

Comparative Evolution of Duplicated *Ddx3* Genes in Teleosts: Insights from Japanese flounder,
Paralichthys olivaceus

Zhongkai Wang, Wei Liu, Huayu Song, Huizhen Wang, Jinxiang Liu, Haitao Zhao, Xinxin Du,
Quanqi Zhang

Key Laboratory of Marine Genetics and Breeding (MGB), Ministry of Education, College
of Marine Life Sciences, Ocean University of China, Qingdao 266003, China

Corresponding author: Quanqi Zhang

Ocean University of China, No 5 Yushan Road, Qingdao 266003, China.

Tel./fax: +86 532 82031806

E-mail: qzhang@ouc.edu.cn

DOI: 10.1534/g3.115.018911

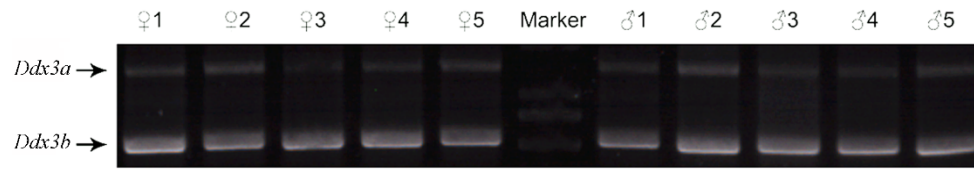


Figure S1 Sex-specific amplification of *Ddx3* genes in Japanese flounder. Using the genomic DNA of females or males as templates, the amplification products of *Ddx3* genes showed no sexual specificity.

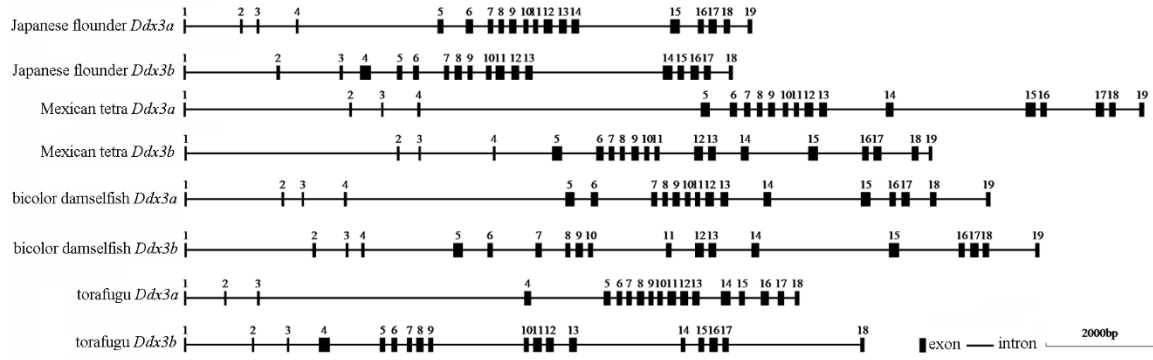


Figure S2 Genomic structure of teleost *Ddx3a* and *Ddx3b* genes.

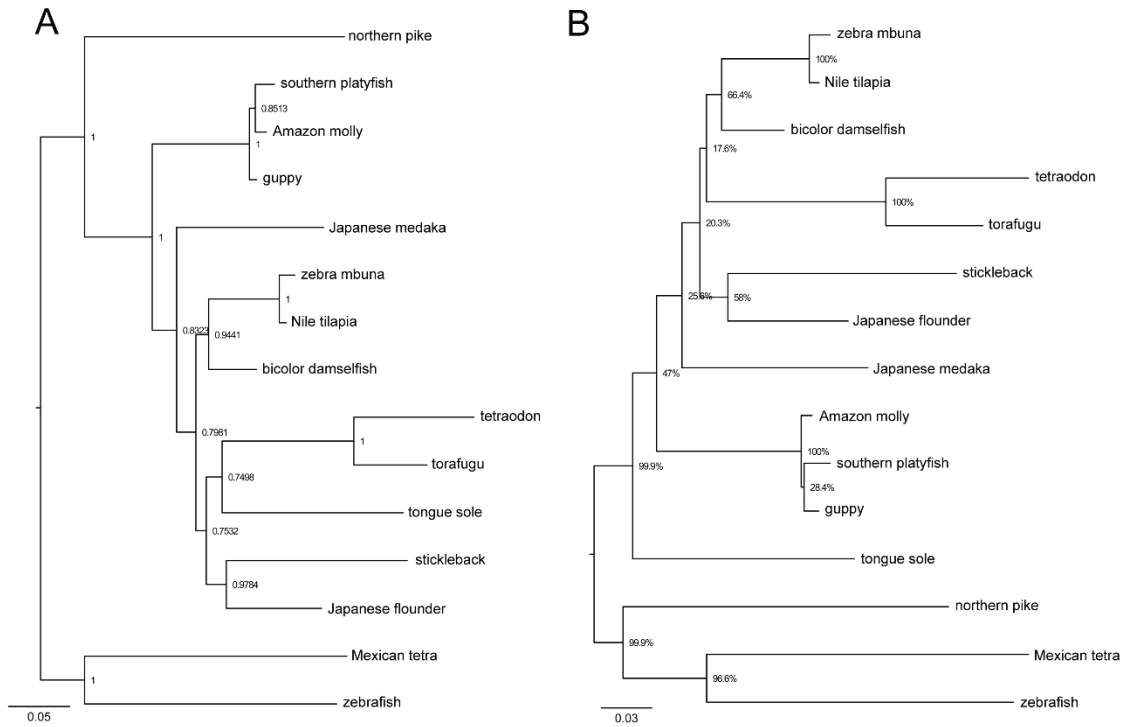


Figure S3 Phylogenetic tree of teleost *Ddx3a* genes used in PAML analysis. (A) Bayesian tree of teleost *Ddx3a* genes. Numbers at the nodes are Bayesian posterior probabilities. Scale bar = 0.05. (B) Maximum likelihood tree of teleost *Ddx3a* genes. Numbers at the nodes are bootstrap support values. Scale bar = 0.03. Phylogenetic reconstructions were based on the coding sequences of *Ddx3a* genes. The accession numbers of these genes at GenBank or Ensembl database are provided in Table S1.

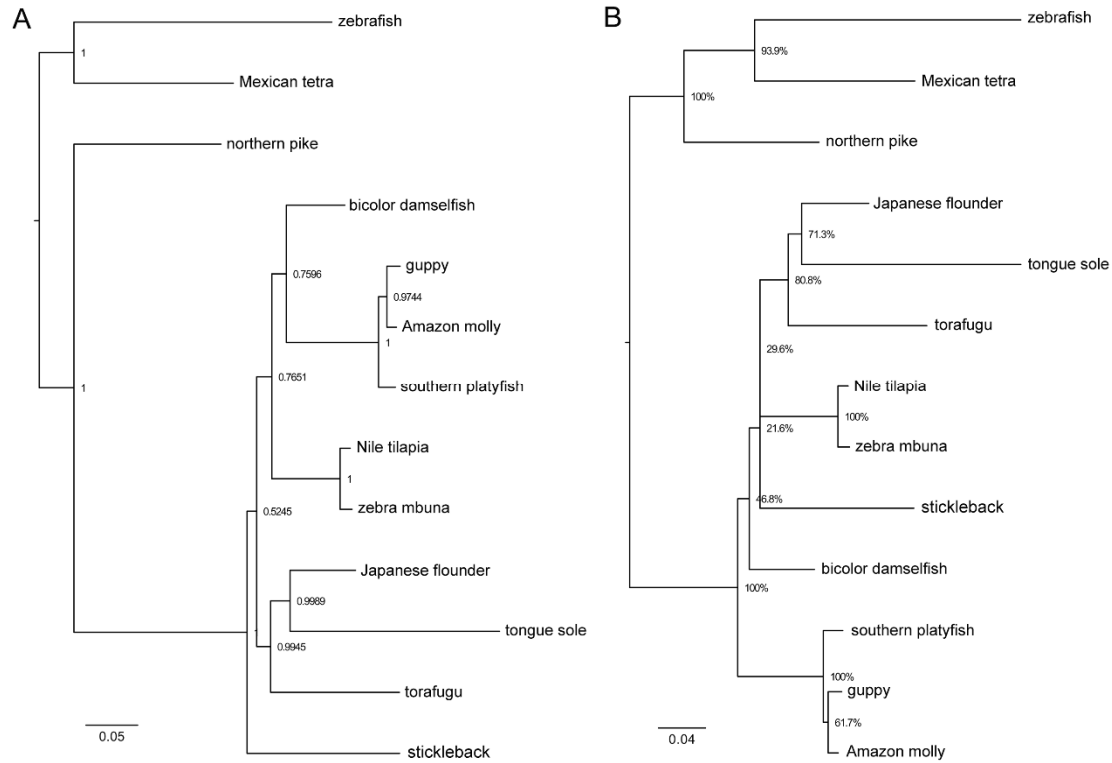


Figure S4 Phylogenetic tree of teleost *Ddx3b* genes used in PAML analysis. (A) Bayesian tree of teleost *Ddx3b* genes. Numbers at the nodes are Bayesian posterior probabilities. Scale bar = 0.05. (B) Maximum likelihood tree of teleost *Ddx3b* genes. Numbers at the nodes are bootstrap support values. Scale bar = 0.04. Phylogenetic reconstructions were based on the coding sequences of *Ddx3b* genes. The accession numbers of these genes at GenBank or Ensembl database are provided in Table S1.

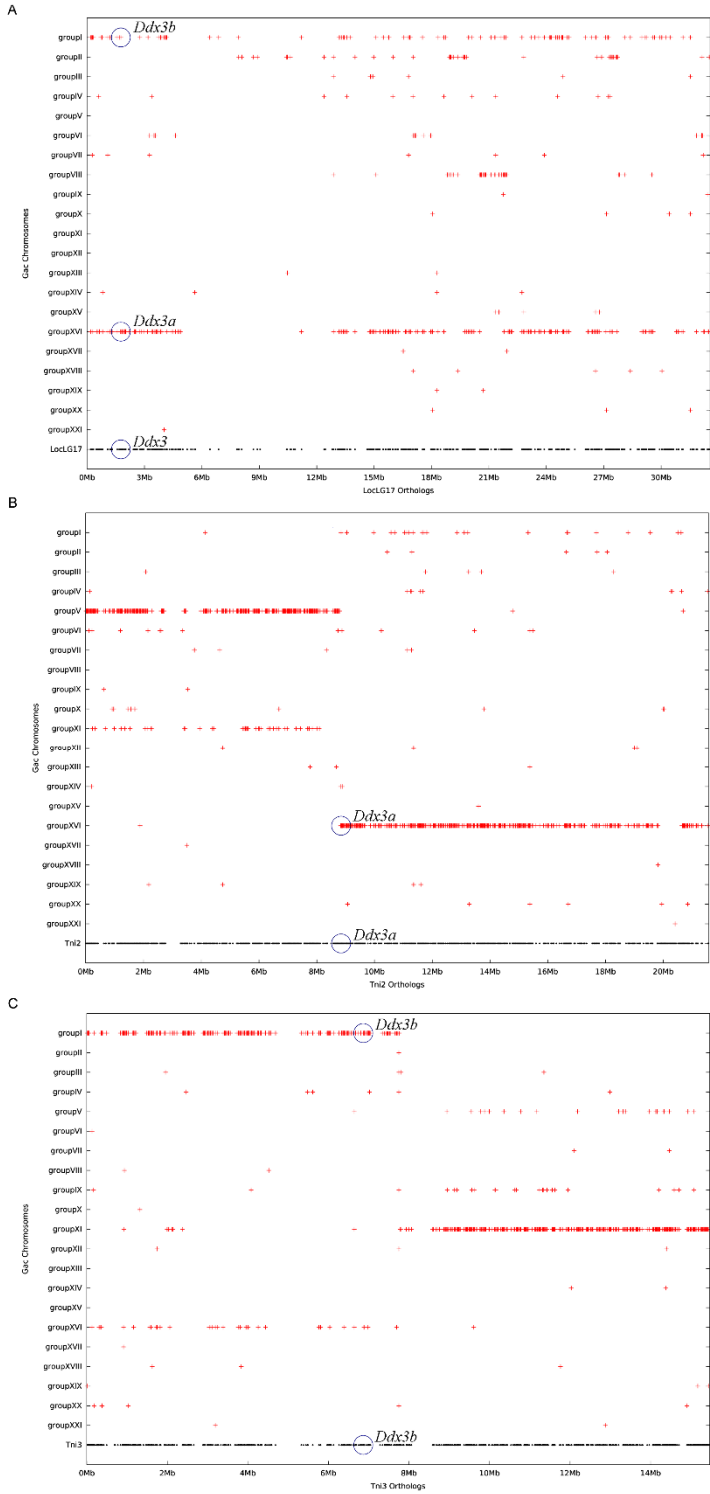


Figure S5 Detection of conserved chromosome among teleosts. Synteny analysis of spotted gar LG17 (A), tetraodon Tni2 (B), and tetraodon Tni3 (C) and stickleback chromosomes. Stickleback *Ddx3a* is in groupXVI and *Ddx3b* is in groupI.

Table S1 List of taxa used in this study.

Organism Name	Common Name	Assembly ID	Gene	Accession No.
<i>Homo sapiens</i>	human	GRCh38	<i>Ddx3X</i>	NM_001356.3
			<i>Ddx3Y</i>	NM_001122665.1
<i>Mus musculus</i>	house mouse	GRCm38.p2	<i>Ddx3X</i>	XM_006527566.1
			<i>Ddx3Y</i>	XM_006531605.1
<i>Bos taurus</i>	cattle	Bos_taurus_UMD_3.1.1	<i>Ddx3X</i>	XM_005228286.2
			<i>Ddx3Y</i>	XM_005228498.2
<i>Pan troglodytes</i>	chimpanzee	Pan_troglodytes-2.1.4	<i>Ddx3X</i>	XM_009438962.1
			<i>Ddx3Y</i>	XM_009439896.1
<i>Monodelphis domestica</i>	gray short-tailed opossum	MonDom5	<i>Ddx3</i>	XM_007493339.1
<i>Ornithorhynchus anatinus</i>	platypus	Ornithorhynchus_ anatinus-5.0.1	<i>Ddx3</i>	XM_007667286.1
<i>Gallus gallus</i>	chicken	Gallus_gallus-4.0	<i>Ddx3</i>	NM_001030800.1
<i>Anolis carolinensis</i>	green anole	AnoCar2.0	<i>Ddx3</i>	XM_008107404.1
<i>Xenopus tropicalis</i>	western clawed frog	Xtropicalis_v7	<i>Ddx3</i>	NM_203865.1
<i>Latimeria chalumnae</i>	coelacanth	LatCha1	<i>Ddx3</i>	XM_006005839.1
<i>Callorhynchus milii</i>	elephant shark	Callorhynchus_ milii-6.1.3	<i>Ddx3</i>	XM_007901579.1
<i>Lepisosteus oculatus</i>	spotted gar	LepOcu1	<i>Ddx3</i>	XM_006638889.1
<i>Danio rerio</i>	zebrafish	GRCz10	<i>Ddx3a</i>	NM_130941.2
			<i>Ddx3b</i>	NM_001126423.1
<i>Astyanax mexicanus</i>	Mexican tetra	Astyanax_ mexicanus-1.0.2	<i>Ddx3a</i>	XM_007227851.1
			<i>Ddx3b</i>	XM_007244224.1
<i>Esox lucius</i>	northern pike	EsoLuc1.0	<i>Ddx3a</i>	XM_010889977.1
			<i>Ddx3b</i>	XM_010886335.1
<i>Stegastes partitus</i>	bicolor damselfish	Stegastes_partitus-1.0.2	<i>Ddx3a</i>	XM_008279959.1
			<i>Ddx3b</i>	XM_008292391.1
<i>Poecilia reticulata</i>	guppy	Guppy_female_1.0+MT	<i>Ddx3a</i>	XM_008437108.1

			<i>Ddx3b</i>	XM_008426133.1
<i>Oreochromis niloticus</i>	Nile tilapia	Orenil1.1	<i>Ddx3a</i>	XM_005475255.1
			<i>Ddx3b</i>	NT_167568.1
<i>Cynoglossus semilaevis</i>	tongue sole	Cse_v1.0	<i>Ddx3a</i>	XM_008327813.1
			<i>Ddx3b</i>	XM_008324681.1
<i>Takifugu rubripes</i>	torafugu	FUGU5	<i>Ddx3a</i>	XM_003961572.1
			<i>Ddx3b</i>	XM_003966411.1
<i>Oryzias latipes</i>	Japanese medaka	ASM31367v1	<i>Ddx3a</i>	XM_004081602.1
			<i>Ddx3b</i>	XM_004085517.1
<i>Paralichthys olivaceus</i>	Japanese flounder	unpublished	<i>Ddx3a</i>	KP205082
			<i>Ddx3b</i>	KP205083
<i>Poecilia formosa</i>	Amazon molly	Poecilia_formosa-5.1.2	<i>Ddx3a</i>	XM_007571428.1
			<i>Ddx3b</i>	XM_007572225.1
<i>Xiphophorus maculatus</i>	southern platyfish	Xiphophorus_ maculatus-4.4.2	<i>Ddx3a</i>	XM_005803788.1
			<i>Ddx3b</i>	XM_005813094.1
<i>Maylandia zebra</i>	zebra mbuna	MetZeb1.1	<i>Ddx3a</i>	XM_004566279.1
			<i>Ddx3b</i>	XM_004573843.1
<i>Tetraodon nigroviridis</i>	tetraodon	TETRAODON 8.0	<i>Ddx3a</i>	ENSTNIG00000011569
			<i>Ddx3b</i>	ENSTNIG00000014514
<i>Gasterosteus aculeatus</i>	stickleback	BROAD S1	<i>Ddx3a</i>	ENSGACG00000008206
			<i>Ddx3b</i>	ENSGACG00000014407

Table S2 Sequences of primers used for cloning Japanese flounder *Ddx3* genes.

Primers	Sequence(5'-3')	Product	usage
ddx3-1_FW	TTTCTGTACCACGAAGGTTACGC	1370 bp	PCR
ddx3-1_RV	CATAGCCGTCATTGCTGTAGAAG		
ddx3-2_FW	ATTGGAACAGGAGCATTTGACC	512 bp	PCR
ddx3-2_RV	GTCAGAGGCTTTGAATGGGGT		
DDX3-1-FW	GATGCTCCAAAAACGATTCAC	128 bp	RT-qPCR
DDX3-1-RV	ATTCCTCTATCATTGCGAGGG		
DDX3-2-FW	ATGCCATTCCAATCGTCAAG	289 bp	RT-qPCR
DDX3-2-RV	TACACAACACAGGGACGCACT		

Table S3 Results of site model analyses on teleost *Ddx3* Bayesian gene trees.

Tree	Mod	lnL	κ	Parameters ^a			Nul	LRT	d	<i>P</i>
				ω/p	ω_1/q	ω_2/ω_p				
<i>Ddx3</i> <i>a</i>	M0	-	1.82	0.077			n/a			
		12032.6	7							
		10								
	M1a	-	1.94	0.042(89.94	1.000(10.06		n/a			
		11821.5	1	%)	%)					
		69								
	M2a	-	1.94	0.042(89.94	1.000(10.06	87.302(0)	M1	0	2	1.00
		11821.5	1	%)	%)		a			0
		69								
	M3	-	1.82	0.013(74.72	0.224(20.60	0.750(4.68	M0	603.50	4	0
	11730.8	9	%)	%)	%)		6			
	57									
M7	-	1.82	0.198	1.828		n/a				
	11734.9	3								
	89									
M8a	-	1.84	0.223	2.257	1.000(3.71	n/a				
	11739.4	9			%)					
	84									
M8	-	1.82	0.207	2.044	2.600(0.35	M7	7.498	2	0.02	
	11731.2	5			%)				4	
	40									
						M8	16.488	1	0	
						a				
<i>Ddx3</i>	M0	-	1.78	0.057			n/a			
		10420.5	2							
		56								
M1a	-	1.85	0.046(95.77	1.000(4.23		n/a				
	10373.0	7	%)	%)						

	22								
M2a	-	1.85	0.046(95.77	1.000(4.23	48.098 (0)	M1	0	2	1.00
	10373.0	7	%)	%)		a			0
	22								
M3	-	1.77	0(52.95%)	0.084(37.85	0.324(9.20	M0	264.91	4	0
	10288.1	5		%)	%)		2		
	00								
M7	-	1.77	0.286	4.080		n/a			
	10288.8	6							
	73								
M8a	-	1.77	0.230	2.738	1.000(0.65	n/a			
	10287.6	4			%)				
	92								
M8	-	1.78	0.306	4.687	1.449(0.41	M7	4.496	2	0.10
	10286.6	2			%)				6
	25								
						M8	2.134	1	0.14
						a			4

Abbreviations: **lnL**, *ln* likelihood; **κ** , transition/transversion ratio; **LRT**, likelihood ratio test statistic; **df**, degrees of freedom; **P**, P-value; **n/a**, not applicable.

^a: ω values of each site class are shown for models M0-M3 (ω_0 - ω_2) with the proportion of each site class in parentheses. For M7-M8, the shape parameters, p and q, which describe the beta distribution are listed. In addition, the ω value for the positively selected site class (ω_p , with the proportion of sites in parentheses) is shown for M8a (where ω_p is constrained to equal one) and M8.

Table S4 Sites selected in *Ddx3a* by BEB in Bayesian tree.

Model	codon site	BEB		
		Pr($\omega > 1$)	post mean	+/-
M8	43	0.513	1.070	0.459
	89	0.583	1.132	0.451
	131	0.578	1.135	0.443
	151	0.577	1.140	0.436
	176	0.860	1.392	0.275
	199	0.571	1.136	0.435
	219	0.562	1.124	0.441
	429	0.962*	1.473	0.147
	607	0.539	1.093	0.458
	609	0.560	1.089	0.483
	616	0.980*	1.486	0.109
	620	0.763	1.306	0.358

*: posterior probability > 0.95

Table S5 Results of site model analyses on teleost *Ddx3* ML gene trees.

Tree	Model	lnL	κ	Parameters ^a			Null	LRT	df	P
				ω_0/p	ω_1/q	ω_2/ω_p				
<i>Ddx3a</i>	M0	- 12069.904	1.870	0.079			n/a			
	M1a	- 11828.656	1.967	0.042(90.37%)	1.000(9.63%)		n/a			
	M2a	- 11828.656	1.967	0.042(90.37%)	1.000(9.63%)	37.503(0)	M1a	0	2	1.000
	M3	- 11746.341	1.872	0.013(74.49%)	0.217(20.62%)	0.813(4.89%)	M0	647.126	4	0
	M7	- 11753.109	1.865	0.190	1.672		n/a			
	M8a	- 11753.656	1.891	0.224	2.438	1.000(4.47%)	n/a			
	M8	- 11745.686	1.873	0.242	3.063	1.000(2.94%)	M7	14.846	2	0
							M8a	15.940	1	0
<i>Ddx3b</i>	M0	- 10427.457	1.799	0.057			n/a			
	M1a	- 10379.321	1.873	0.046(95.89%)	1.000(4.11%)		n/a			
	M2a	- 10379.321	1.873	0.046(95.89%)	1.000(4.11%)	50.201 (0)	M1a	0	2	1.000
	M3	- 10291.089	1.797	0.010(71.32%)	0.178(28.17%)	1.431(0.51%)	M0	272.736	4	0
	M7	- 10293.577	1.793	0.285	4.030		n/a			
	M8a	- 10292.146	1.791	0.230	2.714	1.000(0.67%)	n/a			
	M8	-	1.800	0.306	4.651	1.494(0.42%)	M7	4.892	2	0.087

Abbreviations: **lnL**, *ln* likelihood; **κ** , transition/transversion ratio; **LRT**, likelihood ratio test statistic; **df**, degrees of freedom; **P**, P-value; **n/a**, not applicable.

^a: ω values of each site class are shown for models M0-M3 (ω_0 - ω_2) with the proportion of each site class in parentheses. For M7-M8, the shape parameters, p and q, which describe the beta distribution are listed. In addition, the ω value for the positively selected site class (ω_p , with the proportion of sites in parentheses) is shown for M8a (where ω_p is constrained to equal one) and M8.

Table S6 Sites selected in *Ddx3a* by BEB in ML tree.

Model	codon site	BEB		
		Pr($\omega > 1$)	post mean	+/-
M8	9	0.585	1.127	0.460
	43	0.604	1.157	0.438
	89	0.581	1.129	0.453
	111	0.651	1.198	0.426
	123	0.842	1.368	0.314
	176	0.850	1.383	0.285
	199	0.765	1.309	0.353
	219	0.654	1.211	0.409
	429	0.983*	1.488	0.094
	607	0.791	1.329	0.341
	609	0.544	1.071	0.488
	616	0.984*	1.489	0.092
	620	0.837	1.370	0.301

*: posterior probability > 0.95