Table S1: PCR and sequencing primers used in this study

genetic structure	primer name	primer sequence (5'-3')	Amplicon (bp)	Ref
bla$_{\text{CTX-M}}$	CTX-MU1	ATGTGCAGYACCAGTAARGT	593	13
	CTX-MU2	TGGGTRAARTARGTSACCAGA		
bla$_{\text{SHV}}$	SHV-F	GCCCGGGTTATTCTTATTTGTCGC	1016	13
	SHV-R	TCTTTCCGATGCCGCCGCCAGTCA		
bla$_{\text{TEM}}$	TEM-F	ATGAGTATTCAACATTTCCG	861	13
	TEM-R	TTACCAATGCTTAATCAGTGAG		
bla$_{\text{OXA-9}}$	OXA-9 Fw	CGTCGCTCACCATATCTCCC	315	14
	OXA-9 Rv	CCTCTCGTGCTTTAGACCCG		
bla$_{\text{IMP}}$	IMP-F	GGAATAGAGTGGCTTAAYTC	232	15
	IMP-R	TCGGTTTAAYAAAACAACCACC		
bla$_{\text{VIM}}$	VIM-F	GATGGTGTTTGGTCGCATA	390	15
	VIM-R	CGAATGCGCAGCACCAG		
bla$_{\text{NDM}}$	NDM-F	GGTTTGGCGATCTGGTTTTC	621	16
	NDM-R	CGGAATGGCTCATCACGATC		
bla$_{\text{OXA-48}}$	OXA-48-F	GCGTGGTTAAGGATGAACAC	438	16
	OXA-48-R	CATCAAGTTCAACCCAACCG		
bla$_{\text{KPC}}$	KPC F	CGTCTAGTTCTGCTGTCTTG	798	16
	KPC-R	CTTGTCATCCTTGTTAGGCG		
internal probe	KPC For	CTTGCTGCCGCTGTGCTG	450	20
	KPC-Rev	CTTGTCATCCTTGTTAGGCG		
whole gene	KPC1F	GCTACACCTAGCTCCACCTTC	950	17
	KPC1R	ACAGTGGTTGGTAATCCATGC		

upstream bla_{KPC}	3098U	TGACCCTGAGCGGCGAAAGC		11
outward Tn*4401* region				
outward primers	EcoRIout	CACCCGACCTGGACGAACTA	2700	11
	141R-6	TCACCGGCCCTCACCTTTGG		
sequencing by primer walking	Bu13-1	CGTCCTGCTGCTCATTCA		This study
	Bu13-2	ATGTTATGGCTAACTCTAGGA		

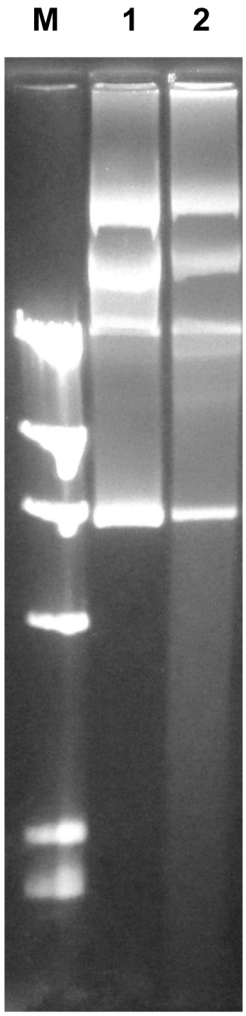

Fig. S1. Plasmid DNA from *E. coli* JM101 transformants.

Separation on 1 % agarose gel of plasmid extracts from JM101 transformed with the entire plasmid content of Kbu-1 (lane 1) and with the 13 kb band alone (lane 2). Lane M: Molecular Weight marker II (Roche).