

Supplemental Experimental Procedures

Animals

The targeting construct harboring mutations in the miR-155 target site in the 3' UTR of the SOCS1 gene was generated using recombineering. For detailed information see the National Cancer Institute recombineering website (<http://redrecombineering.ncifcrf.gov/>). Mice on a B6 genetic background with the germ-line transmission were bred to Cre deleter mice to remove the neomycin resistance cassette. miR-155^{KO} and C57BL/6J mice were purchased from the Jackson Laboratories. *SOCS1*^{fl} mice and *Klra8*^{KO} (Ly49H-deficient) mice were kindly provided by A. Yoshimura and S. Vidal, respectively. All the mice were bred and housed in the specific-pathogen-free animal facility at the Memorial Sloan-Kettering Cancer Center and used in accordance with institutional guidelines.

EAE

Mice were immunized s.c. with 100µg MOG₃₅₋₅₅ emulsified with complete Freund's adjuvant plus 200ng pertussis toxin in 500ul PBS. Animals were monitored 3 times a week for disease symptoms until the experiment terminated, usually 25 days post induction and classified into one of five grades of severity: grade 1 - no tail tone and the tail drops when the mouse is picked up by the base of the tail; grade 2 - tail drop and weakness in the rear limbs, usually evident by an uneven gait; grade 3 - rear limb paralysis; grade 4 - rear limb paralysis and weakness in the forelimbs; grade 5 - moribund. In some experiments, mice were harvested at the peak of the disease (~14 days after EAE induction) followed by FACS analysis.

In Vitro T cell Proliferation and Th17 Polarization Assays

To measure the recall response to MOG antigen, splenic CD4⁺ T cells from mice subjected to EAE induction were co-cultured with CD11c bead-sorted DCs pulsed with different concentrations of MOG₃₅₋₅₅ peptides for 3 days. T cell proliferation was

assessed by with ^3H -TdR incorporation (cpm) in triplicate during the last 8 hr of culture. For *in vitro* Th17 polarization assay, FACS sorted $\text{CD4}^+\text{CD25}^-\text{CD62L}^{\text{hi}}$ naïve T cells (2×10^5) were stimulated with αCD3 ($1 \mu\text{g/ml}$) in the presence of irradiated T cell-depleted splenocytes (1×10^6) as APCs plus $\text{TGF}\beta$ (2ng/ml) and IL-6 (20ng/ml) for 4 days. IL-17 production was assessed by intracellular FACS analysis.

Quantitative PCR and Gene Expression Profiling Analysis

Total RNA was extracted with TriZOL reagent (Invitrogen) from FACS-purified cells. cDNA was synthesized with Superscript III Reverse Transcriptase (Invitrogen), followed by real-time PCR analysis (SYBR green; Applied Biosystems).

For gene expression profiling analysis, CD45.2^+ $\text{miR-155}^{\text{KO}}$ or SOCS1^{KI} or WT OT-1 TCR-tg CD8^+ T cells or NK cells were adoptively transferred into CD45.1^+ WT or Ly49H-deficient hosts, respectively, prior to infection with MCMV-OVA or MCMV. OT-1 CD8^+ T cells or Ly49H $^+$ NK cells were FACS-sorted on d4 post infection and poly-A RNA-sequencing was performed using 3 biological replicates for each cell type analyzed using an Illumina HiSeq2500 platform. Genes with $\text{FDR} < 5\%$ and the absolute value of \log_2 fold change more than 0.75 under both conditions were considered as similarly regulated in $\text{miR-155}^{\text{KO}}$ and SOCS1^{KI} cells compared to WT controls, while genes with $\text{FDR} < 5\%$ in only one condition and absolute value of \log_2 fold change less than 0.75 in the other condition were considered as $\text{miR-155}^{\text{KO}}$ specific or SOCS1^{KI} specific. Each type of regulated genes can be further separated into up-regulated and down-regulated categories based on the direction of regulation. RNA-sequencing data are available from NCBI under accession number [GSE68511](https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE68511).

Figure S1 (related to Figure 1). Generation of mice harboring mutations in the putative miR-155 target site in the 3' UTR of the SOCS1. Schematic representation of the SOCS1 targeting strategy.

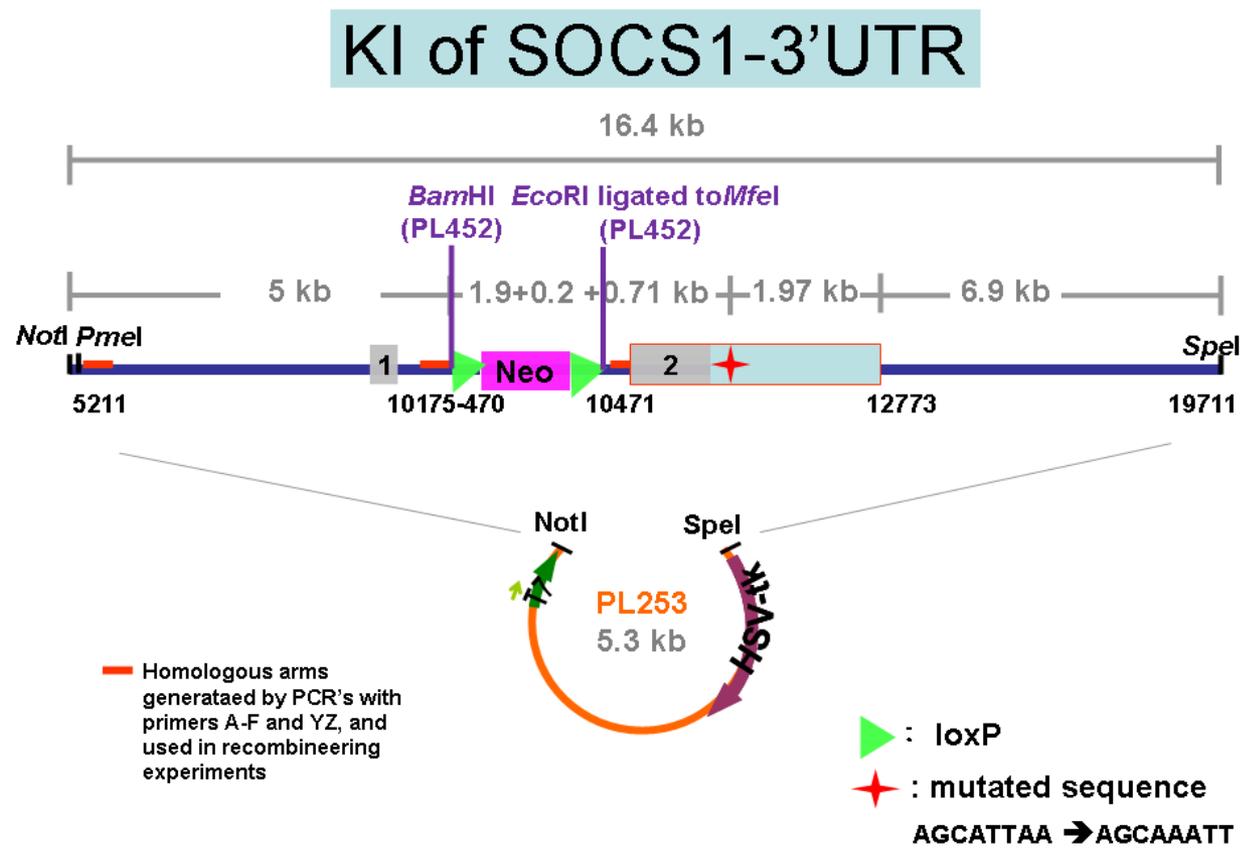


Figure S2 (related to Figure 1). Specific disruption of miR-155-mediated regulation of SOCS1 by mutations in the 3' UTR of the *Socs1* gene. (A) Retroviral miR-155, miR-19 or empty vectors equipped with a GFP reporter were expressed in T cells from SOCS1^{KI}, miR-155^{KO} or WT mice. GFP⁺ cells were sorted 4 days after retroviral transduction and the amounts of SOCS1 were assessed. **(B)** Relative reductions in SOCS1 protein amounts in T cells overexpressed with different miRNAs compared to corresponding T cell populations with empty vector transduction were shown. The data are representative of 2 independent experiments (n = 2).

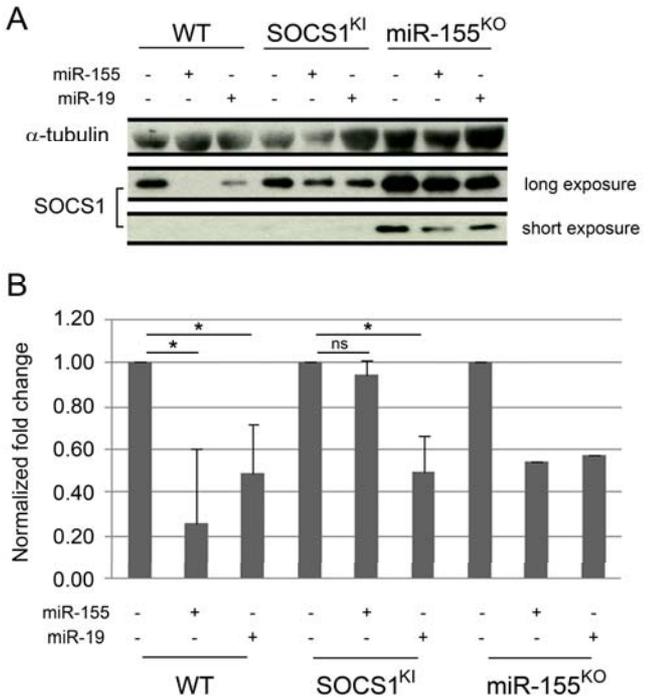


Figure S3 (related to Figure 2). SOCS1 deficiency does not rescue reduced miR-155^{KO} Treg cell numbers. FACS analysis of **(A)** thymus and **(C)** spleen from WT, miR-155^{KO}, *Foxp3*^{cre}SOCS1^{fl/fl} (SOCS1^{ckKO}) or miR-155^{KO}SOCS1^{ckKO} (DKO) mice. **(B,D)** Cellularity of the thymus and spleen and the proportion and absolute numbers of thymic and splenic Foxp3⁺CD4⁺ T cells in indicated mice are shown. The data are shown as mean \pm SD and are representative of 4 independent experiments (n = 5–12).

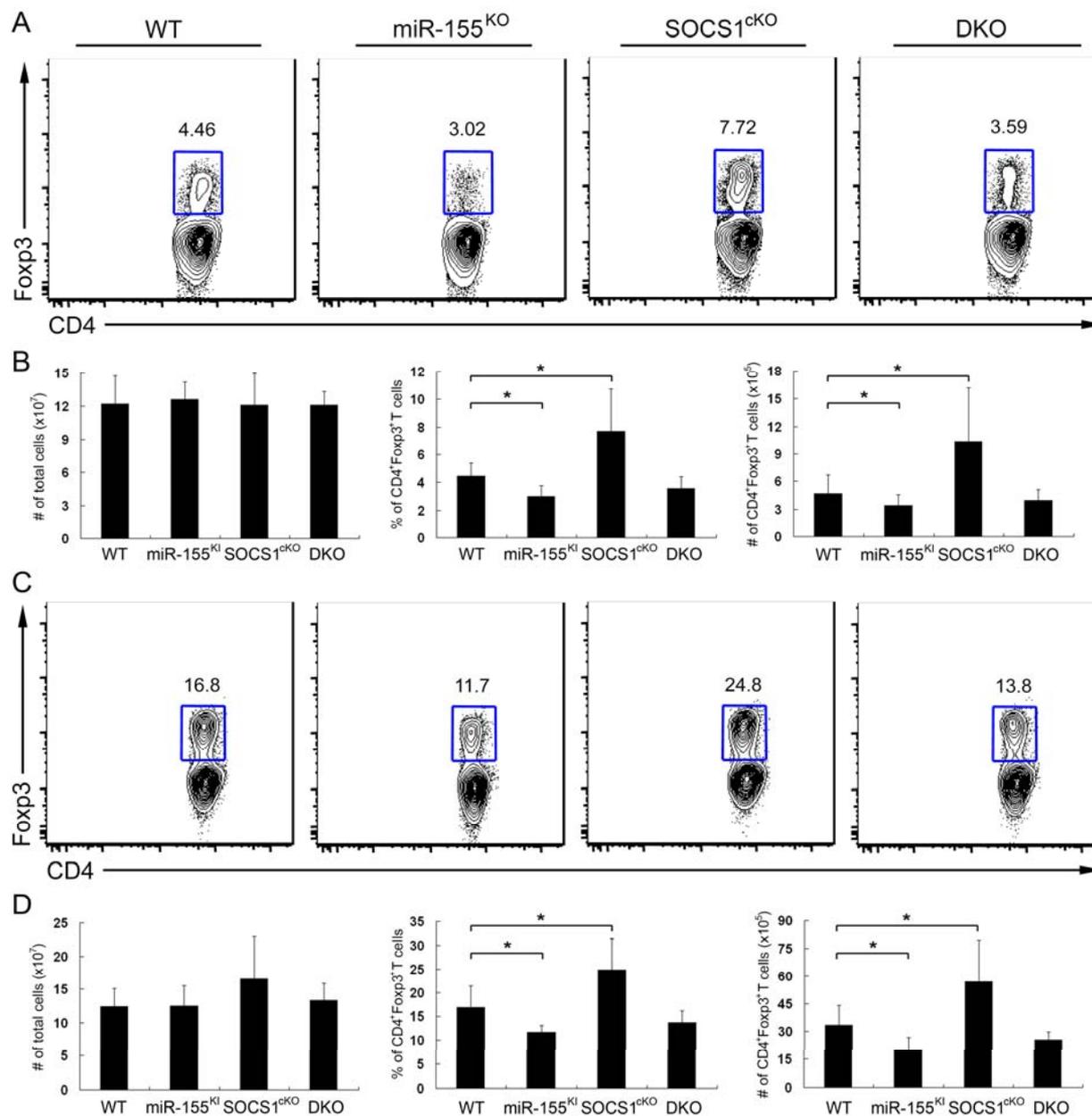


Figure S4 (related to Figure 4). Comparable production of proinflammatory cytokines by SOCS1^{KI} DCs. qPCR analysis of IL-6 expression in CD11c+ DCs isolated from WT, SOCS1^{KI} or miR-155^{KO} mice in the presence or absence of LPS stimulation. The data are shown as mean \pm SD and are pooled of 4 independent experiments (n = 7–8).

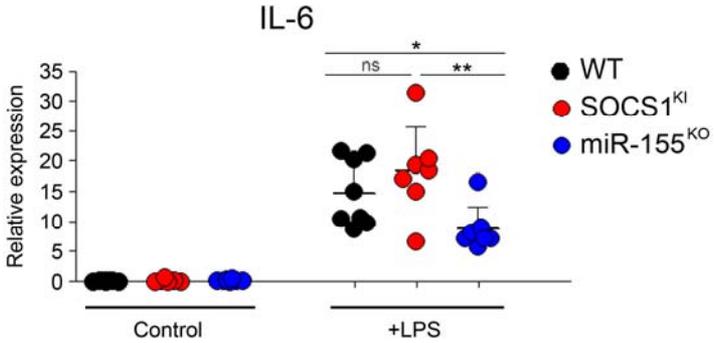


Figure S5 (related to Figure 4). Lack of miR-155-mediated SOCS1 regulation did not diminish Th17 polarization *in vitro*. CD4⁺CD25⁻CD62L^{hi} naive T cells isolated from WT, SOCS1^{KI} and miR-155^{KO} mice were cultured *in vitro* under Th17 polarizing condition. **(A)** FACS analysis and **(B)** frequencies of IL-17⁺ in CD4⁺ T cells were shown. The data are shown as mean \pm SD and are representative of three independent experiments (n = 9).

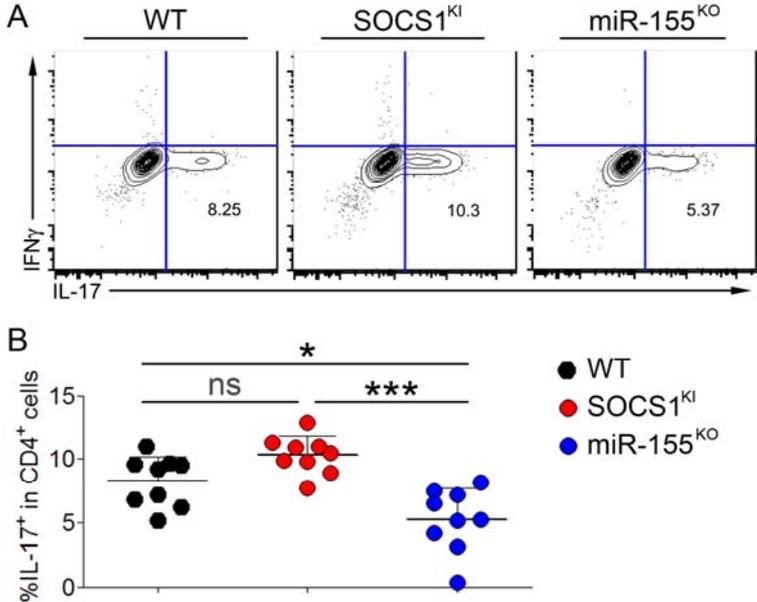


Figure S6 (related to Figure 7). Severe defect of miR155-deficient T cell response in both acute and chronic LCMV infection. CD45.2⁺ miR-155^{KO} or SOCS1^{KI} and CD45.1⁺/CD45.2⁺ naïve CD8⁺ P14 TCR-tg T cells (10⁴ each) were cotransferred into CD45.1⁺ wt hosts prior to infection with LCMV Armstrong (Arm) or Clone 13. Figure shows representative FACS plots and ratios of P14 T cells on d8 pi.

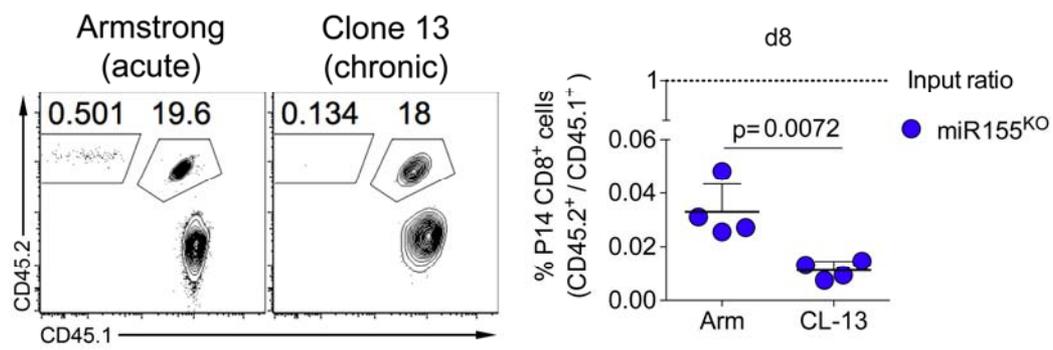


Figure S7 (related to Figure 7)..Unimpeded terminal differentiation and memory inflation of MCMV-specific SOCS1^{KI} T cells.
Mixed BM chimeras were infected with MCMV and MCMV-m38 tetramer binding CD8⁺ T cells were analyzed on d40 pi for expression of differentiation markers CD127 and KLRG-1

