGAGTGGGCGTCATCACGGGGCCTGTGATTCCGGGGATCCGTCGACC	To generate the Δ*dbv3* strain
dbv3 apra rev	CCTCACGCGGCTCGCGGCCGGCTGTGGCCGTCGCCTTCTTGTAGGCTGGAGCTGCTTC	
dbv4-apra for	GTGGACCCGACGGGAGTTGACATAGCCACTCTCCCTGTCATTCCGGGGATCCGTCGACC	To generate the Δ*dbv4* strain
dbv4 apra rev	AGCGGCCAGATCGGTCGCCCGCCCCTCCAGGCGATCCGCTGTAGGCTGGAGCTGCTTC	
dbv6-apra for	ATGCGCGTTCTGGTGGTGGAGGACCAAGTCGACCTGGCCATTCCGGGGATCCGTCGACC	To generate the Δ*dbv6* strain
dbv6 apra rev	ATGCGATATCCCTCGCGCGGGACGGTTTCGTCGCCTTCTTGTAGGCTGGAGCTGCTTC	
dbv3 tar for	GATGAGCACGAGTGGATGAG	Δ*dbv3* verification
dbv3 tar rev	TCACAGCAGATTCCGGTACA	
dbv4 tar for	ACTTGGCCGATCGATTTATG	Δ*dbv4* verification
dbv4 tar rev	GAATCGAGCAACCTCGTCAG	
dbv6 tar for	ACGCGAGCTATGGTGTCAG	Δ*dbv6* verification
dbv6 tar rev	GCTTCTCCTCATCCCTCTCC	
dbv3 over for	AAAAAAAACATATGCTGTTCGGGCGAGATCG	To generate a *NdeI-dbv3-BglII* fragment
dbv3 over rev	AAAAAGATCTCTACAGCCGCACTGCCTCAC	

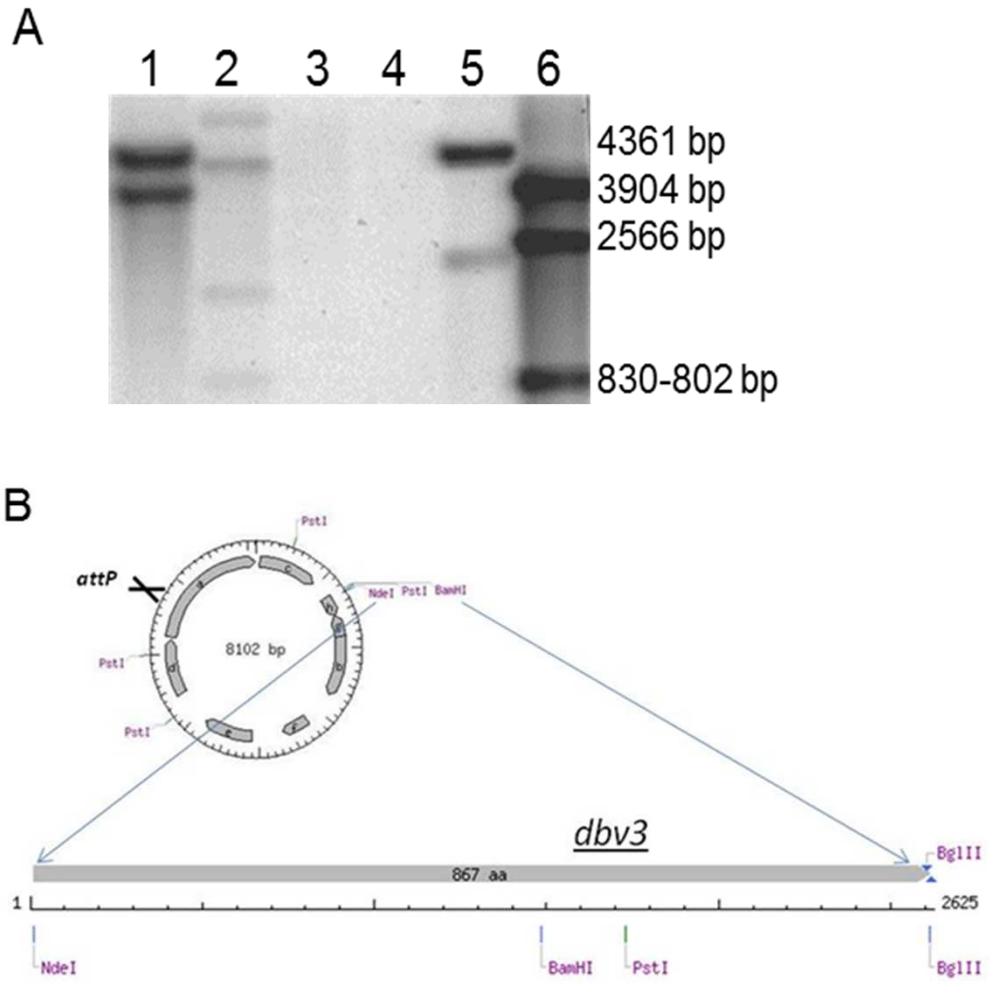

Fig. S1 Integration of pIJ8600-*dbv*3 into the *Nonomuraea* chromosome.

(a) Southern hybridization analysis of *Bam*HI-digested genomic DNA of the Oe-*dbv3* strain (lane 1), PstI-cleaved genomic DNA of the Oe-*dbv3* strain (lane 2), *Bam*HI-digested genomic DNA of the wild type strain (lane 3), *Pst*I-cleaved genomic DNA of the wild type strain (lane 4). *Pst*I-digested pIJ8600 was used as a probe (lane 6). Lane 5: Marker II (Invitrogen).

(b) Schematic representation of the plasmid used to generate the Oe-*dbv3* strain. The *dbv3* gene was ligated to *Nde*I- and *Bam*HI-digested pIJ8600.

*Pst*I, *Bgl*I, *Nde*I and *Bam*HI rectriction sites are indicated.

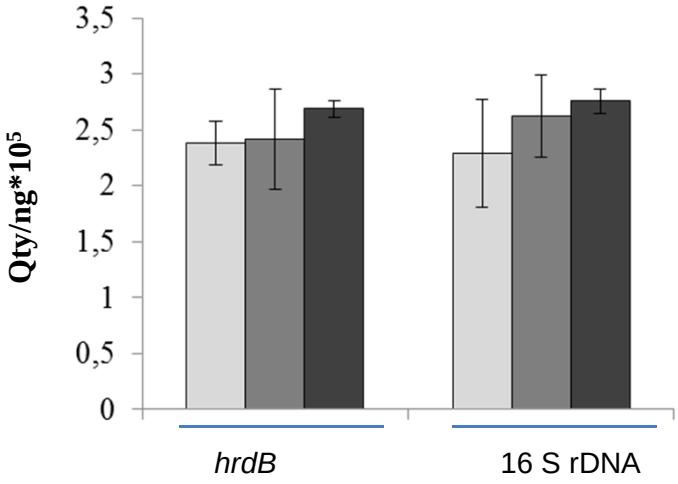

Fig. S2 Absolute Quantitative RT-PCR of *hrdB* and 16S rDNA after 24 (white bars), 48 (gray bars) and 72h (black bars) of *Nonomuraea sp. ATCC39727* grown in R3 medium. Error bars are calculated from three independent experiments.