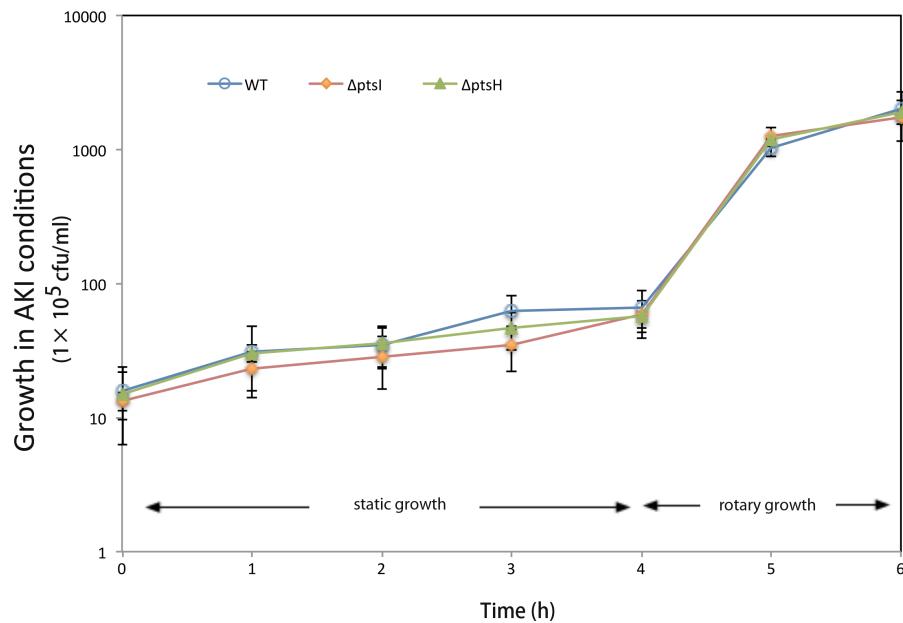


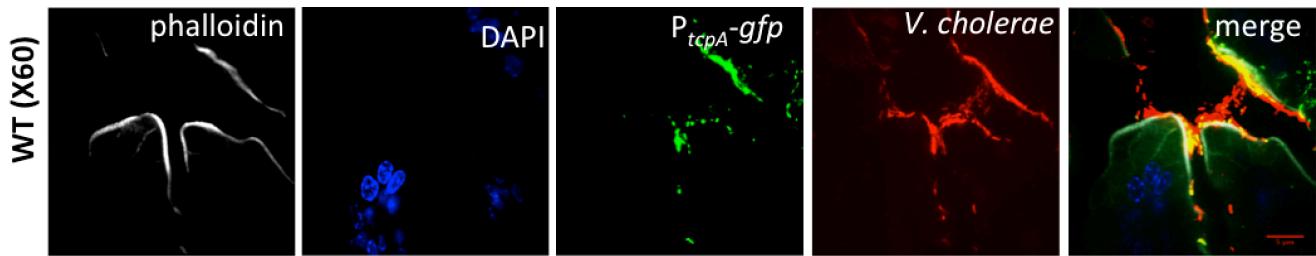
**Figure S1**

**Fig. S1** *aspK* and *cafA* deletion mutants show similar levels of TcpA compared to the wt in AKI conditions. TcpA expression detected by immunoblot in the indicated strains cultivated in AKI conditons; numbers correspond to densitometry measurements normalized to the value of the wt strain. Levels of RNAP alpha serve as a loading control. Densitometry measurements derived from triplicate experiments are shown.



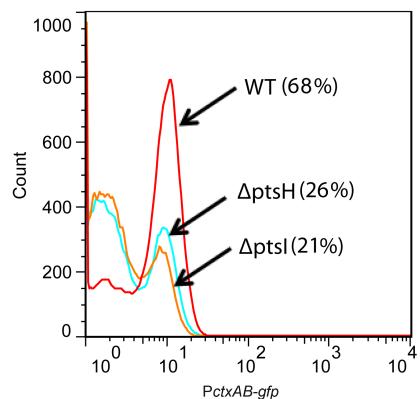
**Figure S2**

**Fig. S2** Growth curve of *V. cholerae* strains grown in AKI induction conditions. The cultures were sampled at indicated time points and plated on the LB agar plates with Streptomycin (200µg/ml) after serial dilutions. Three independent experiments were performed in triplicate.-



**Figure S3**

**Fig. S3** Heterogeneous expression of *tcpA* in wt *V. cholerae* is seen during infection of suckling mice. *V. cholerae* cells (red) in the small intestine were detected by immunofluorescence, and *tcpA* expression (green) was detected from the chromosomal  $P_{tcpA}$ -*gfp* reporter. Tissue sections were counterstained with phalloidin (gray) and DAPI (blue) to visualize actin and nuclei, respectively, and observed under 60× objectives.



**Figure S4**

**Fig. S4** GFP expression from a chromosomal  $P_{ctxAB}$ -*gfp* in the indicated strains after growth in AKI conditions and monitored by flow cytometry

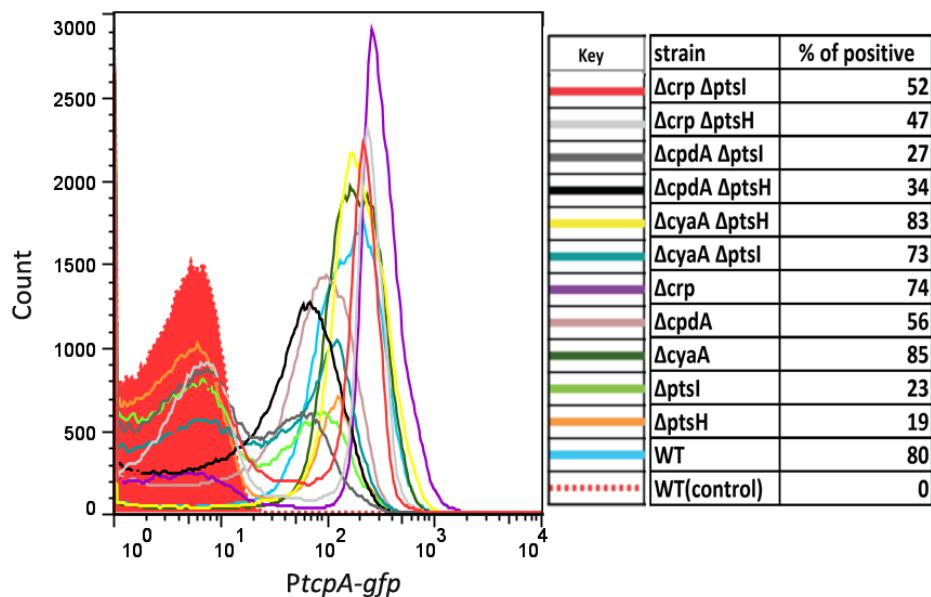


Figure S5

**Fig. S5** Flow cytometry assay of TcpA expression after the indicated strains were grown in AKI conditions. In the flow cytometry experiments, each strain contains a chromosomal  $P_{tcpA}$ -GFP reporter, except for a wt (control) strain that was used to establish the background level of fluorescence. The percentage of GFP positive cells (% of positive) are shown.

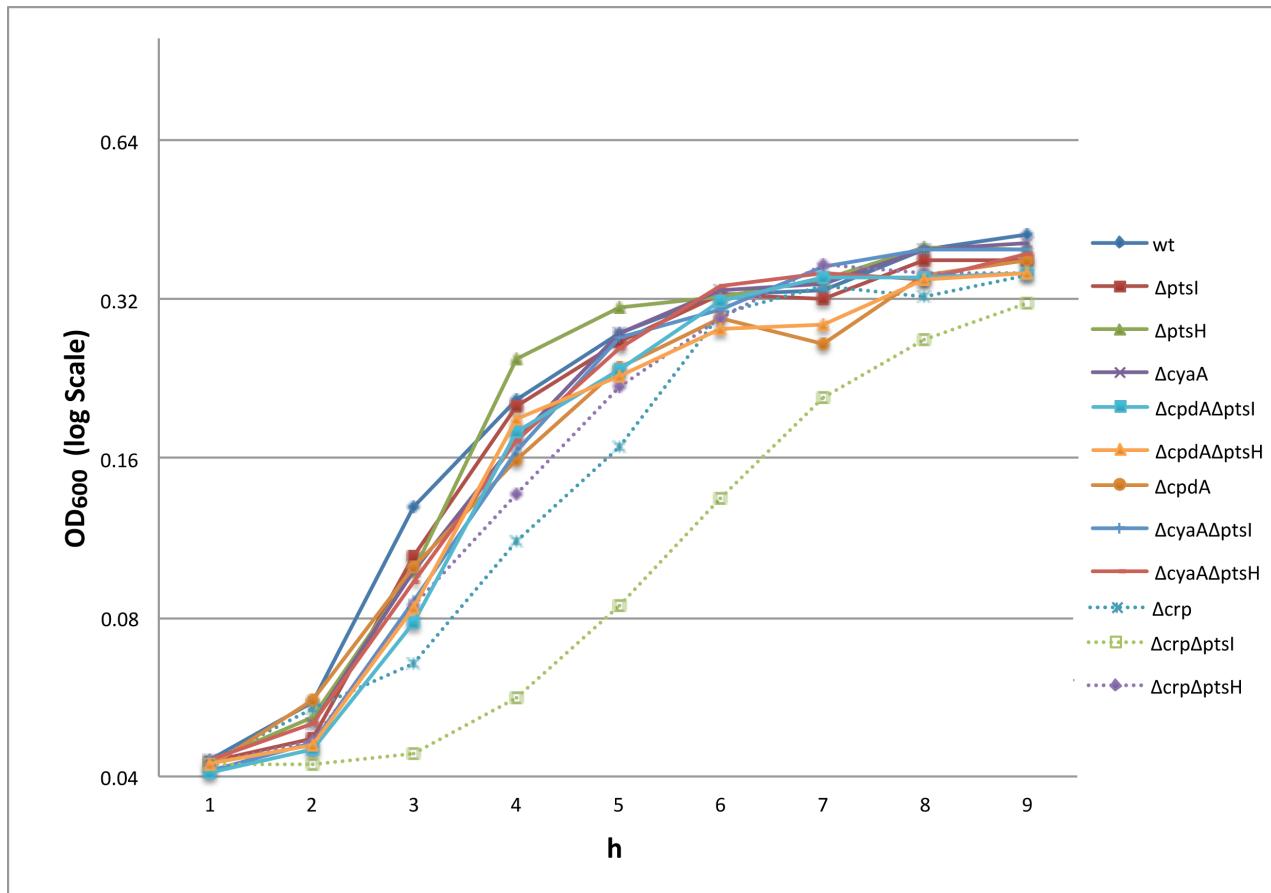


Figure S6

**Fig. S6** Growth curves of the wt and PTS related mutants incubated in LB. The bacterial cultures were sampled at indicated time points and OD<sub>600</sub> were assayed. Data represent three independent assays and values are reported as the mean.

Table S1 Primers used for this study

Construct	primer name	sequence (5'-3')*
$P_{tcpA}$ -bla	PtcpAF1	tggtcgtcggaatgaatcggttgcgtacccgGTCTTATCATGAGCCGCCTA
	PtcpAr1	GGAATAAGGGCGACACGAAATGTTGAATACTCATATTATATAACTCCACCATTGTGTTTAA
	blaF1	ATGAGTATTCAACATTCCGTGTC
	blar1	tgcgtcaactgatacatggcgatgttgcattGCGGCATCAGAGCAGATTG
$P_{tcpA}$ -GFP	PtcpAF2	tggtcgtcggaatgaatcggttgcgtacccgGTCTTATCATGAGCCGCCTA
	PtcpAr2	ACAACCTCCAGTAAAAGTTCTCCTTACTCATATTATATAACTCCACCATTGTGTTTAA
	gfpF1	ATGAGTAAAGGAGAAGAACTTTCACTG
	gpfr1	tgcgtcaactgatacatggcgatgttgcattTATTGTATAGTCATCCATGCCATG
$P_{ctxAB}$ -GFP	PctxF1	tggtcgtcggaatgaatcggttgcgtacccgATGTTCTTATTGTTACGGC
	PctxR1	ACAACCTCCAGTAAAAGTTCTCCTTACTCATATAATGCTCCCTTGTTAACAGAAA
	gfpF1	ATGAGTAAAGGAGAAGAACTTTCACTG
	gpfr1	tgcgtcaactgatacatggcgatgttgcattTATTGTATAGTCATCCATGCCATG
$\Delta$ VC0547	VC0547_UP_R	ACATTGCAGGGCTAACATTAGCATAGATCAGTATTCACCTCCAAGACCAAAGTCT
	VC0547_UP_F	agttatatgtatggtaaaaaggatcgatctGCGGGTGTAAACGTACTGGT
	VC0547_DOWN_F	GAGACTTGGTCTGGAAAGGTGAATACTGATCTATGTAATGTTAGACCTGCAAATGT
	VC0547_DOWN_R	ccggagagctcgatatcgatcggtaccttagTCGGAAAACAACCAACCG
	QY0547Dedo-R	TCTTGGGAGGCACATATTACGATTAA
	QY0547Deup-F	CTGCTGGAGAAAGAGATCCAACAGT
$\Delta$ VC0419	QY0419Dedo-R	AAGAGATCATCGAGCTGTTGACTA
	QY0419Deup-f	CGCCATTGGCATGTGAAATGACAATA
	VC0419_UP_R	TCACGCGCGTAATTGCGCAAATCTCAAGAGCGCACCTCAATGATTAAATGATTAGA
	VC0419_UP_F	agttatatgtatggtaaaaaggatcgatctATCCCCTAGACGTCGTGAAC
	VC0419_DOWN_F	TCTAACATTAAATCATTGAGGTGCGCTCTGAGATTTGCGCAAATTACGCG
	VC0419_DOWN_R	ccggagagctcgatatcgatcggtaccttagTCGCCGTAAATCAAGGACT
pCVD442 primer	CVD442F	ccagccctccgttggaaatg
	CVD442R	ACTGAGAAGCCCTTAGAGCC
$\Delta$ VC0965	0965DELUPF	agttatatgtatggtaaaaaggatcgatctGCAGACATCACTGTGACTTCTAACG
	0965DELUPR	TGGTATCAACCGATAATTACATTTCCTTTGATGAGGGCTAAATC
	0965DELDNF	CATCAAAAAAGAAAAATGTAATTATCGGTGATACCAAGGAGATAGG
	0965DELDOR	ccggagagctcgatatcgatcggtaccttagCCTACGGTCACTGAACCAGAAAG
	0965delTupf	TGTACGAGAAGCAAGTAGAAATCACT
	0965delTdor	TTACTTGGTCACCGTAGGATC
	0965delTinF	GCAACAGATATCCGTGATATCGG
	0965delTinR	AGATGCCGCTATTGAGAACT
	0966DELUPF	agttatatgtatggtaaaaaggatcgatctCGGTTGAACCGGCTGAATCA
$\Delta$ VC0966	0966DELUPR	TTGAAAGGAAGACGGATTACATGTTTATACCCAATGAGTTATTTTG
	0966DELDNF	TCATTGGGTATAAACATGTAATCCGTCTCCTTCAAAGCC
	0966DELDOR	ccggagagctcgatatcgatcggtaccttagTCGCTCAGCGATACGATGTG
	0966delTupf	AAGGCACTCAAGGTAAAGCTATTCT
	0966delTdor	TAGGCAACAAGGACAACCTCTTGAT
	0966delTinF	ACGAGAAGCAAGTAGAAATCACTGC

	0966delTinR	GTGAAGTTGGTCCATCAGAGCA
ptsI complement	VC0965pbadcomf	agcgaattcgagctcggtacccgggatccTTGACCAGTTAAGGTAAGGCTATGAT
	VC0965pbadcomr	aacagccaagcttgcattgcgactcgatgcaggcgCTATCTCCTTGGTATCAACCGATAATTACG
ptsH complement	VC0966pbadcomf	gccaattcgagctcggtacccgggatccACAAACTTACAAAAAATAAAACTCATTGGGG
	VC0966pbadcomr	aaacagccaagcttgcattgcgactcgatgcaggcgCATTTCGCGTAACAAAATGGCTTG
$\Delta cyaA$	cyaAdelTOutF	AATCACATGCCGCACGTGA
	cyaAdelToutR	ACTTTGTTTGACATCATCAGGCATATACA
	cyaAdelTinF	GCCAATTCTCATCATGAATGAAGAGCGT
	CyaAdelTinR	CAGTTGATAAACCAAGTTACGCATCACG
	cyaAdelUPF	aggtatatgtatggtaaaaaggatcgatccCTATAGTCACCAACTCCACCTGCA
	CyaAdelupR	ACGTCAACGAGTTACAAGTTGCTTCCCTGATATGACAGATAAAA
	CyaAdeldoF	ATCAGGGAAAGCAAACCTGTAACTCGTTGACGTCTCAGGTTTCTA
	CyaAdelDOR	cgggagagctcgatatcgatcggtaccttagGTGGCCCAGTTGCCGCAT
	crpdelToutF:	TTTCAGCGTGTCTACGGTCGA
$\Delta crp$	CrpdelToutR:	AAGGTTTCATTAGTGAGCTGAAAGTGGA
	CrpdelTinF:	TAAACCTCAACCGATCCAACACTAGA
	CrpdelTinR:	ATCAGGTTCTGCTCTCCAGCAT
	crpdelupF:	aggtatatgtatggtaaaaaggatcgatccTACTTCTAACCCACTCAATGCCAAG
	CrpdelupR:	TTATCGGGGCACTTAcatATAATCTCACTTCTCTGCAGGGTAC
	CrdeldoF:	CAGAGGAAGTGAGATTATTatgTAAGTGCCCCGATAACCCGTCTC
	CrdeldoR:	cgggagagctcgatatcgatcggtaccttagAAGAGTGTTCACCTGAGCTGCAAG
	cpdAdelToutF	GTGATGAGTCGGACTGAAGCG
$\Delta cpdA$	cpdAdelToutR	CCCTCTTCCACGGTCAGTTAG
	cpdAdelTinF	TCGATCAAGCTTATTCAAGATCACGG
	cpdAdelTinR	AAATCAGGTAAAATCGGCCTTGT
	cpdAdelupF	aggtatatgtatggtaaaaaggatcgatccTATCAATTGGTCAAAGATGGCGAA
	cpdAdelUPR	CGAGCAGTCATATCACAAATCGTAAACCTAACTCTGTTATT
	cpdAdeldoF	GGTTTACCGATTTGTGATATGACTGCTCGTCTGCT
	cpdAdeldoR	cgggagagctcgatatcgatcggtaccttagCCGCTTGGTACTTCTCAACCG
	0964delTUPF	AACTGCGACCGAAGGTCAT
$\Delta VC0964$	0964delTdor	TATTAAAGTGTGCAACACCGGTGCTT
	0964DELUPF	aggtatatgtatggtaaaaaggatcgatccGCTTCGATGAAAACATCGAAATCGG
	0964DELUPR	AATCGAACCAAGCGATTACATTGTGTCATGCTCTAACGTTT
	0964DELDOF	GGAGCATGACACAATGTAATCGCTTGGTTCGATTAAAACGT
	0964DELDOR	cgggagagctcgatatcgatcggtaccttagTCCCATTAAACGCCCGTCAT
	0672delTUPF	CGGCTAAATGTCGGATTGTGA
$\Delta VC0672$	0672delTdor	AACCAATCATGCCGATATCGC
	0672DELUPF	aggtatatgtatggtaaaaaggatcgatccTTTGTAAACAACCATGGTCAGGTTT
	0672DELUPR	TTCACATTGGTTCCTACATGCTTAACCTCTGTGAGTGT
	0672DELDOF	AGAGGTTAACGATGTAGGAACCAAATGTGAATTGAACTCTAG
	0672DELDOR	cgggagagctcgatatcgatcggtaccttagAATCGCAAAACACAGACCGA

	A0518delTUPF	GCGAGTTCTGTCAGTAATTGGCC
	A0518delTdor	ATTCATGCTGCGCAGTTGGCA
ΔVCA0518	A0518DELUPF	agttatgtatggtaaaaaggatcgatctAAACGCGCATAACTGGTGTTC
	A0518DELUPR	ATGTGATGCCCTTACATTCTTAACCTCTGTCTGCCTCTA
	A0518DELDOF	CAGGAGTTAAGAATGTAAGGGGCATCACATGACAAAAAAA
	A0518DELDOR	ccggagagctcgatcatgcggtaccttagTGAGATACTTGTGGCAAACCTGC
ΔVC1820	Vc1820delTUPF	GGCAGGATCTGGATCAACTGTTAC
	Vc1820delTdor	CCGTCATACCACCAAGAACATCGC
	Vc1820DELUPF	agttatgtatggtaaaaaggatcgatctATGCCCTTGGCTATAA
	Vc1820DELUPR	TAATTCACGCCCTACATTATTGGCTCATAAAGTATTAACGTGATGA
ΔVC2013	Vc1820DELDOF	TGAGCCAATAATGTAAGCGTGAATTATGAGTACCCCTAAC
	Vc1820DELDOR	ccggagagctcgatcatgcggtaccttagACCTAACACCACCTTAGAAGCGCC
	Vc2013delTUPF	TCCGACAAGCAACCGATCTTAAAG
	Vc2013delTdor	ATGGCCAGCTTTGGTGGAA
ΔVC1822	Vc2013DELUPF	agttatgtatggtaaaaaggatcgatctAGTCGTGAAAGCCTTACCTCT
	Vc2013DELUPR	GGGAAAGAATGAGATTACACCCAGTGAAAATGTAACCACCTT
	Vc2013DELDOF	ACATTTCACTGGGTGTAATCTCATTCTCCCCTAGAGAAAAG
	Vc2013DELDOR	ccggagagctcgatcatgcggtaccttagCTGGTATTGACTTGGCTATTGAAGA
ΔVC0995	Vc1822delTUPF	AGACAGCGGTTGGCAACATG
	Vc1822delTdor	ATCGCACTAGGCTCACTCACCA
	Vc1822DELUPF	agttatgtatggtaaaaaggatcgatctGACGCTGGCATTACATCATTCA
	Vc1822DELUPR	TGATTCAAGATGTTACACAATTGTTTACCTTACTGTTGTTGAA
ΔVCA1045	Vc1822DELDOF	AAGGTAAAACAATTGTAACATTCTGAATCACTACACTTGAGATAGA
	Vc1822DELDOR	ccggagagctcgatcatgcggtaccttagCTGCACTCATCGTGGCTCTG
	Vc0995delTUPF	GTGTAGATTTGCAGTCGTTAGCGC
	Vc0995delTdor	CCTTTTGAATCAGCTCAATCGCGTATT
ΔVC0911	Vc0995DELUPF	agttatgtatggtaaaaaggatcgatctATACATAATCAGCTAATCCTTGTGAT
	Vc0995DELUPR	GTAAACTCGATTACACCTTAAGTTCCCCTATAGGATT
	Vc0995DELDOF	GGGAACTTAAGGTGTAATCGAGTTAACCCCTAGCCTGAATC
	Vc0995DELDOR	ccggagagctcgatcatgcggtaccttagCATAACTGTCAAATAGTCAGAGGAGT
ΔVCA1045	Vca1045delTUPF	AACACCTTCATCTGTAATGTGTAGGT
	Vca1045delTdor	TGCAACGGTGGCACATCGCA
	Vca1045DELUPF	agttatgtatggtaaaaaggatcgatctCTTCCTACTTACGTATAGTGTACGTTCAT
	Vca1045DELUPR	GGAGCAAAACGTTACATCGCGTCCCCCTGGAT
ΔVC0911	Vca1045DELDOF	CGGGGGACGCGATGTAACGTTTGCTCCTGAGGCAA
	Vca1045DELDOR	ccggagagctcgatcatgcggtaccttagTCGACTGCGGAATCCACAAAGC
	Vc0911delToutF	TGATTGGTCGACGCTGTTGG
	Vc0911delToutR	CTATAGGGGTGACTCAATCAGGATGATG
ΔVC0911	Vc0911delupF	agttatgtatggtaaaaaggatcgatctCTTCATGTATGCTCCTCTCGTGTGATTAC
	Vc0911delupR	AGACACCTTGTGCTcatACTACTCTCACTACTCAAAAAATTGACT
	Vc0911delDOF	AAGTGAGAGTAGTatgAGCAACAAGGTGCTCTGAAAGTG

	Vc0911delDOR	ccggagagctcgatatcgcatcggtaccttagCGTGTTCGTATGGACGTAACACT
$\Delta V C 0910$	Vc0910delToutF	CGCCTATCTCAATTGGTGTGTTGA
	Vc0910delToutR	CACCACCGTTAACCGGCTTACG
	Vc0910delupF	aggatatgtatgggtaaaaaggatcgatccGCATGAGATTGTGGCGAATTACCA
	Vc0910delupR	AATCGGGTAGAGAttaCACCTATTGAATTATTATCATTGCAGCA
	Vc0910delDOF	TTCAATAGGGTGtaaTCTCTACCCGATTGAGGAGTGG
	Vc0910delDOR	ccggagagctcgatatcgcatcggtaccttagCCAAATATAGTAATCACGGTAAGGGCTAT
$\Delta V C A 0245$	Vca0245delToutF	GTTACGCGCCGAAGAAGATGC
	Vca0245delToutR	CGAGTGCCATTGCGGAGATCTC
	Vca0245delupF	aggatatgtatgggtaaaaaggatcgatccACAAGGTAGTATCACGGTCAGTGA
	Vca0245delupR	TACATACTCCCTCctaCATGGTTCTATCTCCAGAGTAGAGCT
	Vca0245delDOF	TGGAGATAGAACCATGtagGAGGGAGTATGTACAGCGAAGT
	Vca0245delDOR	ccggagagctcgatatcgcatcggtaccttagTGTGCACCGCATCTCGGCATT
qRT-PCR	toxTf	GCGTGGCAGATATTGTGG
	toxTR	GAAACGCTAGCAAACCCAGAC
	tcpAf	CTACCGCAAACGCAAATGCT
	tcpAR	GGTCAAGCCACCGACTGTAA
	ctxAf	TTGGAGCATTCCCACAACCC
	ctxAR	GCTCCAGCAGCAGATGGTTA
	ToxRf	GGTTTGGCTCGGGTTAGTGA
	toxrR	CAGCCAGCCAATGTTGTGAC
	toxSf	AGTGACGTCTACCCGACTGA
	toxSr	CTGCTGCTCAGTAGCTGGTT
	gyrBf	GTGGTTTCGGTGAAAGTGCC
	gyrBr	GTTTTCGCTTCGCTTGGGT
	AphAf	CGATGCAACCGGGTACGATA
	AphAr	TGGTTTGCCTTCCTGTGGTT
	AphBf	CTTGGCTGTTGGTCAGTTGC
	AphBr	GGTTGCCAGCCCTCAATACT
	cyaAf	TCATCCTCTGATCCCCGGTT
	cyaAr	ACAAGCCTAAAATGCCCGA
	Crpf	TCTTGATGCGCCTTCAGGT
	crpR	TCTGGCTGACGTGCAAGATT
	cpdAf	CATTACTGTGCTTGGCGACG
	cpdAr	GCAAACGTGAGTGAGCAGCAG
	tcpPf	GGGGTATGTCCCGTGTGATT
	tcpPr	GCTTGGGTACCAATCAGCC
	TCPHF	TAACGATCATCGCACTCCCG
	TCPHR	GTGGATCGGTCTGGGTAAGC
PtsI(H189A)	VC0965H189AUPR	AGCCATGATAGAAGTTGCAGAGGTACGGCCGCAA
	VC0965H189ADOF	GGCGGCCGTACCTCTGCAACTTCTATGGCTCGCTCT

PtsH(H15A)	VC0966H15AUPR	TGCAGCAGGACGAGTTGCAAGGCCGTTCTGCAGT
	VC0966H15ADOF	GCAGAAAACGGCCTTGCAACTCGTCCTGCTGCACAGT
VC0964(H91 D)	VC0964H91Dupr Vc0964H91Ddof	CGTGTGATAACGAAGTCAACAAACAGCTCAACGCCGTCA GTTGAGCTGTTGTTGACTTCGGTATCGACACGGTTGA

\*Lowercase nucleotides indicate the overhangs used for isothermal assembly in the construction of the plasmids.

Table S2 Genes found by TIS to be required for expression and Carb resistance from *PtcpA-bla*

Classification	Locus	Gene name	MannU ( <i>p</i> -value)*	TIGR annotation	Reference
Carb-response related genes	VC2635	<i>pbp1A</i>	1.0×10 <sup>-10</sup>	penicillin-binding protein 1A	1
	VC1887	<i>csiV</i>	0.00026	hypothetical protein	2
	VC2300	<i>ampG</i>	0.00004	AmpG protein, putative	3
	VC0581	<i>lpoA</i>	1.0×10 <sup>-10</sup>	lipoprotein, putative	1
Established TCP regulators	VC0838	<i>tcpN/toxT</i>	0.00001	TCP pilus virulence regulatory protein	4
	VC1049	<i>aphB</i>	1.0×10 <sup>-10</sup>	transcriptional regulator, LysR family	5
	VC2647	<i>aphA</i>	1.0×10 <sup>-10</sup>	PadR family transcriptional regulator	6
	VC0826	<i>tcpP</i>	1.0×10 <sup>-10</sup>	toxin co-regulated pilus biosynthesis protein P	7
	VC0827	<i>tcpH</i>	1.0×10 <sup>-10</sup>	toxin co-regulated pilus biosynthesis protein H	8
	VC0984	<i>toxR</i>	0.00006	regulatory protein ToxR	9
	VC1021	<i>luxO</i>	1.0×10 <sup>-10</sup>	LuxO repressor protein	10
	VC0347	<i>hfq</i>	0.00073	host factor-I, putative	11
	VC2529	<i>rpoN</i>	1.0×10 <sup>-10</sup>	RNA polymerase sigma-54 factor	12
Established factors involved in <i>in vivo</i> colonization	VC0034	<i>tcpG/dsbA</i>	0.00025	thiol:disulfide interchange protein	13
	VC0965	<i>ptsI</i>	1.0×10 <sup>-10</sup>	phosphoenolpyruvate-protein phosphotransferase, EI	14
	VC2433	<i>cpdA</i>	0.00419	cyclic AMP phosphodiesterase	14
	VC1097	<i>pta</i>	0.00014	phosphate acetyltransferase	14
	VC0724	<i>pstC-I</i>	1.0×10 <sup>-10</sup>	phosphate ABC transporter, permease protein	14
	VC0725	<i>pstA-I</i>	0.00014	phosphate ABC transporter, permease protein	14
	VC1839	<i>tolQ</i>	0.00074	TolQ protein	14,15
	VC1716	<i>mukF</i>	0.00094	MukF protein	14
	VC1714	<i>mukB</i>	1.0×10 <sup>-10</sup>	cell division protein MukB	14
Other putative TCP regulators	VC0966	<i>ptsH</i>	0.00009	Phosphocarrier protein HPr	
	VC0547	<i>aspK</i>	0.00103	aspartokinase, alpha and beta subunits	
	VC0419	<i>cafA</i>	1.0×10 <sup>-10</sup>	ribonuclease G/cytoplasmic axial filament protein	
	VC2736	<i>hslO/hsp33</i>	0.00005	33 kDa chaperonin (Heat shock protein 33 ) (HSP33)	
	VC2153	VC2153	0.00128	D,D carboxypeptidase-related protein	
	VC0346	<i>miaA</i>	0.00026	tRNA delta(2)-isopentenylpyrophosphate transferase	
	VC2775	<i>mnmG/gidA</i>	0.00372	tRNA uridine 5-carboxymethylaminomethyl modification enzyme	
	VC0556	<i>gshA</i>	0.00001	glutamate--cysteine ligase	
	VC2033	<i>adhE</i>	1.0×10 <sup>-10</sup>	alcohol dehydrogenase/acetaldehyde dehydrogenase	
	VC1866	<i>pflB</i>	0.00012	formate acetyltransferase	
	VC0003	<i>mnmE/trmE/thdF</i>	0.0008	thiophene and furan oxidation protein ThdF	
	VCA0511	<i>nrdD</i>	1.0×10 <sup>-10</sup>	anaerobic ribonucleoside-triphosphate reductase	

\*Limit of detection and cutoff are *p*-value <1.0×10<sup>-10</sup> and *p*-value <0.005, respectively, with the Mann Whitney U-based analysis of the TIS data.

## References

- 1 **Dörr T, Moll A, Chao MC, Cava F, Lam H, Davis BM, Waldor MK.** 2014. Differential requirement for PBP1a and PBP1b in *in vivo* and *in vitro* fitness of *Vibrio cholerae*. *Infect. Immun.* **82**:2115-2124.
- 2 **Dörr T, Lam H, Alvarez L, Cava F, Davis BM, Waldor MK.** 2014. A novel peptidoglycan binding protein crucial for PBP1A-mediated cell wall biogenesis in *Vibrio cholerae*. *PLoS Genet.* **10**:e1004433.
- 3 **Lindquist S, Weston-Hafer K, Schmidt H, Pul C, Korfmann G, Erickson J, Sanders C, Martin HH, Normark S.** 1993. AmpG, a signal transducer in chromosomal β-lactamase induction. *Mol. Microbiol.* **9**:703-715.
- 4 **Higgins DE, Nazarend E, DiRita VJ.** 1992. The virulence gene activator ToxT from *Vibrio cholerae* is a member of AraC family of transcriptional activators. *J. Bacteriol.* **174**:6974-6980.
- 5 **Kovacikova G, Skorupski K.** 1999. A *Vibrio cholerae* LysR homolog, AphB, cooperates with AphA at the *tcpPH* promoter to activate expression of the ToxR virulence cascade. *J. Bacteriol.* **181**:4250-4256.
- 6 **Skorupski K, Taylor RK.** 1999. A new level in the *Vibrio cholerae* ToxR virulence cascade: AphA is required for transcriptional activation of the *tcpPH* operon. *Mol. Microbiol.* **31**:763-771.
- 7 **Beck NA, Krukonis ES, DiRita VJ.** 2004. TcpH influences virulence gene expression in *Vibrio cholerae* by inhibiting degradation of the transcription activator TcpP. *J. Bacteriol.* **186**:8309- 8316.
- 8 **Carroll PA, Tashima KT, Rogers MB, DiRita VJ, Calderwood SB.** Phase variation in *tcpH* modulates expression of the ToxR regulon in *Vibrio cholerae*. *Mol. Microbiol.* 1997, **25**:1099-1111.
- 9 **Peterson KM, Mekalanos JJ.** 1988. Characterization of the *Vibrio cholerae* ToxR regulon: identification of novel genes involved in intestinal colonization. *Infect. Immun.* **56**:2822-2829.
- 10 **Miller MB, Skorupski K, Lenz DH, Taylor RK, Bassler BL.** 2002. Parallel quorum sensing systems converge to regulate virulence in *Vibrio cholerae*. *Cell* **110**:303-314.
- 11 **Lenz DH, Mok KC, Lilley BN, Kulkarni RV, Wingreen NS, Bassler BL.** 2004. The small RNA chaperone Hfq and multiple small RNAs control quorum sensing in *Vibrio harveyi* and *Vibrio cholerae*. *Cell* **118**:69-82.

- 12 **Lilley BN, Bassler BL.** 2000. Regulation of quorum sensing in *Vibrio harveyi* by LuxO and sigma-54. *Mol. Microbiol.* **36**:940-954.
- 13 **Peek JA, Taylor RK.** 1992. Characterization of a periplasmic thiol: disulfide interchange protein required for the functional maturation of secreted virulence factors of *Vibrio cholerae*. *Proc. Natl. Acad. Sci. U. S. A.* **89**:6210-6214.
- 14 **Merrell DS, Hava DL, Camilli A.** 2002. Identification of novel factors involved in colonization and acid tolerance of *Vibrio cholerae*. *Mol. Microbiol.* **43**:1471-1491.
- 15 **Bina JE, Mekalanos JJ.** 2001. *Vibrio cholerae tolC* is required for bile resistance and colonization. *Infect. Immun.* **69**: 4681-4685.