

Characterization of wall teichoic acid degradation by the bacteriophage ϕ 29 appendage protein GP12 using synthetic substrate analogs

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Supplementary Information

Table S1. Strains, plasmids and oligonucleotides used in this study

Strains	Description	Source
<i>E. coli</i> BL21-CodonPlus (DE3)-RIL	<i>E. coli</i> protein overexpression strain (F ⁻ <i>ompT hsdS</i> (<i>r_Bm_b</i>) ⁻ <i>dcm</i> ⁺ Tet ^r <i>gal endA Hte</i> [<i>argU ileY leuW Cam</i> ^r])	Agilent
Plasmids		
pET28b-GP12	Expression plasmid for N-terminal hexahistidine-GP12	Ref. 27
pΔN88GP12	Expression plasmid for N-terminal hexahistidine-ΔN88GP12	This study
Oligonucleotides		
ΔN88GP12-Forward	CTAGTAGCCATGGGCCACCATCATCATCATCATGATGAAAACC TGTATTTTCAGGGCGACCTCGTTATCCAAGTTATC	
GP12-Reverse	CTAGTAGTTAGTTCAAGAATAGTTGTTTTGGAGACGATTGAGA CCGTTTCTAGTAAGCTTCAAAGGAAAAAGGATACGGTG	

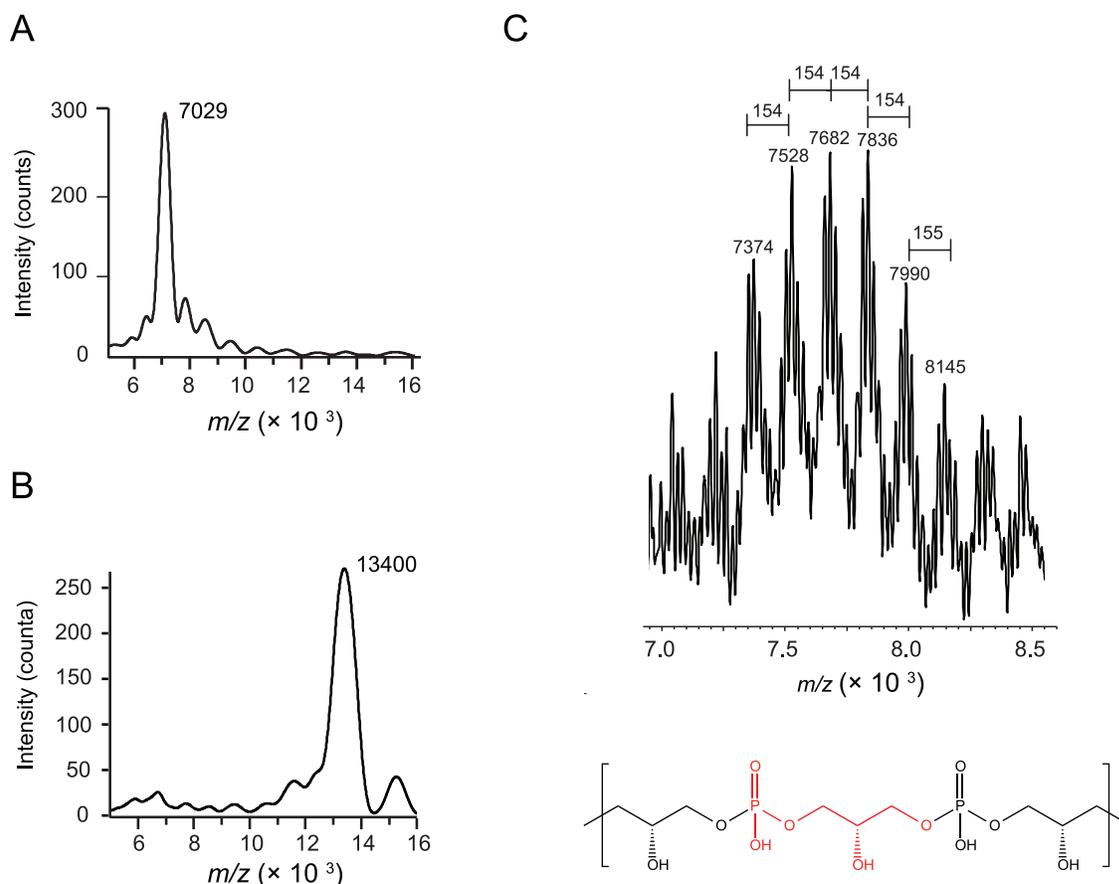


Figure S1. MALDI-TOF mass spectrometry of WTA analogs. Synthetic reactions for WTA analogs were performed as described in the main text. Samples were prepared for analysis by mixing a 5 μL aliquot of the reaction mixture with an equal volume of a saturated matrix solution (2,5-dihydroxybenzoic acid in 50% acetonitrile and 0.1% trifluoroacetic acid). 1 μL of this mixture was spotted onto a stainless steel MALDI plate and spectra were acquired in positive ion linear mode, using a Bruker UltrafleXtreme mass spectrometer, calibrated with external standards. Spectra are shown for *A*, the non-glycosylated analog, m/z 7029: $M + 4 \text{ Na}^+$; M corresponds to the mass of a polymer containing 40 GroP units and *B*, the glycosylated analog, m/z 13400 corresponds to addition of 39 glucose molecules to the non-glycosylated analog. *C*, MALDI-MS analysis of a larger scale (200 μM) synthetic reaction for the lipid $\phi.n$ ($n = 43$) analog shows the products generated differ by the mass of the repeating glycerol phosphate polymer unit, colored red in the structure below the spectrum.

