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Supplementary Figures: 6

Supplementary Tables: 1

Supplementary Videos:

Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read Reporting Life Sciences Research.

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

▶ Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).

- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.

- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.

- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.

- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

	TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
example 1a	one-way ANOVA	Fig. legend	9, 9, 10, 15	mice from at least 3 litters/group	Methods para 8	error bars are mean +/- SEM	Fig. legend	p = 0.044	Fig. legend	$F_{(3, 36)} = 2.97$	Fig. legend
example results, para 6	unpaired t-test	Results para 6	15	slices from 10 mice	Results para 6	error bars are mean +/- SEM	Results para 6	p = 0.0006	Results para 6	$t_{(28)} = 2.808$	Results para 6
+ − 1b	one-way ANOVA	Fig. legend	17	mice from different litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 3.3794e-07	Fig. legend	F = 5.5675	n/a

	FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
		TEST USED			**n**		**DESCRIPTIVE STATS (AVERAGE, VARIANCE)**		**P VALUE**		**DEGREES OF FREEDOM & F/t/z/R/ETC VALUE**	
+ -	1c	one-way ANOVA	Fig. legend	17	mice from >3 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 2.0305e-06	Fig. legend	F = 5.0486	n/a
+ -	1e top	one-way ANOVA	Fig. legend	17	mice from >3 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 3.7932e-06	Fig. legend	F = 4.8838	n/a
+ -	1e bottom	one-way ANOVA	Fig. legend	17	mice from >3 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.0007	Fig. legend	F = 3.1505	n/a
+ -	Fig. 2c,d distal	two-way ANOVA	Fig. legend	7,5	mice from >3 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 1.5584e-09	Fig. legend	F = 45.5973	n/a
+ -	Fig 2c,d basal	two-way ANOVA	Fig. legend	4,5	mice from >2 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.246	Fig. legend	F = 1.37	n/a
+ -	Fig. 2e	one-tailed bootstrap test	Fig. legend	5	mice from >3 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	n/a
+ -	Fig. 2f	one-tailed bootstrap test	Fig. legend	5	mice from >3 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.05	Fig. legend	n/a	n/a
+ -	Fig. 3c	one-way ANOVA	Fig. legend	6	mice from >3 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.6	Fig. legend	F = 0.7624	n/a
+ -	Fig. 3d	two-way ANOVA	Fig. legend	6,5	mice from >2 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 9.7265e-19	Fig. legend	F = 132.6432	n/a
+ -	Fig. 4c	one-way ANOVA	Fig. legend	6	mice from >3 litters	EDF legend	error bars are mean +/- SEM	Fig. legend	p= 0.8	Fig. legend	F = 0.5004	n/a
+ -	Fig. 4d	two-way ANOVA	Fig. legend	6,5	mice from >2 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 1.10595174283291e-06	Fig. legend	F = 27.56	n/a
+ -	Fig. 5d left	one-way ANOVA	Fig. legend	3	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.627	Fig. legend	F = 0.7393	n/a
+ -	Fig. 5d middle	one-way ANOVA	Fig. legend	3	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.9997	Fig. legend	F = 0.0371	n/a
+ -	Fig. 5d right	one-way ANOVA	Fig. legend	3	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 0.9139	Fig. legend	F = 0.3232	n/a
+ -	Fig. 6c	two-way ANOVA	Fig. legend	3,4	mice from >3 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 4.1150e-08	Fig. legend	F = 40.9944	n/a
+ -	Fig. 6c	one-tailed bootstrap test	Fig. legend	3,4	mice from >3 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	n/a
+ -	Fig. 7c	two-way ANOVA	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 8.6135e-28	Fig. legend	F = 171.8326	n/a
+ -	Fig. 7c	two-way ANOVA	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 6.2483e-19	Fig. legend	F = 114.5475	n/a
+ -	Fig. 7e	one-tailed bootstrap test	Fig. legend	5, 12	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	

+ -	Fig. 7e	one-tailed bootstrap test	Fig. legend	5,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.01	Fig. legend	n/a	
+ -	Fig. 7e	one-tailed bootstrap test	Fig. legend	5,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.01	Fig. legend	n/a	
+ -	Fig. 7f	logrank test	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	n/a		p<0.001	Fig. legend	n/a	
+ -	Fig. 7f	logrank test	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	n/a		p<0.001	Fig. legend	n/a	
+ -	Fig. 7g left	one-tailed bootstrap test	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.01	Fig. legend	n/a	
+ -	Fig. 7g left	one-tailed bootstrap test	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.05	Fig. legend	n/a	
+ -	Fig. 7g right	one-tailed bootstrap test	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.05	Fig. legend	n/a	
+ -	Fig. 7g right	one-tailed bootstrap test	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	
+ -	Fig. 8b	one-tailed bootstrap test	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	
+ -	Fig. 8b	one-tailed bootstrap test	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	
+ -	Fig. 8c	one-tailed bootstrap test	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	
+ -	Fig. 8c	one-tailed bootstrap test	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	
+ -	Fig. 8d top	two-way ANOVA	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 4.1697e-08	Fig. legend	F = 32.7603	n/a
+ -	Fig. 8d top	two-way ANOVA	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 1.1432e-06	Fig. legend	F = 25.2351	n/a
+ -	Fig. 8d bottom	two-way ANOVA	Fig. legend	12,5	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 2.6380e-08	Fig. legend	F = 34.1425	n/a
+ -	Fig. 8d bottom	two-way ANOVA	Fig. legend	12,6	mice from >1 litters / group	Fig. legend	error bars are mean +/- SEM	Fig. legend	p = 4.3686e-08	Fig. legend	F = 32.7827	n/a
+ -	Fig. 8e	one-tailed bootstrap test	Fig. legend	5	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	
+ -	Fig. 8e	one-tailed bootstrap test	Fig. legend	5	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.001	Fig. legend	n/a	
+ -	Fig. 8e	one-tailed bootstrap test	Fig. legend	5	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p>0.1	Fig. legend	n/a	
+ -	Fig. 8f	one-tailed bootstrap test	Fig. legend	5	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.05	Fig. legend	n/a	

+ −	Fig. 8f	one-tailed bootstrap test	Fig. legend	5	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p<0.05	Fig. legend	n/a	
+ −	Fig. 8f	one-tailed bootstrap test	Fig. legend	5	mice from >2 litters	Fig. legend	error bars are mean +/- SEM	Fig. legend	p>0.1	Fig. legend	n/a	
+ −	SFig. 2c	one-way ANOVA	SFig. legend	5	mice from >2 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.5177	SFig. legend	F = 0.9274	n/a
+ −	SFig. 2d	one-way ANOVA	SFig. legend	4	mice from >2 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.0633	SFig. legend	F = 2.0196	n/a
+ −	SFig. 5a	one-way ANOVA	SFig. legend	5,5,5	mice from >3 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.06	SFig. legend	F = 3.0187	n/a
+ −	SFig. 5b	chi-square test	SFig. legend	5,5,	mice from >3 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 1.8245e-05	SFig. legend	n/a	
+ −	SFig. 5b	chi-square test	SFig. legend	5,5,	mice from >3 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 8.4314e-05	SFig. legend	n/a	
+ −	SFig. 5b	one-way ANOVA	SFig. legend	5,5,5	mice from >3 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.0044	SFig. legend	F = 5.8049	n/a
+ −	SFig. 6a	one-way ANOVA	SFig. legend	5	mice from >2 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 1.0662e-06	SFig. legend	F = 7.3406	n/a
+ −	SFig. 6a	two-way ANOVA	SFig. legend	5,5	mice from >1 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 7.6037e-16	SFig. legend	F = 92.7820	n/a
+ −	SFig. 6a	two-way ANOVA	SFig. legend	5,12	mice from >1 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.1982	SFig. legend	F = 1.6686	n/a
+ −	SFig. 6a	two-way ANOVA	SFig. legend	5,5	mice from >1 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.24	SFig. legend	F = 1.2921	n/a
+ −	SFig. 6b	one-way ANOVA	SFig. legend	5	mice from >1 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 3.5639e-12	SFig. legend	F = 17.3055	n/a
+ −	SFig. 6b	two-way ANOVA	SFig. legend	5,5	mice from >1 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 3.5539e-19	SFig. legend	F = 124.7922	n/a
+ −	SFig. 6b	two-way ANOVA	SFig. legend	5,5	mice from >1 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.08	SFig. legend	F = 4.2382	n/a
+ −	SFig. 6c	one-way ANOVA	SFig. legend	4	mice from >1 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.9737	SFig. legend	F = 0.3082	n/a
+ −	SFig. 6c	two-way ANOVA	SFig. legend	5,4	mice from >1 litters	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.77	SFig. legend	F = 0.0827	n/a
+ −	SFig. 6d	one-way ANOVA	SFig. legend	5,5,5,4,5,5,	mice from >1 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 1.7098e-09	SFig. legend	F = 30.9934	n/a
+ −	SFig. 6e left	one-way ANOVA	SFig. legend	12,4,5	mice from >1 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.63	SFig. legend	F = 0.4724	SFig. legend
+ −	SFig. 6e right	one-way ANOVA	SFig. legend	12,4,5	mice from >1 litters / group	SFig. legend	error bars are mean +/- SEM	SFig. legend	p = 0.14	SFig. legend	F = 2.3386	SFig. legend

▸ Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

   If so, what figure(s)?

   Representative images are shown in Fig. 2,3,4,6,7, and Supplementary Figure 1, 3,4,

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

   If so, where is this reported (section, paragraph #)?

   > Experimental repetitions are reported and specified for all experiments in the figure legends

▸ Statistics and general methods

1. Is there a justification of the sample size?

   If so, how was it justified?

   Where (section, paragraph #)?

   Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

   > We did not do any statistical tests to predetermine the sample size. Our sample size is similar to other previously reported studies. This is stated in the methods section

2. Are statistical tests justified as appropriate for every figure?

   Where (section, paragraph #)?

   > Yes, we describe this in every figure

   a. If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?

   > Yes, we describe this in every claim

   b. Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?

   Where is this described (section, paragraph #)?

   > Our scoring results in general are normally distributed. This is stated in the methods section

   c. Is there any estimate of variance within each group of data?

   Is the variance similar between groups that are being statistically compared?

   Where is this described (section, paragraph #)?

   > We report the variance within each group of data as the standard error of the mean (SEM) for all samples in all experiments, within the figure panel corresponding to the statistically analyzed data. We do not specify this, but report it as the SEM within the figure panels.

   d. Are tests specified as one- or two-sided?

   > All tests are two-sided

   e. Are there adjustments for multiple comparisons?

   > Yes; for all multiple comparisons we utilized the appropriate Bonferroni correction or post hoc Tukey's test

3. Are criteria for excluding data points reported?

   Was this criterion established prior to data collection?

   Where is this described (section, paragraph #)?

   > We did not exclude any data points

4. Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data.

   If no randomization was used, state so.

   Where does this appear (section, paragraph #)?

   > No specific randomization was used for data collection or analysis, Animals were assigned to particular experimental groups with absolutely no bias and by different experimenters. This is stated in the methods section

5. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included?

   If no blinding was done, state so.

   Where (section, paragraph #)?

> In all the optogenetics experiments, data collection and analysis were performed blind to the conditions of the experiment. For other experiments, the scorer was blinded to the session number of each image, which was randomized. This is stated in the methods section

6. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included?

   Where (section, paragraph #)?

> Yes, all procedures used were in accordance with protocols approved by UCSD institutional Animal Care and Use Committee and guidelines of the National Institute of Health. This is stated in the methods section

7. Is the species of the animals used reported?

   Where (section, paragraph #)?

> Yes, all the experiments were conducted in mice. This is stated in the methods section

8. Is the strain of the animals (including background strains of KO/ transgenic animals used) reported?

   Where (section, paragraph #)?

> Yes, all the transgenic mice are reported. This is stated in the methods section

9. Is the sex of the animals/subjects used reported?

   Where (section, paragraph #)?

> All experiments were conducted in all genders. This is stated in the manuscript

10. Is the age of the animals/subjects reported?

    Where (section, paragraph #)?

> Yes, all experiments were performed on mice over 6 weeks old. This is stated in the figure.

11. For animals housed in a vivarium, is the light/dark cycle reported?

    Where (section, paragraph #)?

> Yes, the light/dark cycle is reported in the methods section

12. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported?

    Where (section, paragraph #)?

> No, the housing group is not reported

13. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)?

    Where (section, paragraph #)?

> Yes, experiments were typically performed during the dark period. This is stated in the methods section

14. Is the previous history of the animals/subjects (e.g. prior drug administration, surgery, behavioral testing) reported?

    Where (section, paragraph #)?

> Yes, time line of surgery and behavioral testing is reported in Figure 2,5,6,8

   a. If multiple behavioral tests were conducted in the same group of animals, is this reported?

      Where (section, paragraph #)?

> N/A

15. If any animals/subjects were excluded from analysis, is this reported?

    Where (section, paragraph #)?

> N/A

a. How were the criteria for exclusion defined?

> N/A

Where is this described (section, paragraph #)?

b. Specify reasons for any discrepancy between the number of animals at the beginning and end of the study.

> N/A

Where is this described (section, paragraph #)?

▶ Reagents

1. Have antibodies been validated for use in the system under study (assay and species)?

> Yes, the antibodies that we used have previously been reported in other papers

a. Is antibody catalog number given?

> Yes, this is stated in the methods section

Where does this appear (section, paragraph #)?

b. Where were the validation data reported (citation, supplementary information, Antibodypedia)?

> The validation data is reported as the catalog number for each antibody in the methods section

Where does this appear (section, paragraph #)?

2. Cell line identity

> N/A

a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by ICLAC and NCBI Biosample?

Where (section, paragraph #)?

b. If yes, include in the Methods section a scientific justification of their use--indicate here in which section and paragraph the justification can be found.

> N/A

c. For each cell line, include in the Methods section a statement that specifies:
    - the source of the cell lines
    - have the cell lines been authenticated? If so, by which method?
    - have the cell lines been tested for mycoplasma contamination?

Where (section, paragraph #)?

> N/A

▶ Data deposition

Data deposition in a public repository is mandatory for:

    a. Protein, DNA and RNA sequences
    b. Macromolecular structures
    c. Crystallographic data for small molecules
    d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available here. We encourage the provision of other source data in supplementary information or in unstructured repositories such as Figshare and Dryad.

We encourage publication of Data Descriptors (see Scientific Data) to maximize data reuse.

1. Are accession codes for deposit dates provided? Where (section, paragraph #)?	N/A

▶ Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

1. Identify all custom software or scripts that were required to conduct the study and where in the procedures each was used.	N/A
2. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.	N/A

▶ Human subjects

1. Which IRB approved the protocol? Where is this stated (section, paragraph #)?	N/A
2. Is demographic information on all subjects provided? Where (section, paragraph #)?	N/A
3. Is the number of human subjects, their age and sex clearly defined? Where (section, paragraph #)?	N/A
4. Are the inclusion and exclusion criteria (if any) clearly specified? Where (section, paragraph #)?	N/A

5. How well were the groups matched?

   Where is this information described (section, paragraph #)?

   N/A

6. Is a statement included confirming that informed consent was obtained from all subjects?

   Where (section, paragraph #)?

   N/A

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?

   Where (section, paragraph #)?

   N/A

▶ fMRI studies

For papers reporting functional imaging (fMRI) results please ensure that these minimal reporting guidelines are met and that all this information is clearly provided in the methods:

1. Were any subjects scanned but then rejected for the analysis after the data was collected?

   N/A

   a. If yes, is the number rejected and reasons for rejection described?

      Where (section, paragraph #)?

      N/A

2. Is the number of blocks, trials or experimental units per session and/or subjects specified?

   Where (section, paragraph #)?

   N/A

3. Is the length of each trial and interval between trials specified?

   N/A

4. Is a blocked, event-related, or mixed design being used? If applicable, please specify the block length or how the event-related or mixed design was optimized.

   N/A

5. Is the task design clearly described?

   Where (section, paragraph #)?

   N/A

6. How was behavioral performance measured?

   N/A

7. Is an ANOVA or factorial design being used?

   N/A

8. For data acquisition, is a whole brain scan used?

   If not, state area of acquisition.

   N/A

   a. How was this region determined?

      N/A

9. Is the field strength (in Tesla) of the MRI system stated?

> N/A

   a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated?

> N/A

   b. Are the field-of-view, matrix size, slice thickness, and TE/TR/ flip angle clearly stated?

> N/A

10. Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated?

> N/A

11. Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)?

> N/A

12. If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)?

> N/A

13. How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.?

> N/A

14. Were any additional regressors (behavioral covariates, motion etc) used?

> N/A

15. Is the contrast construction clearly defined?

> N/A

16. Is a mixed/random effects or fixed inference used?

> N/A

   a. If fixed effects inference used, is this justified?

> N/A

17. Were repeated measures used (multiple measurements per subject)?

> N/A

   a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated?

> N/A

18. If the threshold used for inference and visualization in figures varies, is this clearly stated?

> N/A

19. Are statistical inferences corrected for multiple comparisons?

> N/A

   a. If not, is this labeled as uncorrected?

> N/A

20. Are the results based on an ROI (region of interest) analysis?

N/A

    a. If so, is the rationale clearly described?

N/A

    b. How were the ROI's defined (functional vs anatomical localization)?

N/A

21. Is there correction for multiple comparisons within each voxel?

N/A

22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined?

N/A

▶ Additional comments

Additional Comments