

Cell Reports

Supplemental Information

**Cooperative Transcriptional Activation
of Antimicrobial Genes by STAT and NF- κ B Pathways
by Concerted Recruitment of the Mediator Complex**

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Figure S1

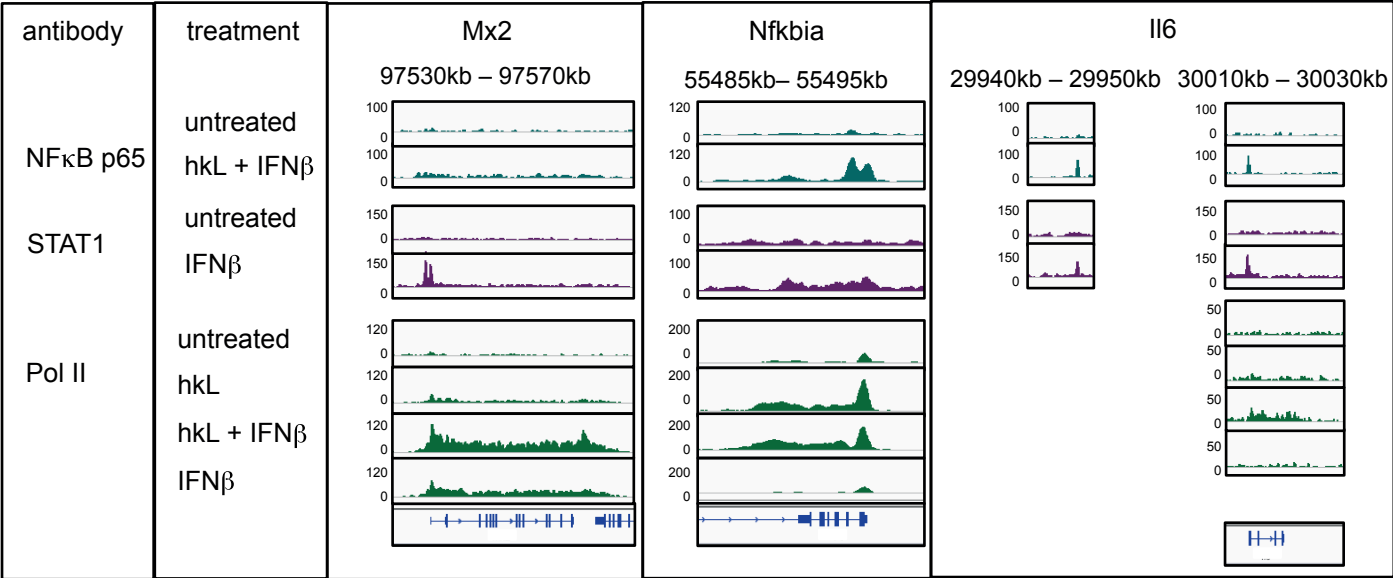
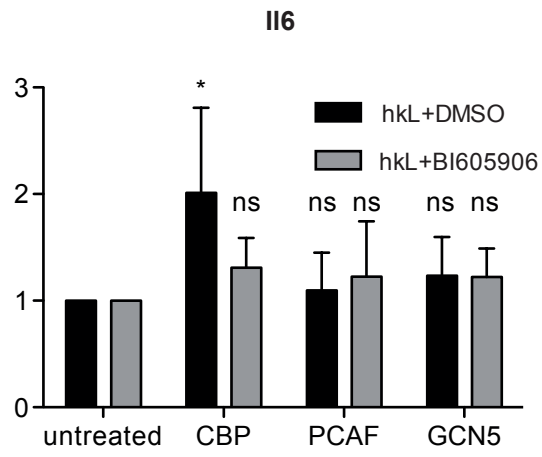
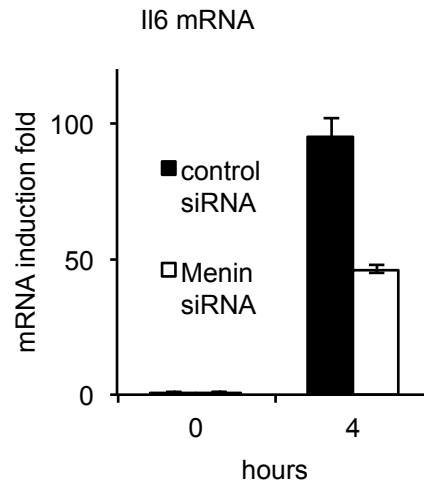


Figure S2

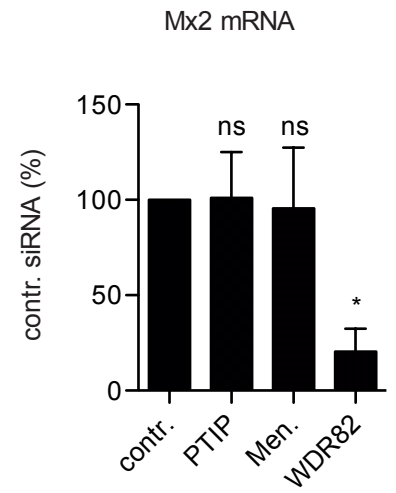
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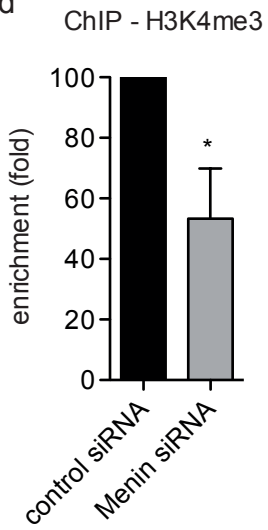
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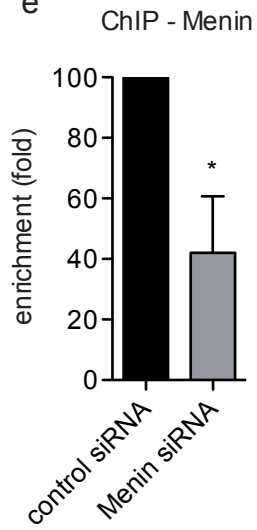
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e



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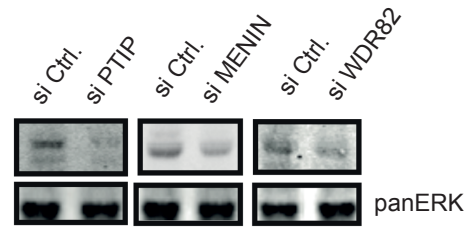


Figure S3

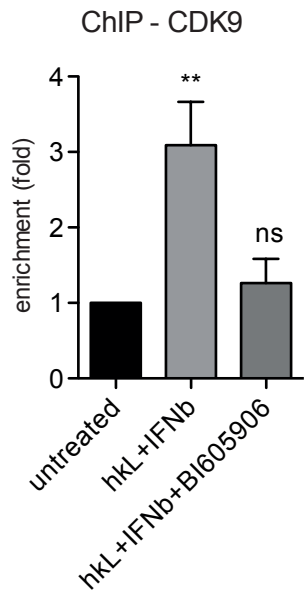


Figure S4

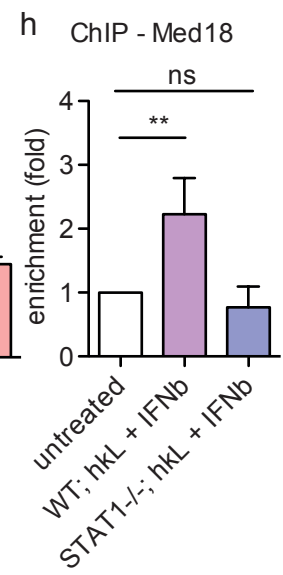
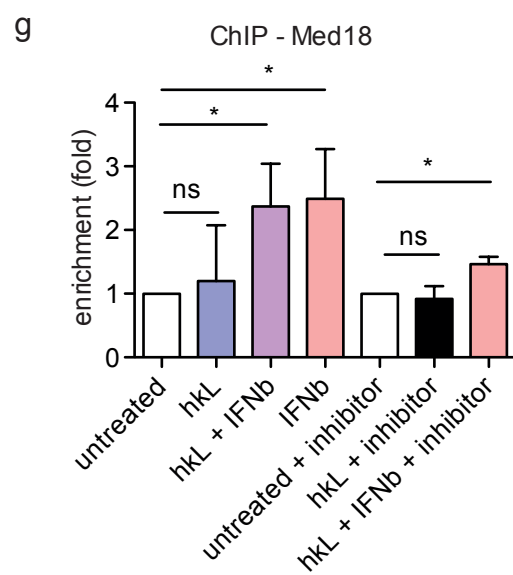
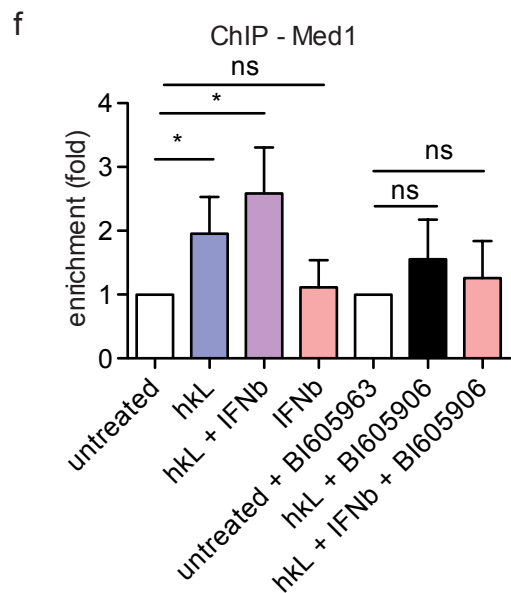
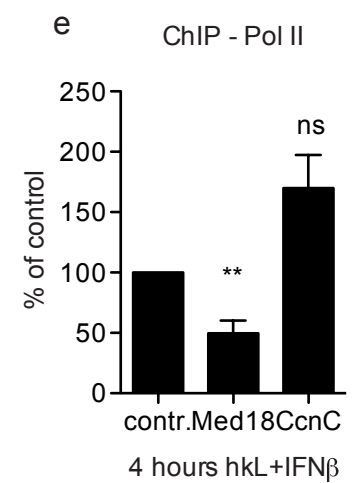
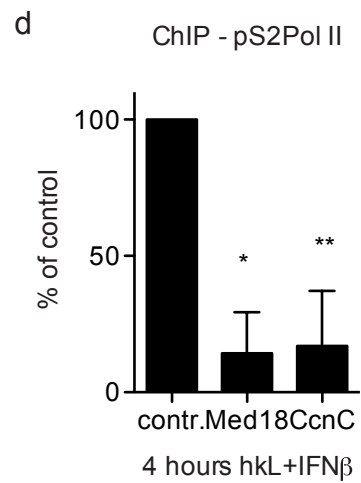
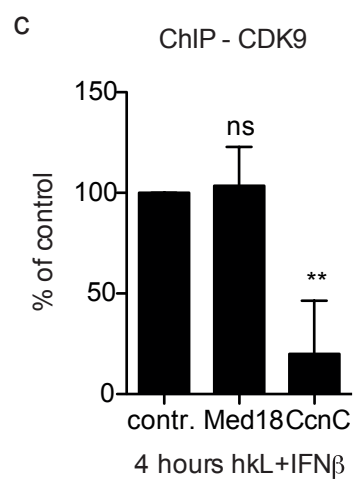
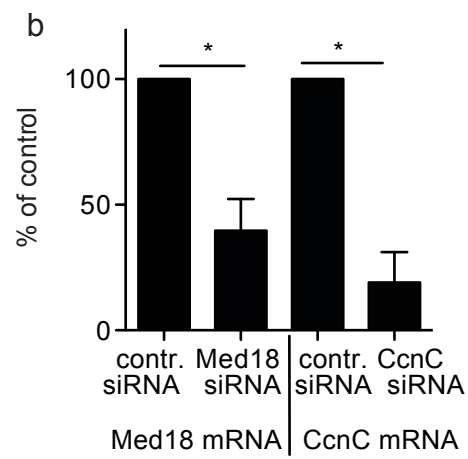
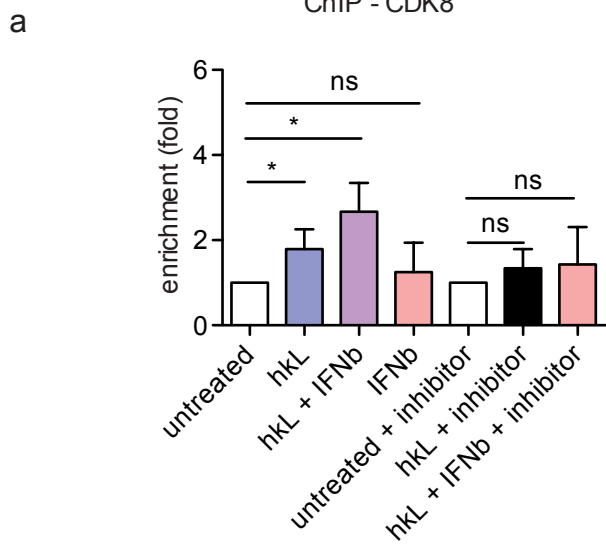


Figure S5

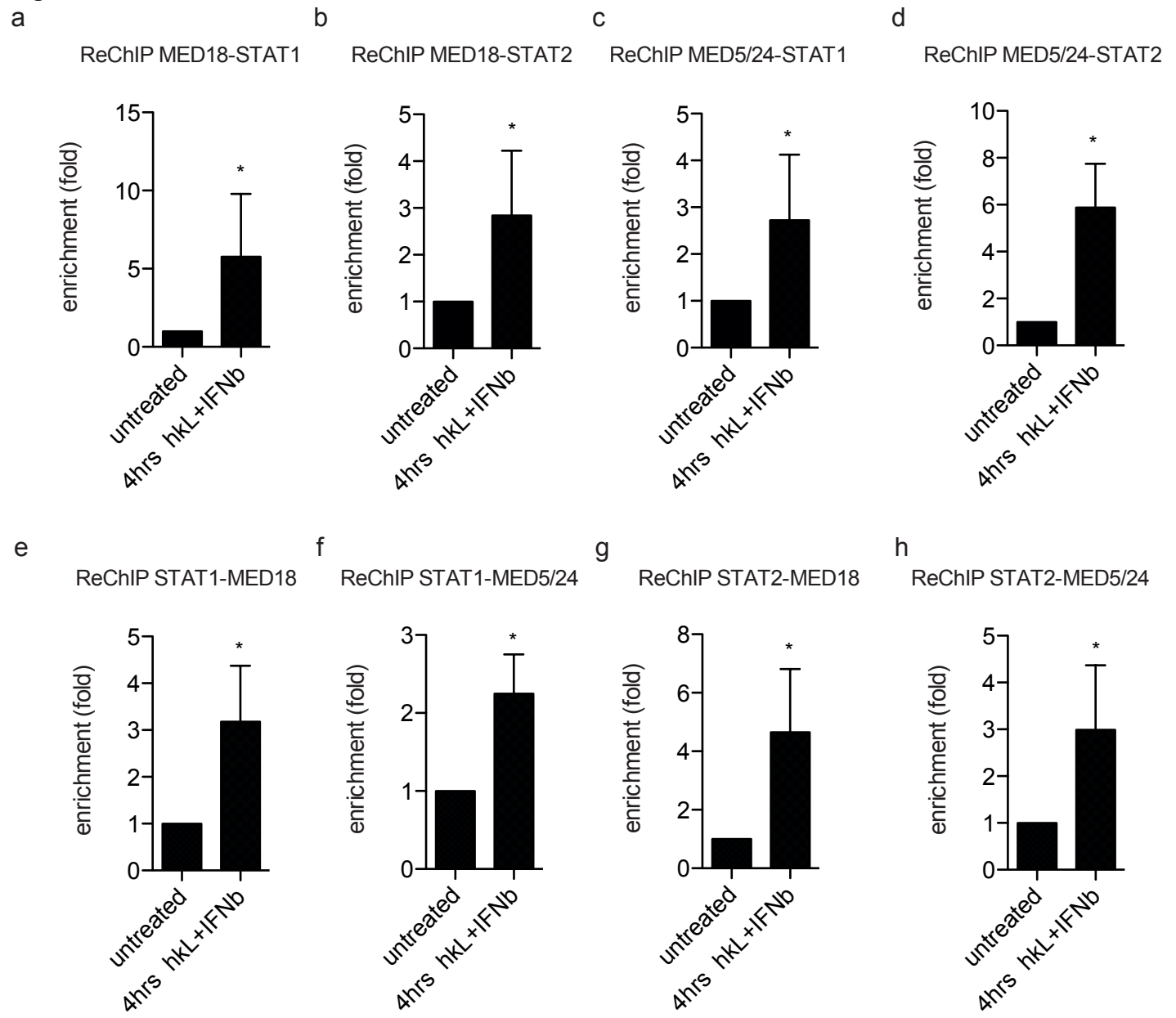
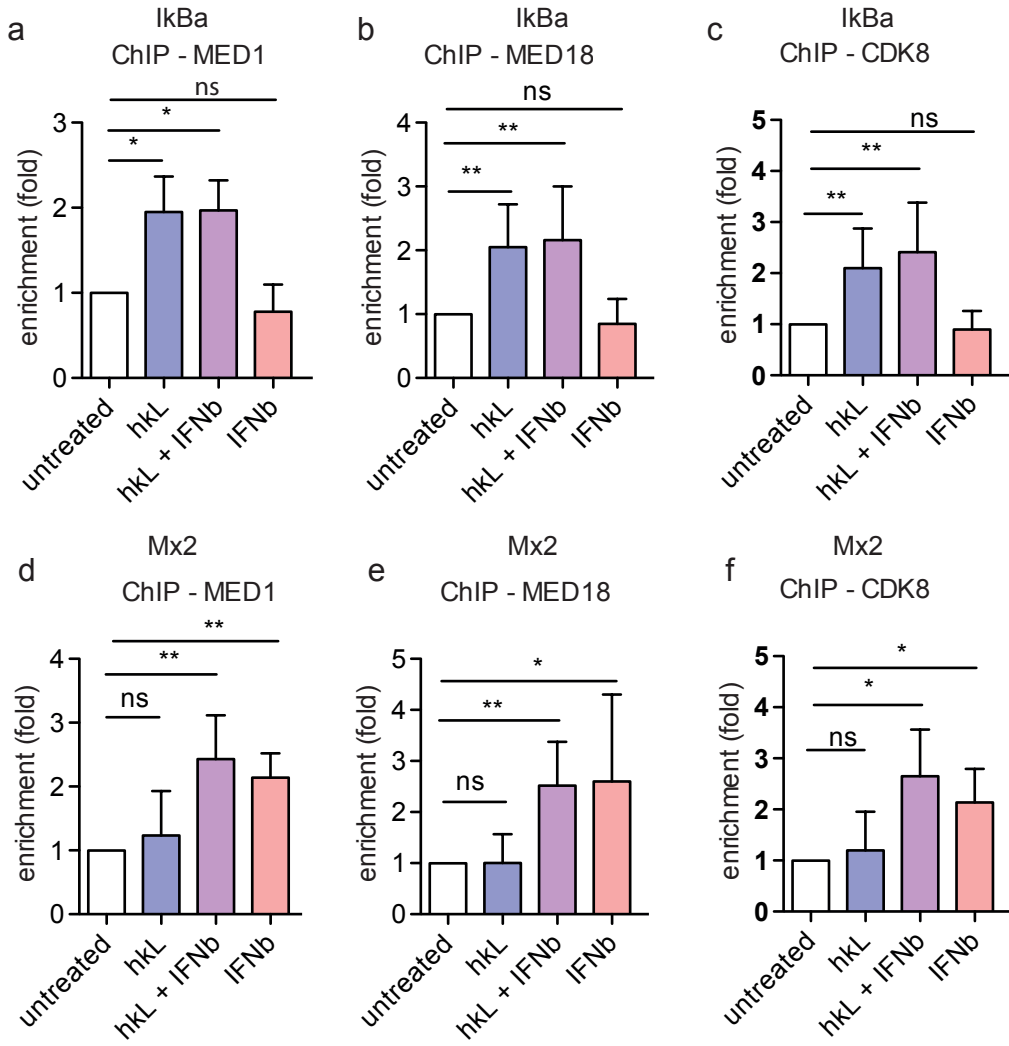


Figure S6



Supplementary figure legends

Figure S1, related to figure 2: ChIP-Seq analysis NF κ B and ISGF3 target genes.

BMDMs were treated with heat-killed *Listeria* (hkL), IFN β , or with a combination of both for 4 hours. For STAT1 ChIP treatment was with IFN β for two hours. Cells were processed for ChIP with indicated antibodies (Pol II, NF κ Bp65, STAT1), followed by next generation sequencing of the precipitate (ChIP-Seq). Genome browser views of the results obtained for the *Mx2* (left panel), *Nfkb1a* (middle panel) and *Il6* (right panel) genes are shown.

Figure S2, related to figure 3: Histone modifiers at the *Il6* promoter. (a) BMDMs were treated with hkL with or without 10 μ M IKK β inhibitor BI605906 for 4 hours followed by ChIP with antibodies to CBP, PCAF and GCN5. DNA encompassing the TSS was amplified. (b-e) BMDM were transfected with Menin siRNA and treated with hkL+IFN β for 4 hours. *Il6* (b) and *Mx2* (c) mRNA expression was analyzed by Q-PCR (b, c), H3K4me3 modification (d) and Menin binding (e) were determined by ChIP, amplifying the region encompassing the TSS. (f) western blot to assess the knock-down efficiency (right panel shows the quantification by densitometry).

Figure S3, related to figure 4: CDK9 recruitment to the *Il6* promoter. BMDMs were treated with hkL+IFN β for 4 hours with (dark grey) or without (light grey) 10 μ M IKK β inhibitor BI605906 followed by ChIP using antibody to CDK9. DNA encompassing the TSS was amplified

Figure S4, related to figure 4, 5 and 6: Knockdown of MED18 and CcnC. (a) WT BMDMs were treated with hkL or IFN β or a combination of both for 4 hours, with and without 10 μ M IKK β inhibitor BI605906 as indicated, followed by ChIP analysis with antibody to CDK8 and Q-PCR for promoter region of *Il6*. (b) Knockdown was performed in BMDMs using siRNA as described in material and methods. The cells were lysed and mRNA analysis (b) was performed to analyze the knockdown efficiency. A knockdown of MED18 or CcnC in BMDMs was performed and cells were treated with hkL and IFN β for 4 hours followed by ChIP with antibodies to CDK9 (c), pS2Pol II (d) and Pol II (e). WT and STAT1 $-/-$ BMDMs were treated with hkL or IFN β or a combination of both for 4 hours, with and without 10 μ M IKK β inhibitor BI605906 as indicated, followed by ChIP analysis with antibody to MED1 (f) and MED18 (g, h) and Q-PCR for promoter region of *Il6*. For all ChIP experiments DNA encompassing the TSS was amplified. Values represent means and standard errors of at least three independent biological replicates. *, P<0,05; **, P<0,01; ***, P<0,001

Figure S5, related to figure 6. ChIP-re-ChIP analysis of Stat1, Stat2 and core mediator binding to the Nos2 promoter. WT BMDMs were treated with hκL and IFNβ for 4 hours, followed by ChIP with the antibodies to MED18 (a, b), MED5/24 (c, d), STAT1 (e, f), and STAT2 (g, h). The samples were re-precipitated with STAT1 (a, c), STAT2 (b, d), MED18 (e, g) and MED5/24 (f, h). The re-precipitates were analyzed by Q-PCR, amplifying the 209bp of the Nos2 promoter including the IFN response region (imperfect ISRE sequences at -952/-940 and -924/-911 with respect to the TSS).

Figure S6, related to figures 4, 5 and 6: Mediator subunit recruitment to the IκBα and Mx2 genes. BMDMs were treated with hκL+IFNβ for 4 hours followed by ChIP analysis with antibodies to MED1, MED18 and CDK8 (a,b). IκBα (a) and Mx2 proximal promoter regions (b) including the TSS were analyzed for protein binding. Values represent means and standard errors of at least four independent biological replicates. *, P<0,05; **, P<0,01; ***, P<0,001

Gene	NF κ B binding site	STAT1 binding site
1190002H23Rik	-	-
1190003J15Rik	-	-
Adora2b	+	-
Arg2	-	-
Ccl7	+	+
Ccr12	+	+
Cd40	-	+
Cd83	+	+
Ch25h	+	+
Cxcl9	-	+
Dgka	-	-
Dusp2	-	+
Edn1	+	+
Egr1	-	-
Gpr109a	-	-
Hdc	-	+
Il10	+	+
Il15ra	+	+
Il19	-	-
Il1a	-	-
Il1rn	+	+
Il27	+	+
Il6	+	+
Indo	-	-
Inhba	+	-
Irf8	-	+
Mxd1	-	+
Nos2	+	+
Nrg1	-	+
Prokr1	-	-
Ptx3	-	-
S100a8	-	-
Serpinb2	-	+
Slamf1	+	+
Slc28a2	+	+
Slc6a4	-	-
Tagap	+	+
Thbs1	-	-
Tmtc2	+	+
Tpbp	-	-
Upp1	-	-
Vcam1	-	-

Table S1, related to figures 1 and 2: mRNAs displaying synergistic induction by type I IFN and heat-killed *Listeria* (hKL) as determined by microarray, related to figures 1 and 2. The presence of binding sites for NF κ B/p65 and STAT1 within 50kB of the transcription start (TSS) of the corresponding genes was determined by ChIP-Seq. For further explanation see text.

a) Contingency tables were computed for the NF κ B_STAT1 pair, depending on the presence or absence of at least one CHIP-Seq peak 50kb upstream of the transcription start or within the gene, and on the presence or absence of at least 2-fold synergistic enrichment of Pol II with the double treatment versus single treatment samples.

NF κ B	STAT1	Pol II syn.	Pol II not syn.	Total
absent	absent	103	26950	27053
present	absent	35	568	603
absent	present	123	5964	6087
present	present	127	1089	1216
Total		388	34571	34959

b) Many hypotheses tests are possible with this table; the two used in the main text are presented. For genes with Pol II synergism, the presence or absence of NF κ B and STAT1 peaks produce the following 2x2 contingency table:

	STAT1 absent	STAT1 present	Total
NF κ B absent	103	123	226
NF κ B present	35	127	162
Total	138	250	388

c) Analysis of genes without and with synergy for the presence or absence of concomitant NF κ B and STAT1 peaks results in the following 2x2 contingency table:

	Pol II syn.	Pol II not syn.	Total
NF κ B or STAT1 absent	261	33482	33743
both NF κ B and STAT1 present	127	1089	1216
Total	388	34571	34959

Table S2, related to figure 2. Association of STAT1 and NF κ B binding with synergistic gene induction by heat-killed *L. monocytogenes* and type I IFN.

Table S3, related to figure 2: GO-analysis and KEGG pathway of 127 genes with STAT1 and NF κ B binding sites showing synergistic NF κ B and ISGF3 activity according to RNA polymerase II signals in CHIP-Seq. Untreated cells are compared with cells subjected to single IFN β or hkL treatment, or with cells treated with combined hkL+IFN β .

Antibody	Catalog number	Company
STAT1	sc-346	Santa Cruz
CDK9	sc-484	Santa Cruz
H3	ab1791	Abcam
H4ac	06-866	Upstate
H4K5ac	Kind gift from Christian Seiser	
H4K8ac	Kind gift from Christian Seiser	
H4K12ac	Kind gift from Christian Seiser	
H4K16ac	Kind gift from Christian Seiser	
Med1	sc-5334	Santa Cruz
Med4	Ab129170	Abcam
Med7	sc-12457	Santa Cruz
CDK8	sc-1521	Santa Cruz
Med18	A300-777A	Bethyl
Med24	A301-472A	Bethyl
Med26	sc-48776	Santa Cruz
CcnC	sc-1061	Santa Cruz
Pol II	sc-899	Santa Cruz
pS2Pol II	A300-654A	Bethyl
CBP	sc-369	Santa Cruz
GCN5	sc-20698x	Santa Cruz
PCAF	sc-8999x	Santa Cruz

Table S4, related to figures 2-6. Antibodies used for ChIP

Q-PCR

Nos2 for	GCTTGCCCCAACAGGAGAAG
Nos2 rev	GCTGCCCGGAAGGTTTGTAC
Il6 for	CTGCAAGAGACTTCCATCCAG
Il6 rev	AGTGGTATAGACAGGTCTGTTGG
Il1rn for	GTGCTACTGGGGCTCATTTGT
Il1rn rev	GGAGTAAGAGGACACTTGCGAAT
Ccl2 for	CCCCGGACGATGAATATGATG
Ccl2 rev	CACCAAGATAAACACCGCCAG
HPRT for	GTTGGATACAGGCCAGACTTTGTTG
HPRT rev	GAGGGTAGGCTGGCCTATTGGCT

ChIP

Nos2 prx for	GGTCCCAGTTTTGAAGTGACTIONACG
Nos2 prx rev	GTTGTGACCCTGGCAGCAG
Nos2 dis for	CCAACACTATTGAGGCCACACAC
Nos2 dis rev	GCTTCCAATAAAGCATTACACA
Mx2 for	ACCCAGCCAAGGCCCCCTTA
Mx2 rev	GCAGCTGCCAGGGCTCAGAC
IkBα for	AAGAAGGGTTCTTGCAGAGGGCT
IkBα rev	TCGTCCTCCACTGAGAAGCCTAAA
Il6 for	ATCCAGTTGCCTTCTTGGGACTGA
Il6 rev	ATCAGTTTCACAGCCTACCCACCT
Nos2 30kb distal for	CTGGAGTCTGTTCTTCTGACTG
Nos2 30kb distal rev	CAAGAGGCCACAAGAGAAT

Table S5, related to figures 1-6. Primer sequences used for gene expression and ChIP analysis by Q-PCR in 5' -> 3' polarity