

## Supporting Information

Janouškovec et al.: Plastid dependency and the early evolution of apicomplexans and their free-living relatives.

### SI Materials and Methods:

#### 1. Environmental sequencing and data analysis:

Environmental samples were collected in sterile Eppendorfs or Falcon tubes at several locations around Vancouver, BC, Canada (Table S1). DNA was extracted as soon as possible on the day of collection using using 0.5 - 1 g of sample material. Corals were healthy specimen sampled at the Birch Aquarium, Scripps Institution of Oceanography, San Diego (records were not available to length of time in captivity or place of origin; photos of each are available from A. T. Howe). Coral mucus was sampled by a 10 ml syringe and concentrated by centrifugation at 10,000 g for 15 min in 1.5 ml Eppendorf tubes. Coral tissue DNA was extracted from a small 0.5 g crushed coral fragment (tissue, skeleton and mucus). DNA from all samples was extracted by the standard protocol of the PowerSoil DNA Isolation Kit (MoBio Laboratories). The V4 and its flanking regions of 18S rDNA was PCR-amplified by using different combinations of the following custom primers sets: Apicomplexa-wide primers: 346F-mix (CADCGACGGGTAACGGGGAATTA; CAGYGACGGGTAACGGGGAATTA; CAGYGACGGGTAACGGGGAATTA; CAGYGACGGGTAACGGGGAATTA) and 785R-mix (IIITATTCCATGCTGIAGTATTCA; IIITATTCCATGCTAAASTATTCA). *Colpodella angusta* group-specific primers: Ctet151 (GAATTCTAGGGCTAATACATCGC); *Alphamonas edax* group-specific primers: Cedax122 (GATGGTYRCTCACTACWTGGATAA); *Voromonas pontica* group-specific primers: Cpont123 (TGGTCACTTACTACATGGATAACCGT); Genotype N group-specific primers: GN123 (TGGCTATATTACTACATGGATAACCG); *Chromera velia*-specific primers: Chrom219 (CCAATCCATCTCTGATGGTWT); *Chromera* and *Vitrella*-specific primers: ChrRMF (TCAMGRCCYGACTTCATGGAAG). PCR products were purified using the Polyethylene glycol (PEG) protocol and cloned using StrataClone PCR Cloning Kit (Agilent) following the instructions of the manufacturer. Clones were PCR-screened using the standard M13F and M13R primers and those containing the corresponding inserts were purified using the PEG protocol and Sanger-sequenced. The total of 78 apicomplexan-related partial 18S rDNA sequences were generated following the removal of duplicates (3), chimeric or low quality sequences (3), and sequences belonging to eukaryotes other than apicomplexans (7 dinoflagellate, 7 ciliate, and 2 rhizarian-like). Among the 78 sequences, 52 branched outside established apicomplexan clades (Figure 1) and 26 branched within them (Figure S2). All sequences were deposited in GenBank under accessions KP213181-KP213258 (Table S1).

Environmental 18S rDNAs sequences were obtained from GenBank by iterative BLASTn searches using both organismal and environmental queries. Hits shorter than 350 nucleotides were removed. Alignment-based check identified several chimaeric entries which were either removed completely (GIs: 309753092, 356459426, 529105829, 529105876, 408773533, 401668321, 213058553) or where the apicomplexan-related part of the sequence was removed (GIs: 89572015, 213058464, 171199596, 355346671, 213058384, 393405443, 60476916, 118420061, 374532687). Several entries that did not well overlap well with the rest of the alignment were removed including 9 sequences from the *Vitrella brassicaformis* clade (Gis: 300088848, 300088897, 300088909, 300088918, 300088921, 300088938, 300088949, 300088950, 300088963). This resulted in 174 environmental sequences from GenBank (generated in 41 studies, 36 published and 9 unpublished; Table S1) that were

subsequently combined with the 78 sequences generated by us. The divergent ends of some sequences were trimmed upon alignment. The apicomplexan affiliation of all 252 sequences was verified in a Maximum Likelihood phylogeny (ML) using a broad representative sampling of other eukaryotes (those distantly related to alveolates were not used further). Another ML analysis was used to verify that none of the 226 was specifically related to alveolates with very fast-evolving sequences (gregarines from crustacean hosts, *Trichotokara*, *Lecudina - Veloxidinium* clade, *Paralecudina polymorpha*, *Filipodium*, *Selenidium vivax/serpulae* cf. *echinatum*, ellobiopsids; none were used further).

## 2. Culturing and transcriptome sequencing:

*Chromera velia* CCMP2878 and *Vitrella brassicaformis* CCMP3155 were cultured in the marine f/2 medium at 22°C. *Voromonas pontica* isolate G-3, *Alphamonas edax* isolate BE-2, and *Colpodella angusta* isolates Spi-2 and BE-6 were cultured on enrichments of the marine bodonid *Procryptobia sorokini* strain B-69, the freshwater heterotrophic chrysophyte *Spumella* sp. isolate OF-40, respectively, and the freshwater bodonid *Parabodo caudatus* strain BAS-1, respectively. Prey organisms were raised on the suspension of *Pseudomonas fluorescens* (strain ICISC19) in the freshwater Pratt medium or marine Schmalz-Pratt medium at room temperature (see (1) for details). Total RNA was extracted using the RNAqueous kit (Ambion). Chromerid cells were first pelleted, the medium was removed, and the cells were broken by grinding in liquid nitrogen before being resuspended in the Lysis buffer. Colpodellid cells were separated from the medium by a gentle centrifugation (1000 rpm or less) on a 0.7 micrometer filter column and subsequently resuspended in the Lysis buffer. Chromerid transcriptomes were sequenced and assembled as described in Text S1 of Keeling et al. 2014 (2) at <http://www.plosbiology.org/article/info%3Adoi%2F10.1371%2Fjournal.pbio.1001889> and the data is available at: <http://camera.crbs.ucsd.edu/mmetsp/list.php>. Total RNA from colpodellid cultures was reverse-transcribed and amplified using the SMARTer Pico PCR cDNA Synthesis Kit (Clontech). Transcriptome of *Spumella* sp. prey was prepared using the same protocol as those of colpodellids. Transcriptomes were generated by Illumina HiSeq sequencing (100bp paired-end reads; multiplexing 0.25 lane per each of the four samples) producing the following number of raw reads (forward+reverse) 177,414,112 raw reads (*A. edax* BE-2); 50,933,148 (*V. pontica* G-3), 42,068,699 raw reads (*C. angusta* Spi-2), and 53,155,624 raw reads (*C. angusta* BE-6). Cutadapt 1.2.1 was used to trim SMARTer adaptors and 3' Illumina sequencing adaptor (in short fragments). Trimmed reads were assembled in Inchworm (Trinity r2012-06-08) using the default settings (K25 L25) and the minimum contig length of 150 nucleotides. Contamination by *Spumella* prey sequence in *A. edax* transcriptome and cross-contamination between colpodellid transcriptomes (a likely consequence of sequencing all samples in a single lane) was removed by using a custom script as follows: for any two contigs that were  $\geq 95\%$  similar between two samples, contig kmer coverages (produced by Inchworm) were compared; if  $\geq 50$  times difference in coverage was found then the low-coverage contig was discarded as a probable contaminant, otherwise both contigs were discarded. Contigs highly similar (95% or more) to known bacterial sequences were also removed by using a custom script, however, many bacterial contaminants were retained due to the lack of reference sequences. Purified contigs from the four colpodellid transcriptomes were deposited in GenBank Sequence Read Archive. Contigs of the two *C. angusta* strains (Spi-2 and BE-6) were pooled in further analysis after being found to be nearly identical at the nucleotide level (differences if present were typically limited to non-coding terminal regions). *Chromera velia* and *Voromonas pontica* contigs were pooled with those generated in independent studies (3, 4). Open reading frames (ORFs) were six-frame translated into amino acids using GetORF (EMBOSS). DNA surveys from *A. edax* and

*C. angusta* cells were prepared by Epicentre DNA extraction kit, followed by sequencing and assembly as above.

### 3. Analysis of plastid dependency:

In order to study distribution of diverse characteristics in apicomplexans and relatives, a custom BLAST database was built. This included protein sequences predicted from the 5 chrompodellid transcriptomes, 11 apicomplexan genomes (*Babesia bovis* T2Bo, *Babesia microti* RI, *Cryptosporidium muris* RN66, *Cryptosporidium parvum* Iowa II, *Eimeria maxima*, *Eimeria tenella* Houghton, *Neospora caninum* LIV, *Plasmodium berghei* ANKA, *Plasmodium falciparum* 3D7, *Plasmodium vivax* SaI1, *Theileria annulata* Ankara, *Theileria parva* Muguga, *Toxoplasma gondii* ME49), the small-scale genome sequence survey of *Ascogregarina taiwanensis*, the unpublished genome draft from *Gregarina niphandrodes*, 11 dinoflagellate transcriptomes (*Alexandrium catenella*, *A. minutum*, *A. ostenfeldii*, *A. tamarense*, *Amphidinium carterae*, *Heterocapsa triquetra*, *Karenia brevis*, *Karlodinium micrum*, *Lingulodinium polyedrum*, *Oxyrrhis marina*, *Symbiodinium* sp. kb8), the unpublished draft genome of *Perkinsus marinus* ATCC50983 and the draft genome of the dinoflagellate *Symbiodinium minutum*, and 53 genomes from representative eukaryotes (*Acanthamoeba castellanii* Neff, *Albugo laibachii* Nc14, *Allomyces macrogynus* ATCC 38327, *Aplanochytrium kerguelense* PBS07, *Arabidopsis thaliana*, *Aspergillus fumigatus* A1163, *Aureococcus anophagefferens* CCMP1984, *Bathycoccus prasinus* RCC1105, *Bigelowiella natans* CCMP2755, *Blastocystis hominis*, *Calliarthron tuberculosum*, *Capsaspora owczarzakii* ATCC30864, *Chlamydomonas reinhardtii*, *Chlorella variabilis* NC64A, *Chlorokybus atmophyticus* UTEX2591, *Chondrus crispus* PCG, *Coccomyxa subellipsoidea* C169, *Cyanidioschyzon merolae* 10D, *Cyanophora paradoxa*, *Dictyostelium discoideum* AX4, *Ectocarpus siliculosus* Ec32, *Emiliana huxleyi* CCMP1516, *Entamoeba dispar*, *Euglena gracilis*, *Galdieria sulphuraria*, *Giardia intestinalis*, *Glaucocystis nostochinearum*, *Guillardia theta* CCMP2712, *Ichthyophthirius multifiliis*, *Leishmania major* Friedlin, *Mesostigma viride*, *Micromonas* sp. RCC299, *Monosiga brevicollis* MX1, *Naegleria gruberi* NEG-M, *Nannochloropsis gaditana* CCMP526, *Ostreococcus* sp. RCC809, *Paramecium tetraurelia*, *Phaeodactylum tricornutum* CCAP1055-1, *Physcomitrella patens patens*, *Porphyridium purpureum*, *Pyropia yezoensis*, *Pythium ultimum* BR144, *Rhizopus oryzae*, *Saprolegnia parasitica* ATCC90214, *Spizelomyces punctatus* DAOM BR117, *Sterkiella histriomuscorum*, *Tetrahymena thermophila*, *Thalassiosira pseudonana* CCMP1335, *Thecamonas trahens* ATCC50062, *Trichomonas vaginalis*, *Trichoplax adhaerens* Grell-BS-1999, *Trypanosoma brucei gambiense*, *Ustilago maydis*). Homologs to eukaryotic queries were retrieved based on BLASTp searches with a variable cutoff. Prokaryotic homologs were retrieved from a custom database (5-10 representatives per phylum) and GenBank (additional sequences most closely related to apicomplexan or chrompodellid queries). Plastid targeting signals were analysed in selected plastid proteins with complete N-termini. Proteins that screened positively for signal peptides in SignalP 4.1 were further tested for the presence of transit peptides in ChloroP 1.1 and listed in Table S3.

### 4. Analysis of the distribution of apicomplexan-specific proteins:

a) 21 bacterial genes predicted to have been acquired in the apicomplexan ancestor (Fig. 4A) were identified from published literature and during preliminary analyses in this study (Table S5). Out of the 23 bacterial transfers previously identified in *Cryptosporidium* ((5); malate and lactate dehydrogenase represent the same HGT event), 12 were examined here (Table S5). The remaining 11 were either specific to *Cryptosporidium* (EAK89684.1,

AAU15128.1, EAK88388.1, EAK88222.1, AAS47699.1, EAK87293.1, AAL83208.1, EAK87294.1, EAK89739.1, EAK89015.1) or excluded because of the divergent character of the sequences (acetyltransferase, EAK87438.1; a homolog was identified in *Alphamonas*, however, suggesting that neither this gene represents an apicomplexan-specific HGT). Retrieval of eukaryotic and bacterial sequences was done as described above. Alignments, removal of variable sites, RAxML, and PhyML phylogenies were done as described in Materials and Methods (in final alignments shorter than 150 amino acids only aLRT supports were calculated; Fig. S5). Results were summarized in Table S5 and Fig. S5.

b-d) 185 OrthoMCL clusters (OG5 edition; (6)) were downloaded from <http://www.orthomcl.org>. 527 Apiloc proteins (representing 382 OrthoMCL clusters) were downloaded from <http://apiloc.biochem.unimelb.edu.au/apiloc>. 173 Wasmuth et al. clusters were downloaded from the Supplement of the paper (7). A query representing each cluster or Apiloc protein (most frequently the sequence from *Toxoplasma gondii*) was then used to search homologs in our eukaryotic database using BLASTp searches, as described in Materials and Methods. All hits were evaluated by eye to prevent misassignment owing to the presence of repeats and amino acid bias both of which are common in secreted proteins. Proteins that contained any regions or domains with significant hits ( $<1e-5$  evaluate similarity) to eukaryotes other than apicomplexans or other myzozoans were removed from further searches as 'eukaryotic' (all control proteins localized to Golgi belonged here). Proteins that were limited to certain apicomplexan subgroups only were likewise removed (many Apiloc proteins; Table S5). These strict homology assignments identified 102 protein cluster candidates that appeared to have originated during early apicomplexan or myzozoan evolution. Further in-depth similarity searches by using these proteins as queries and even stricter criteria (all hits  $1e-3$  to  $1e-5$  were thoroughly evaluated) led to removal of some of the candidates and produced the final count of 74 protein clusters that appeared specific to either apicomplexans ( $n=19$ ), apicomplexans and chrompodelids ( $n=29$ ), or all myzozoans ( $n=26$ ). Alveolins and oocyst wall proteins (OWPs) clusters identified through this process did not share significant similarity to other eukaryotes, however, they belonged to larger families, and some members of these families shared sequence similarity with ciliates (8), and fungi/insects/bacteria (9), respectively. In the case of alveolins, the pattern is consistent with the origin in early alveolates followed by diversification prior to myzozoan divergence (Fig. 4B). In the case of OWPs, the similarity may result from convergent evolution on the same function (9). The distribution of additional proteins (e.g., MyoA, subtilisin proteases, ribonucleotide reductase, proteins lost in apicomplexans) was determined in an analogous fashion and their phylogenies were done as described in Materials and Methods.

## SI References:

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## SI Figure Legends:

**Figure S1:** Phylogeny of the reference 18S rDNA alignment. Maximum likelihood phylogeny (RAxML; GTR+GAMMA+4 model) of 109 eukaryotic 18S rDNA sequences with sequence gi accessions after the @ symbol. Branch support values correspond to 300 non-parametric bootstraps. Supports  $\geq 60$  are shown. Full black circles correspond to complete support (100%).

**Figure S2:** Phylogeny of environmental sequences generated in this study that cluster with known apicomplexan clades. Maximum likelihood phylogeny (RAxML; GTR+GAMMA+4 model) of eukaryotic and environmental 18S rDNA sequences with non-parametric bootstrap supports (300 replicates) and environment types listed according to the key. Supports  $\geq 60$  are shown. Full black circles correspond to complete support (100%). Sequence accessions (gi) are specified following the @ sign.

**Figure S3:** SufB phylogeny. Maximum likelihood phylogeny (RAxML; LG+GAMMA+4 model) with non-parametric bootstraps (300 replicates) / PhyML aLRT supports at relevant nodes. Upper values correspond to the dataset shown, lower values correspond to dataset with fast-evolving sequences excluded (*Vitrella* and *Colpodella*, indicated by asterisks). The position of apicomplexan plastid-encoded SufB was established in a separate analysis and is shown by

a dashed line.

**Figure S4:** ClpC phylogeny. Maximum likelihood phylogeny of ClpC and the closest ClpB paralogs (RAxML; LG+GAMMA+4 model) with PhyML aLRT supports at relevant nodes. First values correspond to the dataset shown, second values correspond to dataset with fast-evolving sequences excluded (*Chromera* and apicomplexans, indicated by asterisks).

**Figure S5:** Phylogenies of 13 bacterial proteins in apicomplexans and relatives. Maximum likelihood phylogeny (RAxML; LG+GAMMA+4 model) with non-parametric bootstraps (300 replicates) / PhyML aLRT supports at relevant nodes (datasets shorter than 150 amino acid positions were not bootstrapped). Predicted horizontal gene transfer (HGT) events are highlighted.

**Figure S6:** Phylogeny of ribonucleotide reductase. Maximum likelihood phylogeny (RAxML; LG+GAMMA+4 model) reveals that the R2e2 type (previously thought to be unique to apicomplexans and not closely related to the R2e1 type) is also found in chrompodellids and originated by lineage-specific duplication from the R2e1 type.

**Figure S7:** Phylogeny of subtilisin proteases. Maximum likelihood phylogeny (RAxML; LG+GAMMA+4 model) shows that the majority of apicomplexan subtilisin proteases (e.g. SUB1-12 in *Toxoplasma*) share an ortholog in chrompodellids (or other myzozoans).

**Table S1:** Environmental sequences related to apicomplexans that were used in this study.

**Table S2:** Abbreviations and full names of proteins and pathways used in Fig. 3.

**Table S3:** Chrompodellid plastidic proteins with complete N-terminal sequence.

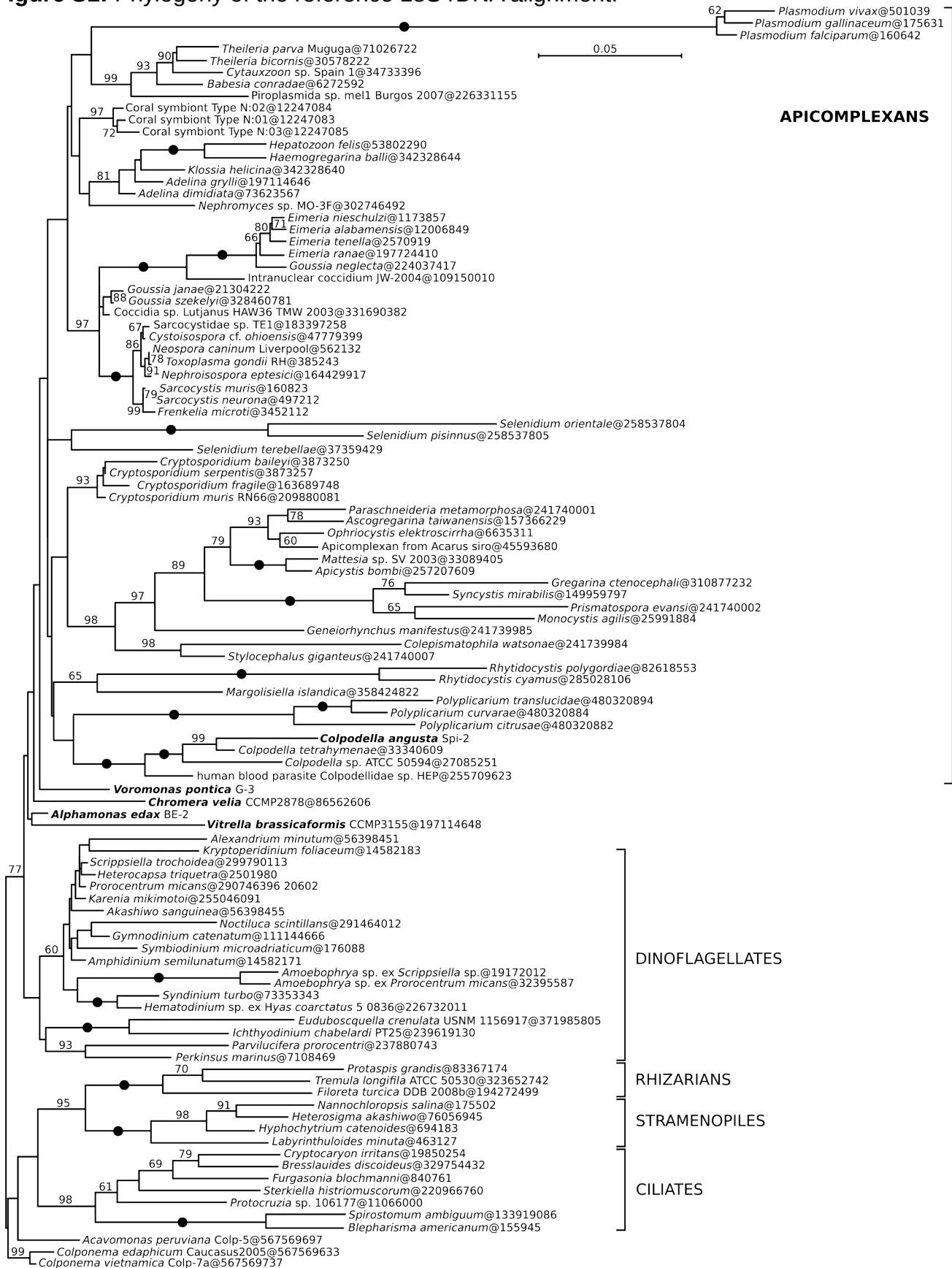
**Table S4:** Classification of 527 proteins from Apiloc (corresponding to 382 OrthoMCL clusters) based on their putative origin.

**Table S5:** Classification of 21 bacterial genes in apicomplexans and relatives acquired by horizontal gene transfer (HGT).

**Table S6:** Classification of 74 unique proteins specific to apicomplexans, apicomplexans and chrompodellids, and myzozoans.

**Table S7:** Domain superfamilies in chrompodellid and dinoflagellate PKSII/FASII proteins identified by NCBI Conserved Domains searches.

Figure S1: Phylogeny of the reference 18S rDNA alignment.





**Figure S2: Phylogeny of environmental sequences generated in this study that cluster with known apicomplexan clades.**

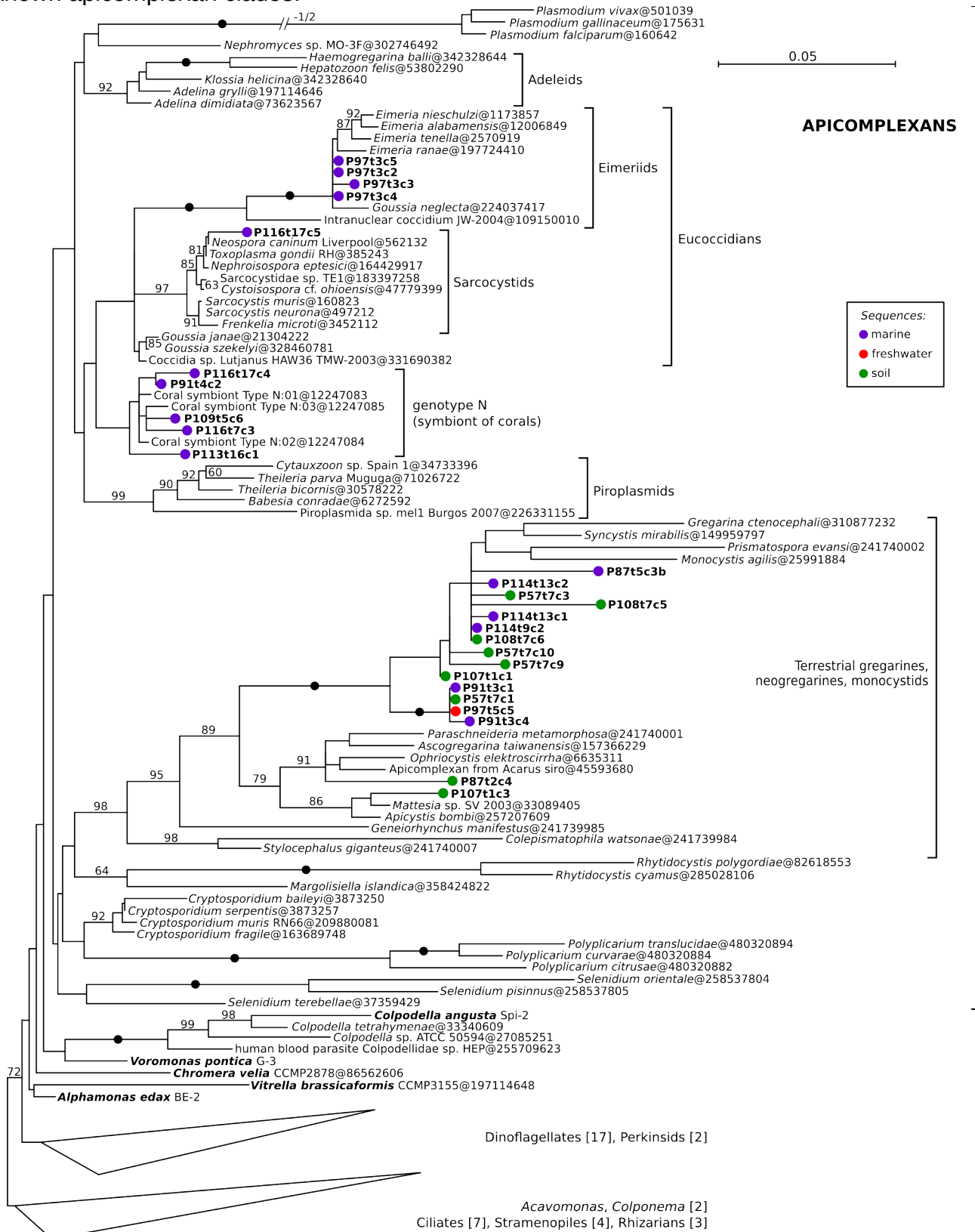
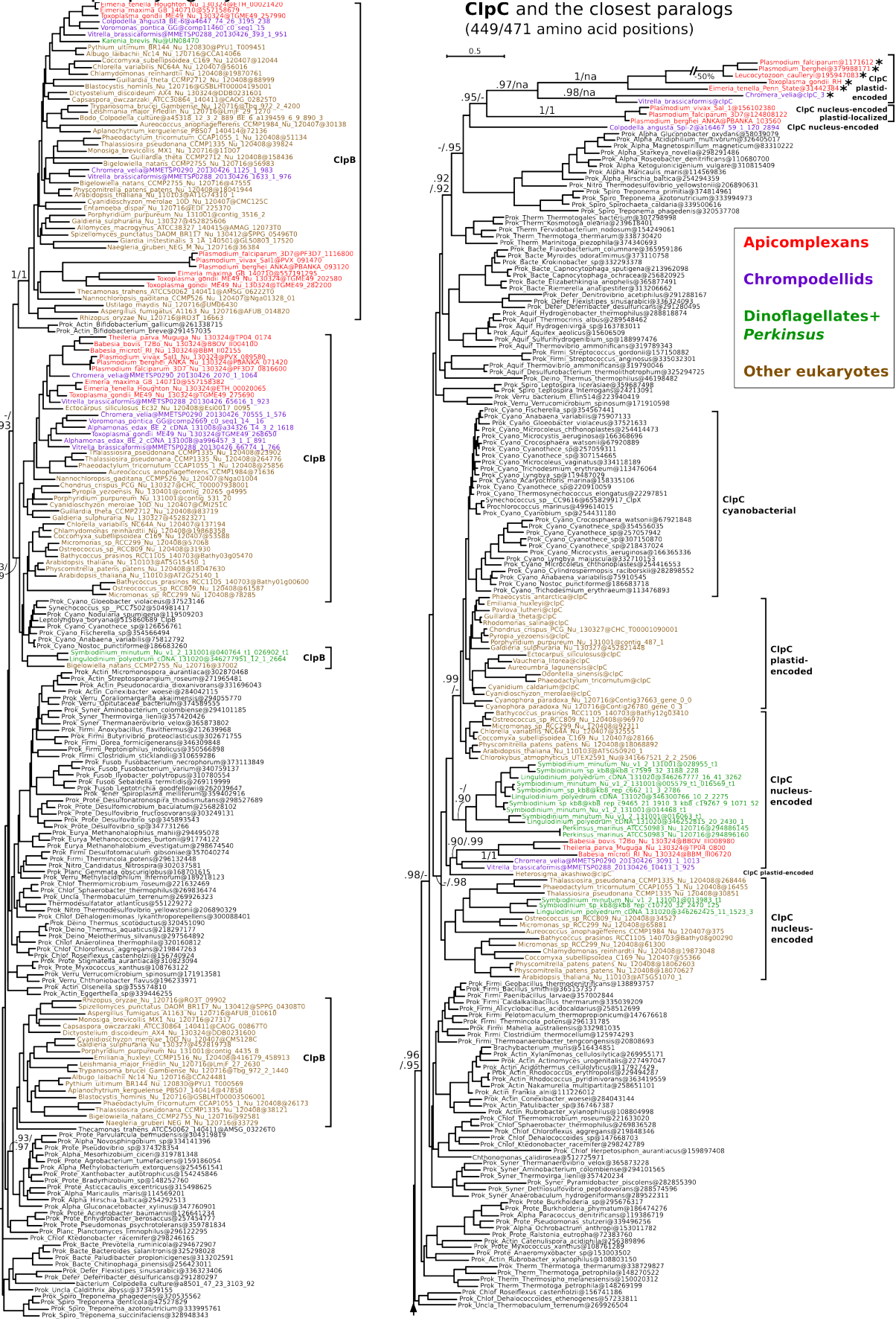


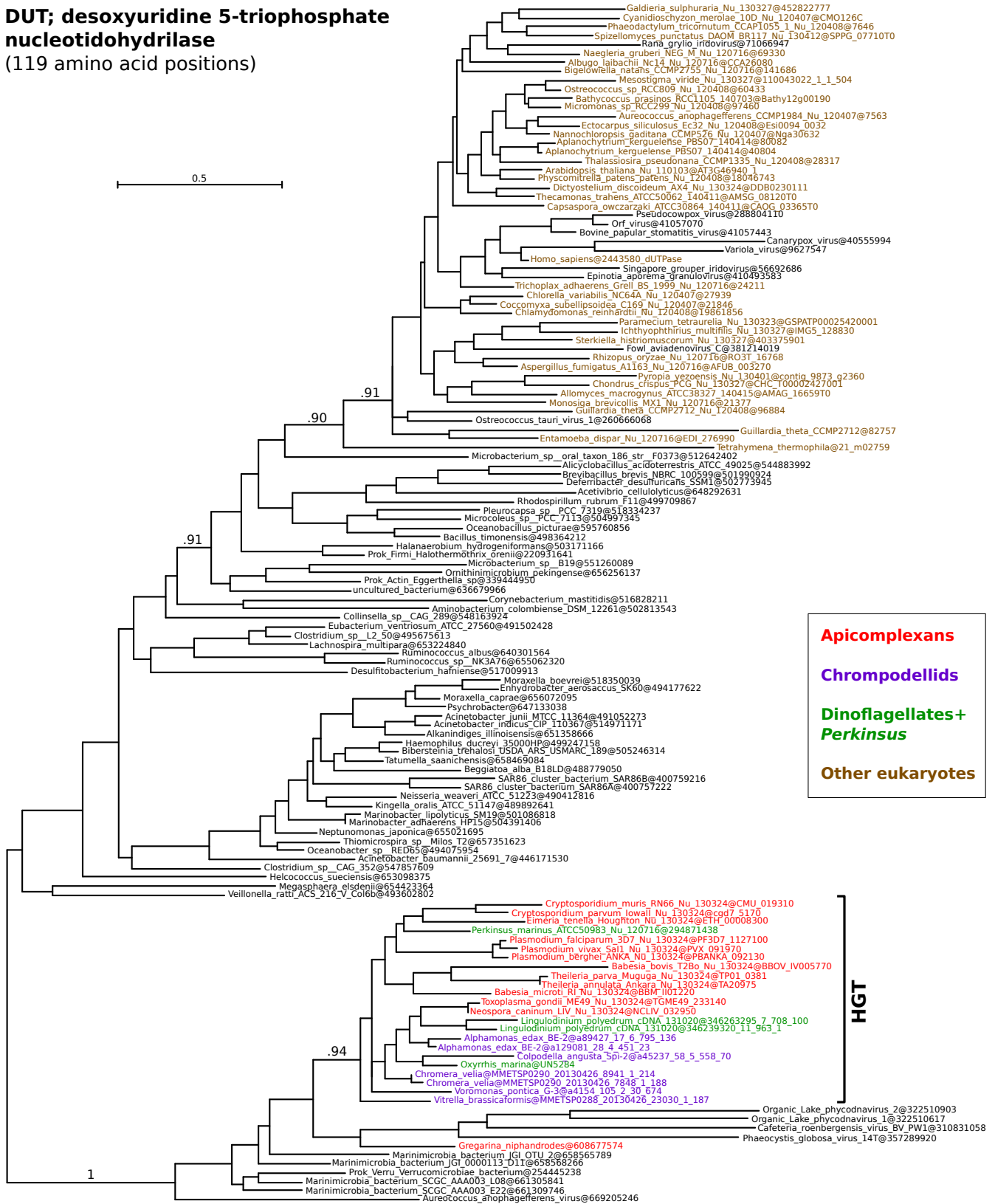


Figure S4: ClpC phylogeny.

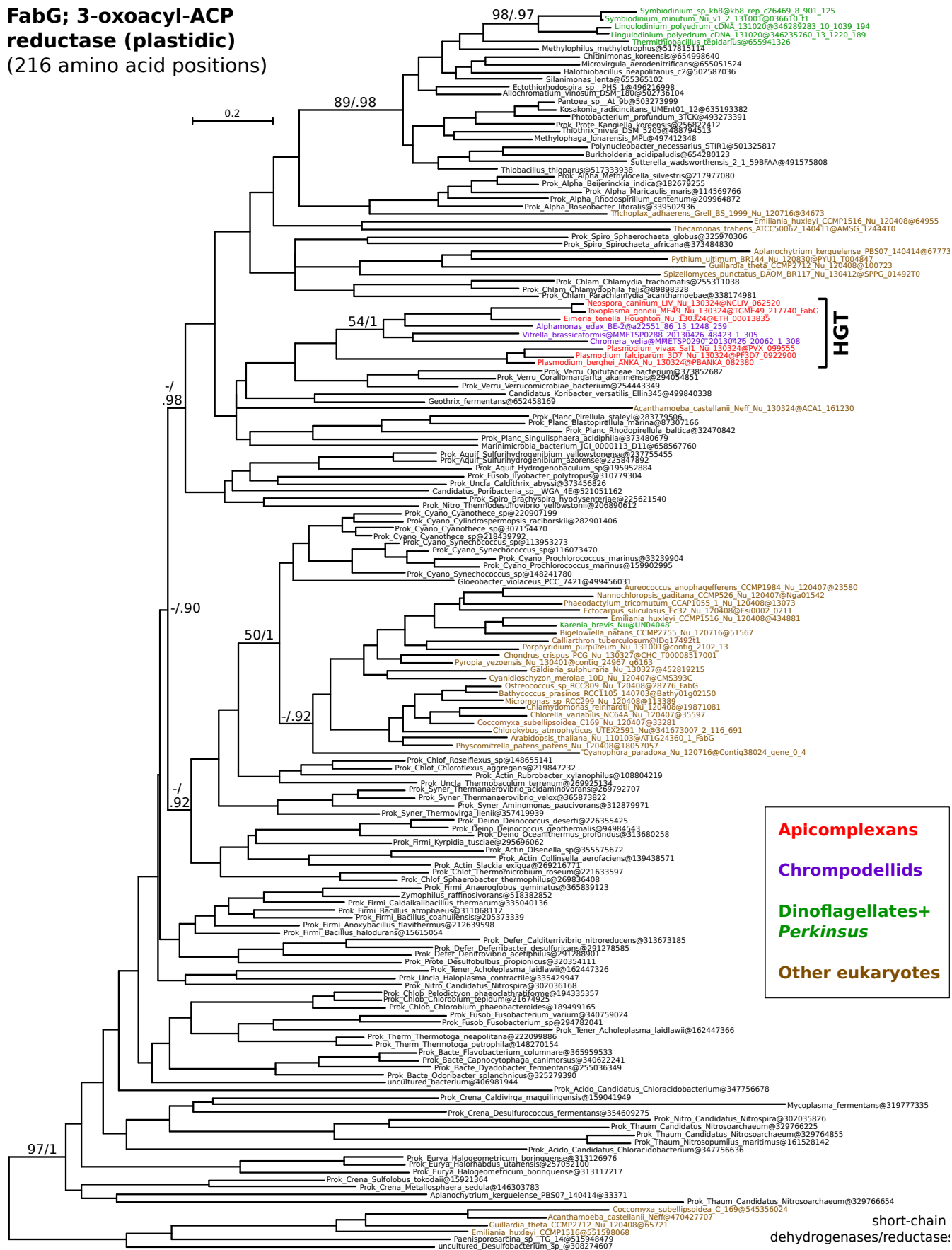


**Figure S5: Phylogenies of 13 bacterial proteins in apicomplexans and relatives.**

**DUT; desoxyuridine 5-triophosphate nucleotidohydrilase**  
(119 amino acid positions)



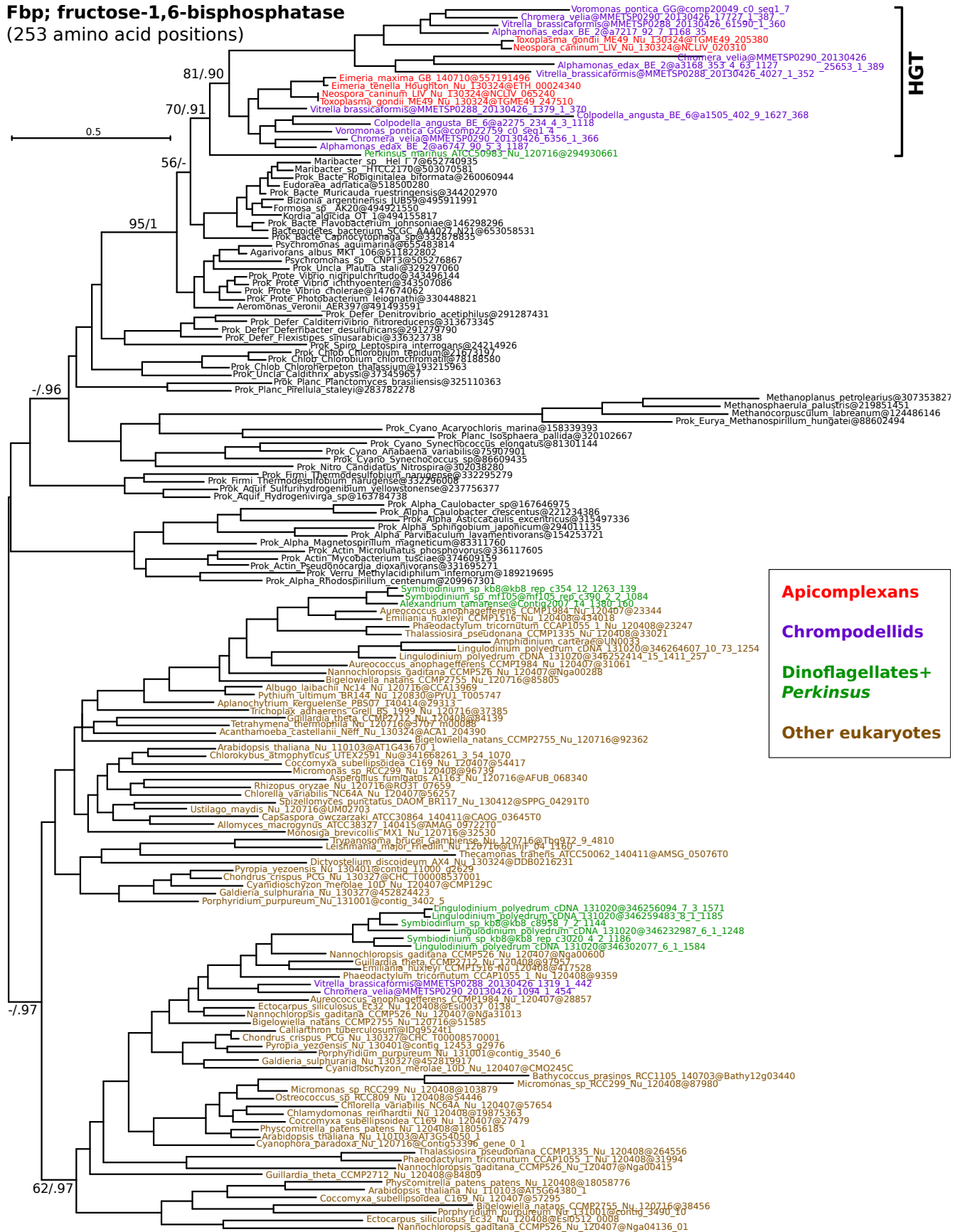
**FabG; 3-oxoacyl-ACP reductase (plastidic)**  
(216 amino acid positions)



**Apicomplexans**  
**Chromodellids**  
**Dinoflagellates+ Perkinsus**  
**Other eukaryotes**

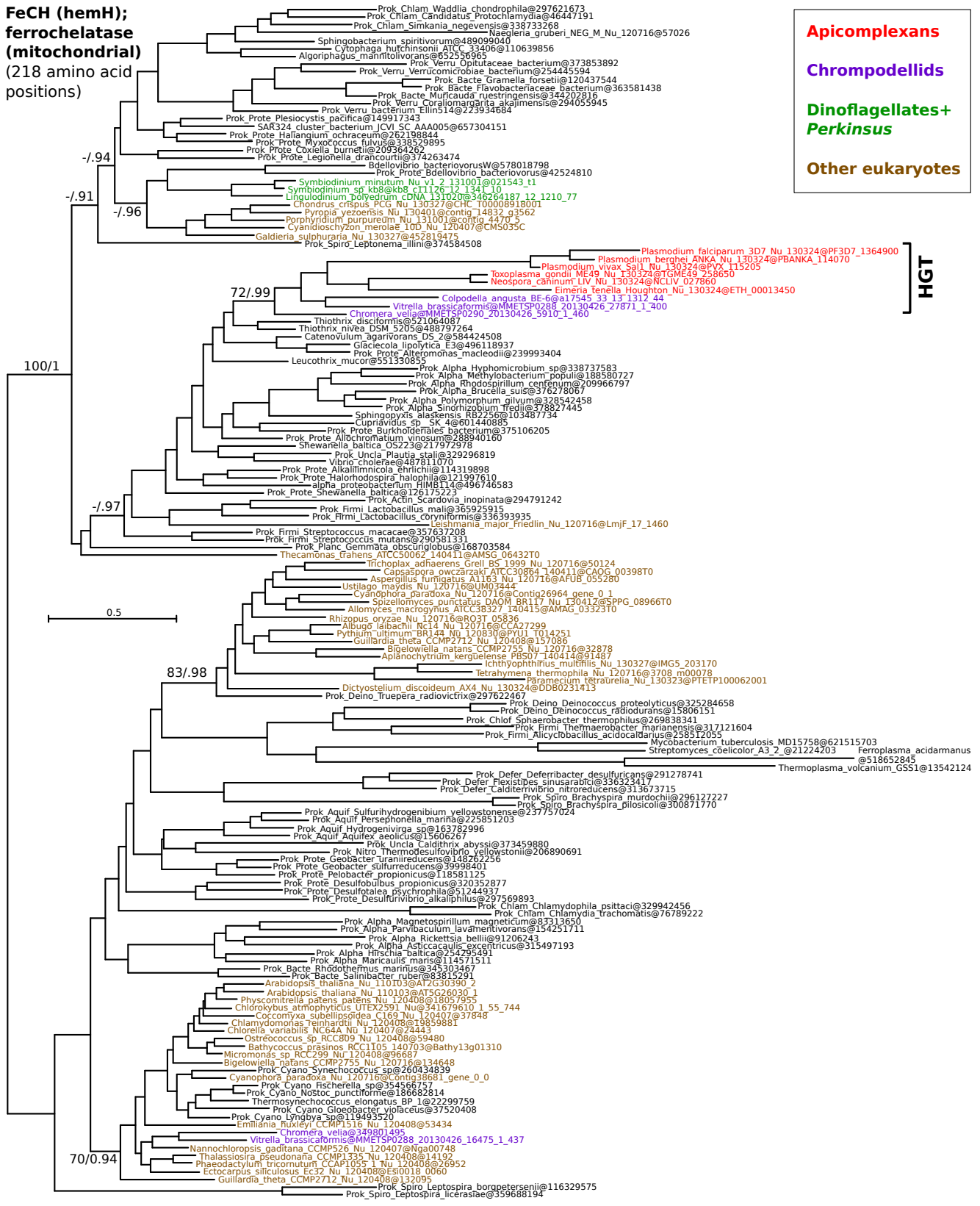
short-chain dehydrogenases/reductase:

**Fbp; fructose-1,6-bisphosphatase**  
(253 amino acid positions)

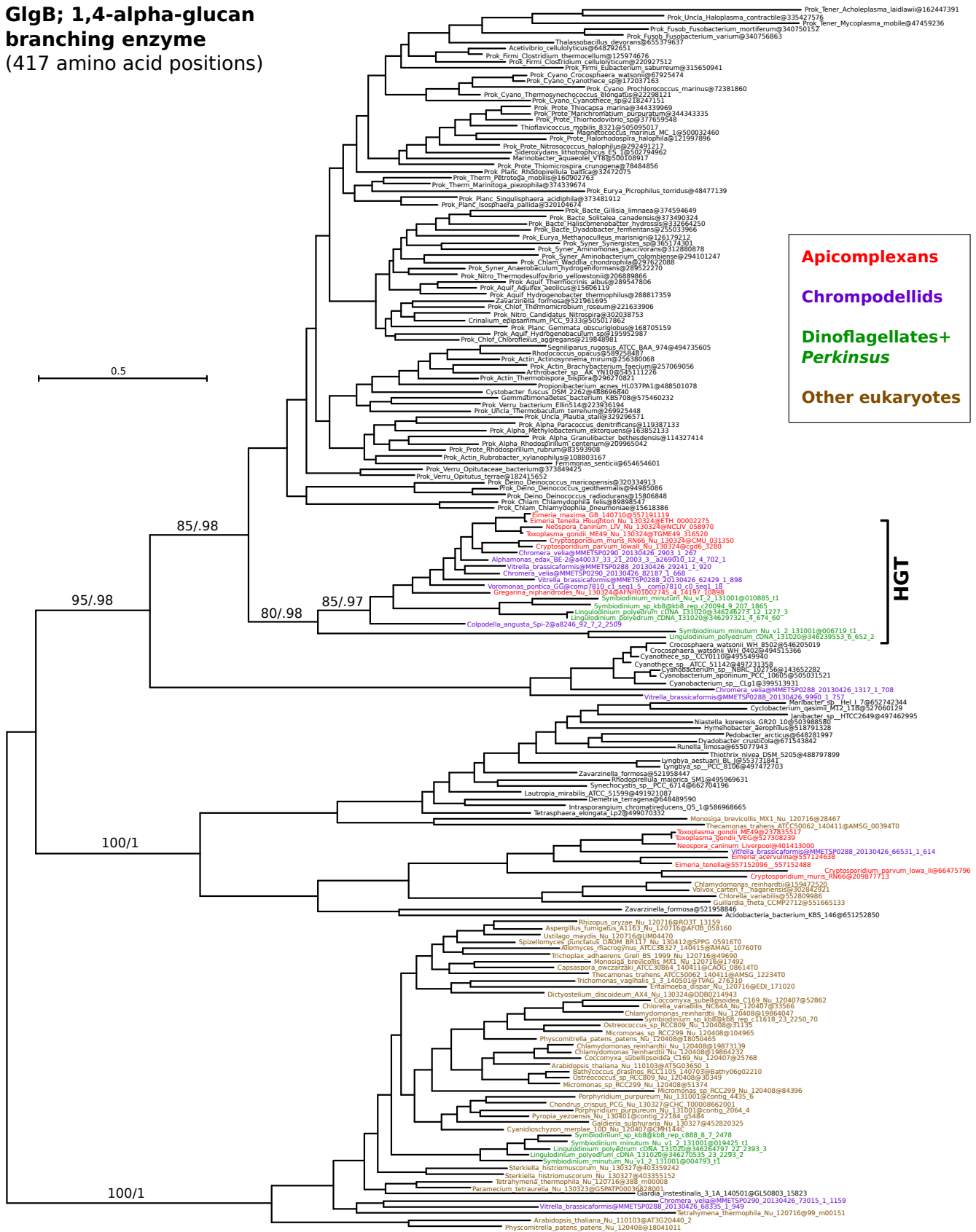


**FeCH (hemH);  
ferrochelatase  
(mitochondrial)**  
(218 amino acid  
positions)

**Apicomplexans**  
**Chromodellids**  
**Dinoflagellates +  
Perkinsus**  
**Other eukaryotes**



**GlgB; 1,4-alpha-glucan branching enzyme**  
(417 amino acid positions)

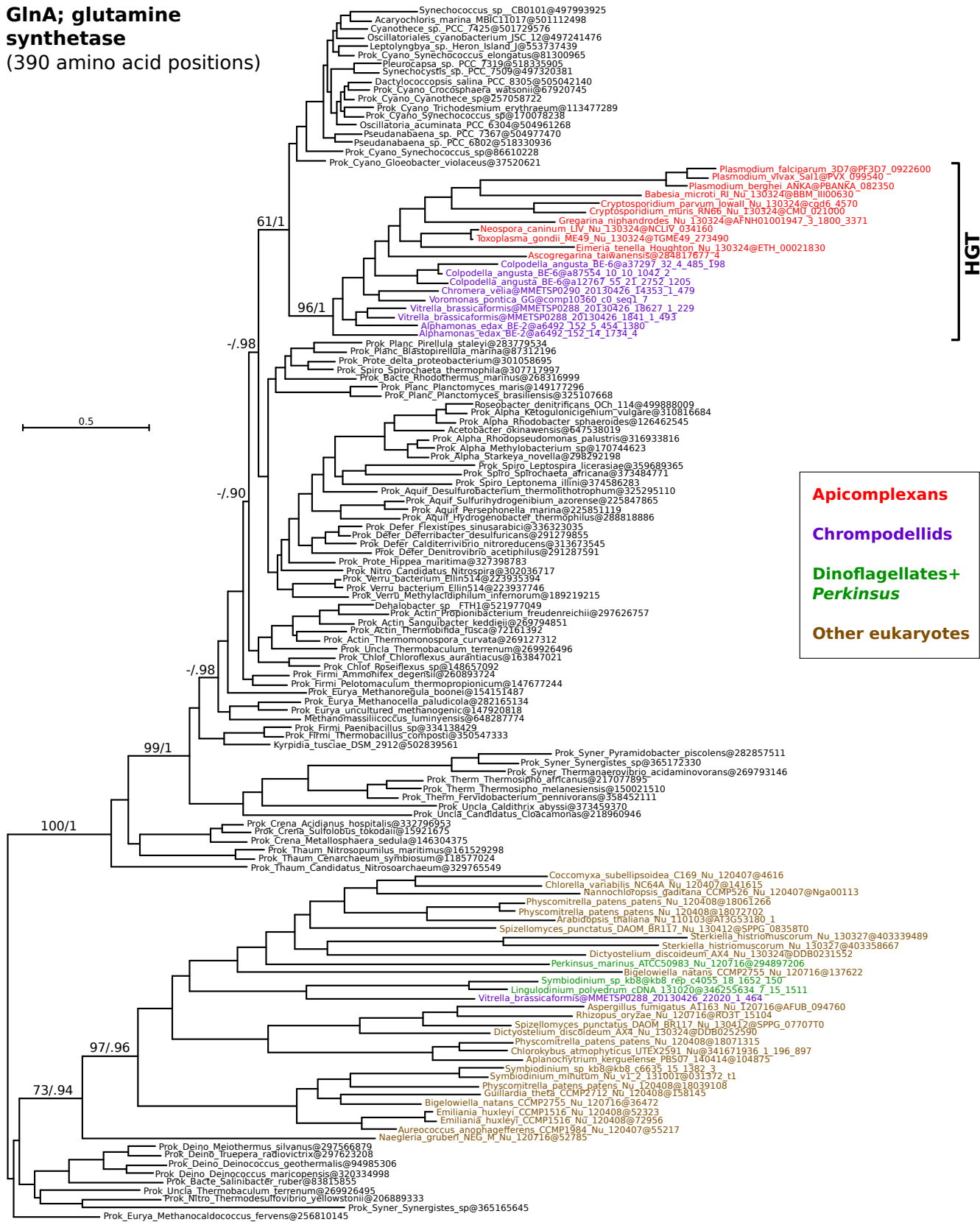


**Apicomplexans**  
**Chromodellids**  
**Dinoflagellates+**  
**Perkinsus**  
**Other eukaryotes**

**HGT**



**GlnA; glutamine synthetase**  
(390 amino acid positions)

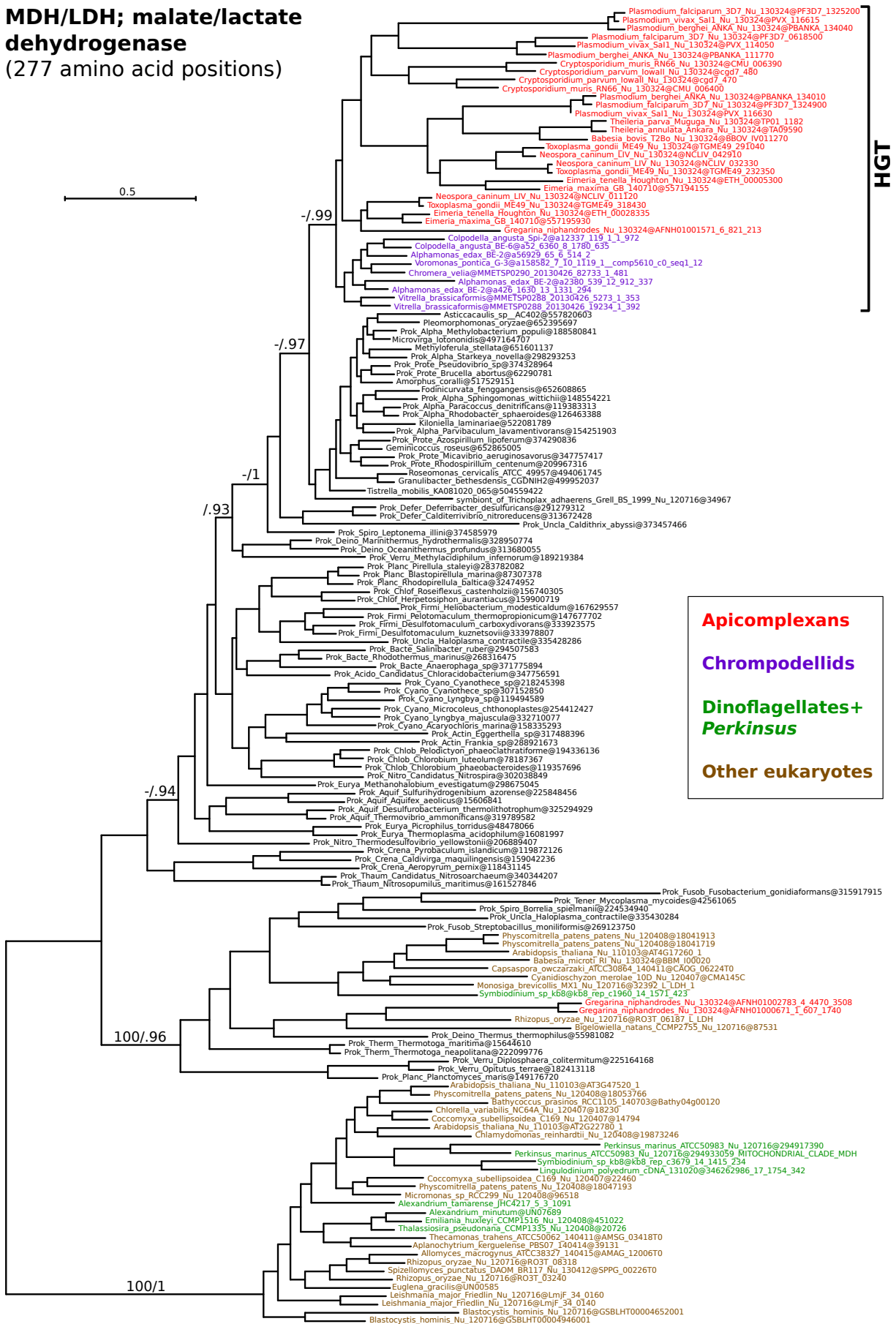


**IspH (LytB); 4-hydroxy-3-methylbut-2-enyl diphosphate reductase (plastidic)**  
(235 amino acid positions)

**Apicomplexans**  
**Chromodellids**  
**Dinoflagellates+ Perkinsus**  
**Other eukaryotes**

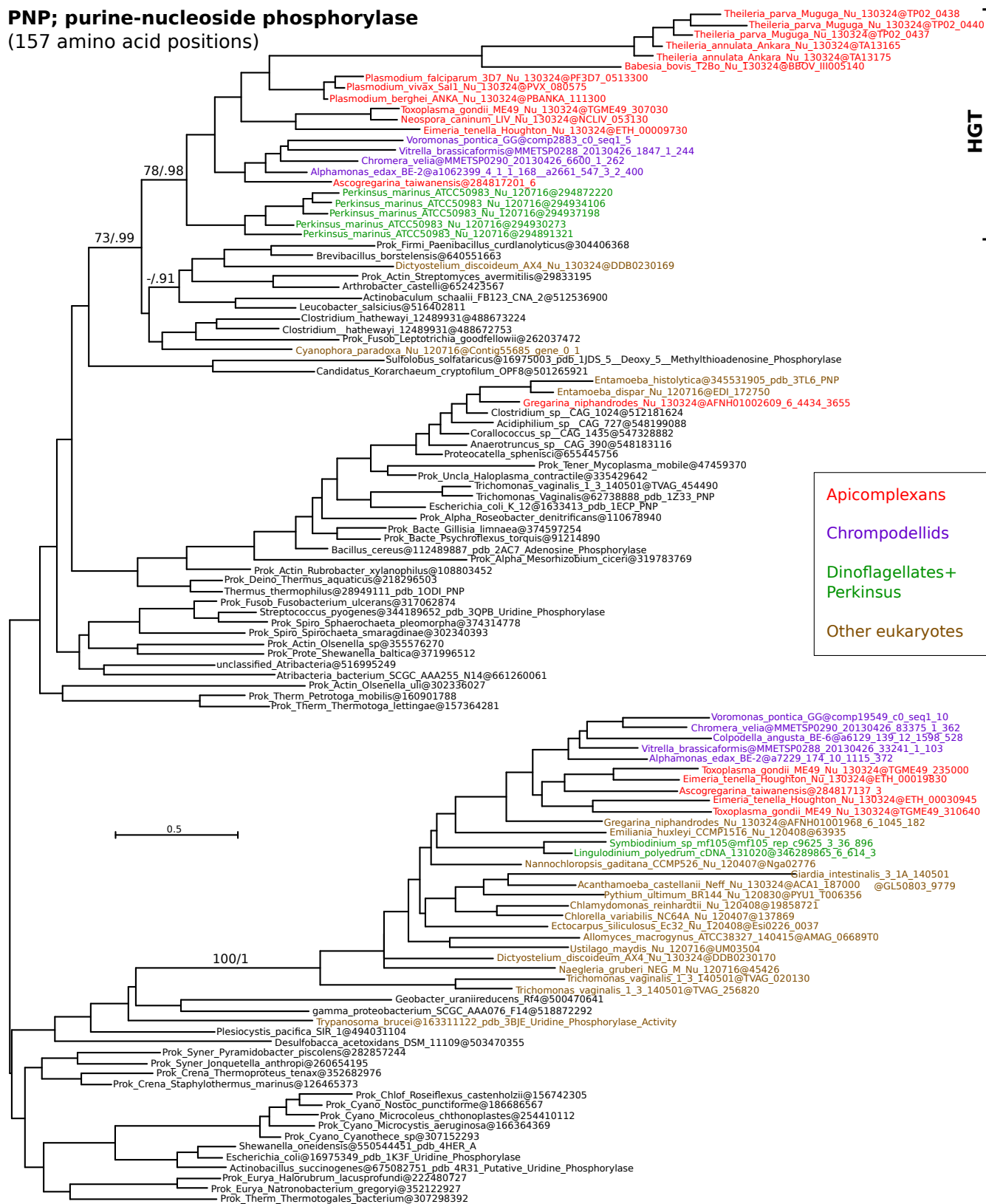


**MDH/LDH; malate/lactate dehydrogenase**  
(277 amino acid positions)



# PNP; purine-nucleoside phosphorylase

(157 amino acid positions)

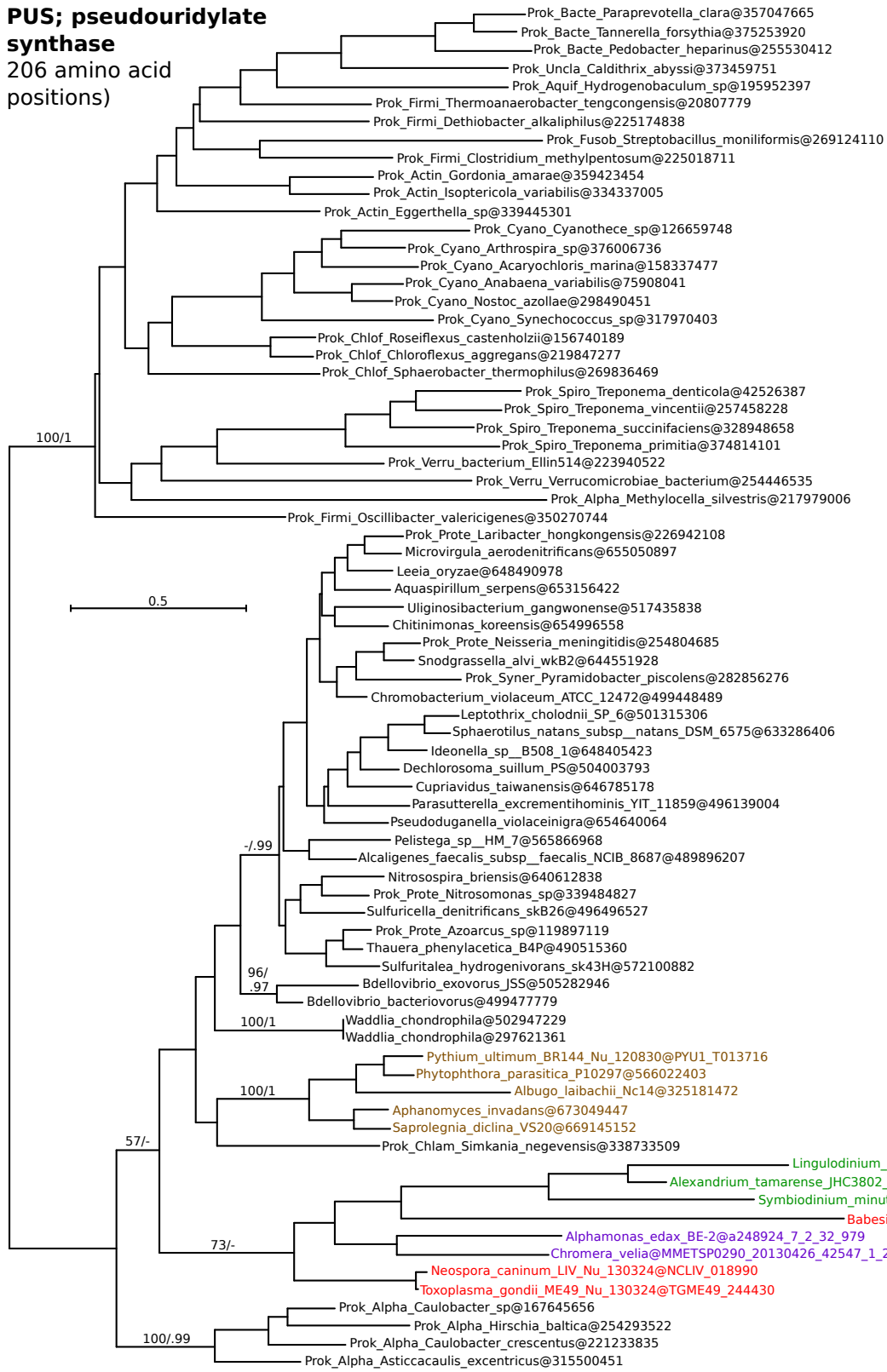


HGT

- Apicomplexans
- Chrompodellids
- Dinoflagellates+Perkinsus
- Other eukaryotes



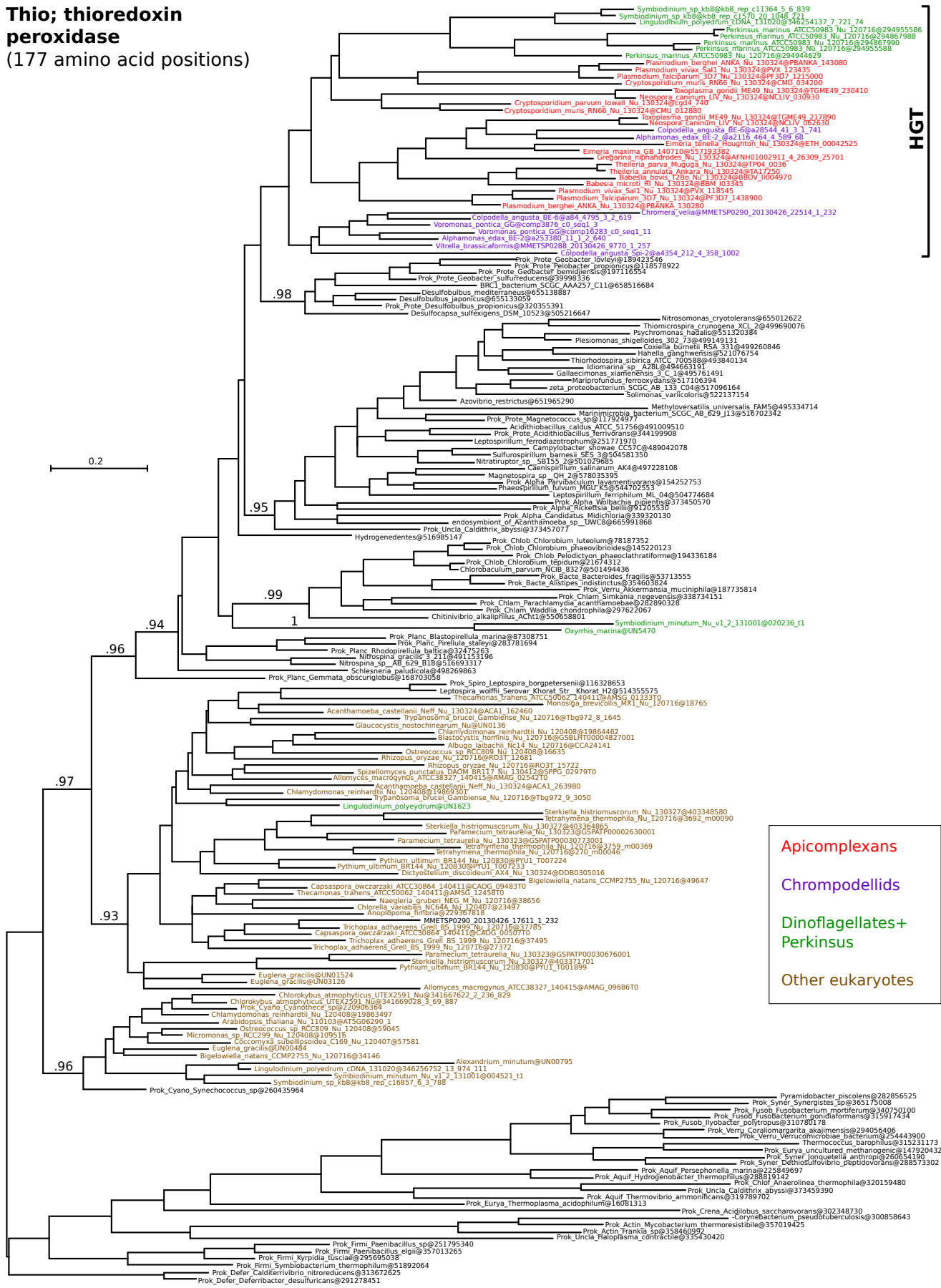
**PUS; pseudouridylate synthase**  
206 amino acid positions)



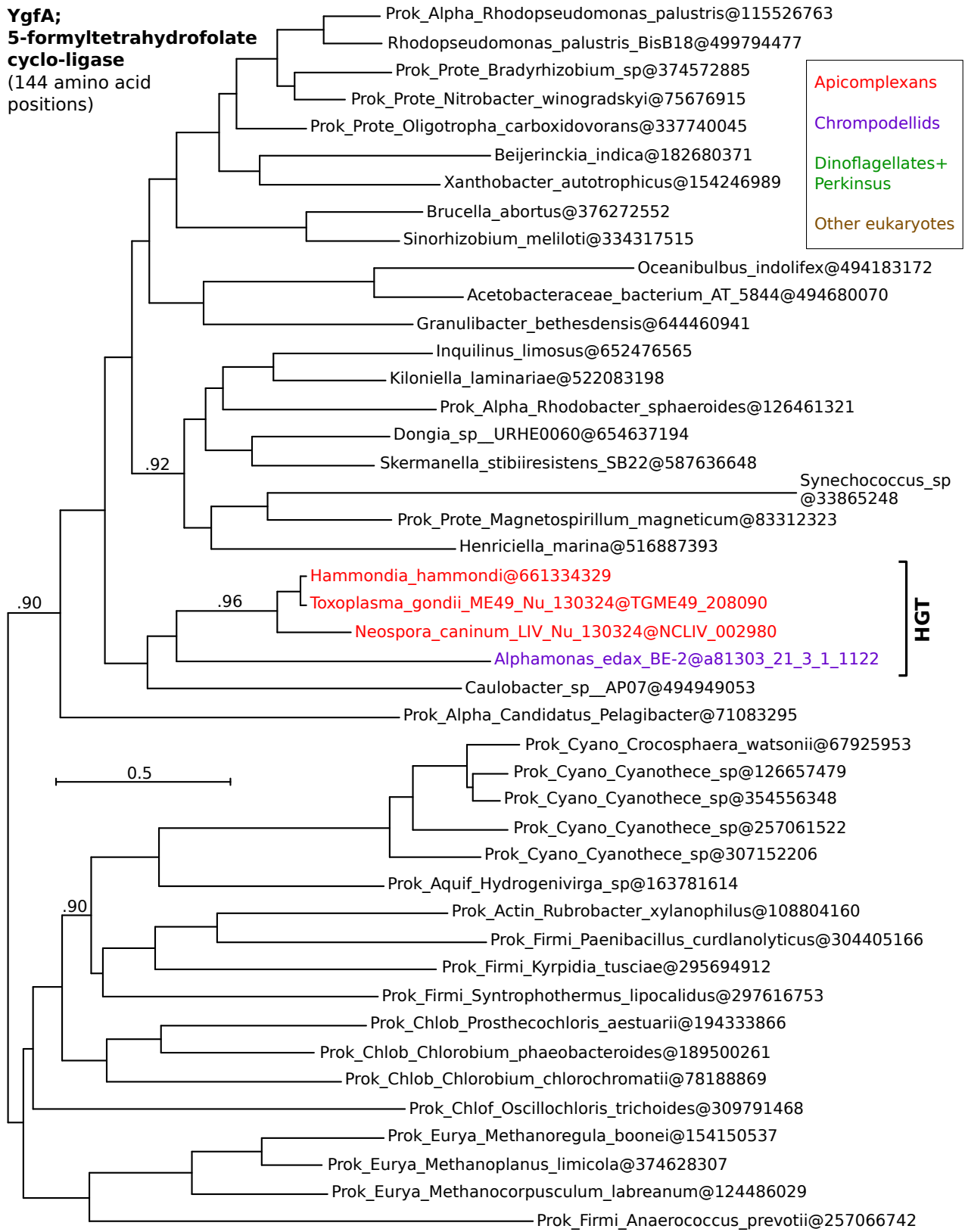
- Apicomplexans
- Chromodellids
- Dinoflagellates+ Perkinsus
- Other eukaryotes

HGT

**Thio; thioredoxin peroxidase**  
(177 amino acid positions)

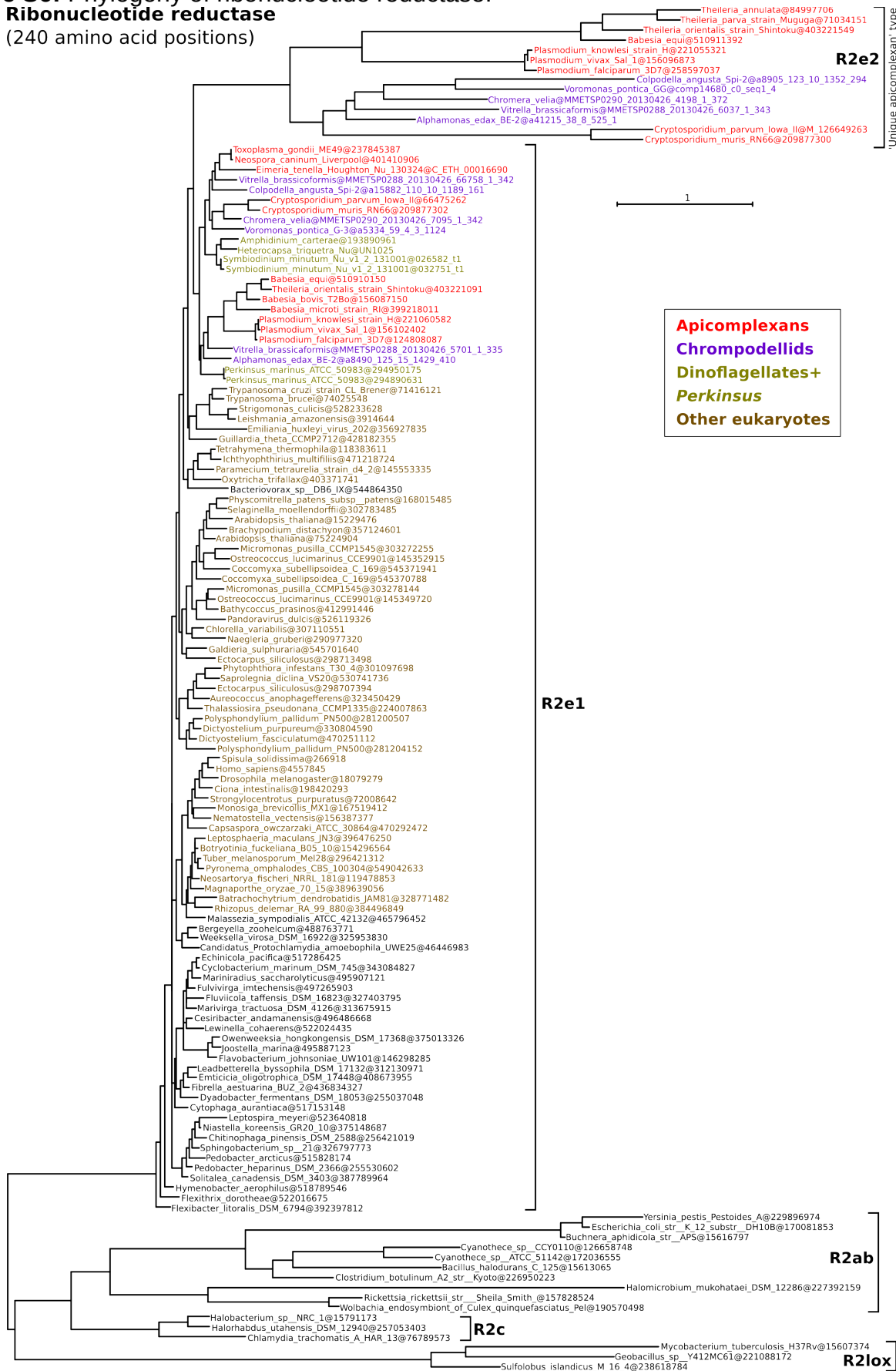


**YgfA;  
5-formyltetrahydrofolate  
cyclo-ligase**  
(144 amino acid  
positions)





**Figure S6: Phylogeny of ribonucleotide reductase.**  
**Ribonucleotide reductase**  
 (240 amino acid positions)



**Figure S7: Phylogeny of subtilisin proteases.**



**Table S1: Environmental sequences related to apicomplexans that were used in this study.**

Figure	Clone name	NCBI GI or Accession	Environment type <sup>1</sup>	Low O <sub>2</sub> <sup>2</sup>	Isolation source; Location	Study <sup>3</sup>	Clade affiliation <sup>4</sup>
Fig. 1	P107t1c2	KP213182	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade IX
Fig. 1	P107t1c6	KP213184	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade IX
Fig. 1	P107t6c1	KP213185	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Colpodella angusta
Fig. 1	P107t6c6	KP213186	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Colpodella angusta
Fig. 1	P108t7c1	KP213187	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Colpodella angusta
Fig. 1	P108t7c2	KP213188	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade IX
Fig. 1	P109t4c2	KP213191	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Colpodella angusta
Fig. 1	P109t4c6	KP213192	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Colpodella angusta
Fig. 1	P113t2c1	KP213195	marine		woodchip on the beach; Canada: Boundary Bay, BC	THIS STUDY	Colpodella angusta
Fig. 1	P113t2c6	KP213196	marine		woodchip on the beach; Canada: Boundary Bay, BC	THIS STUDY	Colpodella angusta
Fig. 1	P114t10c3	KP213197	marine		mucus from Acropora formosa; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Colpodella angusta
Fig. 1	P114t13c5	KP213200	marine		coral Acropora yongei; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Env. clade IX
Fig. 1	P114t9c5	KP213202	marine		mucus from Acropora yongei; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Env. clade IX
Fig. 1	P115t1c2	KP213203	marine		woodchip on the beach; Canada: Boundary Bay, BC	THIS STUDY	Chromera velia
Fig. 1	P115t1c3	KP213204	marine		woodchip on the beach; Canada: Boundary Bay, BC	THIS STUDY	Colpodella angusta
Fig. 1	P116t19c4	KP213207	marine		JAR_04_2010_ARMS_A_Filter corners	THIS STUDY	Env. clade VIII
Fig. 1	P116t19c5	KP213208	marine		JAR_04_2010_ARMS_A_Filter corners	THIS STUDY	Env. clade VIII
Fig. 1	P116t19c7	KP213209	marine		JAR_04_2010_ARMS_A_Filter corners	THIS STUDY	Unresolved: Api.
Fig. 1	P122t3c2	KP213211	marine		coral Echinophyllia sp.; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Env. clade V
Fig. 1	P122t3c3	KP213212	marine		coral Echinophyllia sp.; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Env. clade V
Fig. 1	P122t3c4	KP213213	marine		coral Echinophyllia sp.; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Env. clade V
Fig. 1	P23t21c1	KP213214	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api. relatives
Fig. 1	P23t21c2	KP213215	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api. relatives
Fig. 1	P23t21c5	KP213216	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api. relatives
Fig. 1	P23t5c3	KP213217	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api. relatives
Fig. 1	P23t5c7	KP213218	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api. relatives
Fig. 1	P23t5c8	KP213219	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api. relatives
Fig. 1	P43t2c8	KP213220	marine		rocky intertidal spit; Canada: Spanish Banks, Vancouver, BC	THIS STUDY	Voromonas pontica
Fig. 1	P46t3c16	KP213221	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api.
Fig. 1	P56t2c1	KP213222	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade VI
Fig. 1	P57t7c8	KP213226	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade VI
Fig. 1	P71t4c5	KP213228	marine		rocky intertidal spit; Canada: Spanish Banks, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P71t4c7	KP213229	marine		rocky intertidal spit; Canada: Spanish Banks, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P79t3c2	KP213230	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Unresolved: Api. relatives
Fig. 1	P79t3c8	KP213231	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Colpodella angusta

Fig. 1	P83t1c3	KP213232	marine		rocky intertidal spit; Canada: Spanish Banks, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P85t4c2	KP213233	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Colpodella angusta
Fig. 1	P85t4c5	KP213234	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Colpodella angusta
Fig. 1	P87t1c4	KP213235	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Chromera velia
Fig. 1	P87t2c1	KP213236	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade VI
Fig. 1	P87t2c5	KP213238	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Chromera velia
Fig. 1	P87t3c4	KP213239	marine		rocky intertidal spit; Canada: Spanish Banks, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P87t4c1	KP213240	marine		rocky intertidal spit; Canada: Spanish Banks, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P87t4c2	KP213241	marine		rocky intertidal spit; Canada: Spanish Banks, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P87t5c2	KP213242	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P87t5c3a	KP213243	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P87t5c4	KP213245	marine		damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Chromera velia
Fig. 1	P87t8c3	KP213246	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade VI
Fig. 1	P87t8c4	KP213247	soil		soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Env. clade VI
Fig. 1	P97t1c1	KP213251	marine		Dry high-beach sand; Canada: Boundary Bay, BC	THIS STUDY	Env. clade IX
Fig. 1	P97t5c2	KP213256	freshwater		freshwater duck pond near the beach; Canada: Boundary Bay, BC	THIS STUDY	Chromera velia
Fig. 1	P97t6c3	KP213258	marine		woodchip on the beach; Canada: Boundary Bay, BC	THIS STUDY	Voromonas pontica
Fig. 1	6	472401464	animal feces/manure	YES	Megalapteryx didinus coprolite, sample 10198a; New Zealand: Dart River Valley	PLoS ONE 8(2): E57315 (2013)	Colpodella angusta
Fig. 1	7	472401465	animal feces/manure	YES	Megalapteryx didinus coprolite, sample 10198a; New Zealand: Dart River Valley	PLoS ONE 8(2): E57315 (2013)	Colpodella angusta
Fig. 1	56059	6606066	marine		European oyster, Ostrea edulis	Unpublished	Unresolved: Api.
Fig. 1	1_133	171199596	marine		marine sediment; East Sea	J. Microbiol. 46 (3), 244-249 (2008)	Env. clade I
Fig. 1	1_41	156466089	marine		marine sediment; East Sea	J. Microbiol. 46 (3), 244-249 (2008)	Env. clade I
Fig. 1	1_52	156466099	marine		marine sediment; East Sea	J. Microbiol. 46 (3), 244-249 (2008)	Env. clade I
Fig. 1	1_73	156466110	marine		marine sediment; East Sea	J. Microbiol. 46 (3), 244-249 (2008)	Env. clade I
Fig. 1	9_12	156466122	marine		marine sediment; East Sea	J. Microbiol. 46 (3), 244-249 (2008)	Env. clade I
Fig. 1	9_39	156466143	marine		marine sediment; East Sea	J. Microbiol. 46 (3), 244-249 (2008)	Env. clade I
Fig. 1	9_61	156466155	marine		marine sediment; East Sea	J. Microbiol. 46 (3), 244-249 (2008)	Env. clade VIII
Fig. 1	A_1_87	304361468	freshwater		paddy field soil; Japan: Aomori	Microbes Environ. 25 (4), 281-287 (2010)	Unresolved:Api. relatives
Fig. 1	A2_E039	18025924	marine	YES	hydrothermal sediment; Guaymas Basin	PNAS 99 (11), 7658-62 (2002)	Unresolved: Api. relatives
Fig. 1	AA5F13RJ1A10	291497875	marine		oxygenated water column; Cariaco Basin, Caribbean Sea	ISME J 5 (8), 1344-1356 (2011)	Unresolved:Api. relatives
Fig. 1	AE-2012_FL1	411025944	freshwater		laboratory dishes with mosquito larvae; USA: Rutgers university	Aquat.Ecol.:10.1007/s10452-014-9474-3(2014)	Colpodella angusta
Fig. 1	ANEK03F02	345648463	soil		soil in front of the Brazilian Antarctic Station; Antarctica	Unpublished	Colpodella angusta
Fig. 1	BAQA40	20377996	marine	YES	anoxic marine sediment; USA: Berkeley Aquatic Park, Berkeley, CA	PNAS 99 (12), 8324-29 (2002)	Env. clade VII
Fig. 1	BCB5F13RM1D02	291498055	marine	YES	anoxic water column sample; Caribbean Sea: Cariaco Basin	ISME J 5 (8), 1344-1356 (2011)	Api.: genotype N
Fig. 1	BCB3F14RJ3H04	291495817	marine	YES	anoxic water column sample; Caribbean Sea: Cariaco Basin	ISME J 5 (8), 1344-1356 (2011)	Env. clade X
Fig. 1	BCB5F14RJ1F07	291495355	marine	YES	anoxic water column sample; Caribbean Sea: Cariaco Basin	ISME J 5 (8), 1344-1356 (2011)	Env. clade X
Fig. 1	BIO10_B6	270381369	marine	YES	deep water, depth 500m; Pacific Ocean: South Pacific	Environ. Microbiol. 12 (11), 2946-64 (2010)	Voromonas pontica
Fig. 1	BIO3_B2	270381403	marine	YES	deep water, depth 500m; Pacific Ocean: South Pacific	Environ. Microbiol. 12 (11), 2946-64 (2010)	Voromonas pontica

Fig. 1	BOLA176	20378006	marine	YES	anoxic marine sediment; USA: Bolinas Tidal Flat, Bolinas, CA	PNAS 99 (12), 8324-29 (2002)	Colpodella angusta
Fig. 1	BOLA553	20378005	marine	YES	anoxic marine sediment; USA: Bolinas Tidal Flat, Bolinas, CA	PNAS 99 (12), 8324-29 (2002)	Env. clade V
Fig. 1	BOLA566	20378000	marine	YES	anoxic marine sediment; USA: Bolinas Tidal Flat, Bolinas, CA	PNAS 99 (12), 8324-29 (2002)	Env. clade VI
Fig. 1	BOLA914	20377992	marine	YES	anoxic marine sediment; USA: Bolinas Tidal Flat, Bolinas, CA	PNAS 99 (12), 8324-29 (2002)	Env. clade IV
Fig. 1	Buttons_D03	300088832	marine (*)		intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	C1_E039	18025812	marine	YES	hydrothermal sediment; Guaymas Basin	PNAS 99 (11), 7658-62 (2002)	Api.:Polyplicarum -related
Fig. 1	C1_E047	18025817	marine	YES	hydrothermal sediment; Guaymas Basin	PNAS 99 (11), 7658-62 (2002)	Api.:Polyplicarum -related
Fig. 1	C17_250P117_0239Tplate02_P2	389549566	marine (*)		marine sediment from shallow water hydrothermal system; Papua New Guinea	Pacific Science 64 (3), 391-404 (2010)	Vitrella brassicaformis
Fig. 1	CCI31	27802552	marine	YES	oxygen-depleted marine env.; USA: Great Sippewisset salt marsh, Cape Cod, MA	Appl. Environ. Microbiol. 69(5),2657-63(2003)	Api.:Polyplicarum -related
Fig. 1	CPSGM-1	133778656	marine	YES	deep-sea methane cold seep sediment; Japan: Sagami Bay	Extremophiles 11 (4), 563-576 (2007)	Unresolved: Api.
Fig. 1	'Cryptosporidium struthionis'	58371885	animal feces/manure	YES	oocyst in Struthio camelus feces; Spain	Unpublished	Colpodella angusta
Fig. 1	'Cryptosporidium sp.' NEV10	342918948	animal feces/manure	YES	feces of calves with diarrhea; Turkey: Nevsehir	Unpublished	Colpodella angusta
Fig. 1	D1P01G01	118420045	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Api.:Polyplicarum -related
Fig. 1	D1P01H03	118420061	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Voromonas pontica
Fig. 1	D1P02A01	118420063	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Api.:Polyplicarum -related
Fig. 1	D1P02B08	118420240	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Colpodella angusta
Fig. 1	D1P02C06	118420241	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Api.:Polyplicarum -related
Fig. 1	D1P02D12	118420066	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Colpodella angusta
Fig. 1	D1P02E03	118420067	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade VII
Fig. 1	D1P02F05	118420046	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Api.:Polyplicarum -related
Fig. 1	D2P03B10	118420105	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade VII
Fig. 1	D2P03D02	118420082	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Colpodella angusta
Fig. 1	D2P03D06	118420106	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade I
Fig. 1	D2P03E11	118420245	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Api.:Polyplicarum -related
Fig. 1	D3P05C07	118420125	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade XI
Fig. 1	D3P05F04	118420152	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade VII
Fig. 1	D3P06B02	118420161	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade XI
Fig. 1	D4P07B05	118420253	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade VII
Fig. 1	D4P07C10	118420192	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade XI
Fig. 1	D4P07G08	118420173	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade I
Fig. 1	D4P07H09	118420191	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade XI
Fig. 1	D4P08D06	118420260	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade VII
Fig. 1	D4P08E10	118420178	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Api.:Polyplicarum -related
Fig. 1	D4P08E11	118420179	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade I
Fig. 1	D4P08G08	118420181	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade XI

Fig. 1	D5P09C09	118420206	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Colpodella angusta
Fig. 1	D5P10A10	118420214	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Api.:Polyplicarum-related
Fig. 1	D5P10D11	118420226	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Env. clade VII
Fig. 1	D5P10G04	118420223	marine	YES	oxygen-depleted intertidal marine sediment, upper 2 cm; Greenland: Arctic	PLoS ONE 2(8): e728 (2007)	Colpodella angusta
Fig. 1	DGGE_gel_band_3mFR1L9B9	544587044	animal feces/manure	YES	composting dairy manure	Unpublished	Colpodella angusta
Fig. 1	DGGE_gel_band_3mFR11011	544587051	animal feces/manure	YES	composting dairy manure	Unpublished	Colpodella angusta
Fig. 1	He000427_15535	89572115	marine		marine sample; Germany: Helgoland	Microb. Ecol. 52 (1), 53-71 (2006)	Colpodella angusta
Fig. 1	He000427_20	89572015	marine		marine sample; Germany: Helgoland	Microb. Ecol. 52 (1), 53-71 (2006)	Colpodella angusta
Fig. 1	dpeuk39	60592951	freshwater	YES	freshwater anaerobic pond; USA: Zodlone Spring, Oklahoma	Appl. Environ. Microbiol. 71(10), 6175-84(2005)	Unresolved:Api. relatives
Fig. 1	DSGM-12	133778560	marine	YES	deep-sea methane cold seep sediment; Japan: Sagami Bay	Extremophiles 11 (4), 563-576 (2007)	Unresolved: Api.
Fig. 1	DSGM-13	133778561	marine	YES	deep-sea methane cold seep sediment; Japan: Sagami Bay	Extremophiles 11 (4), 563-576 (2007)	Api.:Polyplicarum-related
Fig. 1	DSGM-8	133778556	marine	YES	deep-sea methane cold seep sediment; Japan: Sagami Bay	Extremophiles 11 (4), 563-576 (2007)	Unresolved: Api.
Fig. 1	E08_250P220_0239Tplate03_P3	389549864	marine (*)		marine sediment from shallow water hydrothermal system; Papua New Guinea	Pacific Science 64 (3), 391-404 (2010)	Vitrella brassicaformis
Fig. 1	E12_180P126_0239Uplate02_P2	389549895	marine (*)		marine sediment from shallow water hydrothermal system; Papua New Guinea	Pacific Science 64 (3), 391-404 (2010)	Vitrella brassicaformis
Fig. 1	E1aD71	168203738	marine		marine plankton; South China Sea, northern part	J. Plankton Res. 33 (3), 445-456 (2011)	Voromonas pontica
Fig. 1	E6	30144456	marine	YES	permanently anoxic deep-sea basin; Caribbean Sea: Cariaco Basin	Appl. Environ. Microbiol. 69(9), 5656-63(2003)	Api.:Polyplicarum-related
Fig. 1	EBA140.9	213058639	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Env. clade II
Fig. 1	EBA43.25	213058601	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Env. clade IV
Fig. 1	EBA47.26	213058602	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Env. clade IV
Fig. 1	EBA52.36	213058604	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Env. clade IV
Fig. 1	EBA9.8	213058584	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Unresolved:Api. relatives
Fig. 1	EBF1.128	213058470	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Env. clade IV
Fig. 1	EBF1.90	213058469	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Unresolved:Api. relatives
Fig. 1	EBM3.51	213058518	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Unresolved:Api. relatives
Fig. 1	EC12	55831924	animal feces/manure	YES	pig manure storage pit	FEMS Microbiol. Ecol. 52(2), 229-42(2005)	Colpodella angusta
Fig. 1	EC54	148743803	animal feces/manure	YES	pig manure storage pit	FEMS Microbiol. Ecol. 52(2), 229-42(2005)	Colpodella angusta
Fig. 1	EC67	148743812	animal feces/manure	YES	pig manure storage pit	FEMS Microbiol. Ecol. 52(2), 229-42(2005)	Colpodella angusta
Fig. 1	elb6-t0-otu7	294829215	freshwater		ice-covered lake; Antarctica	ISME J 5, 1559-1564 (2011)	Unresolved:Api. relatives
Fig. 1	Elev_18S_5171	166084485	soil		trembling aspen rhizosphere, elevated CO2 conditions; USA: UMBS, Michigan	Environ. Microbiol. 10 (4), 926-941 (2008)	Colpodella angusta
Fig. 1	ESS270706.033	299789590	freshwater		lake water filtered through 3 um from lake Esch sur Sure, depth 0m; Luxembourg	Unpublished	Unresolved:Api. relatives
Fig. 1	H11C09cA	261290035	marine (*)		stromatolite (marine microbial mat); Bahamas: Highborne Cay	Environ. Microbiol. 11 (10), 2710-19 (2009)	Vitrella brassicaformis
Fig. 1	He000427_1	89572118	marine		marine sample; Germany: Helgoland	Microb. Ecol. 52 (1), 53-71 (2006)	Colpodella angusta
Fig. 1	I10_180P53_0239Rplate06_P6	389550613	marine (*)		marine sediment from shallow water hydrothermal system; Papua New Guinea	Pacific Science 64 (3), 391-404 (2010)	Vitrella brassicaformis
Fig. 1	I9S18K_ML_148	223034114	freshwater		Industrial Canal, water, greater than 3 micron; USA: New Orleans, LA	Environ. Sci. Technol. 42(24), 9072-8(2008)	Env. clade IV
Fig. 1	K1OCT2011	430726581	freshwater		freshwater lake; Japan: Gunma, lake Kusaki	Unpublished	Env. clade IV

Fig. 1	K2MAY2011	430726574	freshwater		freshwater lake; Japan: Gunma, lake Kusaki	Unpublished	Env. clade IV
Fig. 1	K5AUG2009	327247873	freshwater		freshwater lake; Japan: Gunma, lake Kusaki	Unpublished	Env. clade IV
Fig. 1	KRL01E25	353255973	freshwater		freshwater lake; Greece: Lake Karla	ScientificWorldJournal 2012, 504135 (2012)	Unresolved:Api. relatives
Fig. 1	KRL03E03	449890141	freshwater		freshwater lake; Greece: Lake Karla	ScientificWorldJournal 2012, 504135 (2012)	Env. clade IV
Fig. 1	KRL03E57	449890156	freshwater		freshwater lake; Greece: Lake Karla	Hydrobiologia 718, 73-83 (2013)	Env. clade III
Fig. 1	KRL09E06	449890170	freshwater		freshwater lake; Greece: Lake Karla	Hydrobiologia 718, 73-83 (2013)	Env. clade III
Fig. 1	KRL09E21	449890174	freshwater		freshwater lake; Greece: Lake Karla	Hydrobiologia 718, 73-83 (2013)	Env. clade IV
Fig. 1	L07_250P64_023 9DSNP001F_P3	389551156	marine (*)		marine sediment from shallow water hydrothermal system; Papua New Guinea	Pacific Science 64 (3), 391-404 (2010)	Vitrella brassicaformis
Fig. 1	LG02-01	60476886	freshwater		oligotrophic freshwater lake; USA: Lake George, Adirondack Park	Environ. Microbiol. 7 (9), 1413-1425 (2005)	Env. clade II
Fig. 1	LG05-01	60476898	freshwater		oligotrophic freshwater lake; USA: Lake George, Adirondack Park	Environ. Microbiol. 7 (9), 1413-1425 (2005)	Env. clade II
Fig. 1	LG09-02	60476916	freshwater		oligotrophic freshwater lake; USA: Lake George, Adirondack Park	Environ. Microbiol. 7 (9), 1413-1425 (2005)	Env. clade II
Fig. 1	LG25-01	60476973	freshwater		oligotrophic freshwater lake; USA: Lake George, Adirondack Park	Environ. Microbiol. 7 (9), 1413-1425 (2005)	Env. clade II
Fig. 1	LG32-03	60476996	freshwater		oligotrophic freshwater lake; USA: Lake George, Adirondack Park	Environ. Microbiol. 7 (9), 1413-1425 (2005)	Env. clade II
Fig. 1	LT19_H1	529105019	hypersaline		hypersaline lake; Australia: Lake Tyrrell, Victoria	Front Microbiol 4, 115 (2013)	Unresolved:Api. relatives
Fig. 1	LT35_C14	529105426	hypersaline		hypersaline lake; Australia: Lake Tyrrell, Victoria	Front Microbiol 4, 115 (2013)	Unresolved:Api. relatives
Fig. 1	LT37_G4	529105789	hypersaline		hypersaline lake; Australia: Lake Tyrrell, Victoria	Front Microbiol 4, 115 (2013)	Chromera velia
Fig. 1	LT41_e05_2	529106034	hypersaline		hypersaline lake; Australia: Lake Tyrrell, Victoria	Front Microbiol 4, 115 (2013)	Unresolved:Api. relatives
Fig. 1	LT50_P1	529106341	hypersaline		hypersaline lake; Australia: Lake Tyrrell, Victoria	Front Microbiol 4, 115 (2013)	Unresolved:Api. relatives
Fig. 1	LT62_D14	529106413	hypersaline		hypersaline lake; Australia: Lake Tyrrell, Victoria	Front Microbiol 4, 115 (2013)	Unresolved:Api. relatives
Fig. 1	Mat131	356459394	marine (*)	YES	anoxic sink; Gulf of Mexico: Jewfish Sink	Hydrobiologia 677 (1), 65-87 (2011)	Vitrella brassicaformis
Fig. 1	Mat140	356459402	marine (*)	YES	anoxic sink; Gulf of Mexico: Jewfish Sink	Hydrobiologia 677 (1), 65-87 (2011)	Vitrella brassicaformis
Fig. 1	Mat179	356459439	marine (*)	YES	anoxic sink; Gulf of Mexico: Jewfish Sink	Hydrobiologia 677 (1), 65-87 (2011)	Vitrella brassicaformis
Fig. 1	ML1_dino18S-9	309753096	freshwater		5 micron fraction of surface freshwater lake; USA: Connecticut, Storrs, Mirror Lake	PNAS 107 (46), 20033-38 (2010)	Env. clade IV
Fig. 1	MLBA54.107	213058446	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Unresolved:Api. relatives
Fig. 1	MLBA74.128	213058464	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Unresolved:Api. relatives
Fig. 1	MLBM116.37	213058384	freshwater		freshwater lake; China	Acta Protozool. 48, 245-264 (2009)	Unresolved:Api. relatives
Fig. 1	MPE1-2	374532643	freshwater		aquatic moss pillars, lake; Antarctica: Skarvsnes, Hotoke-Ike lake	Polar Biol. 35, 1495-1504 (2012)	Unresolved:Api. relatives
Fig. 1	MPE1-46	374532687	freshwater		aquatic moss pillars, lake; Antarctica: Skarvsnes, Hotoke-Ike lake	Polar Biol. 35, 1495-1504 (2012)	Unresolved:Api. relatives
Fig. 1	MPE2-2	374532695	freshwater		aquatic moss pillars, lake; Antarctica: Skarvsnes, Hotoke-Ike lake	Polar Biol. 35, 1495-1504 (2012)	Unresolved:Api. relatives
Fig. 1	Out-556	356459667	marine		anoxic sink; Gulf of Mexico: Jewfish Sink	Hydrobiologia 677 (1), 65-87 (2011)	Vitrella brassicaformis
Fig. 1	Out-623	356459729	marine		anoxic sink; Gulf of Mexico: Jewfish Sink	Hydrobiologia 677 (1), 65-87 (2011)	Vitrella brassicaformis
Fig. 1	Out-678	356459781	marine		anoxic sink; Gulf of Mexico: Jewfish Sink	Hydrobiologia 677 (1), 65-87 (2011)	Vitrella brassicaformis
Fig. 1	PAA22AU2004	82470184	freshwater		deep freshwater meromictic lake; France: Lake Pavin, Massif Central	Environ. Microbiol. 9 (1), 61-71 (2007)	Env. clade IV
Fig. 1	PAF8AU2004	82470188	freshwater		deep freshwater meromictic lake; France: Lake Pavin, Massif Central	Environ. Microbiol. 9 (1), 61-71 (2007)	Unresolved:Api. relatives
Fig. 1	PC04.G3_Pla_1	393405643	freshwater		high mountain lake; Spain: Pyrenees, Lake Plan	Environ. Microbiol. 14 (9), 2445-56 (2012)	Env. clade II
Fig. 1	PC05.B12_Ger_1 2	393405440	freshwater		high mountain lake; Spain: Pyrenees, Lake Plan	Environ. Microbiol. 14 (9), 2445-56 (2012)	Env. clade II
Fig. 1	PC05.F10_Ger_1	393405443	freshwater		high mountain lake; Spain: Pyrenees, Lake Plan	Environ. Microbiol. 14 (9), 2445-56 (2012)	Env. clade II

Fig. 1	PC06.B8_Ger_1	393405451	freshwater	high mountain lake; Spain: Pyrenees, Lake Plan	Environ. Microbiol. 14 (9), 2445-56 (2012)	Env. clade II
Fig. 1	PC06.D4_Ger_9	393405454	freshwater	high mountain lake; Spain: Pyrenees, Lake Plan	Environ. Microbiol. 14 (9), 2445-56 (2012)	Env. clade II
Fig. 1	PC06.H4_Ger_6	393405472	freshwater	high mountain lake; Spain: Pyrenees, Lake Plan	Environ. Microbiol. 14 (9), 2445-56 (2012)	Env. clade II
Fig. 1	Pink_A01	300088886	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_C09	300088917	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_D06	300088926	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_D07	300088927	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_D12	300088932	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_E09	300088940	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_E10	300088941	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_F04	300088947	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_F05	300088948	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_F11	300088953	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_G07	300088960	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	Pink_G11	300088964	marine (*)	intertidal thrombolites; Bahamas: Highborne Cay	Geobiology 8 (4), 337-354 (2010)	Vitrella brassicaformis
Fig. 1	PR2_3E_116	316926548	freshwater	water from Praz Rodet peat bog; Switzerland: Jura Mountains	Protist 162, 14–32 (2011)	Unresolved: Api.
Fig. 1	PR2_3E_74	299778455	freshwater	water from Praz Rodet peat bog; Switzerland: Jura Mountains	Protist 162, 14–32 (2011)	Unresolved: Api. relatives
Fig. 1	PR3_4E_33	299778456	freshwater	water from Praz Rodet peat bog; Switzerland: Jura Mountains	Protist 162, 14–32 (2011)	Env. clade VI
Fig. 1	PS13A10	262357995	marine (*)	Lakshwadeep coral reef sand; Arabian Sea	FEMS Microbiol. Ecol. 71(3),399–412(2010)	Vitrella brassicaformis
Fig. 1	PS13C9	262358012	marine (*)	Lakshwadeep coral reef sand; Arabian Sea	FEMS Microbiol. Ecol. 71(3),399–412(2010)	Vitrella brassicaformis
Fig. 1	PS13D9	262358015	marine (*)	Lakshwadeep coral reef sand; Arabian Sea	FEMS Microbiol. Ecol. 71(3),399–412(2010)	Vitrella brassicaformis
Fig. 1	SB16_2010	387912501	freshwater	water purification plant; Japan:Gunma, Kiryu	Unpublished	Voromonas pontica
Fig. 1	SB19_2010	387912504	freshwater	water purification plant; Japan:Gunma, Kiryu	Unpublished	Chromera velia
Fig. 1	SB21_2010	387912506	freshwater	water purification plant; Japan:Gunma, Kiryu	Unpublished	Unresolved: Api. relatives
Fig. 1	SB7_2010	387912492	freshwater	water purification plant; Japan:Gunma, Kiryu	Unpublished	Unresolved: Api. relatives
Fig. 1	SC8051102_25	225548510	freshwater	freshwater reservoir; South Korea: Sangcheon reservoir	J. Microbiol. Methods 81 (1), 61-68 (2010)	Unresolved: Api. relatives
Fig. 1	SEO81101_35	225548493	freshwater	freshwater reservoir; South Korea: Seoho reservoir	J. Microbiol. Methods 81 (1), 61-68 (2010)	Env. clade IV
Fig. 1	SEO81107_36	225548503	freshwater	freshwater reservoir; South Korea: Seoho reservoir	J. Microbiol. Methods 81 (1), 61-68 (2010)	Env. clade IV
Fig. 1	Sey061	49356800	freshwater	river sediment; Switzerland: Seymaz River, Geneva	BMC Biol. 2, 13 (2004)	Env. clade VII
Fig. 1	TB4-1	259906333	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved: Api. relatives
Fig. 1	TB4-15	259906347	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved: Api. relatives
Fig. 1	TB4-24	259906354	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved: Api. relatives
Fig. 1	TB4-27	259906357	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved: Api. relatives
Fig. 1	TB4-39	259906369	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved: Api. relatives
Fig. 1	TB4-43	259906371	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved: Api. relatives



Fig. 1	TB4-44	259906372	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	TB4-45	259906373	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	TB4-48	259906376	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	TB4-51	259906379	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	V10-5	355346671	freshwater	freshwater reservoir; France, Villerest reservoir	Unpublished	Env. clade IV
Fig. 1	VP1	100289290	freshwater	lacustrine community in hyper-eutrophic conditions; France, Villerest reservoir	FEMS Microbiol. Ecol. 61(3), 483-95(2007)	Unresolved:Api. relatives
Fig. 1	WD0-1	259906092	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Env. clade II
Fig. 1	WD0-50	259906128	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Env. clade IV
Fig. 1	WD4-1	259906134	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	WD4-110	259906236	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	WD4-128	259906254	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	WD4-138	259906264	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	WD4-66	259906196	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. 1	WD4-80	259906208	freshwater	freshwater lake; China: Lake Taihu	FEMS Microbiol. Ecol. 74 (1), 19-31 (2010)	Unresolved:Api. relatives
Fig. S2	P107t1c1	KP213181	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P107t1c3	KP213183	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P108t7c5	KP213189	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P108t7c6	KP213190	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P109t5c6	KP213193	marine	coral <i>Porites compressa</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.: genotype N
Fig. S2	P113t16c1	KP213194	marine	Coral 3, identified as <i>Seriatopora hystrix</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.: genotype N
Fig. S2	P114t13c1	KP213198	marine	coral <i>Acropora yongei</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P114t13c2	KP213199	marine	coral <i>Acropora yongei</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P114t9c2	KP213201	marine	mucus from <i>Acropora yongei</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P116t17c4	KP213205	marine	Coral 3, identified as <i>Seriatopora hystrix</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.: genotype N
Fig. S2	P116t17c5	KP213206	marine	Coral 3, identified as <i>Seriatopora hystrix</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.: Sarcocystids
Fig. S2	P116t7c3	KP213210	marine	coral <i>Seriatopora hystrix</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.: genotype N
Fig. S2	P57t7c1	KP213223	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P57t7c10	KP213224	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P57t7c3	KP213225	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P57t7c9	KP213225	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P87t2c4	KP213237	soil	soil from UBC endowment lands; Canada: Vancouver	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P87t5c3b	KP213244	marine	Damp woodchip and surface sand; Canada: Locarno Beach, Vancouver, BC	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P91t3c1	KP213248	marine	coral <i>Porites astreoides</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P91t3c4	KP213249	marine	coral <i>Porites astreoides</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.:Terrestrial gregarines
Fig. S2	P91t4c2	KP213250	marine	coral <i>Porites compressa</i> ; USA: Birch Aquarium, San Diego, CA	THIS STUDY	Api.: genotype N

Fig. S2	P97t5c5	KP213257	freshwater	Freshwater duck pond near the beach; Canada: Boundary Bay, BC	THIS STUDY	Api.: Terrestrial gregarines
Fig. S2	P97t3c2	KP213252	marine	Damp mid-beach sand under broken shell and debris; Canada: Boundary Bay, BC	THIS STUDY	Api.: Eimeriids
Fig. S2	P97t3c3	KP213253	marine	Damp mid-beach sand under broken shell and debris; Canada: Boundary Bay, BC	THIS STUDY	Api.: Eimeriids
Fig. S2	P97t3c4	KP213254	marine	Damp mid-beach sand under broken shell and debris; Canada: Boundary Bay, BC	THIS STUDY	Api.: Eimeriids
Fig. S2	P97t3c5	KP213255	marine	Damp mid-beach sand under broken shell and debris; Canada: Boundary Bay, BC	THIS STUDY	Api.: Eimeriids

<sup>1</sup> Marine, CaCO<sub>3</sub>-dominated environments specifically associated with sequences from the *Vitrella brassicaformis* clade are indicated by the (\*) symbol. These include coral reef sediment (Arabian Sea), thrombolites and stromatolites at Higborne Cay (Bahamas), CaCO<sub>3</sub>-precipitating sink off the coast of Florida, and coastal carbonate sediment in Papua New Guinea. See the respective publications for further details.

<sup>2</sup> Low oxygen environment (anoxic or microaerophilic)

<sup>3</sup> THIS STUDY = sequences generated in this study by apicomplexan-specific primers

<sup>4</sup> Based on Fig. 1; Env. = Environmental; Api. = Apicomplexa

**Table S2: Abbreviations and full names of proteins and pathways used in Fig. 3.**

Pathway	Localization	Short names	E.C.	Protein name			
Isoprenoid precursor biosynthesis (Isopentenyl diphosphate= IPP/ Dimethylallyl diphosphate= DMAP)	Cytosolic (mevalonate pathway = MEV)	HMGCS	2.3.3.10	hydroxymethylglutaryl-CoA synthase			
		HMGCR	1.1.1.34	hydroxymethylglutaryl-CoA reductase			
		MVK	2.7.1.36	mevalonate kinase			
	Plastidic (non-mevalonate pathway = MEP/DOXP)		PMVK	2.7.4.2	phosphomevalonate kinase		
			MVD	4.1.1.33	diphosphomevalonate decarboxylase		
			DXS	2.2.1.7	1-deoxy-D-xylulose-5-phosphate synthase		
			IspC	1.1.1.267	1-deoxy-D-xylulose-5-phosphate reductoisomerase		
			IspD	2.7.7.60	2-C-methyl-D-erythritol 4-phosphate cytidyltransferase		
			IspE	2.7.1.148	4-diphosphocytidyl-2-C-methyl-D-erythritol kinase		
			IspF	4.6.1.12	2-C-methyl-D-erythritol 2,4-cyclodiphosphate synthase		
			IspG	1.17.7.1	(E)-4-hydroxy-3-methylbut-2-enyl-diphosphate synthase		
			IspH (LytB)	1.17.1.2	4-hydroxy-3-methylbut-2-enyl diphosphate reductase		
			Tetrapyrrole biosynthesis (heme, chlorophyll, etc.)	Mitochondrial (C4 pathway)	ALAS	2.3.1.37	5-aminolevulinatase synthase
ALAD (HemB)	4.2.1.24	5-aminolevulinatase dehydratase/ porphobilinogen synthase					
PBGD (HemC)	2.5.1.61	porphobilinogen deaminase hydroxymethylbilane synthase					
Cytosolic/ Mitochondrial (C4 pathway) or Plastidic (C5 pathway)		UROS (HemD)		4.2.1.75	uroporphyrinogen-III synthase		
		UROD (HemE)		4.1.1.37	uroporphyrinogen decarboxylase		
		CPOX (HemF)		1.3.3.3	coproporphyrinogen III oxidase		
		PPOX (HemY)		1.3.3.4	protoporphyrinogen oxidase		
		FECH (HemH)		4.99.1.1	ferrochelatase		
		Plastidic (C5 pathway)			GTS (HemA)	1.2.1.70	glutamyl-tRNA reductase
					GTR (HemL)	5.4.3.8	glutamate-1-semialdehyde 2,1-aminomutase
FAAL	2.3.1.86				fatty acid synthase type I, fatty acyl-AMP ligase domain		
Fatty acid biosynthesis and elongation	Cytosolic (polyketide synthase/ fatty acid synthase type I pathway = PKSII/FASI)	KS		2.3.1.86	fatty acid synthase type I, ketoacyl synthase domain		
		AT		2.3.1.86	fatty acid synthase type I, acyl transferase domain		
		DH	2.3.1.86	fatty acid synthase type I, dehydrase domain			
		ER	2.3.1.86	fatty acid synthase type I, enoyl reductase domain			
		KR	2.3.1.86	fatty acid synthase type I, ketoacyl reductase domain			
		ACP	2.3.1.86	fatty acid synthase type I, acyl carrier protein domain			
		SDR	2.3.1.86	fatty acid synthase type I, terminal reductase domain			
		Endoplasmic reticulum (ER) fatty acid elongation pathway)		ELO	2.3.1.199	beta-ketoacyl-CoA synthase	
				KCR	1.1.1.330	beta-ketoacyl-CoA reductase	
				PHS	4.2.1.134	beta-hydroxyacyl-CoA dehydratase	
				TECR	1.3.1.93	trans-2-enoyl-CoA reductase	
				FabD	2.3.1.39	malonyl-CoA-acyl carrier protein transacylase	
				FabG	1.1.1.100	beta-ketoacyl-acyl carrier protein reductase	
	FabH			2.3.1.180	beta-ketoacyl-acyl carrier protein synthase III		
	Plastidic (fatty acid synthase type II pathway = FASII)		FabZ	4.2.1.59	D-3-hydroxyoctanoyl-acyl carrier protein dehydratase		
			FabI	1.3.1.9	enoyl acyl carrier protein reductase		
			FabB/F	2.3.1.41	beta-ketoacyl-acyl carrier protein synthetase		
			ACP		acyl-carrier protein		
			SufB		cysteine desulfurase activator complex subunit SufB		
			ClpC		ATP-dependent Clp protease ATP-binding subunit ClpC		
Other functions	Plastidic (plastid-encoded)	Ycf93		plastid membrane protein of unknown function			

**Table S3: Chromodellid plastidic proteins with complete N-terminal sequence.**

Chromodellid species	Short name	Contig/Protein reference	SP <sup>1</sup>	SP score <sup>1</sup>	SP length <sup>1</sup>	CTP <sup>2</sup>	CTP score <sup>2</sup>	CTP length <sup>2</sup>
Alphamonas edax BE-2	FabF	a7716_61_4_2_1591	Y	0.81	23		0.46	16
Alphamonas edax BE-2	FabG	a22551_86_13_1248_259	Y	0.9	24		0.49	38

Alphamonas edax BE-2	FabZ	a263933_12_2_2_448	Y	0.88	20		0.47	35
Alphamonas edax BE-2	SufE	a110568_24_3_80_688	Y	0.94	20	Y	0.52	34
Chromera velia	ALAD	MMETSP0290_20130426_17951_1_402	Y	0.55	15	Y	0.55	47
Chromera velia	ClpC	MMETSP0290_20130426_3091_1_1013	Y	0.76	18	Y	0.56	92
Chromera velia	CPOX	MMETSP0290_20130426_4127_1_409	Y	0.8	17	Y	0.53	36
Chromera velia	CPOX	MMETSP0290_20130426_4154_1_411	Y	0.7	23		0.49	51
Chromera velia	FabD	MMETSP0290_20130426_21839_1_349	Y	0.85	19	Y	0.51	35
Chromera velia	FabF	MMETSP0290_20130426_1976_1_463	Y	0.59	19	Y	0.55	51
Chromera velia	FabG	MMETSP0290_20130426_20062_1_308	Y	0.88	18		0.49	54
Chromera velia	FabH	MMETSP0290_20130426_17765_1_393	Y	0.77	21	Y	0.55	57
Chromera velia	FabI	MMETSP0290_20130426_14303_1_355	Y	0.82	19	Y	0.54	38
Chromera velia	FabZ	MMETSP0290_20130426_4631_1_213	Y	0.89	17		0.49	54
Chromera velia	Fdx	MMETSP0290_20130426_1982_1_149	Y	0.75	15	Y	0.54	50
Chromera velia	FECH	MMETSP0290_20130426_5910_1_460	Y	0.79	33	Y	0.51	63
Chromera velia	IspD	MMETSP0290_20130426_22696_1	Y	0.82	19	Y	0.51	64
Chromera velia	IspE	MMETSP0290_20130426_14275_1	Y	0.8	18	Y	0.57	53
Chromera velia	PBGD	MMETSP0290_20130426_12195_1_411	Y	0.85	18	Y	0.57	56
Chromera velia	PPOX	MMETSP0290_20130426_18890_1_615	Y	0.58	19	Y	0.52	15
Chromera velia	SufB	MMETSP029020130426_15825_1_566	Y	0.7	18		0.48	25
Chromera velia	SufS	contig12900_JO801918_blast_full	Y	0.72	19	Y	0.5	34
Chromera velia	UROS	MMETSP0290_20130426_22563_1_399	Y	0.48	17	Y	0.52	24
Colpodella angusta BE-6	ALAD	a1810_243_9_1431_172	Y	0.65	19		0.45	14
Colpodella angusta BE-6	CPOX	a5011_127_9_1611_262	Y	0.76	19		0.44	13
Colpodella angusta BE-6	Fdx	a63516_19_2_1_489	Y	0.7	15		0.44	14
Colpodella angusta BE-6	FECH	a17545_33_13_1312_44	Y	0.73	17		0.5	16
Colpodella angusta BE-6	PPOX	a34610_19_5_1_1764	Y	0.7	25		0.45	20
Colpodella angusta BE-6	SufB	a7216_97_2_21570	Y	0.74	19		0.43	15
Colpodella angusta BE-6	SufC	a30602_28_6_898_62	Y	0.61	15		0.43	26
Colpodella angusta BE-6	UROD	a13302_39_9_1650_67	Y	0.54	22		0.47	40
Colpodella angusta Spi-2	FNR	a13692_81_9_976_65	Y	0.58	22		0.45	19
Colpodella angusta Spi-2	IspC	a44398_21_7_1598_168	Y	0.8	20		0.47	57
Colpodella angusta Spi-2	IspD	a18350_62_2_44_820	Y	0.62	18		0.44	25
Colpodella angusta Spi-2	IspE	a31756_25_7_1128_46	Y	0.72	16		0.44	26
Colpodella angusta Spi-2	PBGD	a53819_34_11_1198_62	Y	0.81	21		0.45	6
Vitrella brassicaformis	ClpC	MMETSP0290_20130426_10413_1_925	Y	0.58	19	Y	0.51	74
Vitrella brassicaformis	CPOX	MMETSP0290_20130426_65767_1_398	Y	0.83	19		0.48	41
Vitrella brassicaformis	CPOX	MMETSP0290_20130426_28340_1_400	Y	0.82	23	Y	0.51	71
Vitrella brassicaformis	DXS	MMETSP0290_20130426_5902_1	Y	0.87	19	Y	0.55	73
Vitrella brassicaformis	FabG	MMETSP0290_20130426_48423_1_305	Y	0.82	24	Y	0.54	47
Vitrella brassicaformis	FabZ	MMETSP0290_20130426_8033_1_242	Y	0.7	22	Y	0.55	67
Vitrella brassicaformis	Fdx	MMETSP0290_20130426_11262_1_158	Y	0.82	17		0.48	59
Vitrella brassicaformis	Fdx	MMETSP0290_20130426_12802_1_160	Y	0.62	17		0.49	14
Vitrella brassicaformis	Fdx	MMETSP0290_20130426_5440_1_197	Y	0.9	24		0.46	50









Eukaryotic	OG5_128905	PF3D7_1235200	Plasmodium falciparum 3D7	V-type K -independent H -translocating inorganic pyrophosphatase (VP2)	1057		y			y	y	y	
Eukaryotic	OG5_128905	PF3D7_1456800	Plasmodium falciparum 3D7	V-type H(-)-translocating pyrophosphatase, putative (VP1)	717		y			y	y	y	
Eukaryotic	OG5_128905	TGME49_248670	Toxoplasma gondii ME49	V-type H(+)-translocating pyrophosphatase VP1	816	y	y			y	y	y	
Eukaryotic	OG5_128917	TGME49_259530	Toxoplasma gondii ME49	GalNac	751				y	y	y	y	
Eukaryotic	OG5_128972	PF3D7_1031200	Plasmodium falciparum 3D7	MORN repeat-containing protein 1 (MORN1)	364	y				y	y	y	
Eukaryotic	OG5_128972	TGME49_310440	Toxoplasma gondii ME49	membrane occupation and recognition nexus protein MORN1 (MORN1)	363	y		y		y	y	y	
Eukaryotic	OG5_128979	cgd4_2720	Cryptosporidium parvum Iowa II	P-type ATPase involved in cation transport	1528	y				y	y	y	
Eukaryotic	OG5_129142	TGME49_234570	Toxoplasma gondii ME49	sterol carrier protein-2 HAD-2SCP-2 (HAD2SCP2)	625	y				y	y	y	
Eukaryotic	OG5_129186	PBANKA_020930	Plasmodium berghei ANKA	actin-related protein (ARP1)	384	y				y	y	y	
Eukaryotic	OG5_129627	PF3D7_0918000	Plasmodium falciparum 3D7	secreted acid phosphatase (GAP50)	396	y		y		y	y	y	
Eukaryotic	OG5_129627	TGME49_219320	Toxoplasma gondii ME49	acid phosphatase GAP50 (GAP50)	431			y		y	y	y	
Eukaryotic	OG5_129645	BBOV_IV007730	Babesia bovis T2Bo	cysteine protease 2	445		y			y	y	y	
Eukaryotic	OG5_129767	PF3D7_1229400	Plasmodium falciparum 3D7	macrophage migration inhibitory factor (MIF)	116		y			y	y	y	
Eukaryotic	OG5_130026	PF3D7_1211400	Plasmodium falciparum 3D7	heat shock protein DNAJ homologue Pfj4 (PFJ4)	244		y			y	y	y	
Eukaryotic	OG5_130494	TGME49_289620	Toxoplasma gondii ME49	cathepsin CPC1 (CPC1)	733	y				y	y	y	
Eukaryotic	OG5_130542	PBANKA_141030	Plasmodium berghei ANKA	M1-family aminopeptidase, putative	1064	y				y	y	y	
Eukaryotic	OG5_131119	TGME49_248830	Toxoplasma gondii ME49	phosphoinositide phospholipase PIPLC (PIPLC)	1097			y		y	y	y	
Eukaryotic	OG5_131558	PF3D7_0718300	Plasmodium falciparum 3D7	cysteine repeat modular protein 2 (CRMP2)	2543	y	y			y	y	y	
Eukaryotic	OG5_131558	PF3D7_0911300	Plasmodium falciparum 3D7	cysteine repeat modular protein 1 (CRMP1)	3303	y				y	y	y	
Eukaryotic	OG5_132039	TGME49_242720	Toxoplasma gondii ME49	aspartyl protease ASP5 (ASP5)	1012			y		y	y	y	
Eukaryotic	OG5_132295	PF3D7_1343000	Plasmodium falciparum 3D7	phosphoethanolamine N-methyltransferase (PMT)	266			y		y	y	y	
Eukaryotic	OG5_132467	PBANKA_030490	Plasmodium berghei ANKA	serine repeat antigen 3 (SERA3)	1096	y				y	y	y	
Eukaryotic	OG5_132467	PBANKA_030500	Plasmodium berghei ANKA	serine repeat antigen 2 (SERA2)	1122	y				y	y	y	
Eukaryotic	OG5_132803	TGME49_315560	Toxoplasma gondii ME49	ATP-binding cassette G family transporter ABCG77 (ABCG77)	1163			y	y	y	y	y	
Eukaryotic	OG5_133090	TGME49_315220	Toxoplasma gondii ME49	rhoprtry protein ROP14 (ROP14)	1061	y				y	y	y	
Eukaryotic	OG5_133546	PF3D7_0808200	Plasmodium falciparum 3D7	plasmepsin X	573	y				y	y	y	
Eukaryotic	OG5_133546	TGME49_246550	Toxoplasma gondii ME49	aspartyl protease ASP3 (ASP3)	643			y		y	y	y	
Eukaryotic	OG5_134280	PBANKA_082420	Plasmodium berghei ANKA	perforin like protein 3 (PPLP3)	815	y				-	y	y	
Eukaryotic	OG5_134280	PBANKA_100630	Plasmodium berghei ANKA	perforin like protein 1,sporozoite micronemal protein essential for cell traversal (SPECT2)	810	y				-	y	y	
Eukaryotic	OG5_134280	PY00454	Plasmodium yoelii yoelii 17XNL	MAC/Perforin domain, putative	727	y				-	y	y	
Eukaryotic	OG5_134280	TGME49_204130	Toxoplasma gondii ME49	perforin-like protein PLP1 (PLP1)	1161	y				-	-	y	
Eukaryotic	OG5_134639	TGME49_259200	Toxoplasma gondii ME49	Na+/H+ exchanger NHE1	2288		y			y	y	y	
Eukaryotic	OG5_134918	PF3D7_0522600	Plasmodium falciparum 3D7	inner membrane complex protein	483			y		y	-	y	
Eukaryotic	OG5_134971	gij5708122	Eimeria tenella	microneme protein 5	932	y	y			y	-	-	X
Eukaryotic	OG5_135032	PBANKA_103520	Plasmodium berghei ANKA	LCCL domain-containing protein (CCp3)	1304	y	y			y	y	y	X
Eukaryotic	OG5_135032	PF3D7_1407000	Plasmodium falciparum 3D7	LCCL domain-containing protein (CCp3)	1272	y	y			y	y	y	X
Eukaryotic	OG5_135662	PF3D7_0109100	Plasmodium falciparum 3D7	LCCL domain-containing protein (CCp5)	1029	y				y	y	y	
Eukaryotic	OG5_135662	PF3D7_1451600	Plasmodium falciparum 3D7	LCCL-like protein (FNPA)	865	y				y	y	y	
Eukaryotic	OG5_135731	PF3D7_0506900	Plasmodium falciparum 3D7	rhomboid protease ROM4 (ROM4)	759	y				y	y	y	
Eukaryotic	OG5_135731	TGME49_268590	Toxoplasma gondii ME49	rhomboid protease ROM4 (ROM4)	665	y				y	y	y	
Eukaryotic	OG5_136613	PF3D7_1479000	Plasmodium falciparum 3D7	acyl-CoA synthetase (ACS1a)	820	y				y	y	y	
Eukaryotic	OG5_136898	PF3D7_1227200	Plasmodium falciparum 3D7	potassium channel protein (K1)	1966	y				y	y	y	
Eukaryotic	OG5_137241	PBANKA_093350	Plasmodium berghei ANKA	rhomboid protease ROM1 (ROM1)	278	y	y			y	y	y	
Eukaryotic	OG5_137241	TGME49_200290	Toxoplasma gondii ME49	rhomboid protease ROM1 (ROM1)	293	y		y		y	y	y	
Eukaryotic	OG5_137241	TGME49_263290	Toxoplasma gondii ME49	rhomboid protease ROM2 (ROM2)	283	y		y		y	y	y	
Eukaryotic	OG5_138369	NCLIV_021050	Neospora caninum Liverpool	unspecified product	865	y				y	y	y	
Eukaryotic	OG5_138369	PF3D7_0507500	Plasmodium falciparum 3D7	subtilisin-like protease 1 (SUB1)	688	y	y			y	y	y	
Eukaryotic	OG5_138369	TGME49_204050	Toxoplasma gondii ME49	subtilisin SUB1 (SUB1)	761	y				y	y	y	
Eukaryotic	OG5_138427	gij110681634	Plasmodium gallinaceum	circumsporozoite protein/thrombospondin-related anonymous protein-related protein	1875	y				y	y	y	
Eukaryotic	OG5_138427	gij156257452	Eimeria tenella	microneme 1 precursor	675	y				y	y	y	





Eukaryotic	OG5_165382	PF3D7_0104200	Plasmodium falciparum 3D7	conserved Plasmodium protein, unknown function	466	y					y	-	y	
Eukaryotic	OG5_165464	TGME49_204420	Toxoplasma gondii ME49	oocyst wall protein OWP1 (OWP1)	499				y	y	-	y		
Eukaryotic	OG5_166914	PF3D7_0410000	Plasmodium falciparum 3D7	conserved Plasmodium protein, unknown function	833	y					y	-	y	
Eukaryotic	OG5_168383	cgd1_3500	Cryptosporidium parvum Iowa II	thrombospondin related adhesive protein	687	y					y	y	y	
Eukaryotic	OG5_171965	TGME49_206510	Toxoplasma gondii ME49	toxolysin TLN4 (TLN4)	2340	y					y	y	y	
Eukaryotic	OG5_171990	TGME49_268310	Toxoplasma gondii ME49	oocyst wall protein OWP3 (OWP3)	640				y	y	-	y		
Eukaryotic	OG5_175386	TGME49_206580	Toxoplasma gondii ME49	formin FRM2 (FRM2)	4709			y			y	y	y	
Eukaryotic	OG5_193087	TGME49_259020	Toxoplasma gondii ME49	bradyzoite antigen BAG1 (BAG1)	229		y				y	-	y	
Eukaryotic	OG5_195418	cgd6_780	Cryptosporidium parvum Iowa II	CpTSP8, predicted extracellular protein with 3 TSP1 repeats, an EGF domain and a C-terminal transmembrane domain. adjacent TSP domain containing gene.	625	y		y			y	y	y	
Eukaryotic	OG5_204795	TGME49_293770	Toxoplasma gondii ME49	chitinase-like protein CLP1 (CLP1)	714	y					-	-	y	
Eukaryotic	OG5_205532	PF3D7_0902500	Plasmodium falciparum 3D7	serine/threonine protein kinase, FIKK family (FIKK9.6)	591	y					y	-	y	
Eukaryotic	OG5_209056	TGME49_247540	Toxoplasma gondii ME49	ATP-binding cassette G family transporter ABCG107 (ABCG107)	981		y				y	y	y	
Eukaryotic	OG5_225241	TGME49_246930	Toxoplasma gondii ME49	calmodulin CAM1 (CAM1)	179	y		y			y	y	y	
Eukaryotic	OG5_231889	TGME49_245490	Toxoplasma gondii ME49	microneme protein MIC8 (MIC8)	684	y					y	y	y	
Eukaryotic	OG5_239228	TGME49_294690	Toxoplasma gondii ME49	rhomboid protease ROM5 (ROM5)	841		y				y	y	y	
Eukaryotic	OG5_239269	TGME49_208030	Toxoplasma gondii ME49	microneme protein MIC4 (MIC4)	580	y					y	-	-	X
Eukaryotic	OG5_239269	gij7108680	Sarcocystis muris	microneme protein SML2	138	y					y	-	-	X
Eukaryotic	OG5_239636	TGME49_286150	Toxoplasma gondii ME49	PAN/Apple domain-containing protein	407	y					y	y	-	X
Eukaryotic	OG5_239946	TGME49_282055	Toxoplasma gondii ME49	protein phosphatase PP2C-hn (PP2CHN)	479	y	y				y	y	y	
Eukaryotic	OG5_239994	TGME49_262730	Toxoplasma gondii ME49	rhoptry protein ROP16 (ROP16)	707	y					y	y	y	
Eukaryotic	OG5_240010	TGME49_262010	Toxoplasma gondii ME49	calmodulin CAM2 (CAM2)	142	y		y			y	y	y	
Eukaryotic	OG5_240014	TGME49_261780	Toxoplasma gondii ME49	microneme protein MIC7 (MIC7)	340	y					y	y	y	
Eukaryotic	OG5_241405	gij32816824	Sarcocystis neurona	nucleoside triphosphate hydrolase	714	y					-	-	y	
Eukaryotic	OG5_243453	PF3D7_0424500	Plasmodium falciparum 3D7	serine/threonine protein kinase, FIKK family (FIKK4.1)	622	y					y	-	y	
Eukaryotic	OG5_143115	PF3D7_0414900	Plasmodium falciparum 3D7	armadillo-domain containing rhoptry protein (ARO)	275	y					y	y	y	X,C
Eukaryotic	OG5_136403	TGME49_231630	Toxoplasma gondii ME49	alveolin domain containing intermediate filament IMC4 (ALV4)	452	y		y			y	y	y	X,C
Eukaryotic	OG5_139161	TGME49_275670	Toxoplasma gondii ME49	alveolin domain containing intermediate filament IMC15 (ALV5)	692		y				y	y	y	X
Eukaryotic	OG5_141746	NCLIV_000610	Neospora caninum Liverpool	putative profilin family protein	163	y					y	y	y	X
Eukaryotic	OG5_144120	TGME49_216000	Toxoplasma gondii ME49	alveolin domain containing intermediate filament IMC3 (ALV3)	538		y				y	y	y	X,C
Eukaryotic	OG5_146323	PF3D7_0525800	Plasmodium falciparum 3D7	membrane skeletal protein IMC1-related	300	y		y			y	y	y	X,C
Eukaryotic	OG5_156798	PF3D7_0304000	Plasmodium falciparum 3D7	inner membrane complex protein 1a, putative (IMC1a)	861		y				y	y	y	X,C
Eukaryotic	OG5_156798	TGME49_231640	Toxoplasma gondii ME49	alveolin domain containing intermediate filament IMC1 (ALV1)	609		y				y	y	y	X,C
Eukaryotic	OG5_240413	TGME49_268900	Toxoplasma gondii ME49	dense granular protein GRA10 (GRA10)	894	y					y	y	y	X
Myzozoans	OG5_153786	PBANKA_090710	Plasmodium berghei ANKA	inner membrane complex protein 1b (IMC1b)	535	y					-	-	-	X
Myzozoans	OG5_165465	PF3D7_1321900	Plasmodium falciparum 3D7	conserved Plasmodium protein, unknown function	292	y					y	y	-	X,W
Myzozoans	OG5_147397	TA20205	Theileria annulata strain Ankara	schizont protein e	441	y					y	y	-	X,W
Myzozoans	OG5_144710	PF3D7_1003600	Plasmodium falciparum 3D7	membrane skeletal protein IMC1-related (ALV5)	281			y			y	y	-	X
Myzozoans	OG5_191678	TA19865	Theileria annulata strain Ankara	surface protein d precursor	178	y	y				y	y	-	X
Piroplasmids	OG5_139232	TP04_0051	Theileria parva strain Muguga	polymorphic immunodominant molecule	480		y				-	-	-	R
Piroplasmids	OG5_153115	PF3D7_1233600	Plasmodium falciparum 3D7	asparagine and aspartate rich protein 1 (AARP1)	5439	y					-	-	-	R
Piroplasmids	OG5_156194	PY02159	Plasmodium yoelii yoelii 17XNL	Drosophila melanogaster CG15040 gene product	908	y					-	-	-	R
Piroplasmids	OG5_191821	BBOV_IV009860	Babesia bovis T2Bo	rhoptry-associated protein 1 (RAP-1)	565	y					-	-	-	W
Piroplasmids	OG5_191821	gij255708246	Babesia gibsoni	rhoptry-associated protein-1b	484	y					-	-	-	
Piroplasmids	OG5_191821	gij255708248	Babesia gibsoni	rhoptry-associated protein-1c	437	y					-	-	-	
Piroplasmids	OG5_191821	gij88770794	Babesia gibsoni	rhoptry-associated protein	474	y	y				-	-	-	W
Plasmodium	NO_GROUP	PF3D7_0202200	Plasmodium falciparum 3D7	Plasmodium exported protein, unknown function	290	y					-	-	-	
Plasmodium	NO_GROUP	PF3D7_0424100	Plasmodium falciparum 3D7	reticulocyte binding protein homologue 5 (RH5)	526	y		y			-	-	-	
Plasmodium	NO_GROUP	PF3D7_0424200	Plasmodium falciparum 3D7	reticulocyte binding protein homologue 4 (RH4)	1716	y					-	-	-	
Plasmodium	NO_GROUP	PF3D7_0501200	Plasmodium falciparum 3D7	parasite-infected erythrocyte surface protein (PIESP2)	408	y					-	-	-	R
Plasmodium	NO_GROUP	PF3D7_0935900	Plasmodium falciparum 3D7	ring-exported protein 1 (REX1)	713	y					-	-	-	
Plasmodium	NO_GROUP	PF3D7_0936800	Plasmodium falciparum 3D7	Plasmodium exported protein (PHISTc), unknown function	383	y					-	-	-	R









**Table S5:** Classification of 21 bacterial genes in apicomplexans and relatives acquired by horizontal gene transfer (HGT).

Origin <sup>1</sup>	EuPathDb accession	Short name	Protein name	EC	Pathway	Original reference for HGT
API+CHP	TGME49_261850	PREX	DNA polymerase complex (plastid)	2.7.7.7	DNA replication	Seow et al., 2005 (10)
API+CHP	TGME49_227420	IspH, (LytB)	4-hydroxy-3-methylbut-2-enyl diphosphate reductase (plastid)	1.17.1.2	Isoprenoid biosynthesis	Identified in this study
API+CHP	TGME49_258650	FeCH (HemH)	ferrochelatase (mitochondrial)	4.99.1.1	Tetrapyrrole biosynthesis	Sato & Wilson, 2003 (11)
API+CHP	TGME49_217740	FabG	3-oxoacyl-ACP reductase (plastid)	1.1.1.100	Fatty acid biosynthesis	Identified in this study
API+CHP	cgd7_480	MDH/LDH	malate/lactate dehydrogenase	1.1.1.37	Carbohydrate metabolism	Madern et al., 2003 (12)
API+CHP	cgd6_4570	GlnA	glutamine synthetase	6.3.1.2	Amino acid and nitrogen metabolism	Huang et al., 2004a (5)
API+CHP <sup>2</sup>	TGME49_208090	YgfA	5-formyltetrahydrofolate cyclo-ligase	6.3.3.2	Folate metabolism	Identified in this study
MYZ	TGME49_307030	PNP	purine-nucleoside phosphorylase	2.4.2.1	Purine metabolism	Kicska et al., 2003 (13)
MYZ	TGME49_244430	PUS	pseudouridylate synthase	4.2.1.70	Pyrimidine metabolism	Identified in this study
MYZ	TGME49_233140	DUT	desoxyuridine 5-triophosphate nucleotidohydrolase	3.6.1.23	Pyrimidine metabolism	Identified in this study
MYZ	cgd4_740	Thio	thioredoxin peroxidase	1.11.1.15	Redox reactions	Huang et al., 2004a (5)
MYZ	TGME49_247510	Fbp	fructose-1,6-bisphosphatase	3.1.3.11	Carbohydrate metabolism	Huang et al., 2004b (14)
MYZ	cgd6_3280	GlgB	1,4-alpha-glucan branching enzyme	2.4.1.18	Carbohydrate metabolism	Huang et al., 2004a (5)
EUK	cgd2_570	TrmH	tRNA/rRNA methyltransferase	2.1.1.34	RNA modification	Huang et al., 2004a (5)
EUK	cgd6_3750	AMY	alpha-amylase	3.2.1.1	Carbohydrate metabolism	Huang et al., 2004a (5)
EUK	cgd1_730	QPCT	glutamine cyclotransferase	2.3.2.5	Peptide modification	Huang et al., 2004a (5)
EUK	cgd8_3430	ANPEP	aminopeptidase N	3.4.11.2	Peptide modification	Huang et al., 2004a (5)
EUK	cgd2_2020		hypothetical protein		Unknown	Huang et al., 2004a (5)
EUK	cgd5_3230	SOD	superoxide dismutase	1.15.1.1	Superoxide conversion	Huang et al., 2004a (5)
EUK	cgd8_350	NARS	asparaginyl-tRNA synthetase	6.3.5.6	Aminoacyl-tRNA synthesis	Huang et al., 2004a (5)
EUK <sup>3</sup>	cgd6_2130	RsmE	16S RNA methyltransferase	2.1.1.193	RNA modification	Huang et al., 2004a (11)

<sup>1</sup> Acquired in the ancestor of apicomplexans and chrompodellids (API+CHP), myzozoans (MYZ) or shares a common origin with one or more other eukaryotes (EUK).

<sup>2</sup> YgfA is absent in dinoflagellates and most other eukaryotes.

<sup>3</sup> The relationship of apicomplexan RsmE to that of bacteria and other eukaryotes is unresolved in phylogenies (however a related homolog is found in chrompodellids suggesting that neither this gene represents an apicomplexan-specific HGT).

**Table S6:** Classification of 74 unique proteins specific to apicomplexans, apicomplexans and chrompodellids, and myzozoans.

Origin	Localization	Protein EuPathDb accession	Organism	Protein annotation	Len gth	OrthoMCL Group	Note	Putative function (reference)
Apicomplexan-specific	Apical	PF3D7_1318000	Plasmodium falciparum 3D7	conserved Plasmodium membrane protein, unknown function	192	OG5_145102		Apical protein linked to merozoite invasion (15)
Apicomplexan-specific	Apical	PF3D7_1030200	Plasmodium falciparum 3D7	conserved Plasmodium protein, unknown function	450	OG5_143206	R	Apical protein linked to merozoite invasion (15)
Apicomplexan-specific	Apical	PF3D7_0210600	Plasmodium falciparum 3D7	conserved Plasmodium protein, unknown function	446	OG5_141751		Apical protein linked to merozoite invasion (15)
Apicomplexan-specific	Apical	PF3D7_0214900	Plasmodium falciparum 3D7	rhostry neck protein 6 (RON6)	950	OG5_150575	R	Secreted from rhostrys during merozoite invasion (16)
Apicomplexan-specific	Apical	TGME49_310010	Toxoplasma gondii ME49	rhostry neck protein RON1 (RON1)	1158	OG5_222773	R	GPI anchor-containing rhostry membrane protein (17)
Apicomplexan-specific	Apical	PBANKA_141830	Plasmodium berghei ANKA	rhostry protein 2, putative (PRP2)	1272	OG5_166877	R	Essential rhostry protein possibly involved in rhostry assembly (18)
Apicomplexan-specific	Exported	PF3D7_1463900	Plasmodium falciparum 3D7	conserved Plasmodium membrane protein, unknown function	1071	OG5_141738		Exported to erythrocyte from merozoite (19)
Apicomplexan-specific		TGME49_261750	Toxoplasma gondii ME49	rhostry neck protein RON10 (RON10)	835	OG5_171370		Possibly involved in sorting proteins to rhostrys (20)
Apicomplexan-specific		cgd7_4030	Cryptosporidium parvum Iowa II	hypothetical low complexity protein	586	OG5_160601		Unknown
Apicomplexan-specific		TGME49_200330	Toxoplasma gondii ME49	hypothetical protein	589	OG5_142982		Unknown

Apicomplexan-specific		TGME49_202120	Toxoplasma gondii ME49	hypothetical protein	1322	OG5_142642	Unknown
Apicomplexan-specific		TGME49_222350	Toxoplasma gondii ME49	hypothetical protein	410	OG5_143198	Unknown
Apicomplexan-specific		TGME49_222940	Toxoplasma gondii ME49	hypothetical protein	489	OG5_171982	Unknown
Apicomplexan-specific		TGME49_235670	Toxoplasma gondii ME49	hypothetical protein	558	OG5_156810	Unknown
Apicomplexan-specific		TGME49_244080	Toxoplasma gondii ME49	hypothetical protein	376	OG5_171957	Unknown
Apicomplexan-specific		TGME49_274160	Toxoplasma gondii ME49	hypothetical protein	525	OG5_145098	Unknown
Apicomplexan-specific		TGME49_282020	Toxoplasma gondii ME49	hypothetical protein	394	OG5_180667	Unknown
Apicomplexan-specific		TGME49_318390	Toxoplasma gondii ME49	hypothetical protein	1278	OG5_171978	Unknown
Apicomplexan-specific		TGME49_202290	Toxoplasma gondii ME49	membrane protein, putative	349	OG5_141728	Unknown
Apicomplexan-specific	IMC	TGME49_202500	Toxoplasma gondii ME49	GAPM1a	305	OG5_140542	Six-pass transmembrane IMC protein with IMC-anchoring role in gliding (21)
Apicomplexan-specific	IMC	TGME49_206690	Toxoplasma gondii ME49	glideosome-associated protein with multiple-membrane spans GAPM2B (GAPM2B)	426	OG5_141755	Six-pass transmembrane IMC protein with IMC-anchoring role in gliding (21)
Apicomplexan-specific	IMC	TGME49_271970	Toxoplasma gondii ME49	glideosome-associated protein with multiple-membrane spans GAPM3	310	OG5_143227	Six-pass transmembrane IMC protein with IMC-anchoring role in gliding (21)
Apicomplexan-specific	Oocyst wall	cgd6_200	Cryptosporidium parvum Iowa II	oocyst wall protein 8	457	OG5_225269	Oocyst wall-specific protein (9)
Apicomplexan-specific	Oocyst wall	TGME49_209610	Toxoplasma gondii ME49	oocyst wall protein OWP2 (OWP2)	462	OG5_239307	Oocyst wall-specific protein (22)
Apicomplexan-specific	Apical	PF3D7_0414600	Plasmodium falciparum 3D7	conserved protein, unknown function	235	OG5_145116	Apical protein linked to merozoite invasion (15)
Api+Chromp-specific		TGME49_200460	Toxoplasma gondii ME49	hypothetical protein	436	OG5_171961	Unknown
Api+Chromp-specific		TGME49_202040	Toxoplasma gondii ME49	hypothetical protein	1047	OG5_143225	Unknown
Api+Chromp-specific		TGME49_205680	Toxoplasma gondii ME49	hypothetical protein	805	OG5_156805	R Unknown
Api+Chromp-specific		TGME49_209490	Toxoplasma gondii ME49	hypothetical protein	505	OG5_171967	Unknown
Api+Chromp-specific		TGME49_209600	Toxoplasma gondii ME49	hypothetical protein	451	OG5_171975	Unknown
Api+Chromp-specific		TGME49_217480	Toxoplasma gondii ME49	hypothetical protein	1589	OG5_169915	Unknown
Api+Chromp-specific		TGME49_219160	Toxoplasma gondii ME49	hypothetical protein	349	OG5_145100	Unknown
Api+Chromp-specific		TGME49_240470	Toxoplasma gondii ME49	hypothetical protein	409	OG5_171976	Unknown
Api+Chromp-specific		TGME49_245520	Toxoplasma gondii ME49	hypothetical protein	235	OG5_169642	Unknown
Api+Chromp-specific		TGME49_255410	Toxoplasma gondii ME49	hypothetical protein	165	OG5_143229	Unknown
Api+Chromp-specific		TGME49_258920	Toxoplasma gondii ME49	hypothetical protein	481	OG5_150770	Unknown
Api+Chromp-specific		TGME49_262890	Toxoplasma gondii ME49	hypothetical protein	555	OG5_144475	Unknown
Api+Chromp-specific		TGME49_263440	Toxoplasma gondii ME49	hypothetical protein	600	OG5_163433	Unknown
Api+Chromp-specific		TGME49_264020	Toxoplasma gondii ME49	hypothetical protein	647	OG5_171222	Unknown
Api+Chromp-specific		TGME49_266630	Toxoplasma gondii ME49	hypothetical protein	582	OG5_141750	Unknown
Api+Chromp-specific		TGME49_268690	Toxoplasma gondii ME49	hypothetical protein	577	OG5_160615	Unknown
Api+Chromp-specific		TGME49_273040	Toxoplasma gondii ME49	hypothetical protein	311	OG5_143217	Unknown
Api+Chromp-specific		TGME49_273705	Toxoplasma gondii ME49	hypothetical protein	424	OG5_145099	Unknown
Api+Chromp-specific		TGME49_311880	Toxoplasma gondii ME49	hypothetical protein	728	OG5_160577	Unknown
Api+Chromp-specific		TGME49_313380	Toxoplasma gondii ME49	hypothetical protein	274	OG5_141745	Unknown
Api+Chromp-specific		TGME49_309850	Toxoplasma gondii ME49	insulin-degrading enzyme	456	OG5_147636	Unknown
Api+Chromp-specific		cgd8_5160	Cryptosporidium parvum Iowa II	uncharacterized apicomplexan-specific protein	170	OG5_143199	Unknown
Api+Chromp-specific		cgd7_1550	Cryptosporidium parvum Iowa II	uncharacterized protein	840	OG5_195587	Unknown
Myzozoan specific	Exported, Membrane	TA20205	Theileria annulata strain Ankara	schizont protein e	441	OG5_147397	W Exported protein associated with host cell microtubules (23)
Myzozoan specific	Exported, Membrane	TA19865	Theileria annulata strain Ankara	surface protein d precursor	178	OG5_191678	Exported membrane protein (24)
Myzozoan specific	Apical	PF3D7_1321900	Plasmodium falciparum 3D7	conserved Plasmodium protein, unknown function	292	OG5_165465	W Apical protein linked to merozoite invasion (15)
Myzozoan specific	IMC	TGME49_237820	Toxoplasma gondii ME49	IMC sub-compartment protein ISP2 (ISP2)	160	OG5_141732	IMC protein involved in cell division (25)
Myzozoan specific	Apical	PBANKA_090710	Plasmodium berghei ANKA	inner membrane complex protein 1b, putative (IMC1b)	504	OG5_153786	Membrane skeleton protein involved in cell shape maintenance, gliding motility, and infection (26)
Myzozoan specific	IMC	PF3D7_1003600	Plasmodium falciparum 3D7	IMC-related (ALV5)	281	OG5_144710	Alveolin domain-containing protein linked to merozoite invasion (15)
Myzozoan specific		cgd8_5030	Cryptosporidium parvum Iowa II	articulin family protein, Pfs77 protein-related	324	OG5_147639	Alveolin domain-containing protein
Myzozoan specific		TGME49_260500	Toxoplasma gondii ME49	COPI associated protein, putative	270	OG5_143228	Unknown
Myzozoan specific		cgd8_2990	Cryptosporidium parvum Iowa II	hypothetical protein	180	OG5_151232	Unknown
Myzozoan specific		TGME49_203358	Toxoplasma gondii ME49	hypothetical protein	395	OG5_143204	Unknown
Myzozoan specific		TGME49_205320	Toxoplasma gondii ME49	hypothetical protein	655	OG5_141752	Unknown
Myzozoan specific		TGME49_207240	Toxoplasma gondii ME49	hypothetical protein	108	OG5_171986	Unknown
Myzozoan specific		TGME49_209890	Toxoplasma gondii ME49	hypothetical protein	193	OG5_138808	Unknown
Myzozoan specific		TGME49_215430	Toxoplasma gondii ME49	hypothetical protein	244	OG5_143223	Unknown
Myzozoan specific		TGME49_216410	Toxoplasma gondii ME49	hypothetical protein	188	OG5_143208	Unknown
Myzozoan specific		TGME49_225840	Toxoplasma gondii ME49	hypothetical protein	686	OG5_156809	Unknown
Myzozoan specific		TGME49_244360	Toxoplasma gondii ME49	hypothetical protein	383	OG5_145121	Unknown



Myzozoon specific	TGME49_258820	Toxoplasma gondii ME49	hypothetical protein	129	OG5_141758	Unknown
Myzozoon specific	TGME49_265510	Toxoplasma gondii ME49	hypothetical protein	272	OG5_146989	Unknown
Myzozoon specific	TGME49_267510	Toxoplasma gondii ME49	hypothetical protein	346	OG5_150760	Unknown
Myzozoon specific	TGME49_269310	Toxoplasma gondii ME49	hypothetical protein	968	OG5_141231	Unknown
Myzozoon specific	TGME49_293720	Toxoplasma gondii ME49	hypothetical protein	336	OG5_147633	Unknown
Myzozoon specific	TGME49_297870	Toxoplasma gondii ME49	hypothetical protein	872	OG5_171974	Unknown
Myzozoon specific	TGME49_306980	Toxoplasma gondii ME49	hypothetical protein	226	OG5_147631	Unknown
Myzozoon specific	TGME49_310210	Toxoplasma gondii ME49	hypothetical protein	788	OG5_150590	Unknown
Myzozoon specific	TGME49_310270	Toxoplasma gondii ME49	hypothetical protein	1357	OG5_141163	Unknown

**Table S7:** Domain superfamilies in chrompodellid and dinoflagellate PKS1/FAS1 proteins identified by NCBI Conserved Domains searches.

Organism	Contig translation or Protein name	Start	End	E-value	Bit Score	Super-family	Superfamily description	Domain n <sup>1</sup>	Not e <sup>2</sup>
Colpodella angusta Spi-2	a2699_83_57_7353-79	1	44	1.97E-015	74.98	cl09936	PP-binding	ACP	N
Colpodella angusta Spi-2	a2699_83_57_7353-79	77	474	2.07E-160	502.86	cl09938	cond_enzymes	KS	
Colpodella angusta Spi-2	a2699_83_57_7353-79	527	826	3.96E-107	346.7	cl08282	Acyl_transf_1	AT	
Colpodella angusta Spi-2	a2699_83_57_7353-79	1235	1456	8.06E-048	178.25	cl09931	NADB_Rossmann	KR	N
Colpodella angusta Spi-2	a2699_83_57_7353-79	1504	1564	1.51E-012	66.5	cl09936	PP-binding	ACP	N
Colpodella angusta Spi-2	a2699_83_57_7353-79	1629	1926	6.30E-069	236.78	cl09931	NADB_Rossmann	SDR	
Colpodella angusta Spi-2	a6216_94_7_1-3726	120	209	1.99E-005	46.58	cl00509	hot_dog	DH	N
Colpodella angusta Spi-2	a6216_94_7_1-3726	610	903	3.61E-083	274.06	cl16912	MDR	ER	
Colpodella angusta Spi-2	a6216_94_7_1-3726	950	1121	6.87E-035	133.03	cl09931	NADB_Rossmann	KR	
Colpodella angusta Spi-2	a6216_94_7_1-3726	1184	1239	3.77E-007	49.11	cl09936	PP-binding	ACP	
Colpodella angusta BE-6	a1114_130_47_6309-79	1	126	9.03E-044	166.97	cl09938	cond_enzymes	KS	N
Colpodella angusta BE-6	a1114_130_47_6309-79	179	478	7.33E-108	348.24	cl08282	Acyl_transf_1	AT	
Colpodella angusta BE-6	a1114_130_47_6309-79	887	1108	3.48E-048	179.02	cl09931	NADB_Rossmann	KR	N
Colpodella angusta BE-6	a1114_130_47_6309-79	1156	1216	1.16E-012	66.5	cl09936	PP-binding	ACP	N
Colpodella angusta BE-6	a1114_130_47_6309-79	1281	1578	3.73E-069	237.16	cl09931	NADB_Rossmann	SDR	
Colpodella angusta BE-6	a1273513_2_3_336-1	1	44	2.66E-008	46.86	cl09936	PP-binding	ACP	N
Colpodella angusta BE-6	a1273513_2_3_336-1	64	111	3.31E-014	66.81	cl09938	cond_enzymes	KS	C
Colpodella angusta BE-6	a172836_7_22_2511-1	1	209	1.18E-052	185.08	cl16912	MDR	ER	N
Colpodella angusta BE-6	a172836_7_22_2511-1	222	453	5.26E-067	228.71	cl09931	NADB_Rossmann	KR	N
Colpodella angusta BE-6	a172836_7_22_2511-1	484	562	3.24E-022	92.7	cl09936	PP-binding	ACP	
Colpodella angusta BE-6	a172836_7_22_2511-1	595	833	6.71E-095	306.02	cl09938	cond_enzymes	KS	C
Colpodella angusta BE-6	a490947_4_4_1-738	1	124	8.98E-044	153.87	cl09938	cond_enzymes	KS	N
Colpodella angusta BE-6	a490947_4_4_1-738	153	224	2.27E-014	69.3	cl00509	hot_dog	DH	C
Colpodella angusta BE-6	a511308_3_4_2-496	1	79	1.15E-032	120.36	cl09938	cond_enzymes	KS	N
Colpodella angusta BE-6	a511308_3_4_2-496	112	165	5.45E-012	60.88	cl08282	Acyl_transf_1	AT	C
Colpodella angusta BE-6	a950_137_16_23-7138	53	497	1.80E-154	486.3	cl09938	cond_enzymes	KS	
Colpodella angusta BE-6	a950_137_16_23-7138	701	1006	5.75E-045	167.58	cl08282	Acyl_transf_1	AT	
Colpodella angusta BE-6	a950_137_16_23-7138	1098	1383	4.51E-012	67.76	cl00509	hot_dog	DH	
Colpodella angusta BE-6	a950_137_16_23-7138	1740	2033	2.01E-082	275.6	cl16912	MDR	ER	
Colpodella angusta BE-6	a950_137_16_23-7138	2080	2251	2.05E-034	133.03	cl09931	NADB_Rossmann	KR	
Colpodella angusta BE-6	a950_137_16_23-7138	2314	2369	3.82E-007	49.88	cl09936	PP-binding	ACP	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_11567_1_305	1	64	2.50E-009	56.04	cl16912	MDR	ER	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_11567_1_305	82	305	7.82E-078	244.12	cl09931	NADB_Rossmann	KR	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_3923_1_1196	2	101	3.17E-028	118.16	cl09931	NADB_Rossmann	KR	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_3923_1_1196	134	212	9.58E-019	83.45	cl09936	PP-binding	ACP	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_3923_1_1196	370	669	2.32E-074	249.11	cl09931	NADB_Rossmann	SDR	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_6061_1_544	43	442	0.00E+000	534.45	cl09938	cond_enzymes	KS	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_6061_1_544	514	544	5.07E-011	62.42	cl08282	Acyl_transf_1	AT	C
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	25	442	2.42E-171	536.37	cl09938	cond_enzymes	KS	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	502	797	1.96E-109	354.02	cl08282	Acyl_transf_1	AT	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	847	1156	5.94E-050	182.17	cl00509	hot_dog	DH	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	1402	1512	3.90E-011	66.54	cl09931	NADB_Rossmann	KR	C
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	1635	1949	6.75E-081	271.75	cl16912	MDR	ER	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	1977	2211	1.28E-061	219.08	cl09931	NADB_Rossmann	KR	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	2252	2335	1.47E-019	87.31	cl09936	PP-binding	ACP	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_65964_1_2899	2496	2796	2.12E-079	267.6	cl09931	NADB_Rossmann	SDR	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66433_1_1647	52	1010	7.86E-129	415.06	cl17068	AFD_class_I	FAAL	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66433_1_1647	1105	1176	2.48E-009	56.49	cl09936	PP-binding	ACP	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66433_1_1647	1248	1547	5.09E-047	172.45	cl09931	NADB_Rossmann	KR	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66484_1_586	1	115	2.49E-043	158.99	cl09931	NADB_Rossmann	KR	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66484_1_586	218	273	1.07E-015	73.05	cl09936	PP-binding	ACP	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66484_1_586	473	585	5.66E-035	133.16	cl09931	NADB_Rossmann	SDR	C
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66893_1_2173	2	53	5.26E-006	46.86	cl09936	PP-binding	ACP	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66893_1_2173	82	604	1.95E-150	473.58	cl09938	cond_enzymes	KS	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66893_1_2173	636	961	1.69E-105	341.69	cl08282	Acyl_transf_1	AT	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66893_1_2173	1242	1740	2.81E-069	240.65	cl09931	NADB_Rossmann	KR	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66893_1_2173	1791	1874	3.16E-013	68.43	cl09936	PP-binding	ACP	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_66893_1_2173	1910	2169	3.56E-025	107.91	cl19140	Hydrolase_4		
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_67019_1_1077	3	291	8.22E-030	120.92	cl00509	hot_dog	DH	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_67019_1_1077	402	563	4.11E-008	55.37	cl09931	NADB_Rossmann	KR	C
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_67019_1_1077	599	896	2.89E-102	325.29	cl16912	MDR	ER	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_67019_1_1077	930	1076	1.93E-036	137.27	cl09931	NADB_Rossmann	KR	C
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_67680_1_938	605	653	4.63E-008	54.87	cl09938	cond_enzymes	KS	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_67680_1_938	681	820	3.27E-040	151.02	cl08282	Acyl_transf_1	AT	C

Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_68652_1_1566	122	182	6.80E-008	54.5	cl16912	MDR	ER	C
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_68652_1_1566	342	591	9.27E-063	220.62	cl09931	NADB_Rossmann	KR	N
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_68652_1_1566	696	778	7.57E-019	84.22	cl09936	PP-binding	ACP	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_68652_1_1566	837	1161	2.14E-078	262.59	cl09931	NADB_Rossmann	SDR	
Vitrella brassicaformis CCMP3155	MMETSP0288-20130426_68652_1_1566	1310	1493	3.87E-008	54.75	cl19140	Hydrolase_4		
Chromera velia CCMP2878	MMETSP0290-20130426_10064_1_839	2	195	3.77E-041	155.23	cl09931	NADB_Rossmann	KR	N
Chromera velia CCMP2878	MMETSP0290-20130426_10064_1_839	261	345	1.75E-020	87.69	cl09936	PP-binding	ACP	
Voromonas pontica GG	comp4108_c1_seq1_4	3	157	4.83E-026	101.79	cl09931	NADB_Rossmann	KR	N
Voromonas pontica GG	comp4108_c1_seq1_4	234	295	2.58E-009	52.57	cl09936	PP-binding	ACP	
Voromonas pontica GG	comp7328_c0_seq1_8	1	51	2.34E-010	56.91	cl09931	NADB_Rossmann	KR	N
Voromonas pontica GG	comp7328_c0_seq1_8	93	166	1.95E-022	86.92	cl09936	PP-binding	ACP	
Voromonas pontica GG	comp7328_c2_seq1_7	1	120	1.98E-026	103.03	cl16912	MDR	ER	N
Voromonas pontica GG	comp7328_c2_seq1_7	135	209	6.99E-010	54.45	cl09931	NADB_Rossmann	KR	C
Lingulodinium polyedrum	346235801_9_1-2160	35	175	1.58E-016	80.79	cl09931	NADB_Rossmann	KR	C
Lingulodinium polyedrum	346235801_9_1-2160	207	501	5.60E-099	309.11	cl16912	MDR	ER	-
Lingulodinium polyedrum	346235801_9_1-2160	510	720	2.56E-053	189.03	cl09931	NADB_Rossmann	KR	N
Lingulodinium polyedrum	346254191_19_1415-3	1	254	1.83E-096	299.47	cl09938	cond_enzymes	KS	N
Lingulodinium polyedrum	346254191_19_1415-3	273	471	1.67E-084	264.65	cl08282	Acyl_transf_1	AT	C
Lingulodinium polyedrum	346254191_19_1415-3	349	785	7.38E-111	356.1	cl09938	cond_enzymes	KS	-
Lingulodinium polyedrum	346305629_13_1-2883	65	496	1.96E-123	384.6	cl09938	cond_enzymes	KS	-
Lingulodinium polyedrum	346305629_13_1-2883	593	628	1.66E-006	47.21	cl09101	E_set		C
Symbiodinium minutum	003781.t1	285	334	2.41E-012	68.35	cl00615	Membrane-FADS-like		NC
Symbiodinium minutum	003781.t1	554	906	2.24E-108	356.87	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	003781.t1	927	1234	1.61E-082	277.36	cl08282	Acyl_transf_1	AT	-
Symbiodinium minutum	003781.t1	1272	1558	1.19E-033	135.17	cl00509	hot_dog	DH	-
Symbiodinium minutum	003781.t1	1640	1757	1.36E-014	77.71	cl09931	NADB_Rossmann	KR	C
Symbiodinium minutum	003781.t1	1785	2060	1.10E-074	254.8	cl16912	MDR	ER	-
Symbiodinium minutum	003781.t1	2068	2305	7.17E-067	235.26	cl09931	NADB_Rossmann	KR	N
Symbiodinium minutum	003781.t1	2344	2390	2.37E-005	45.7	cl09936	PP-binding	ACP	C
Symbiodinium minutum	003781.t1	3477	3792	3.35E-017	84.79	cl19095	Cyt_C5_DNA_methylase		-
Symbiodinium minutum	003781.t1	4109	4158	9.76E-007	50.46	cl19088	ANK		NC
Symbiodinium minutum	008781.t1	1	114	1.93E-027	116.89	cl09938	cond_enzymes	KS	N
Symbiodinium minutum	008781.t1	142	440	4.58E-026	111.29	cl00509	hot_dog	DH	-
Symbiodinium minutum	008781.t1	511	649	4.79E-009	59.22	cl09931	NADB_Rossmann	KR	C
Symbiodinium minutum	008781.t1	677	1000	6.24E-082	274.06	cl16912	MDR		-
Symbiodinium minutum	008781.t1	1011	1240	1.32E-058	209.45	cl09931	NADB_Rossmann	KR	N
Symbiodinium minutum	008781.t1	1279	1360	2.42E-016	77.29	cl09936	PP-binding	ACP	-
Symbiodinium minutum	008781.t1	1367	1441	1.09E-015	75.36	cl09936	PP-binding	ACP	-
Symbiodinium minutum	008781.t1	1477	1880	2.15E-160	501.7	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	008781.t1	1924	2080	8.14E-020	92.03	cl00509	hot_dog	DH	C
Symbiodinium minutum	008782.t1	9	336	2.44E-080	272.24	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	008782.t1	375	449	2.97E-016	76.91	cl09936	PP-binding	ACP	-
Symbiodinium minutum	008782.t1	476	888	8.22E-149	467.42	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	008782.t1	999	1374	3.45E-050	184.8	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	008782.t1	1407	1484	9.88E-015	72.28	cl09936	PP-binding	ACP	-
Symbiodinium minutum	008782.t1	1521	1840	1.65E-118	381.91	cl09938	cond_enzymes	KS	C
Symbiodinium minutum	008783.t1	15	283	8.37E-032	125.16	cl00509	hot_dog	DH	-
Symbiodinium minutum	008783.t1	376	492	8.68E-013	69.24	cl09931	NADB_Rossmann	KR	C
Symbiodinium minutum	008783.t1	521	557	1.37E-009	58.35	cl16912	MDR	ER	C
Symbiodinium minutum	008784.t1	2	87	1.94E-011	63.74	cl16912	MDR	ER	N
Symbiodinium minutum	008784.t1	89	331	1.29E-059	203.29	cl09931	NADB_Rossmann	KR	N
Symbiodinium minutum	008784.t1	383	424	1.74E-005	43.01	cl09936	PP-binding	ACP	C
Symbiodinium minutum	008784.t1	452	514	2.85E-007	48.4	cl09936	PP-binding	ACP	C
Symbiodinium minutum	012436.t1	88	303	3.77E-041	158.72	cl08282	Acyl_transf_1	AT	N
Symbiodinium minutum	012436.t1	441	723	7.46E-014	75.42	cl19241	Condensation	KS	-
Symbiodinium minutum	012436.t1	884	1295	1.79E-130	424.7	cl17068	AFD_class_I	FAAL	-
Symbiodinium minutum	012436.t1	1795	2124	3.78E-073	256.33	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	012436.t1	2149	2425	6.60E-023	103.97	cl00509	hot_dog	DH	-
Symbiodinium minutum	012436.t1	2518	2846	1.14E-054	200.97	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	012436.t1	2907	2958	1.14E-005	47.56	cl09936	PP-binding	ACP	N
Symbiodinium minutum	012436.t1	3054	3457	3.32E-124	404.25	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	012436.t1	3749	3976	4.90E-016	82.4	cl00509	hot_dog	DH	-
Symbiodinium minutum	012436.t1	4156	4254	4.97E-013	70.1	cl17173	AdoMet_MTases		-
Symbiodinium minutum	012436.t1	4267	4576	5.15E-036	145.6	cl09931	NADB_Rossmann	KR	N
Symbiodinium minutum	012436.t1	4706	5110	2.51E-095	320.66	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	012436.t1	5220	5265	1.90E-003	43.55	cl08282	Acyl_transf_1	AT	C
Symbiodinium minutum	012436.t1	5536	5844	1.60E-043	165.56	cl19241	Condensation	KS	-
Symbiodinium minutum	012436.t1	5870	5929	6.09E-013	68.8	cl07150	NRPS		-
Symbiodinium minutum	012436.t1	6040	6470	1.42E-143	461.14	cl17068	AFD_class_I	FAAL	-
Symbiodinium minutum	012436.t1	6829	6868	2.81E-007	56.4	cl17068	AFD_class_I	FAAL	N
Symbiodinium minutum	012436.t1	6979	7249	1.77E-042	162.48	cl19241	Condensation	KS	-
Symbiodinium minutum	012436.t1	7273	7375	1.25E-007	54.03	cl16371	HxxPF_rpt		-
Symbiodinium minutum	012436.t1	7467	7723	1.45E-069	246.59	cl17068	AFD_class_I	FAAL	N
Symbiodinium minutum	012436.t1	7852	8257	6.08E-100	334.14	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	012436.t1	8331	8571	8.52E-017	84.71	cl00509	hot_dog	DH	-
Symbiodinium minutum	012436.t1	8667	9037	2.53E-067	237.95	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	012436.t1	9082	9142	2.62E-003	40.63	cl09936	PP-binding	ACP	-
Symbiodinium minutum	012436.t1	9172	9582	1.62E-128	416.96	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	012436.t1	9730	10068	5.56E-051	189.9	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	012436.t1	10213	10427	4.85E-019	91.73	cl19140	Hydrolase_4		-
Symbiodinium minutum	015786.t1	1	207	3.45E-061	209.73	cl16912	MDR	ER	N
Symbiodinium minutum	015786.t1	233	452	3.01E-047	173.24	cl09931	NADB_Rossmann	KR	N

Symbiodinium minutum	015786.t1	498	581	5.97E-021	89.23	cl09936	PP-binding	ACP	-
Symbiodinium minutum	015786.t1	625	893	6.91E-020	90.19	cl19140	Hydrolase_4		-
Symbiodinium minutum	015788.t1	30	195	4.03E-048	169.2	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	015788.t1	288	363	3.06E-015	72.67	cl09936	PP-binding	ACP	-
Symbiodinium minutum	015788.t1	393	805	1.60E-148	447.39	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	015789.t1	104	503	1.75E-070	241.42	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	015789.t1	541	615	1.13E-015	74.59	cl09936	PP-binding	ACP	-
Symbiodinium minutum	015789.t1	639	1067	7.74E-144	443.92	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	026959.t1	1	150	1.78E-011	65.07	cl00509	hot_dog	DH	N
Symbiodinium minutum	026959.t1	280	373	4.65E-004	42.27	cl09931	NADB_Rossmann	KR	C
Symbiodinium minutum	026959.t1	404	694	1.52E-096	310.66	cl16912	MDR	ER	-
Symbiodinium minutum	026959.t1	701	930	7.00E-058	205.31	cl09931	NADB_Rossmann	KR	N
Symbiodinium minutum	026959.t1	973	1074	1.02E-014	71.51	cl09936	PP-binding	ACP	-
Symbiodinium minutum	029496.t1	1252	1316	3.11E-007	51.09	cl15299	VCBS		-
Symbiodinium minutum	029496.t1	1305	1371	3.71E-006	48.01	cl15299	VCBS		-
Symbiodinium minutum	029496.t1	1633	1733	2.87E-006	48.96	cl19082	TNFR		-
Symbiodinium minutum	029496.t1	1716	1812	9.09E-009	56.67	cl19082	TNFR		-
Symbiodinium minutum	029496.t1	2220	2618	5.65E-123	399.63	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	029496.t1	2651	2928	1.46E-034	137.87	cl00509	hot_dog	DH	-
Symbiodinium minutum	029496.t1	3030	3389	3.77E-070	244.89	cl09931	NADB_Rossmann	KR	-
Symbiodinium minutum	029496.t1	3422	3495	1.19E-015	76.52	cl09936	PP-binding	ACP	-
Symbiodinium minutum	029496.t1	3523	3931	1.81E-134	432.75	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	029496.t1	4078	4378	7.99E-037	146.66	cl09931	NADB_Rossmann	KR	N
Symbiodinium minutum	029496.t1	4409	4489	1.25E-016	79.22	cl09936	PP-binding	ACP	-
Symbiodinium minutum	029496.t1	4533	4940	4.89E-140	448.55	cl09938	cond_enzymes	KS	-
Symbiodinium minutum	039996.t1	1	253	2.61E-085	279.83	cl09938	cond_enzymes	KS	C
Symbiodinium minutum	039996.t1	289	591	3.51E-111	343.62	cl08282	Acyl_transf_1	AT	-
Symbiodinium minutum	039996.t1	627	780	1.07E-020	92.8	cl00509	hot_dog	DH	C
Symbiodinium minutum	040601.t1	53	347	1.41E-071	230.15	cl16912	MDR	ER	-
Symbiodinium minutum	040601.t1	359	448	8.97E-017	77.52	cl09931	NADB_Rossmann	KR	C

<sup>1</sup> See Fig. 3A and Table S2 for domain organization and names, respectively. Note that FAAL domains in *Symbiodinium* are highly divergent and phylogenetically distant to those in *Cryptosporidium* and chrompodellids.

<sup>2</sup> N=incomplete at N-terminus; C=incomplete at C-terminus