Supplementary Information

Difficidin and bacilysin from *Bacillus amyloliquefaciens* FZB42 have antibacterial activity against *Xanthomonas oryzae* rice pathogens

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Strain	Description	Source or reference
B. amyloliquefaciens		
FZB42	Wild type, producer of lipopeptides, polyketides and bacilysin	8
AK1	$\triangle bmyA$::Em ^r , deficient in bacillomycin D synthesis	8
AK2	<i>△fenA</i> ::Cm ^r , deficient in fengycin synthesis	8
CH1	$\triangle srfA$::Em ^r , deficient in surfactin synthesis	8
CH3	$\triangle sfp$::Em ^r , deficient in lipopeptides and polyketides synthesis	10
CH6	$\triangle bae$::Cm ^r , no synthesis of bacillaene	10
CH7	$\triangle mln$::Cm ^r , no synthesis of macrolactin	10
CH8	$\triangle dfn$::Em ^r , no synthesis of difficidin	10
RS2	$\triangle bac::Cm^r \ \triangle dfn::Em^r$, deficient in bacilysin and difficidin	26
RS6	$\triangle sfp$::Em ^r $\triangle bac$::Cm ^r , no lipopeptides, polyketides and bacilysin	21
Rice pathogenic bacteria		
PXO99 ^A	Xanthomonas oryzae pv. oryzae, causal agent of bacterial blight	27
RS105	Xanthomonas oryzae pv. oryzicola, causal agent of bacterial leaf streak,	2
	Rif ^r	

Table S1. Bacterial strains used in this study.

Xanthomonas oryzae pv. oryzae		Xanthomonas oryzae pv. oryzicola	
Name	Sequence (5 -3)	Name	Sequence (5 -3)
16 S -F	TGGCAACTAAGGACAAGGG	16S-F	AATGGGCGCAAGCCTGATC
16S-R	AAGCGGTGGAGTATGTGG	16S-R	AACCACCACCTACGCACGC
rpfF-F	GTTGGTCTGATAGCGGGTGA	rpfF-F	TAGCGGGTGATGTCGTCG
rpfF-R	TGAAGAACCGCAGCGTGAG	rpfF-R	TGAAGAACCGCAGCGTGAG
gumD-F	TTTCATAGTGCGTGTTGTGCT	gumD-F	AACGACGCACGCATCACG
gumD-R	AACGACGCACGCATCACG	gumD-R	ATCCAGCCACAGCGACCAAC
ftsZ-F	CTGCTGGACGATGTGAACCTG	ftsZ-F	CCGTGATGTCGGAAATGGG
ftsZ-R	ATAATCGTTGGGCAGGTCAGC	ftsZ-R	GCCAGGTTCACATCGTCCAG
glmS-F	CCCCAGCGATTTGGCGTAC	glmS-F	ACGATCAGATGCTGTGGGTTC
glmS-R	GCGTATGCGAACCCACAGC	glmS-R	GCCAAGGGTTTCCCGTCAT
rrlA-F	ACAGGGTGGTATTTCAAGG	rrlA-F	GCAAACTCCGAATACCAG
rrlA-R	GCCGCTGCGACTGTTTAT	rrlA-R	CAACCTCCTGGCTGTCTAT

Table S2. DNA primers used in this study.



Fig. S1. HPLC analysis of difficidin from B. amyloliquefaciens FZB42. The retention time of

difficidin is 8.574 min



Fig. S2. Detection of antagonistic action of B. amyloliquefaciens mutants strains against

Xanthomonas oryzae pv. *oryzae* (A) and *Xanthomonas oryzae* pv. *oryzicola* (B) by paper-disc agar diffusion assay. 1, complemented FZB42 \triangle sfp \triangle amyE::sfp; 2, complemented FZB42 \triangle *bmyA\triangleamyE::bmyA*; 3, complemented FZB42 \triangle fenA \triangle amyE::fenA; 4, complemented FZB42 \triangle *srfA\triangleamyE::srfA*; 5, complemented FZB42 \triangle bae \triangle amyE::bae; 6, complemented FZB42 \triangle mln \triangle amyE::mln; 7, complemented FZB42 \triangle dfn \triangle amyE::dfn; 8, complementation of RS6; 9, complementation of RS2.



Fig. S3. Phase-contrast and fluorescence microscope images of *Xanthomonas oryzae* pv. *oryzae* cells in the absence and presence of difficidin or bacilysin after 12 h of cultivation. Staining using the LIVE/DEAD BacLight bacterial viability kit assessed bacterial viability: green areas indicate live cells; red areas indicate dead cells. Untreated *X. oryzae* pv. *oryzae* cells were observed by A, phase-contrast microscopy and B-C, fluorescence microscopy. *X. oryzae* pv. *oryzae* cells after treatments with difficidin (10 μg/ml, 50 μg/ml) were examined by D and G, phase-contrast microscopy and E-F and H-I, fluorescence microscopy. Pictures were taken after treatments with bacilysin (10 μg/ml, 50 μg/ml) from the J to L and M to O. Bars indicate 20 μm.



Fig. S4. The phase-contrast and fluorescence microscope images of Xanthomonas oryzae pv.

oryzicola cells in the absence and presence of difficidin and bacilysin after 12 h of cultivation. Staining using the LIVE/DEAD BacLight bacterial viability kit assessed bacterial viability: green areas indicate live cells; red areas indicate dead cells. Bars: 20 µm.



Fig. S5. Pathogenicity test of *Xanthomonas oryzae* pv. *oryzae* and *Xanthomonas oryzae* pv. *oryzicola* strains on rice. A. Representative result of lesion length symptom tests on the leaves of adult susceptible rice after treatment with *B. amyloliquefaciens* FZB42 mutants. B. Representative result of water-soaking lesion length tests on rice seedling leaves after infiltration with *B. amyloliquefaciens* FZB42 mutants. C. Calculated lesion lengths on the leaves of susceptible adult rice. D. Calculated water-soaking lesion lengths on the leaves of rice seedlings. Data are expressed as means \pm standard deviation (SD); Different letters indicate means that differ statistically at P < 0.01.



Fig. S6. The level of gene *rrlA* and of glucosamine synthase activity (encoded by *glmS*) in *Xanthomonas* spp. A. The *rrlA* activity in *Xanthomonas* spp. treated with 10 µg/ml and 50 µg/ml difficidin for 30 min. β -galactosidase activity was assayed using o-nitrophenyl- β -D-galactopyranoside as the substrate and is reported in Miller units. B. The glucosamine synthase activity of *Xanthomonas* spp. treated with 10 µg/ml and 50 µg/ml bacilysin for 30 min. The measured activity in the sample containing no bacilysin (E₀) was 0.501 units for *X. oryzae* pv. *oryzae* and 0.489 units for *X. oryzae* pv. *oryzicola*. E represents the corresponding relative residual activities in the presence of bacilysin.