

High Precision and High Yield Fabrication of Dense Nanoparticle Arrays onto DNA Origami at Statistically Independent Binding Sites [†]

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S1 – SEQUENCE OF THE STAPLE STRANDS FOR EACH DESIGN

For the DNA nanostructures described in this research (Figure S1), black sequences represent standard sequences, red sequences represent the α sequence, blue sequences represents the β sequence, green sequences represent the filler sequences, bold orange sequences represent the linker strands, and the bold black represents biotin functionalization. The standard DNA nanotube oligos are for synthesizing a DNA nanotube without any binding sites (Table S1). Each design was synthesized by replacing the designated standard DNA nanotube oligos with modified oligos (Table S2). The names of the standard DNA nanotube oligos to be replaced by the modified oligos are listed on the “Replace” column of the table.

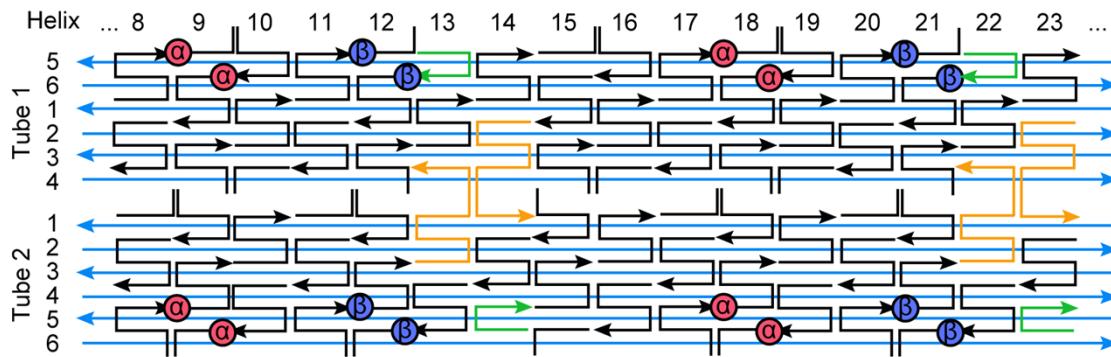


Figure S1: Schematic of a 4x DNA nanostructure. The red and blue circles represent the location of extended α and β tethers, respectively. The orange strands represent the linker strands, which crossover two nanotubes. The green strands represent the filler strands, which fill-in the single stranded DNA regions created by adding the linker strands.

Table S1: List of Standard DNA Nanotube Oligos

Name	Sequence
H1-C2	GCCAGAGGGGGTAAAGACTCCTTATTACAACGCAAAGACACC
H1-C5	CAATACTGCCAATAACGCAATAATAACATAGAAAATTCTATA
H1-C8	AAATGCTTAAACATAAGCAGATAGCCGCACATTCAACCGA
H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCTTTAA
H1-C14	GCGGATTGCATCAACAAGAATTGAGTTAGCCATTGGGAATT
H1-C17	CAAATATCGCGTTAGTCAGAGGTAAATTACCATAGCAAG
H1-C20	GGAAGCAAACCTCCAGAACAGCGCATTAGACATAGCAGCACCGTA
H1-C23	TTGCTCCTTTGATTGAAAATAGCAGCCTAGCGTCAGACTG
H1-C26	GCTTAATTGCTGAACCCAATCCAATAATAGCCCCCTTATT
H1-C29	ATATGCAACTAAAGGCCCTAATTGCCAGTCACCGGAACCGAGA
H1-C32	AACAGTTGATTCCCTTATCCTGAATCTCCGCCACCCCTCAGA
H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCGAGA
H1-C38	TATATTTCATTTGAGGCCTTCTAGCGAACAGGGAGTTAGACT
H1-C41	TCTACTAATAGTAGCAAATCAGATATAGATCCTTGCCGAA
H1-C44	GCAAGGCAAAGAATTTCATCGTATTATCATTGCG

H1-C47	GCATAAAGCTAAATATTAAACCAAGTACATTATCATCATATT
H1-C50	TAATACTTTGCGGATCAATAATCGGCTAATATAATCCTGAT
H1-C53	AAAATTTTAGAACAAAAATAATATCCCAGGGTTAGAACCTA
H1-C56	GTAATGTGTAGGTAAGAACGCGCTGTTAGAAATAAGAAAT
H1-C59	GACAGTCAAATCACTGTCCAGACGACTGAATATACAGTAA
H1-C62	TGATAAATTAATGCAGTAATAAGAGAATAACGGATTGCCCTG
H1-C65	TACAAAGGCTATCAAACAACGCCAACATGCGCAGAGGCGAAT
H1-C68	AAGAGAATCGATGACCAACGCTCACAGAGATGATGAAACAA
H1-C71	CATATGTACCCCGGTTAGTATCATATGTAACAATTTCATT
H1-C74	GAAGATTGTATAAGATAAGAATAAACACATAAATCAATATAT
H1-C77	TTTGTAAAATTCTGAATGGTTGAAATCGTCGCTATTAAATT
H1-C80	TTTAACCAATAGGTTCAAATATTTAGCGATAGCTTAGA
H1-C83	CCTCCTGTAGCCATGATGCAAATCCAATTATCAAATCA
H2-C2	TATCATAACCCCTCGCTTCCAGACGGTACAAACTACAAC
H2-C5	CATAACGCCAAAAGTTGCTAAACAACCTCCAATAGGAACCCA
H2-C8	TCAGTTGAGATTAAAGGAACAACTAAACCACCCCTAGAGCC
H2-C11	AACGAACTAACGGATGAAAATCTCAAAGGTTAGTACCGCC
H2-C14	TATACCAGTCAGGAGTATCGGTTATCAATATAAGTATAGCC
H2-C17	ATCATTGTGAATTAAGCTTGATACCGATTGGCTCAGTAC
H2-C20	CGAGTAGTAAATTGGCCCACGCATAACCAGAGGCTGAGACTC
H2-C23	TCATTCACTGAAATAGAGTTAAAGGCCGCTGCCTATTCGGAA
H2-C26	AGAACCGGATATTCAAAGACAGCATGGGTGCCTTGAGTAAC
H2-C29	GGCGCATAGGCTGGTGAGGACTAAAGAGATGATACAGGAGT
H2-C32	TGACCAACTTGAAGGGTAAACATCGTATCTGAATTACC
H2-C35	GCCGGAACGAGGCGCGAAAGAGGCAAAACAAACAAATAATC
H2-C38	GATAAATTGTGTCGCCAGCGATTATACAGAAGTAGTTGAGG
H2-C41	TTTGCCTATTGGCTCTTCACCACTGTAATAGATTAGAGC
H2-C44	CCAGCTGCATTAATCGCCTGGCCCTGAGTTGAGGAAGGTTAT
H2-C47	GTTGCGCTCACTGCTGCCAGCAGGCAATCAATATCTGGT
H2-C50	AGCCTGGGTGCCTATCGCAAAATCCCCTAAAGCATCAC
H2-C53	CACAATTCCACACAGGTTGAGTGTGCTGCAACAGTGC
H2-C56	ATCATGGTCATAGCAAGAACGTGGACTCAGCAGAACGATAAAA
H2-C59	GTCGACTCTAGAGGCAGGGCGATGCCCTAGCCCTAAACAT
H2-C62	CGTTGTAACGACTTTGGGTCAGCAATATTGAAAT
H2-C65	AGGCGATTAAGTTGAAAGGGAGCCCCCGAGAACCCCTTGAC
H2-C68	TCTCGCTATTACGAACGTGGCGAGAACACACGACCAGTAA
H2-C71	TTCAGGCTCGCAAGCTAGGGCGTGGCAATCGTCTGAAATG
H2-C74	ACCGCTTCTGGTGCACCACACCCCGCAACAGGAAAAACGC
H2-C77	GTATCGCCTCAGGTATGGTTGCTTGACTGCTGGTAATAT
H2-C80	GCATCGTAACCGTGAGAACATCAGAGCGGGAAATAACATCACTTG
H2-C83	GGATTGACCGTAATTTAGACAGGAACGATCACGCAAATTAA
H3-C1	ATCTAAAGTTGTTACCAAGACGGCAAAAGAAGTTT
H3-C4	TTCTGTATGGGATTGAATTACGAGGCATGACTGGATAGCGTC
H3-C7	GGAGTGAGAATAGAGGAATACCAATTCTATTGAAATCCCCCTC
H3-C10	TAATTTTCACTACAACATTATTACAAATGACCATAATC
H3-C13	AGGAGCCTTAATTGTTGGAGAAAATAGTCAGAACAA

H3-C16	TGAATTCTAAACCCTATGCGATTTAGCCGAAAGACTT
H3-C19	TGACACAACCATCGGCTTGAGATGGTAAGCGAACAGACC
H3-C22	CTGAGGCTTGCAGGAGGCTTGCCCTGACGAGAGTACCTTAA
H3-C25	CACCCCTCAGCAGCGATTACCCAATCAAGCGGATGGCTTAGA
H3-C28	CGGCTACAGAGGCTCTGACCTTCATCAATCAACATGTTTAA
H3-C31	AGTTCCATTAAACAGAGGACAGATGAAGTTCATCCATAT
H3-C34	GCACCAACCTAAAACAGACGGTCAATCAGTAGATTAGTTG
H3-C37	ACTCATTTGACCAAATCCGCACCTGATAACCTGTTAGC
H3-C40	AGTACAATGGTTTGCCAGGGCGGAGATAAGGTGGCATCAAT
H3-C43	TGATTGCCCTCACGAATCGGCCAACGCAATAATCATACAG
H3-C46	CGGTCCACGCTGGTCCGCTTCCAGTCGAATAAGCCTCAGA
H3-C49	ATGGTGGTCCGAAAATGAGTGAGCTAAACATTATGACCCTG
H3-C52	AATAGCCCAGAGATAACACATACGAGCCGGATCAACGCAAGGATA
H3-C55	AGAGTCCACTATTATGTTCTGTGTGAAATGCAATGCCCTGA
H3-C58	GAAAAACCGTCTATATCCCCGGGTACCGTGAGAAAGGCCCGA
H3-C61	CACCCAAATCAAGTGGCCAGTGCCAAGCTCAACCCTCTAGC
H3-C64	TAAATCGGAACCCTGGTAACGCCAGGGTATTTTGAGAGATC
H3-C67	GGGGAAAGCCGGCGCCAGCTGGCGAAAGAGTCTGGAGCAAAC
H3-C70	CGAAAGGAGCGGGCCTGTTGGAAAGGGCACTAGCATGTCAAT
H3-C73	CGCTGCGCGTAACCGGAAACCAGGGCAAAGCCCCAAAAACAG
H3-C76	TACAGGGCGCGTACAAGATCGCACTCCAGTAAACGTTAATAT
H3-C79	GTGCTTCCTCGTTCATGCCCAGTTGTTAAATCAGCTCATT
H3-C82	CCGATTAAAGGGATGGGATAGGTACGTTAACATCGCTCTGG
H3-C85	TGAGAAGTGTGTTTCGTCGGATTCTCGTAAATGTGAGCGAG
H4-C1	GCCTGTAGCATCCAAACATATAAAAGAGCAGTATGTTAGCA
H4-C4	TGTACCGTAACACTTTGTCACAATCAGGAATACCCAAAAG
H4-C7	ACCACCTCATTTCAAAGACAAAAGGAACAAAGTTACCAAG
H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAAGCC
H4-C13	CGGAATAGGTGTATCGTCACCGACTTGAAGCCCAATAATAAG
H4-C16	AGCGGATAAGTGCCACCGAGTAGCACCATGAGCGCTAATATC
H4-C19	CTCAAGAGAAGGATCAATGAAACCACATGGGGAGAATTAACTG
H4-C22	CCTATTATTCTGAAAATCAAGTTGCCCTTACAGAGAGAAT
H4-C25	AGTCCCGTATAAACGGCATTTCGGTCGAAACGATTGTTTG
H4-C28	GTACTGGTAATAAGTTCATAATCAAATTACAAATAAAACA
H4-C31	GTTCCAGTAAGCGTCGCCCTCCCTCAGAGTACCAACGCTAACG
H4-C34	CTCATTAAAGCCAGGCCACCAACCTCATAGTTGCTATTG
H4-C37	CAGGTCAAGCAGATTGCCGCCAGCATTGACCTCCGACTTGC
H4-C40	CGTCAATAGATAATACAACACTGTATTAAAAGGCTTATCCGGT
H4-C43	CTAAAATATCTTAAAAGTTGAGTAACAGGAATCATACCG
H4-C46	CAGTTGGCAAATCACCAGAACGGAGCGGACGCACTCATCGAGA
H4-C49	CTTGTGAACCTCAGATGGCAATTCTGCTTCCATTATCA
H4-C52	CACGCTGAGAGCCATTCTGAATAATGGAATCCTAATTACGA
H4-C55	CAGAGGTGAGGCGGTTGCACGTAAAATATCAACAAATAGAT
H4-C58	CGCCATTAAAAATAGGTTAACGTCAAGAGACAATAACAAACA
H4-C61	GGCTATTAGTCTTTCGGAGAAACAATATAAAGTACCGACA
H4-C64	CTGAAAGCGTAAGACAAGTTACAAATCGTAATTAGGCAGA

H4-C67	TAAAAGGGACATTCACCTGAGCAAAAGATAAGGGCTTAATTGA
H4-C70	GATTATTTACATTGAAATTAAATTACATCGTTACAAATTCTC
H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATTAA
H4-C76	CCAGAACAAATTATTGCTCTGAAATACCGACC GTGTGAT
H4-C79	CCTGAGTAGAAGAAATCCTGAAAACATTAGTTAATTTCATC
H4-C82	CCGTTGTAGCAATAAGAGTCATAGTGATCGCAAGACAAAGA
H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGTTATATAA
H5-C3	ACGGAATAAGTTAGAGTTCTGTCACCATTAGTAAATGAATT
H5-C6	TGGTTTACCA CGCGCCAGGGATAGCAAGCTAACAGTT CAGC
H5-C9	TTGAGGGAGGGAAGCACCCTCAGAACCGGGATTGCGAATAA
H5-C12	AGGTGAATTATCACCACCGTACTCAGGAAAAAAGGCTCCAAA
H5-C15	AGAGCCAGAAAATCGTCGAGAGGGTTGGCTTGCTTCAGG
H5-C18	GCCGAAACGTCACTAGGATTAGCGGGAGTTGCGCCGACAA
H5-C21	ATCAGTAGCGACAGACATGAAAGTATTAGATATATT CGGTG
H5-C24	TAGCGCGTTTCATCAGTTAATGCCCTTTGCGGGATCGT
H5-C27	AGCGTTGCCATCTTTAACGGGGCAAACGAGGGTAGCAA
H5-C30	GCCACCACCGAACCATACATGGCTTTCTTTCATGAGGA
H5-C33	ACCGCCACCCTCAGAATGGAAAGCGCAGATGCCACTACGAAG
H5-C36	ACCACCACCA GAGCGCCCTGATATT CAGAATA CACTAAAAC
H5-C39	TTACAAACAATTGACATTGAGGATTCAAGCGGAAACAA
H5-C42	CGTTATTAATT TAGGAGCACTAACACAGACGGCAACAGC
H5-C45	GAACAAAGAAACCAACAGTTGAAAGGAAAGAGTTGAGCAAG
H5-C48	CCTGATTATCAGATAATATCAAACCCCTCGAAAATCCTGTTG
H5-C51	TGTTGGATTATACGCAGCAAATGAAAATTATAATCAAAG
H5-C54	CCATATCAAATTATCAGTATTAAACACCTCCAGTTGGAACA
H5-C57	TGCGTAGATTTCACCGAACGAACCACCCAACGTCAAAGGGC
H5-C60	CAGTACCTTTACAAATCGCGA ACTGA ACTACGTGAACCAT
H5-C63	ATTGCTTGAAATACATA CGTGGCACAGAGTGCCGTAAAGCAC
H5-C66	TATTCATTTCAATTGGCCAACAGAGATATTAGAGCTTGAC
H5-C69	ACATCAAGAAAACAGCAGATT CACCAGTGGAAAGGAAGAAAG
H5-C72	GAATTACCTTTTACATT TGACGCTCAAGTGTAGCGGTCA
H5-C75	GTGAGTGAATAACCCCGCCAGCCATTGCGCTTAATGCCCGC
H5-C78	AATTTCCCTTAGACTCAA ACTATCGGCCGAGCACGTATAAC
H5-C81	TTAAGACGCTGAGACTCTTGATTAGTAGCTAAACAGGAGG
H5-C84	TAGGTCTGAGAGACAAAAGAGTCAGCTGTCCGTACGCCAGAATCC
H6-C3	AACTGGCATGATTATAGTAAATGTTAAGTAAGAGCAACAC
H6-C6	AAGGAAACCGAGGACGT CATAAA ATTCAACTAATGCAGATA
H6-C9	CTTTTAAGAAAAGGTCAGAAAACGAGGGTAGAAAGATTCA
H6-C12	AGCAAGAAACAATGTACCC TGACTATTAAATCTACGTAAATAA
H6-C15	AGAGAGATAACCCAAAAGATTAAGAGGAAAGACTGGCTCAT
H6-C18	AACACCCCTGAACAATAATTGAGCTCATAATTCAACTTTA
H6-C21	AACATAAAAACAGGACAGGT CAGGATTAGAGAAACACCAGAA
H6-C24	TTAACGTAAAAAGAGGT CATTTCGTAACAAAGCTGC
H6-C27	GCCATATTATTATAATGCTGTAGCGAGTAATCTTGACA
H6-C30	AGCGTCTTCCAGATACGGTGTGGAACGGTGTACAGACCA
H6-C33	CACCCAGCTACAATAATTCTCGCGAACGATAAGGGAACCGAAC

H6-C36	GGGAGGTTTGAAGTCGCAAATGGCACTCCATGTTACTTA
H6-C39	ATTCTAAGAACCGGGCGCAGCTGAATTGTATCATGCCT
H6-C42	CGCCCAATAGCAAGTAGCATTACATCCGGGGAGAGGCAG
H6-C45	ACAAGCAAGCGTTAGCAAAATAAGCGGAAACCTGTCGTG
H6-C48	TTCCAAGAACGGGCGGTTACCAAAACTCACATTAATTGC
H6-C51	GCATGTAGAAACCAGAGAAGCCTTATTAGCATAAAGTGTAA
H6-C54	AAGTCCTGAACAAGCCTCATATATTAAATTGTTATCCGCT
H6-C57	TGTTCAGCTAATGCAAGATTCAAAAGGGAGCTCGAATTGTA
H6-C60	AAAGGTAAAGTAATCATCAATATGATATTGCATGCCTGCAG
H6-C63	GGCATTTCGAGCCGGAGAGGGTAGCTTCCCAGTCACGA
H6-C66	GAATGCCATATTGGTCATTGCCTGAGGGGGATGTGCTGCA
H6-C69	TTACCAAGTATAAGACGGTAATCGTAAAGATCGGTGCGGCC
H6-C72	CTAGAAAAGCCTGTTGATAATCAGAAAAGGCCATTGCCA
H6-C75	AAATAAGGCCTAACAAATTTAAATTGCCAGCTTCCGGC
H6-C78	TTCTGACCTAAATTCAATTGTTAGGGGACGACGACA
H6-C81	ACCGAGAAAACCTAACGCCATAAAAATGGTGTAGATGGC
H6-C84	CTATATGTAATGCGCTTCATCAACATTGGAACAAACGGC

Table S2: List of Replacement Oligos for each Design

1x9 _a Replacement Oligos		Sequence
Name	Replace	Sequence
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAAGCC ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAACTG ACCAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATATAATCAAAATTACAAAATAACA ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAGACGATTGCCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGTTGCACGTAAAATTACAAAATAAGAT ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCAATAGTGATCGCAAGACAAAGA ACCAGTGCTCCTACG

2x9 _a Replacement Oligos		Sequence
Name	Replace	Sequence
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCCGACATTCAACCGA ACCAGTGCTCCTACG
H1-C17 α	H1-C17	CAAATATCGCTTAGTCAGAGGGTAATTACCATAGCAAG ACCAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCCAATTAAGCCCCCTTATT ACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCAGA ACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTCATCGTATTATCATTGCG ACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTTTAGAACAAAAATAATATCCAGGGTTAGAACCTA ACCAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTAAATGCGAGTAAAGAGAATAACGGATTGCCGTG ACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCCGTTAGTATCATATGTAACAATTTCATTT ACCAGTGCTCCTACG
H1-C80 α	H1-C80	TTTTAACCAATAGTTCAAATATTTAGCGATAGCTAGA ACCAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAAGCC ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAACTG ACCAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATATAATCAAAATTACAAAATAACA ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAGACGATTGCCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGTTGCACGTAAAATTACAAAATAAGAT ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCAATAGTGATCGCAAGACAAAGA ACCAGTGCTCCTACG

4x9 _a Replacement Oligos		Sequence	
A-Tube	Name	Replace	Sequence
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAAGCC ACCAGTGCTCCTACG	
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAACTG ACCAGTGCTCCTACG	
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATATAATCAAAATTACAAAATAACA ACCAGTGCTCCTACG	
H4-C37 α	H4-C37	CAGGTCAGACGATTGCCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG	
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG	
H4-C55 α	H4-C55	CAGAGGTGAGGCCGTTGCACGTAAAATTACAAAATAAGAT ACCAGTGCTCCTACG	
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG	
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG	
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCAATAGTGATCGCAAGACAAAGA ACCAGTGCTCCTACG	

H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCGCACATTCAACCGA
H1-C17 α	H1-C17	ACCAGTGCTCCTACG
H1-C26 α	H1-C26	CAAATATCGCGTTAGTCAGAGGTAAATTACCATAGCAAG
H1-C35 α	H1-C35	ACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCTTAATTGCTAACCCAATCCAATAATAGCCCCCTTATT
H1-C53 α	H1-C53	ACCAGTGCTCCTACG
H1-C62 α	H1-C62	ACCATTAGATACATCCTTAAATCAAGATGAGGCCACCAGA
H1-C71 α	H1-C71	ACCAGTGCTCCTACG
H1-C80 α	H1-C80	GCAAGGCAAAGAATTTATTCATCGTATTATCATTTGCG
A_H3-C4	H3-C4	ACCAGTGCTCCTACG
A_H3-C13	H3-C13	AAAATTAGAACAAAAATAATATCCCAGGGTTAGAACCTA
A_H3-C22	H3-C22	ACCAGTGCTCCTACG
A_H3-C31	H3-C31	TGATAAAATTAATGCAGTAATAAGAGAATAACGGATTGCGCTG
A_H3-C40	H3-C40	ACCAGTGCTCCTACG
A_H3-C49	H3-C49	CATATGTACCCGGTTAGTATCATATGTAACAATTTCATT
A_H3-C58	H3-C58	ACCAGTGCTCCTACG
A_H3-C67	H3-C67	TTTTAACCAATAGGTTCAAATATATTAGCGATAGCTAGA
A_H3-C76	H3-C76	ACCAGTGCTCCTACG
A_H6-C5	H1-C5	TTCTGTATGGGATTGAATTACGAGGCATGACTGGATAGCGTCTGTACCGTAACACT
A_H6-C14	H1-C14	AGGAGCCTTAATTGTTGGAGAAAAATAGTCAGAACGGAATAGGTGTAT
A_H6-C23	H1-C23	CTGAGGCTTGCAGGAGGCTTGCCTGACGAGAGTACCTTAACCTATTATTCTGAA
A_H6-C32	H1-C32	AGTTCCATTAAACAGAGGACAGATGAAGTTCATCCATATGTTCCAGTAAGCGT
A_H6-C41	H1-C41	AGTACAATGGTTTGCAGGGCGGAGATAAGGTGGCATCAATCGTCAATAGATAAT
A_H6-C50	H1-C50	ATGGTGGTCCGAAAATGAGTGAGCTAAACATTATGACCCCTGCTGCTAACCTCA
A_H6-C59	H1-C59	GAAAAACCGTCTATATCCCCGGTACCGTGAGAAAGGCCGACGCCATAAAAATA
A_H6-C68	H1-C68	GGGGAAAGCCGGGCCAGCTGGCGAAAGAGTCTGGAGCAAACCTAAAGGGACATT
A_H6-C77	H1-C77	TACAGGGCGCGTACAAGATCGCACTCCAGTAAACGTTAATATCCAGAACATATTA
		AACGCAATAATAACATAGAAAATTATA
		CAAGAATTGAGTTAGCCATTGGGATT
		TGAAAATAGCAGCCTAGCGTCAGACTG
		TTTATCCTGAATCTCCGCCACCCCTCAGA
		CAAATCAGATATAGATCCTTGCCCCGAA
		ATCAATAATCGGCTAATATAATCCTGAT
		TCTGTCCAGACGACTGAATATACAGTAA
		CCAACGCTCAACAGAGATGATGAAACAA
		TAATGGTTGAAATCGTCGCTATTAAATT

B-Tube

H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTACCGAAGCC
H4-C19 α	H4-C19	ACCAGTGCTCCTACG
H4-C28 α	H4-C28	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAACTG
H4-C37 α	H4-C37	ACCAGTGCTCCTACG
H4-C46 α	H4-C46	GTACTGGTAATAAGTTCATATCAAATTACAAAATAACA
H4-C55 α	H4-C55	ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CAGGTTCAGACGATTGCCAGCATTGACCTCCGACTTGC
H4-C73 α	H4-C73	ACCAGTGCTCCTACG
H4-C82 α	H4-C82	CAGAGGTGAGGCCAGTACGTAAAACTATCACAAATAGAT
H1-C8 α	H1-C8	ACCAGTGCTCCTACG
H1-C17 α	H1-C17	CTGAAAGCGTAAGACAAGTTACAAATCGTAATTAGGAGA
H1-C26 α	H1-C26	ACCAGTGCTCCTACG
H1-C35 α	H1-C35	TCATGAAATACCTAATGAAACAGTACCGGAATCATAATT
H1-C44 α	H1-C44	ACCAGTGCTCCTACG
H1-C53 α	H1-C53	CCGTTGTAGCAATAAGAGTCATAGTGATCGCAAGACAAAGA
H1-C62 α	H1-C62	AAATGCTTAAACATAAGCAGATAGCCGCACATTCAACCGA
H1-C71 α	H1-C71	ACCAGTGCTCCTACG
		ACCAGTGCTCCTACG
		GCTTAATTGCTAACCCAATCCAATAATAGCCCCCTTATT
		ACCAGTGCTCCTACG
		ACCATTAGATACATCCTTAAATCAAGATGAGGCCACCAGA
		ACCAGTGCTCCTACG
		GCAAGGCAAAGAATTTATTCATCGTATTATCATTTGCG
		ACCAGTGCTCCTACG
		AAAATTAGAACAAAAATAATATCCCAGGGTTAGAACCTA
		ACCAGTGCTCCTACG
		TGATAAAATTAATGCAGTAATAAGAGAATAACGGATTGCGCTG
		ACCAGTGCTCCTACG
		CATATGTACCCGGTTAGTATCATATGTAACAATTTCATT

H1-C80 α	H1-C80	TTTTAACCAATAGGTTCAAATATATTAGCGATAGCTTAGA ACCAAGTGCTCCTACG
B_H2-C14	H2-C14	TTTGTACAATCAGGAATACCCAAAAG
B_H2-C23	H2-C23	CGTCACCGACTTGAAGGCCAATAATAAG
B_H2-C32	H2-C32	AATCAAGTTGCCTTTACAGAGAGAAAT
B_H2-C41	H2-C41	CGCCTCCCTCAGAGTACCAACGCTAACG
B_H2-C5	H2-C5	ACAACTCGTATTAAGGCTTATCGGT
B_H2-C50	H2-C50	GATGGCAATTCATCGTCTTCCTTATCA
B_H2-C59	H2-C59	GGTTAACGTAGAGACAATAACACA
B_H2-C68	H2-C68	ACCTGAGAAAAGATAGGGCTTAATTGA
B_H2-C77	H2-C77	TTGCTTCTGAAATACCGACCCTGTGAT
B_H5-C13	H4-C13	CATAACGCCAAAAGTTGCTAACAACTCCAATAGGAACCCACAATAACTGCGGAAT
B_H5-C22	H4-C22	TATACCAGTCAGGAGTATCGGTTATCAATATAAGTATAGCCGGATTGCATCAA
B_H5-C31	H4-C31	TCATTCACTGAATAGAGTTAAAGGCCGCTGCCATTTCGGAATTGCTCCTTTGAT
B_H5-C4	H4-C4	TGACCAACTTGAAGGGAAAATACGTATCTCTGAATTACCAACAGTTGATTCCC
B_H5-C40	H4-C40	TTTGCCTATTGGGCTCTTCACCAGTGTAAATAGATTAGAGCTCTACTAATAGTAG
B_H5-C49	H4-C49	AGCCTGGGTGCCTATGCCAAAATCCCCTAAAGCATCACTAATACTTTCGCG
B_H5-C58	H4-C58	GTCGACTCTAGAGGCAGGGCGATGCCCTAGCCCTAAACATGACAGTCAAATCAC
B_H5-C67	H4-C67	TCTTCGCTATTACGAACGTGGCGAGAACACACGACCAGTAAAGAGAATCGATGA
B_H5-C76	H4-C76	GTATCGGCCTCAGGTATGGTGCCTTGACTTGCTGGTAATATTGTTAAAATTCG

1x9 β Replacement Oligos

Name	Replace	Sequence
H4-C22 β	H4-C22	CCTATTATTCTGAAAATCAAGTTGCCTTTACAGAGAGAAAT TCTCTACCGCCTACG
H4-C31 β	H4-C31	GTTCCAGTAAGCGTCGCCTCCCTCAGAGTACCAACGCTAACGT TCTCTACCGCCTACG
H4-C40 β	H4-C40	CGTCAATAGATAATACAACCTCGTATTAAGGCTTATCCGGT TCTCTACCGCCTACG
H4-C58 β	H4-C58	CGCCATTAAAATAGGTTAACGTCAAGAGACAATAACACA TCTCTACCGCCTACG
H4-C67 β	H4-C67	TAAAAGGGACATTCACCTGAGCAAAAGATAGGGCTTAATTGAT TCTCTACCGCCTACG
H4-C76 β	H4-C76	CCAGAACAAATTATTGCTCTGAAATACCGACCCTGTGAT TCTCTACCGCCTACG
H4-C13 β	H4-C13	CGGAATAGGTGTACGTACCGACTGAAGCCAATAATAAG TCTCTACCGCCTACG
H4-C49 β	H4-C49	CTTGCTGAACCTCAGATGGCAATTACGTCTTCCTTATCA TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGTTATAA TCTCTACCGCCTACG

2x9 β Replacement Oligos

Name	Replace	Sequence
H1-C11 β	H1-C11	AAAAATCAGGTCTAAATAGCAATAGCTAAATTATTCAATTAA TCTCTACCGCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCCAGAACGCAATTAGACATAGCAGCACCGTA TCTCTACCGCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCCATTGCCCCAGTCACCGGAACCAGA TCTCTACCGCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCCATTAGCGAACAGGAGTTAGACT TCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATATT TCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAAGAACGCGCCTGTTAGAAATAAGAAAT TCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACACGCCAACATCGCAGAGGGGAAT TCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAAATAACACATAATCAATTATT TCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCAAATTATCAAAATCA TCTCTACCGCCTACG
H4-C13 β	H4-C13	CGGAATAGGTGTACGTACCGACTGAAGCCAATAATAAG TCTCTACCGCCTACG
H4-C22 β	H4-C22	CCTATTATTCTGAAAATCAAGTTGCCTTTACAGAGAGAAAT TCTCTACCGCCTACG
H4-C31 β	H4-C31	GTTCCAGTAAGCGTCGCCTCCCTCAGAGTACCAACGCTAACGT TCTCTACCGCCTACG

H4-C40 β	H4-C40	CGTCAATAGATAATAACAACCTGTATTAAAAGGCTATCCGGT TCTCTACCGCCTACG
H4-C49 β	H4-C49	CTTGCTAACCTCAGATGGCAATTATCGTCTTCCTTATCA TCTCTACCGCCTACG
H4-C58 β	H4-C58	CGCCATTAAAAATAGGTTAACGTAGAGACAATAAACAA TCTCTACCGCCTACG
H4-C67 β	H4-C67	TAAAAGGGACATTACCTGAGCAAAGATAAGGGCTTAATTGA TCTCTACCGCCTACG
H4-C76 β	H4-C76	CCAGAACAAATTATTGCTCTGAAATACCGACC GTGAT TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTATATAA TCTCTACCGCCTACG

4x9 β Replacement Oligos**A-Tube**

Name	Replace	Sequence
H1-C11 β	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCAATTAA TCTCTACCGCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCAGAAGCGCATTAGACATAGCAGCACCGTA TCTCTACCGCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCTAATTGCGCAGTCACCGGAACCAGA TCTCTACCGCCTACG
H1-C38 β	H1-C38	TATATTTCATTTGAGGCAGTTAGCGAACAGGAGTTAGACT TCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATATT TCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAAAGAACCGCCTGTTAGAAATAAGAA TCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAAACGCCAACATGCGCAGAGGCGA TCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAAATAACACATAAAATCAATATA TCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAATTATCAAATCAT TCTCTACCGCCTACG
H4-C13 β	H4-C13	CGGAATAGGTGTATCGTCACCGACTTGAAGCCAATAATAAG TCTCTACCGCCTACG
H4-C22 β	H4-C22	CCTATTATTCTGAAAATCAAGTTGCCTTTACAGAGAGAA TCTCTACCGCCTACG
H4-C31 β	H4-C31	GTTCCAGTAAGCGTCGCCTCCCTCAGAGTACCAACGCTAACGT TCTCTACCGCCTACG
H4-C40 β	H4-C40	CGTCAATAGATAATAACAACCTGTATTAAAAGGCTATCCGGT TCTCTACCGCCTACG
H4-C49 β	H4-C49	CTTGTGAACCTCAGATGGCAATTATCGTCTTCCTTATCA TCTCTACCGCCTACG
H4-C58 β	H4-C58	CGCCATTAAAAATAGGTTAACGTAGAGACAATAAACAA TCTCTACCGCCTACG
H4-C67 β	H4-C67	TAAAAGGGACATTACCTGAGCAAAGATAAGGGCTTAATTGA TCTCTACCGCCTACG
H4-C76 β	H4-C76	CCAGAACAAATTATTGCTCTGAAATACCGACC GTGAT TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTATATAA TCTCTACCGCCTACG
A_H3-C13	H3-C13	AGGAGCCTTAATCGTGGAGAAAATAGTCAGAACGGAATAGGTGTAT
A_H3-C22	H3-C22	CTGAGGCTTGCAGGAGGCTTGCCTGACGAGAGTACCTTTAACCTATTATTCTGAA
A_H3-C31	H3-C31	AGTTCCATTAAACAGAGGACAGATGAAGTTCATCCATATGTTCCAGTAAGCGT
A_H3-C40	H3-C40	AGTACAATGGTTTGCAGGGCGGAGATAAGTGGCATCAATCGTAAAGATAAT
A_H3-C49	H3-C49	ATGGTGGTCCGAAAATGAGT GAGCTAACATTATGACCTGCTGCTGAACTCA
A_H3-C58	H3-C58	GAAAAAACGTCTATATCCCCGGGTACCGTGAGAAAGGCCGACGCCATTAAAATA
A_H3-C67	H3-C67	GGGAAAGCCGGGCCAGCTGGCAAAGAGCTGGAGCAAACATAAAAGGGACATT
A_H3-C76	H3-C76	TACAGGGCGGTACAAGATCGCACTCCAGTAAACGTTAATATCCAGAACAAATT
A_H6-C5	H1-C5	AACGCAATAATAACATAGAAAATT CATA
A_H6-C14	H1-C14	CAAGAATTGAGTTAGCCATTGGATT
A_H6-C23	H1-C23	TGAAAATAGCAGCCTTAGCGTCAGACTG
A_H6-C32	H1-C32	TTTATCCTGAATCTCCGCCACCCCTCAGA
A_H6-C41	H1-C41	CAAATCAGATATA GATCCTTGGCCGA
A_H6-C50	H1-C50	ATCAATAATCGGCTAATATAATCCTGAT
A_H6-C59	H1-C59	TCTGTCCAGACGACTGAATATA CAGTAA
A_H6-C68	H1-C68	CCAACGCTAACAGAGATGATGAAACAA
A_H6-C77	H1-C77	TAATGGTTGAAATCGTCGCTATTAATT

B-Tube		
H1-C11 β	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCACTAA TCTCTACCGCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCCAGAACGCATTAGACATAGCAGCACCGTA TCTCTACCGCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCCATTGCGCAGTCACCGAACAGA TCTCTACCGCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCAGTTAGCGAACAGGAGTAGACT TCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAGCTAAATATTAAACCAAGTACATTATCATCATATT TCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAGAACACGCCCTGTTAGAAATAAGAA TCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAACGCCAACATGCGCAGAGGCGA TCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAATAAACACATAAAATCAATAT TCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTCCTGTAGCCATGATGCAAATCCAAATTATCAAAATCA TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATATAA TCTCTACCGCCTACG
B_H2-C14	H2-C14	CGTCACCGACTTGAAGCCAATAATAAGTCTCTACCGCCTACG
B_H2-C23	H2-C23	AATCAAGTTGCCTTTACAGAGAGAATCTCTACCGCCTACG
B_H2-C32	H2-C32	CGCCTCCCTCAGAGTACCAACGCTAACGCTCTACCGCCTACG
B_H2-C41	H2-C41	ACAACCTGTATTAAGGCTTATCCGGTCTCTACCGCCTACG
B_H2-C5	H2-C5	GATGGCAATTATCGTCTTCCTTATCATCTCTACCGCCTACG
B_H2-C50	H2-C50	GGTTAACGTCAAGAGACAATAAACACATCTCTACCGCCTACG
B_H2-C59	H2-C59	ACCTGAGCAAAGATAAGGCTTAATTGATCTCTACCGCCTACG
B_H2-C68	H2-C68	TTGCTTCTGTAAATACCGACCGTGTGATCTCTACCGCCTACG
B_H2-C77	H2-C77	CATAACGCCAAAGTTGCTAAACAACCTTCAATAGGAACCCACAATACTGCGGA AT
B_H5-C13 β	H4-C13	TATACCAAGTCAGGAGTATCGGTTATCAATATAAGTATAGCCGCGATTGCATCAA
B_H5-C22 β	H4-C22	TCATTCACTGAATAGAGTTAAAGGCCGCTGCCTATTCGGAATTGCTCCTTTGAT
B_H5-C31 β	H4-C31	TGACCAACTTGAAGGGTAAAATACGTATCTCTGAATTACCAACAGTTGATTCCC
B_H5-C40 β	H4-C40	TTTGCCTATTGGGCTCTTCACCAGTGTAAAGATTAGAGCTACTAATAGTAG
B_H5-C49 β	H4-C49	AGCCTGGGTCCTATGGCAAATCCATCTAAAGCATCACTAATACTTGC
B_H5-C58 β	H4-C58	GTGCACTCTAGAGGCAGGGCGATGCCCTAGCCCTAAAACATGACAGTC
B_H5-C67 β	H4-C67	ACATGCTTACCGACCGTGGCAGAAACACACAGGACAGTAAAGAGAATCGATGA
B_H5-C76 β	H4-C76	GTATGCCCTCAGGTATGGTGCTTGAATTGCTGGTAATATTGTTAAAATTG

1x14_a Replacement Oligos

Name	Replace	Sequence
H4-C7 α	H4-C7	ACCACCTCATTCAAAGACAAAAGGAAACAAGTACCA ACCAGTGCTCCTACG
H4-C13 α	H4-C13	CGGAATAGGTGTATCGTCACCGACTTGAAGCCCAATAATAAG ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATGGGGAGAATTAACTG ACCAGTGCTCCTACG
H4-C25 α	H4-C25	AGTCCCCGTATAACGGCATTTCGGTCGAAACGATTG ACCAGTGCTCCTACG
H4-C31 α	H4-C31	GTTCCAGTAAGCGTCGCCCTCCCTCAGAGTACCAACGCTAACG ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTACAGCATTGCCGCCAGATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C43 α	H4-C43	CTAAAATATCTTAAAGTTGAGTAACAGGAATCATTACCG ACCAGTGCTCCTACG
H4-C49 α	H4-C49	CTTGCTAACCTCAGATGGCAATTACATCGTCTTCCTTATCA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGTTGCACGTAAAACATCAACAATAGAT ACCAGTGCTCCTACG
H4-C61 α	H4-C61	GGCTATTAGTCTTCGGGAGAAACAATATAAGTACCGACA ACCAGTGCTCCTACG
H4-C67 α	H4-C67	TAAAAGGGACATTACCTGAGCAAAAGATAGGGCTTAATTGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGAAATACCTAATGAAACAGTACCGGAATCATATT ACCAGTGCTCCTACG
H4-C79 α	H4-C79	CCTGAGTAGAAGAAATCCTGAAAACATTAGTTAATTTCATC ACCAGTGCTCCTACG
H4-C85 α	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATATAA ACCAGTGCTCCTACG

2x14_u Replacement Oligos

Name	Replace	Sequence
H4-C7 α	H4-C7	ACCACCCCTCATTTCAAAGACAAAAGGGAACAAAGTACCAAG ACCAGTGCTCCTACG
H4-C13 α	H4-C13	CGGAATAGGTGTATCGTCACCGACTTGAAGCCCAATAATAAG ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAAACTG ACCAGTGCTCCTACG
H4-C25 α	H4-C25	AGTCCCCGTATAAACGGCATTTCGGTGAAACGATTTTTG ACCAGTGCTCCTACG
H4-C31 α	H4-C31	GTTCCAGTAAGCGTCGCCTCCCTCAGAGTACCAACGCTAACG ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAGACGATTGCCGCCAGCATTGACCTCCCGACTTGC ACCAGTGCTCCTACG
H4-C43 α	H4-C43	CTAAAATATCTTAAAAGTTGAGTAACAGGAATCATTACCG ACCAGTGCTCCTACG
H4-C49 α	H4-C49	CTTGCTAACCTCAGATGGCAATTACATCGTCTTCCTTATCA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGGTTGCACGTAAAACATCAACAATAGAT ACCAGTGCTCCTACG
H4-C61 α	H4-C61	GGCTATTAGTCTTCGGGAGAAACAATATAAGTACCGACA ACCAGTGCTCCTACG
H4-C67 α	H4-C67	TAAAAGGGACATTCACCTGAGCAAAAGATAGGGCTTAATTGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C79 α	H4-C79	CCTGAGTAGAAGAAATCCTGAAAACATTAGTTAATTTCATC ACCAGTGCTCCTACG
H4-C85 α	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATATAA ACCAGTGCTCCTACG
H1-C11 α	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCAATTAA ACCAGTGCTCCTACG
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGTAATTACCATAGCAAG ACCAGTGCTCCTACG
H1-C29 α	H1-C29	ATATGCAACTAAAGGCTAATTGCCAGTCACCGGAACCAGA ACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGGCCACCAGA ACCAGTGCTCCTACG
H1-C47 α	H1-C47	GCATAAAGCTAAATATTAAACCAAGTACATTATCATATT ACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTTTAGAACAAAAATAATATCCCAGGGTTAGAACCTA ACCAGTGCTCCTACG
H1-C65 α	H1-C65	TACAAAGGCTATCAAACACGCCAACATGCGCAGAGGCGAAT ACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCGGTTAGTATCATATGTAACAATTTCATT ACCAGTGCTCCTACG
H1-C83 α	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAATTATCAAATCA ACCAGTGCTCCTACG
H1-C5 α	H1-C5	CAAACTGCGGAATAACGCAATAATAACATAGAAAATTATA ACCAGTGCTCCTACG
H1-C23 α	H1-C23	TTGCTCCTTTGATTGAAAATAGCAGCCTAGCGTCAGACTG ACCAGTGCTCCTACG
H1-C41 α	H1-C41	TCTACTAATAGTAGCAAATCAGATATAGATCCTTGCCCGAA ACCAGTGCTCCTACG
H1-C59 α	H1-C59	GACAGTCAAATCACTCTGTCCAGACGACTGAATATACTAGTAA ACCAGTGCTCCTACG
H1-C77 α	H1-C77	TTTGTAAAATTCTGAATGGTTGAAATCGTCGCTATTAAATT ACCAGTGCTCCTACG

4x14_u Replacement Oligos

A-Tube		Sequence
Name	Replace	Sequence
H4-C7 α	H4-C7	ACCACCCCTCATTTCAAAGACAAAAGGGAACAAAGTACCAAG ACCAGTGCTCCTACG
H4-C13 α	H4-C13	CGGAATAGGTGTATCGTCACCGACTTGAAGCCCAATAATAAG ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAAACTG ACCAGTGCTCCTACG
H4-C25 α	H4-C25	AGTCCCCGTATAAACGGCATTTCGGTGAAACGATTTTTG ACCAGTGCTCCTACG
H4-C31 α	H4-C31	GTTCCAGTAAGCGTCGCCTCCCTCAGAGTACCAACGCTAACG ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAGACGATTGCCGCCAGCATTGACCTCCCGACTTGC ACCAGTGCTCCTACG
H4-C43 α	H4-C43	CTAAAATATCTTAAAAGTTGAGTAACAGGAATCATTACCG ACCAGTGCTCCTACG
H4-C49 α	H4-C49	CTTGCTAACCTCAGATGGCAATTACATCGTCTTCCTTATCA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGGTTGCACGTAAAACATCAACAATAGAT ACCAGTGCTCCTACG
H4-C61 α	H4-C61	GGCTATTAGTCTTCGGGAGAAACAATATAAGTACCGACA ACCAGTGCTCCTACG
H4-C67 α	H4-C67	TAAAAGGGACATTCACCTGAGCAAAAGATAGGGCTTAATTGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG

H4-C79 α	H4-C79	CCTGAGTAGAAGAAATCCTGAAAACATTAGTTAATTTCATC ACCAGTGCTCCTACG
H4-C85 α	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTATATAA ACCAGTGCTCCTACG
H1-C11 α	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCACTAA ACCAGTGCTCCTACG
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATTA ACCAGTGCTCCTACG
H1-C29 α	H1-C29	ATATGCAACTAAAGGCCTAACCTGCCAGTCACCGGAACCAGA ACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCAGA ACCAGTGCTCCTACG
H1-C47 α	H1-C47	GCATAAAGCTAAATATTAAACCAAGTACATTATCATCATATT ACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTAGAACAACAAAATAATATCCCAGGGTTAGAACCTA ACCAGTGCTCCTACG
H1-C65 α	H1-C65	TACAAAGGCTATCAAACAACGCCAACATGCGCAGAGGCAGA ACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCGGTTAGTATCATATGTAACAATTCACTTT ACCAGTGCTCCTACG
H1-C83 α	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAAATTTCATCAAAATCA ACCAGTGCTCCTACG
A_H6-C5 α	H1-C5	AACGCAATAATAACATAGAAAATTCTA ACCAGTGCTCCTACG
A_H6-C23 α	H1-C23	TGAAAATAGCAGCCTTAGCGTCAGACTG ACCAGTGCTCCTACG
A_H6-C41 α	H1-C41	CAAATCAGATATAGATCCTTGCCCCGAA ACCAGTGCTCCTACG
A_H6-C59 α	H1-C59	TCTGTCCAGACGACTGAATATACAGTAA ACCAGTGCTCCTACG
A_H6-C77 α	H1-C77	TAATGGTTGAAATCGTCGCTATTAAATT ACCAGTGCTCCTACG
A_H3-C4	H3-C4	TTCTGTATGGGATTGAATTACGAGGCATGACTGGATAGCGTCTGTACCGTAACACT
A_H3-C13	H3-C13	AGGAGCCTTAATTCGTTGGAGAAAATAGTCAGAAGCAAACGGAATAGGTGTAT
A_H3-C22	H3-C22	CTGAGGCTTGCAGGAGGCTGCCCTGACGAGAGTACCTTAACCTATTATTCTGAA
A_H3-C31	H3-C31	AGTTCCATTAAACAGAGGCAGATGAAGTTCATTCCATATGTTCCAGTAAGCGT
A_H3-C40	H3-C40	AGTACAATGGTTTGCCAGGGCGGAGATAAGGTGGCATCAATCGCAATAGATAAT
A_H3-C49	H3-C49	ATGGTGGTCCGAAAATGAGTGAGCTAAACATTATGACCTGCTGCTAACCTCA
A_H3-C58	H3-C58	GAAAAACCGTCTATATCCCCGGTACCGTGAGAAAGGCCGACGCCATTAAAATA
A_H3-C67	H3-C67	GGGGAAAGCCGGGCCAGCTGGCAGAGTCTGGAGCAAACACTAAAGGGACATT
A_H3-C76	H3-C76	TACAGGGCGCGTACAAGATCGCACTCCAGTAAACGTTAATATCCAGAACATATTA
A_H6-C14	H1-C14	CAAGAATTGAGTTAGCCATTGGGATT
A_H6-C32	H1-C32	TTTATCCTGAATCTCGCCACCCCTCAGA
A_H6-C50	H1-C50	ATCAATAATCGGCTAATATAATCCTGAT
A_H6-C68	H1-C68	CCAACGCTCAACAGAGATGATGAAACAA
B-Tube		
H4-C7 α	H4-C7	ACCACCTCATTCAAAGACAAAAGGGAACAAAGTACCA ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGAGAATTAACTG ACCAGTGCTCCTACG
H4-C25 α	H4-C25	AGTCCCCGTATAAACGGCATTTCGGTGAAACGATTTC ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTACAGCATTGCCGCCAGCATTGACCTCCGACTTG ACCAGTGCTCCTACG
H4-C43 α	H4-C43	CTAAAATATCTTAAAGTTGAGTAACAGGAATCATTACCG ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTAGGGCGTTGCACGAAACTATCAACAAATAGAT ACCAGTGCTCCTACG
H4-C61 α	H4-C61	GGCTATTAGTCTTTCGGAGAAACAATATAAAGTACCGACA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C79 α	H4-C79	CCTGAGTAGAAGAAATCCTGAAAACATTAGTTAATTTCATC ACCAGTGCTCCTACG
H4-C85 α	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTATATAA ACCAGTGCTCCTACG
H1-C11 α	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCACTAA ACCAGTGCTCCTACG
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATTA ACCAGTGCTCCTACG
H1-C29 α	H1-C29	ATATGCAACTAAAGGCCTAACCTGCCAGTCACCGGAACCAGA ACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCAGA ACCAGTGCTCCTACG
H1-C47 α	H1-C47	GCATAAAGCTAAATATTAAACCAAGTACATTATCATATT ACCAGTGCTCCTACG

H1-C53 α	H1-C53	AAAATTTAGAACAAAAATAATATCCCAGGGTTAGAACCTA ACCAGTGCTCCTACG
H1-C65 α	H1-C65	TACAAAGGCTATCAAACAAACGCCAACATGCGCAGAGGCGAAT ACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCCGGTTAGTATCATATGTAACAATTTCATT ACCAGTGCTCCTACG
H1-C83 α	H1-C83	CCTCCTGTAGCCATGATGCAAATCCAATTATCAAAATCA ACCAGTGCTCCTACG
H1-C5 α	H1-C5	CAATACTCGGAATAACGCAATAAACATAGAAAATTATA ACCAGTGCTCCTACG
H1-C23 α	H1-C23	TTGCTCCTTGTGATTGAAAATAGCAGCCTAGCGTCAGACTG ACCAGTGCTCCTACG
H1-C41 α	H1-C41	TCTACTAATAGTAGCAAATCAGATATAGATCCTTGCCTGAA ACCAGTGCTCCTACG
H1-C59 α	H1-C59	GACAGTCAAATCACTCTGTCAGACGACTGAATATACTAGTAA ACCAGTGCTCCTACG
H1-C77 α	H1-C77	TTTGTAAAATTCTGAATGGTTGAAATCGTCGCTATTAAATT ACCAGTGCTCCTACG
H4-C13 α	H4-C13	CGTCACCGACTTGAAGCCCCATAATAAG ACCAGTGCTCCTACG
H4-C31 α	H4-C31	CGCCTCCCTCAGAGTACCAACGCTAACG ACCAGTGCTCCTACG
H4-C49 α	H4-C49	GATGGCAATTCTCGTCTTCCTTATCA ACCAGTGCTCCTACG
H4-C67 α	H4-C67	ACCTGAGCAAAGATAAGGGCTTAATTGA ACCAGTGCTCCTACG
B_H2-C14	H2-C14	TTTGTACAATCAGGAATAACCCAAAAG
B_H2-C23	H2-C23	AATCAAGTTGCCTTTACAGAGAGAAAT
B_H2-C32	H2-C32	ACAACCTGTATTAAAAGGCTTATCCGGT
B_H2-C41	H2-C41	GGTTAACGTCAGAGACAATAACAAACA
B_H2-C5	H2-C5	TTGCTCTGTAAATACCGACCGTGTGAT
B_H2-C50	H2-C50	CATAACGCCAAAAGTTGCTAAACAACTCCAATAGGAACCCACAATACTGCGGAAT
B_H2-C59	H2-C59	TATACCAGTCAGGAGTATCGGTTATCAATATAAGTATAGCCGCGGATTGCATCAA
B_H2-C68	H2-C68	TCATTCACTGAATAGAGTTAAAGGCCGCTGCCTATTCCGAATTGCTCCTTTGAT
B_H2-C77	H2-C77	TGACCAACTTGAAGGGTAAATACGTATCTCTGAATTACCAACAGTTGATTCCC
B_H5-C22	H4-C22	TTTGCCTATTGGGCTCTTTCACCAGTGTAAATAGATTAGAGCTCTACTAATAGTAG
B_H5-C4	H4-C4	AGCCTGGGTGCCTATGGCAAAATCCCCTAAAGCATCACTAATACCTTGC
B_H5-C40	H4-C40	GTCGACTCTAGAGGCAGGGCGATGCCCTAGCCCTAAACATGACAGTCAAATCAC
B_H5-C58	H4-C48	TCTTCGCTATTACGAACGTGGCGAGAAACACACGACCAGTAAAGAGAATCGATGA
B_H5-C76	H4-C76	GTATCGGCCTCAGGTATGGTTGCTTTGACTTGCTGGTAATTGTTAAAATTG

2x18 $\alpha\beta$ Replacement Oligos

Name	Replace	Sequence
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCCGACATTCAACCGA ACCAGTGCTCCTACG
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATAGCAAG ACCAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCCAATAATAGCCCCCTATT ACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACTCCTAAATCAAGATGAGCCGCCACAGA ACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTCATCGTATTATCATTGCG ACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTTAGAACAAAAATAATATCCCAGGGTTAGAACCTA ACCAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTATGCACTAAAGAGAATAACGGATTGCCCTG ACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCCGGTTAGTATCATATGTAACAATTTCATT ACCAGTGCTCCTACG
H1-C80 α	H1-C80	TTTAAACCAATAGGTTCAAATATTTAGCGATAGCTTAGA ACCAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAAGCC ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATGGGGAGAATTAACTG ACCAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATATCAAATTACAAAATAACAA ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAAGACGATTGCCGCCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGGTTGCACGTAAAATCAACAAATAGAT ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG

H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATTACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCATAGTATCGAAGACAAAGAACCAGTGCTCCTACG
H1-C11 β	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCAATTACCGCTCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCAGAACGCATTAGACATAGCAGCACCGTAACCAGTGCTCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCCATTAGGCAGTCACCGGAACCAGAACCAGTGCTCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCAGTTAGCGAACAGGAGTTAGACTTACCGCTCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATATTACCGCTCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAGAACGCCTGTTAGAAAATAAGAAATTACCGCTCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAACGCCAACATGCAGAGGCGAACAGTGCTCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAATAAACACATAATCAATATATTACCGCTCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAATTATCAAATCATACCGCTCCTACG
H4-C13 β	H4-C13	CGGAATAGGTGTATCGTACCGACTTGAAGCCAAATAATAAGTACCGCTCCTACG
H4-C22 β	H4-C22	CCTATTATTCTGAAAATCAAGTTGCCTTACAGAGAGAATTACCGCTCCTACG
H4-C31 β	H4-C31	GTTCCAGTAAGCGTCGCCCTCAGAGTACCAACGCTAACGTTACCGCTCCTACG
H4-C40 β	H4-C40	CGTCAATAGATAATAACACTGTATTAAAAGGCTTATCCGGTTACCGCTCCTACG
H4-C49 β	H4-C49	CTTGCTGAACCTCAGATGGCAATTATCGTCTTCTTATCATACCGCTCCTACG
H4-C58 β	H4-C58	CGCCATTAAAAATAGGTTAACGTACAGAGACAATAAACACAACCAGTGCTCCTACG
H4-C67 β	H4-C67	TAAAAGGGACATTACCTGAGCAAAAGATAGGGCTTAATTGATACCGCTCCTACG
H4-C76 β	H4-C76	CCAGAACAAATTATTGCTCTGAAATACCGACCGTGTGATACCGCTCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTATATAATACCGCTCCTACG

4x18 α/β Replacement Oligos

A-Tube Name	Replace	Sequence
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATAGCAAGACCAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCCAATAAGCCCCCTTATTACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGGCCACCAGAACAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTATTATTCATCGTATTATCATTTGCGACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTAGAACAAAAATAATATCCAGGGTTAGAACCTAACAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTATGCAGTAATAAGAGAATAACGGATTGCGCTGACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCGGTTAGTATCATATGTAACAATTTCATTACCAGTGCTCCTACG
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCGCGACATTCAACCGAACAGTGCTCCTACG
H1-C80 α	H1-C80	TTTTAACCAATAGGTTCAAATATATTAGCGATAGCTTAGAACAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAACGCCACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAAACTGACCAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATAATCAAATTACAAATAAACACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTACAGACGATTGCCGCCAGCATTGACCTCCGACTTGCACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAACGGAGCGACGCACCATCGAGAACAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGGTTGCACGTTAAACTATCAACAAATAGATACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAATCGTAATTAGGCAGAACAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATTACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCATAGTATCGAAGACAAAGAACAGTGCTCCTACG
H1-C11 β	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCAATTACCGCTCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCAGAACGCATTAGACATAGCAGCACCGTAACCAGTGCTCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCCATTAGGCAGTCACCGGAACCAGAACAGTGCTCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCAGTTAGCGAACAGGAGTTAGACTTACCGCTCCTACG

H1-C47 β	H1-C47	GCATAAAGCTAAATATTAAACCAAGTACATTATCATATT TCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTAGGTAGAACCGCGCTTTAGAAATAAGAAAT TCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAACGCCAACATGCGCAGAGGCGAAT TCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAATAAACACATAAATCAATATA TCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAATTTCATAAAAT TCTCTACCGCCTACG
H4-C13 β	H4-C13	CGGAATAGGTGTATCGCACCGACTTGAAGCCAAATAAG TCTCTACCGCCTACG
H4-C22 β	H4-C22	CCTATTATTCTGAAAATCAAGTTGCCTTTACAGAGAGAA TCTCTACCGCCTACG
H4-C31 β	H4-C31	GTTCCAGTAAGCGTCGCCCTCCCTCAGAGTACCAACG TCTCTACCGCCTACG
H4-C40 β	H4-C40	CGTCAATAGATAATACAACCTGTATTAAAAGGCTTATCCGG TCTCTACCGCCTACG
H4-C49 β	H4-C49	CTTGCTAACCTCAGATGGCAATTCATCGTCTTCCTTAT TCTCTACCGCCTACG
H4-C58 β	H4-C58	CGCCATTAAAAATAGGTTAACGTCAGAGACAATAAACAA TCTCTACCGCCTACG
H4-C67 β	H4-C67	TAAAAGGGACATTACCTGAGCAAAAGATAGGGCTTAATT TCTCTACCGCCTACG
H4-C76 β	H4-C76	CCAGAACAAATTATTGCTCTGAAATACCGACCCTGTG TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATATA TCTCTACCGCCTACG
A_H3-C4	H3-C4	TTCTGTATGGGATTGAATTACGAGGCATGACTGGATAGCGTCTGTACCGTAACACT
A_H3-C13	H3-C13	AGGAGCCTTAATTGCTGGAGAAAATAGTCAGAACGAAACGAAATAGGTGTAT
A_H3-C22	H3-C22	CTGAGGCTTGAGGAGGCTTGCCTGACGAGAGTACCTTAAACCTATTATTCTGAA
A_H3-C31	H3-C31	AGTTCCATTAAACAGAGGACAGATGAAGTTCATCCATATGTTCCAGTAAGCGT
A_H3-C40	H3-C40	AGTACAATGGTTTGCCAGGGCGGAGATAAGGTGGCATCAATCGCAATAGATAAT
A_H3-C49	H3-C49	ATGGTGGTCCGAAAATGAGTGAGCTAACATTATGACCTGCTGCTGAACCTCA
A_H3-C58	H3-C58	GAAAAACCGTCTATATCCCCGGTACCGTGAGAAAGGCCGACGCCATTAAAATA
A_H3-C67	H3-C67	GGGGAAAGCCGGCGCCAGCTGGCGAAAGAGTCTGGAGGCAAACAAAGGGACATT
A_H3-C76	H3-C76	TACAGGGCGCGTACAAGATCGCACTCCAGTAAACGTTAATATCCAGAACATATTA
A_H6-C5	H1-C5	AACGCAATAATAACATAGAAAATTATA
A_H6-C14	H1-C14	CAAGAATTGAGTTAGCCATTGGGATT
A_H6-C23	H1-C23	TGAAAATAGCAGCCTTAGCGTCAGACTG
A_H6-C32	H1-C32	TTTATCCTGAATCTCGCCACCCCTCAGA
A_H6-C41	H1-C41	CAAATCAGATATAGATCCTTGCCGAA
A_H6-C50	H1-C50	ATCAATAATGGCTAATATAATCCTGAT
A_H6-C59	H1-C59	TCTGTCCAGACGACTGAATATACAGTAA
A_H6-C68	H1-C68	CCAACGCTAACAGAGATGATGAAACAA
A_H6-C77	H1-C77	TAATGGTTGAAATCGTCGCTATTAATT
B-Tube		
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATAGCAAG ACCAAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCCAATAATAAGCCCCCTTATT ACCAAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCAGA ACCAAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTCATTTCATCGTATTATCATTGCG ACCAAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTTCAGAACAAAAATAATATCCCAGGGTTAGAACCTA ACCAAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTATGCAGTAATAAGAGAATAACGGATTGCCTG ACCAAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCGGTTAGTATCATATGTAACAATTTCATT ACCAAGTGCTCCTACG
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCGCGACATTCAACCGA ACCAAGTGCTCCTACG
H1-C80 α	H1-C80	TTTAACCAATAGGTTCAAATATATTAGCGATAGCTTAGA ACCAAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTACCGAAGCC ACCAAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAACTG ACCAAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATATAACAAAATTACAAACAA ACCAAGTGCTCCTACG

H4-C37 α	H4-C37	CAGGTCAAGACGATTGCCGGCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGGTTGCACGTAACAAACTATCAACAATAGAT ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTTAGCAATAAGAGTCAATAGTATCGCAAGACAAAGA ACCAGTGCTCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCAGAAGCGCATTAGACATAGCAGCACCGTA TCTCTACCGCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGCCTAATTGCCAGTCACCGGAACCGA TCTCTACCGCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCCTTTAGCGAACAGGAGTTAGACTTCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAGAACACGCGCTGTTAGAAATAAGAA TCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAAACGCCAACATGCGCAGAGCGA TCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAATAAACACATAAAATCAATATATTCTCTACCGCCTACG
H1-C11 β	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCAATT TCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATCATATT TCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAATTATCAAATCAT TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATATAAT TCTCTACCGCCTACG
B_H2-C5	H2-C5	TATACCAGTCAGGAGTATCGGTTATCAATATAAGTATAGCCGGATTGCATCAA
B_H2-C14	H2-C14	TCATTCACTGAATAGAGTTAAAGGCCGCTGCCTATTCCGAATTGCTCCTTTGAT
B_H2-C23	H2-C23	TGACCAACTTGAAGGGTAAATACGTATCTCTGAATTACCAACAGTTGATTCCC
B_H2-C32	H2-C32	TTTGCCTATTGGGCTCTTTCACCAGTGTAAATAGATTAGAGCTCTACTAATAGTAG
B_H2-C41	H2-C41	CATAACGCCAAAAGTTGCTAAACAACTCCAATAGGAACCCACAATACTGCCGAAT
B_H2-C50	H2-C50	AGCCTGGGTGCCTATCGCAAATCCCCTAAAGCATCACTAAACTTTGCCG
B_H2-C59	H2-C59	GTCGACTCTAGAGGCAGGGCGATGCCCTAGCCCTAAACATGACAGTCAAATCAC
B_H2-C68	H2-C68	TCTTCGCTATTACGAACGTGGCGAGAACACACGACCAGTAAAGAGAATCGATGA
B_H2-C77	H2-C77	GTATCGGCTCAGGTATGGTGTCTGACTTGCTGGTAATATTGTTAAATTG
B_H5-C4	H4-C4	TTTGTACAATCAGGAATAACCCAAAAG
B_H5-C13 β	H4-C13	CGTCACCGACTTGAAGGCCAATAATAAGTCTCTACCGCCTACG
B_H5-C22 β	H4-C22	AATCAAGTTGCCTTTACAGAGAGAA TCTCTACCGCCTACG
B_H5-C31 β	H4-C31	CGCCTCCCTCAGAGTACCAACGCTAACGCTCTACCGCCTACG
B_H5-C40 β	H4-C40	ACAACTCGTATTAAAAGGCTTATCCGGTTCTCTACCGCCTACG
B_H5-C49 β	H4-C49	GATGGCAATTCTCGTCTTCTTATCAT TCTCTACCGCCTACG
B_H5-C58 β	H4-C58	GGTTAACGTCAGAGACAATAACAAACAT TCTCTACCGCCTACG
B_H5-C67 β	H4-C67	ACCTGAGCAAAGATAAGGGCTTAATTG A TCTCTACCGCCTACG
B_H5-C76 β	H4-C76	TTGCTTCTGTAATACCGACCGTGTAT TCTCTACCGCCTACG

H1 Replacement Oligos

A-Tube Name	Replace	Sequence
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCAGA ACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTCATTTCATCGTATTATCATTTGCG ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAAGACGATTGCCGGCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCCTTTAGCGAACAGGAGTTAGACTTCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATATT TCTCTACCGCCTACG
H4-C40 β	H4-C40	CGTCAATAGATAATACAACCTCGTATTAAAAGGCTTATCCGGT TCTCTACCGCCTACG
H4-C49 β	H4-C49	CTTGCTAACCTCAGATGGCAATTCTCGTCTTCTTATCAT TCTCTACCGCCTACG

H4-C43 Bio	H4-C43	CTAAAATATCTTAAAAGTTGAGTAACAGGAATCATTACCGTTTTTTTT/3Bio/
A_H3-C4	H3-C4	TTCTGTATGGGATTGAATTACGAGGCATGACTGGATAGCGTCTGTACCGTAACACT
A_H3-C13	H3-C13	AGGAGCCTTAATTCGTTGGAAAGAAAATAGTCAGAAGCAAACGGAATAGGTGTAT
A_H3-C22	H3-C22	CTGAGGCTTGCAGGAGGCTGCCCTGACGAGAGTACCTTAACCTATTATTCTGAA
A_H3-C31	H3-C31	AGTTCCATTAAACAGAGGACAGATGAAGTTCATTCCATATGTTCCAGTAAGCGT
A_H3-C40	H3-C40	AGTACAATGGTTTGCCAGGGCGGAGATAAGGTGGCATCAATCGTAATAGATAAT
A_H3-C49	H3-C49	ATGGTGGTCCGAAAATGAGTGAGCTAACATTATGACCTGCTGCTAACCTCA
A_H3-C58	H3-C58	GAAAAACCGTCTATATCCCCGGTACCGTGAGAAAGGCCGACGCCATTAAAATA
A_H3-C67	H3-C67	GGGGAAAGCCGGCGCCAGCTGGCAGAAGAGTCTGGAGCAAACTAAAAGGGACATT
A_H3-C76	H3-C76	TACAGGGCGCGTACAAGATCGCACTCCAGTAAACGTTAATATCCAGAACAAATTAA
A_H6-C5	H1-C5	AACGCAATAATAACATAGAAAATTATA
A_H6-C14	H1-C14	CAAGAATTGAGTTAGCCATTGGAAATT
A_H6-C23	H1-C23	TGAAAATAGCAGCCTAGCGTCAGACTG
A_H6-C32	H1-C32	TTTATCCTGAATCTCCGCCACCCCTCAGA
A_H6-C41	H1-C41	CAAATCAGATATAGATCCTTGCCGAA
A_H6-C50	H1-C50	ATCAATAATCGGCTAATATAATCCTGAT
A_H6-C59	H1-C59	TCTGTCCAGACGACTGAATATAACAGTAA
A_H6-C68	H1-C68	CCAACGCTAACAGAGATGATGAAACAA
A_H6-C77	H1-C77	TAATGGTTGAAATCGTCGCTATTAAATT

B-Tube		
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACAGA ACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTCATCGTATTATCATTGGCG ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAGACGATTGCCGCCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG
H1-C38 β	H1-C38	TATATTTCATTTGAGGCGTTTAGCGAACAGGGAGTTAGACTTCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAGCTAAATATTAAACCAAGTACATTATCATCATATT TCTCTACCGCCTACG
H4-C43 Bio	H4-C43	CTAAAATATCTTAAAAGTTGAGTAACAGGAATCATTACCGTTTTTTTT/3Bio/
B_H2-C5	H2-C5	CATAACGCCAAAAGTTGCTAACAACTCCAATAGGAACCCACAATACTGCCGAAT
B_H2-C14	H2-C14	TATACCAGTCAGGAGTATCGGTTATCAATATAAGTATAGCCGGGATTGCATCAA
B_H2-C23	H2-C23	TCATTCACTGAATAGAGTTAAAGGCCGCTGCCTATTCCGAATTGCTCCTTTGAT
B_H2-C32	H2-C32	TGACCAACTTGAAGGGTAAACACGTATCTCTGAATTACCAACAGTTGATTCCC
B_H2-C41	H2-C41	TTTGCCTATTGGGCTCTTTCACCAAGTGAATAGATTAGAGCTCTACTAATAGTAG
B_H2-C50	H2-C50	AGCCTGGGTGCCTATCGCAAAATCCATCTAAAGCATCACTAAACTTTCGG
B_H2-C59	H2-C59	GTCGACTCTAGAGGCAGGGCGATGCCCTAGCCCTAAACATGACAGTCAC
B_H2-C68	H2-C68	TCTTCGCTATTACGAACGTGGCGAGAAACACACGACCAGTAAAGAGAATCGATGA
B_H2-C77	H2-C77	GTATCGGCTCAGGTATGGTGTCTTGAATTGCTGGTAATATTGTTAAAATTG
B_H5-C40 β	H4-C40	ACAACCTCGTATTAAAGGCTTATCGGTTCTACCGCCTACG
B_H5-C49 β	H4-C49	GATGGCAATTCACTCGTCTTCCTTATCATCTACCGCCTACG
B_H5-C4	H4-C4	TTTGTACAATCAGGAATACCCAAAAG
B_H5-C13	H4-C13	CGTCACCGACTTGAAGCCCAATAATAAG
B_H5-C22	H4-C22	AATCAAGTTGCCTTTACAGAGAGAAAT
B_H5-C31	H4-C31	CGCCTCCCTCAGAGTACCAACGCTAACG
B_H5-C58	H4-C58	GGTTAACGTCAAGAGACAATAACAAACA
B_H5-C67	H4-C67	ACCTGAGCAAAGATAGGGCTTAATTGA
B_H5-C76	H4-C76	TTGCTTCTGTAAATACCGACCGTGTGAT

H2 Replacement Oligos

A-Tube		Sequence
Name	Replace	
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATAGCAAG ACCAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCCAATAAATAGCCCCTTATT ACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCAGA ACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAGAACAGAATTTCATCGTATTATCATTTCG ACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTTCAGAACAAAAATAATATCCAGGGTTAGAACCTA ACCAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTAAATGCAGTAATAAGAGAATAACGGATTGCCTG ACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCCGGTTAGTATCATATGTAACAATTTCATT ACCAGTGCTCCTACG
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCGCGACATTCAACCGA ACCAGTGCTCCTACG
H1-C80 α	H1-C80	TTTTAACCAATAGGTTCAAATATATTAGCGATAGCTTAGA ACCAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAAGCC ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAACTG ACCAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATAATCAAATTACAAAATAACA ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTACAGACGATTGCCGCCAGATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACCAGAAGGAGCGGACGCACCATCGAGA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGTTGCACGTAAAATCAACAATAGAT ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCAATAGTGATCGCAAGACAAAGA ACCAGTGCTCCTACG
H1-C11 β	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCAATTAA TCTCTACCGCCTACG
H1-C20 β	H1-C20	GGAAGCCTAACTCCAGAACCGCATTAGACATAGCAGCACCGT TCTCTACCGCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCTAATTGCCAGTCACCGAACCGA TCTCTACCGCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCCTTACGAAACAGGAGTTAGACTTCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATCATATT TCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAAAGAACCGCCTGTTAGAAATAAGAAATT TCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAAACGCCAACATCGCAGAGGCGAAT TCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAAATAACACATAAAATCAATATATTCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCAAATTATCAAATCA TCTCTACCGCCTACG
H4-C13 β	H4-C13	CGGAATAGGTGTACGTACCGACTGAAAGCCAATAAG TCTCTACCGCCTACG
H4-C22 β	H4-C22	CCTATTATTCTGAAAATCAAGTTGCCCTTACAGAGAGAAATTCTCTACCGCCTACG
H4-C31 β	H4-C31	GTTCCAGTAAGCGTCGCCCTCAGAGTACCAACGCTAACG TCTCTACCGCCTACG
H4-C40 β	H4-C40	CGTCAATAGATAATACAACACTGTATTAAAAGGCTTATCCGGT TCTCTACCGCCTACG
H4-C49 β	H4-C49	CTTGCTAACCTCAGATGGCAATTACATCGTCTTCCATTATCA TCTCTACCGCCTACG
H4-C58 β	H4-C58	CGCCATTAAAAATAGGTTAACGTCAAGAGACAATAACA TCTCTACCGCCTACG
H4-C67 β	H4-C67	TAAAAGGGACATTACCTGAGCAAAAGATAGGGCTTAATTGA TCTCTACCGCCTACG
H4-C76 β	H4-C76	CCAGAACAAATTATTGCTCTGTAAATACCGACCGTGTGATT TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATAA TCTCTACCGCCTACG
H4-C43 Bio	H4-C43	CTAAAATATCTTAAAGTTGAGTAACAGGAATCATTACCGTTTTTTT/3Bio/
A_H3-C4	H3-C4	TTCTGTATGGGATTGAATTACGAGGCATGACTGGATAGCGTCTGTACCGTAACACT
A_H3-C13	H3-C13	AGGAGCCTTAAATCGTTGGAGAAAAATAGTCAGAACGAAACGGAATAGGTGTAT
A_H3-C22	H3-C22	CTGAGGCTTGCAGGAGGCTIGCCCTGACGAGAGTACCTTAAACCTATTATTCTGAA
A_H3-C31	H3-C31	AGTTCCATTAAACAGAGGACAGATGAAGTTCATCCATATGTTCCAGTAAGCGT
A_H3-C40	H3-C40	AGTACAATGGTTTGCAGGGCGGAGATAAGGTGGCATCAATCGCAATAGATAAT

A_H3-C49	H3-C49	ATGGTGGTCCGAAAATGAGTGAGCTAACATTATGACCTGCTGCTGAACCTA
A_H3-C58	H3-C58	GAAAAACCGTCTATATCCCCGGTACCGTGAGAAAGGCCGACGCCATTAATA
A_H3-C67	H3-C67	GGGGAAAGCCGGCGCCAGCTGGCGAAAGAGTCTGGAGCAAACATAAAGGGACATT
A_H3-C76	H3-C76	TACAGGGCGCGTACAAGATCGCACTCCAGTAAACGTTAATATCCAGAACAAATATTA
A_H6-C5	H1-C5	AACGCAATAATAACATAGAAAATTATA
A_H6-C14	H1-C14	CAAGAATTGAGTTAGCCATTGGATT
A_H6-C23	H1-C23	TGAAAATAGCAGCCTTAGCGTCAGACTG
A_H6-C32	H1-C32	TTTATCCTGAATCTCCGCCACCCCTCAGA
A_H6-C41	H1-C41	CAAATCAGATATAAGATCCTTGCCCAGA
A_H6-C50	H1-C50	ATCAATAATCGGCTAATATAATCCTGAT
A_H6-C59	H1-C59	TCTGTCCAGACGACTGAATATACAGTAA
A_H6-C68	H1-C68	CCAACGCTCAACAGAGATGATGAAACAA
A_H6-C77	H1-C77	TAATGGTTGAAATCGTCGCTATTAAATT
B-Tube		
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATTAGCAAGACCAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCCAAATAAATAGCCCCCTTATTACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGCCGCCACCAAGAACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTATTTCATCGTATTATCATTTGCGACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTAGAACAAAATAATATCCCAGGGTTAGAACCTAACAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTAATGCAGTAATAAGAGAATAACGGATTGCCTGACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCGGTTAGTATCATATGTAACAATTTCATTACCAGTGCTCCTACG
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCGCGACATTCAACCGAACAGTGCTCCTACG
H1-C80 α	H1-C80	TTTAAACCAATAGGTTCAAATATATTAGCGATAGCTTAGAACAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAACGCCACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAACTGACCAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCTAAATCAAATTACAAAATAAACACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTTCAGACGATTGCCGCCAGCATTGACCTCCGACTTGCACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATACCAGAACCGGAGCGCACCATCGAGAACAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGGTGGCGGTTGCACGTAACACTATCAACAAATAGATACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAAATCGTAATTAGGCAGAACAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATTACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCATAGTGTACGCAAGACAAAGAACAGTGCTCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCAGAACCGCATTAGACATAGCAGCACCGTAACAGTGCTCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCTAATTGCCAGTCACCGAACCGAACAGTGCTCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCCTTGTAGCGAACAGGGATTAGACTTCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAAGAACCGCCTGTTAGAAATAAGAAATTCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACACGCCAACATCGCAGAGGCCAACAGTGCTCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAATAAACACACATAATCAATATTCTCTACCGCCTACG
H1-C11 β	H1-C11	AAAAATCAGGTCTAAATAGCAATAGCTAAATTATTCAATTCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATCATATTCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAAATTATCAAAATCATCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGTTATAATACTCTACCGCCTACG
H4-C43 Bio	H4-C43	CTAAAATATCTTAAAAGTTGAGTAACAGGAATCATTACCGTTTTTTTT/3Bio/
B_H2-C5	H2-C5	CATAACGCCAAAAGTTGCTAACACAACCTCCAATAGGAACCCACAATACTGCGGAAT
B_H2-C14	H2-C14	TATACCAGTCAGGAGTATCGGTTATCAATATAAGTATAGCCGGATTGCATCAA

B_H2-C23	H2-C23	TCATTCA GTGAATAGAGTTAAAGGCCGCTGCCATTTCGGATTGCTCCTTTGAT
B_H2-C32	H2-C32	TGACCA ACTTGAAGGGAAAATACGTATCTGAATTACCAACAGTGATTCCC
B_H2-C41	H2-C41	TTTGC GTATTGGGCTTTACCCAGTGAATAGATTAGAGCTCTACTAATAGTAG
B_H2-C50	H2-C50	AGCCTGGGT GCCATCGCAAATCCCCTAAAGCATCACTAATACCTTGC
B_H2-C59	H2-C59	GTCG ACTCTAGAGGCAGGGGATGCCCTAGCCCTAAACATGACAGTC
B_H2-C68	H2-C68	TCTTC GCTATTACGAACGTGGCGAGAACACGACCAGTAAAGAGAATCGATGA
B_H2-C77	H2-C77	GTAT CGGCTCAGGTATGGTGCCTTGACTTGCTGGTAATATTTGTTAAATT
B_H5-C4	H4-C4	CGT TTCACAATCAGGAATACCCAAAAG
B_H5-C13 β	H4-C13	CGTC ACCGACTTGAAGCCCATAATAAGTCTTACCGCCTACG
B_H5-C22 β	H4-C22	AAT CAAGTTGCCCTTACAGAGAGAATTCTCTACCGCCTACG
B_H5-C31 β	H4-C31	CGC CTCCCTCAGAGTACCAACGCTAACGCTTACCGCCTACG
B_H5-C40 β	H4-C40	ACA ACTCGTATTAAAAGGCTTATCCGGTTCTACCGCCTACG
B_H5-C49 β	H4-C49	GAT GGCAATTACATCGTCTTCCTTATCATCTTACCGCCTACG
B_H5-C58 β	H4-C58	GGTT AACGTCAAGAGACAATAAACAAACATCTTACCGCCTACG
B_H5-C67 β	H4-C67	ACCT GAGAAAAGATAGGGCTTAATTGATCTTACCGCCTACG
B_H5-C76 β	H4-C76	TTG CTCTGTAAATACCGACCGTGTGATTCTTACCGCCTACG

H3 Replacement Oligos**A-Tube**

Name	Replace	Sequence
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCAATTAGCAAG ACCAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCAAATAAGCCCCCTTATT ACCAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATACATCCTTAAATCAAGATGAGGCCACCAGA ACCAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTCATCGTATTATCATTGCG ACCAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTAGAACAAAATAATCCCAGGGTTAGAACCTA ACCAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTAAATGCAGTAATAAGAGAATAACGGATTGCCTG ACCAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCCGGTTAGTATCATATGTAACAATTTCATT ACCAGTGCTCCTACG
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCGACATTCAACCGA ACCAGTGCTCCTACG
H1-C80 α	H1-C80	TTTTAACCAATAGGTTCAAATATATTAGCGATAGCTTAGA ACCAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTTACCGAAGCC ACCAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGAGAATTAACTG ACCAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGATAAAAGTTCTAAATCAAATTACAAAATAACA ACCAGTGCTCCTACG
H4-C37 α	H4-C37	CAGGTCAAGACGATTGCCGCCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATCACAGAAGGAGCGGACGCACTCATGAGA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGTTGCACGTAACAAACTATCAACATAGAT ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGAAATACCTAATGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTGTAGCAATAAGAGTCATAGTATCGCAAGACAAAGA ACCAGTGCTCCTACG
H1-C11 β	H1-C11	AAAAATCAGGTCTTAAATAGCAATAGCTAAATTATTCAATT TCTCTACCGCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCAGAACGCCATTAGACATAGCAGCACCGTA TCTCTACCGCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCTAATTGCCAGTCACCGGAACCAGAT TCTCTACCGCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCCTTACCGAACAGGGAGTTAGACT TCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATAACCAAGTACATTATCATATT TCTCTACCGCCTACG
H1-C56 β	H1-C56	GTAATGTAGGTAAAGAACGCCCTGTTAGAAATAAGAAAT TCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAAACGCCAACATGCGCAGAGGCCAAT TCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAATAAACACATAAAAT TCTCTACCGCCTACG

H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAATTATCAAAATCA TCTCTACCGCCTACG
H4-C13 β	H4-C13	CGGAATAGGTGTATCGTACCGACTGAAGCCAAATAAG TCTCTACCGCCTACG
H4-C22 β	H4-C22	CCTATTATTCTGAAAATCAAGTTGCCTTTACAGAGAGAATTCTCTACCGCCTACG
H4-C31 β	H4-C31	GTTCCAGTAAGCGTCGCCCTCAGAGTACCAACGCTAACGCTCTACCGCCTACG
H4-C40 β	H4-C40	CGTCAATAGATAATACAACCGTATTAAAAGGCTTATCCGGT TCTCTACCGCCTACG
H4-C49 β	H4-C49	CTTGCTAACCTCAGATGGCAATTATCGTCTTCCTTATCA TCTCTACCGCCTACG
H4-C58 β	H4-C58	CGCCATTAAAAATAGGTTAACGTACAGAGACAATAAACACA TCTCTACCGCCTACG
H4-C67 β	H4-C67	TAAAAGGGACATTCACCTGAGCAAAAGATAGGGCTTAATTGA TCTCTACCGCCTACG
H4-C76 β	H4-C76	CCAGAACAAATTATTGCTTCTGAAATACCGACCCTGTGATT TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATATAA TCTCTACCGCCTACG
H4-C16 Bio	H4-C16	AGGC GGATAAGTGCCACCACTAGCACCATGAGCGCTAACATCTTTTTTTT/3Bio/
H4-C34 Bio	H4-C34	CTCATTAAAGCCAGAGCCACCACCCCTCATAGTTGCTATTTGTTTTTTT/3Bio/
H4-C52 Bio	H4-C52	CACGCTGAGAGCCATTCTGAATAATGAAATCTAACCTAAACGATTTTTTTT/3Bio/
H4-C70 Bio	H4-C70	GATTATTACATTGAAATTAAATTACATTGTTACAAATTCTTTTTTT/3Bio/
A_H3-C4	H3-C4	TTCTGTATGGGATTGAATTACGAGGCATGACTGGATAGCGTCTGTACCGTAACACT
A_H3-C13	H3-C13	AGGAGCCTTAAATCGTTGGGAAAGAAAATAGTCAGAACGAAACGGAATAGGTGTAT
A_H3-C22	H3-C22	CTGAGGCTTGCAGGAGGCTTGCCTGACGAGAGTACCTTAAACCTATTATCTGAA
A_H3-C31	H3-C31	AGTTCCATTAAACAGAGGACAGATGAAGTTCATCCATATGTTCCAGTAAGCGT
A_H3-C40	H3-C40	AGTACAATGGTTTGCAGGGCGGAGATAAGGTGGCATCAATCGTAATAGATAAT
A_H3-C49	H3-C49	ATGGTGGTCCGAAAATGAGTGAGCTAACATTATGACCTGCTGCTGAACCTCA
A_H3-C58	H3-C58	GAAAAACCGTCTATATCCCCGGTACCGTGAGAAAGGCCGACGCCATTAAAATA
A_H3-C67	H3-C67	GGGGAAAGCCGGCGCCAGCTGGCGAAAGAGTCTGGAGGCAAACAAAAGGACATT
A_H3-C76	H3-C76	TACAGGGCGCGTACAAGATCGCACTCCAGTAAACGTTAATATCCAGAACATATTA
A_H6-C5	H1-C5	AACGCAATAATAACATAGAAAATTATA
A_H6-C14	H1-C14	CAAGAATTGAGTTAGCCATTGGGAAATT
A_H6-C23	H1-C23	TGAAAATAGCAGCCTTAGCGTCAGACTG
A_H6-C32	H1-C32	TTTATCCTGAATCTCGCCACCCCTCAGA
A_H6-C41	H1-C41	CAAATCAGATATAGATCCTTGCCCGAA
A_H6-C50	H1-C50	ATCAATAATGGCTAATATAATCCTGAT
A_H6-C59	H1-C59	TCTGTCCAGACGACTGAATATACAGTAA
A_H6-C68	H1-C68	CCAACGCTAACAGAGATGATGAAACAA
A_H6-C77	H1-C77	TAATGGTTGAAATCGTCGCTATTAATT
B-Tube		
H1-C17 α	H1-C17	CAAATATCGCGTTAGTCAGAGGGTAATTACCATAGCAAG ACCAAGTGCTCCTACG
H1-C26 α	H1-C26	GCTTAATTGCTGAACCCAATCCAATAAATAGCCCCCTATT ACCAAGTGCTCCTACG
H1-C35 α	H1-C35	ACCATTAGATAACATCCTTAAATCAAGATGAGCCGCCACCAGA ACCAAGTGCTCCTACG
H1-C44 α	H1-C44	GCAAGGCAAAGAATTTCATTTCATCGTATTATCATTGCG ACCAAGTGCTCCTACG
H1-C53 α	H1-C53	AAAATTTCAGAACAAAATAATATCCCAGGGTTAGAACCTA ACCAAGTGCTCCTACG
H1-C62 α	H1-C62	TGATAAAATTATGCAGTAATAAGAGAATAACGGATTGCCTG ACCAAGTGCTCCTACG
H1-C71 α	H1-C71	CATATGTACCCGGTTAGTATCATATGAAACATTTCATT ACCAAGTGCTCCTACG
H1-C8 α	H1-C8	AAATGCTTAAACATAAGCAGATAGCCGCGACATTCAACCGA ACCAAGTGCTCCTACG
H1-C80 α	H1-C80	TTTAACCAATAGGTTCAAATATATTAGCGATAGCTTAGA ACCAAGTGCTCCTACG
H4-C10 α	H4-C10	ACCCTCAGAACCGCGTAAATATTGACGGATCTACCGAAGCC ACCAAGTGCTCCTACG
H4-C19 α	H4-C19	CTCAAGAGAAGGATCAATGAAACCATCGGGGAGAATTAAACTG ACCAAGTGCTCCTACG
H4-C28 α	H4-C28	GTACTGGTAATAAGTTCATACAAATTACAAAATAACA ACCAAGTGCTCCTACG

H4-C37 α	H4-C37	CAGGTCAAGACGATTGCCGCCAGCATTGACCTCCGACTTGC ACCAGTGCTCCTACG
H4-C46 α	H4-C46	CAGTTGGCAAATACCAGAAGGAGCGGACGCACTCATCGAGA ACCAGTGCTCCTACG
H4-C55 α	H4-C55	CAGAGGTGAGGCCGGTTGCACGTAACAAACTATCAACAATAGAT ACCAGTGCTCCTACG
H4-C64 α	H4-C64	CTGAAAGCGTAAGACAAGTTACAAATCGTAATTAGGCAGA ACCAGTGCTCCTACG
H4-C73 α	H4-C73	TCATGGAAATACCTAATGGAAACAGTACCGGAATCATAATT ACCAGTGCTCCTACG
H4-C82 α	H4-C82	CCGTTTAGCAATAAGAGTCAATAGTATCGCAAGACAAAGA ACCAGTGCTCCTACG
H1-C20 β	H1-C20	GGAAGCAAACCTCAGAAGCGCATTAGACATAGCAGCACCGTA TCTCTACCGCCTACG
H1-C29 β	H1-C29	ATATGCAACTAAAGGCCATTGCCCCAGTCACCGGAACCGA TCTCTACCGCCTACG
H1-C38 β	H1-C38	TATATTTCATTGAGGCAGTTAGCGAACAGGAGTTAGACTTCT TACCGCCTACG
H1-C56 β	H1-C56	GTAATGTGTAGGTAGAACACGCGCCTGTTAGAAATAAGAA ATTCTCTACCGCCTACG
H1-C65 β	H1-C65	TACAAAGGCTATCAAACAAACGCCAACATGCGCAGAGCGA ATTCTCTACCGCCTACG
H1-C74 β	H1-C74	GAAGATTGTATAAGATAAGAATAAACACATAAAATCAAT ATTCTCTACCGCCTACG
H1-C11 β	H1-C11	AAAAATCAGGTCTAAATAGCAATAGCTAAATTATTCAATT TCTCTACCGCCTACG
H1-C47 β	H1-C47	GCATAAAAGCTAAATATTAAACCAAGTACATTATCATCATT TCTCTACCGCCTACG
H1-C83 β	H1-C83	CCTTCCTGTAGCCATGATGCAAATCCAATTATCAAAATCAT TCTCTACCGCCTACG
H4-C85 β	H4-C85	TGAGGCCACCGAGTTACCTTTAACCTGTTGGGTTATATAA TCTCTACCGCCTACG
H4-C16 Bio	H4-C16	AGGCAGATAAGTGCACCAAGTAGCACCATGAGCGCTAATAT CTTTTTTTTT/3Bio/
H4-C34 Bio	H4-C34	CTCATTAAAGCCAGAGCCACCACCCCTCATAGTTGCTATT TTTTTTTTTT/3Bio/
H4-C52 Bio	H4-C52	CACGCTGAGAGCCATTCTGAATAATGGAATCTAAATTACGAT TTTTTTTTTT/3Bio/
H4-C70 Bio	H4-C70	GATTATTACATTGAAATTAAATTACATTGTTATACAAATT CTTTTTTTTT/3Bio/
B_H2-C5	H2-C5	CATAACGCCAAAAGTTGCTAAACAACTCCAATAGGAACCCACAATACTGCCGAAT
B_H2-C14	H2-C14	TATACCAGTCAGGAGTATCGGTTATCAATATAAGTATAGCCGGATTGCATCAA
B_H2-C23	H2-C23	TCATTCACTGAATAGAGTTAAAGGCCGCTGCCTATTCCGAATTGCTCCTTTGAT
B_H2-C32	H2-C32	TGACCAACTTGAAGGGTAAAATACGTATCTGAATTACCAACAGTTGATTCCC
B_H2-C41	H2-C41	TTTGCCTATTGGGCTTTACCAAGTGTAAATAGATTAGAGCTACTAATAGTAG
B_H2-C50	H2-C50	AGCCTGGGTGCCTATCGCAAATCCCCTAAAGCATCACTAATACTTTGCCG
B_H2-C59	H2-C59	GTCGACTCTAGAGGCAGGGCGATGCCCTAGCCCTAAACATGACAGTCAAATCAC
B_H2-C68	H2-C68	TCTTCGCTATTACGAACGTGGCGAGAACACAGCACCAGTAAAAGAGAATCGATGA
B_H2-C77	H2-C77	GTATCGGCTCAGGTATGGTTGCTTGAATTGCTGGTAATTGTTAAAATTG
B_H5-C4	H4-C4	TTTGTACAATCAGGAATACCCAAAAG
B_H5-C13 β	H4-C13	CGTCACCGACTTGAAGCCAAATAATAAGTCTCTACCGCCTACG
B_H5-C22 β	H4-C22	AATCAAGTTGCCATTACAGAGAGAATTCTCTACCGCCTACG
B_H5-C31 β	H4-C31	CGCCTCCCTCAGAGTACCAACGCTAACGCTCTACCGCCTACG
B_H5-C40 β	H4-C40	ACAACCTGTATTAAAAGGCTTATCGGTTCTCTACCGCCTACG
B_H5-C49 β	H4-C49	GATGGCAATTACCGTCTTCTTATCATCTCTACCGCCTACG
B_H5-C58 β	H4-C58	GGTTAACGTCAGAGACAATAACACATCTCTACCGCCTACG
B_H5-C67 β	H4-C67	ACCTGAGCAAAGATAGGGCTTAATTGATCTCTACCGCCTACG
B_H5-C76 β	H4-C76	TTGCTTCTGTAAATACCGACCGTGTGATTCTCTACCGCCTACG

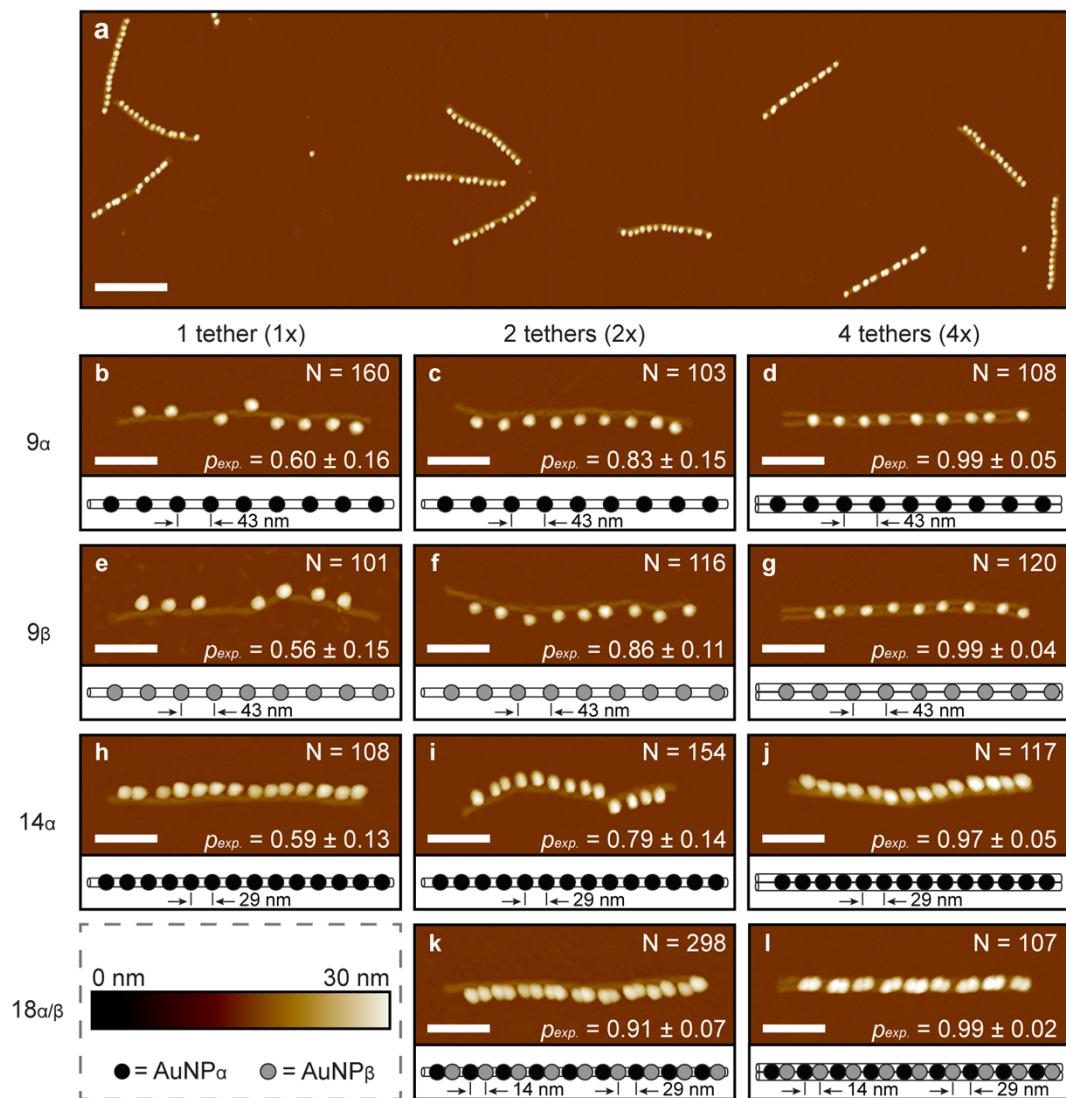
S2 – AFM HEIGHT IMAGES OF FIGURE 3

Figure S2: AFM height images of DNA nanostructures functionalized with gold nanoparticles (AuNPs). The scale bars are 300 nm for (a) and 100 nm for (b - l). (a) Low-resolution AFM height image of 4x14 α nanostructures. (b-l) High-resolution AFM height images of DNA nanostructures with corresponding nanoparticle schematics. DNA nanostructure designs included: (b) 1x9 α , (c) 2x9 α , (d) 4x9 α , (e) 1x9 β , (f) 2x9 β , (g) 4x9 β , (h) 1x14 α , (i) 2x14 α , (j) 4x14 α , (k) 2x18 α/β , and (l) 4x18 α/β . The columns from left to right represent increasing numbers of tethers per binding site including one tether (1x), two tethers (2x), and four tethers (4x). The rows from top to bottom represent increasing numbers of binding sites including 9 sites, 14 sites, and 18 sites for α (5'-ACCAAGTGCTCCTACG-3') and/or β (5'-TCTCTACCGCCTACG-3') tethers. N is the total number of nanostructures counted to determine the average probability of AuNP site-occupation, $p_{exp.}$. The designed diameter and length of the 1x and 2x nanostructures was 6 nm and 412 nm, respectively. The designed width, height, and length of the cross-linked, 4x nanostructure were 12 nm, 6 nm, and 412 nm, respectively.

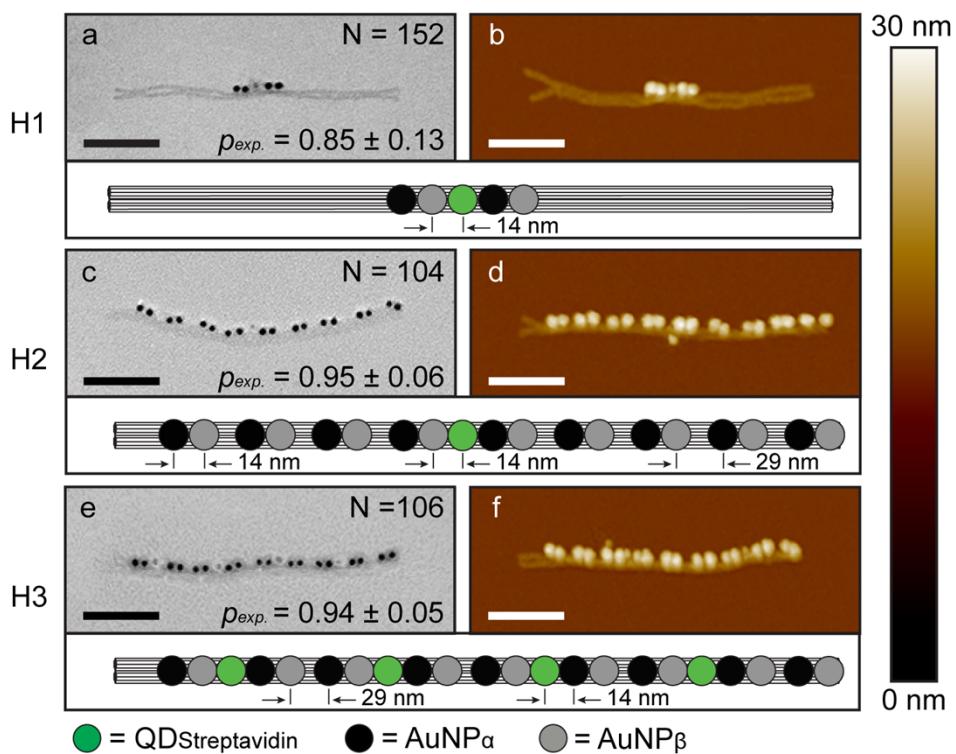
S2 – AFM HEIGHT IMAGES OF FIGURE 7

Figure S3: TEM (a, c, e) and AFM (b, d, f) height images of H1, H2, and H3 heterostructures, including schematics for each design. The images show successful site-specific attachment of AuNPs and QDs. The number of samples evaluated, N, and the experimentally measured average probability of overall site-occupation, p_{exp} , are shown in the upper and lower right-hand corners for the TEM images, respectively. The EDS analysis of the QD and the AuNP for H1, shown in Support Information S3, confirmed that the nanoparticle attached in the middle of H1 is a QD and the two nanoparticles on either side of the QD are AuNPs. Based on the QD and AuNP mass contrast difference in TEM, successful synthesis of H2 and H3 is also implied. The scale bars are 100 nm.

S3 – EDS LINE-SCAN OF H1

Due to the small interparticle distance, an extremely small probe size was required for EDS analysis. A TEM with a field emission gun (Tecnai TF30-FEG STwin STEM) was utilized at the Center for Advanced Energy Studies (CAES) in Idaho. Images of the sample were taken in scanning transmission electron microscopy (STEM) mode at 300 kV and the EDS line-scan across the AuNPs and the QDs were performed (Figure S4). The EDS spectra of AuNPs and QDs clearly showed the elemental difference between two nanoparticles. Although the QDs consisted of CdSe cores and ZnS shells, the EDS spectrum only showed the distinct sulfur peak. The peaks of the other elements did not appear in the spectrum because of the low concentration of the elements. In addition, some of their peaks overlapped with the background signals. An EDS line-scan of H1 with a missing QD was also performed for comparison. The EDS spectrum of the empty QD site showed only the background signal, which confirms that the sulfur peak was generated by the QD. In addition, uranium was detected because of the staining.

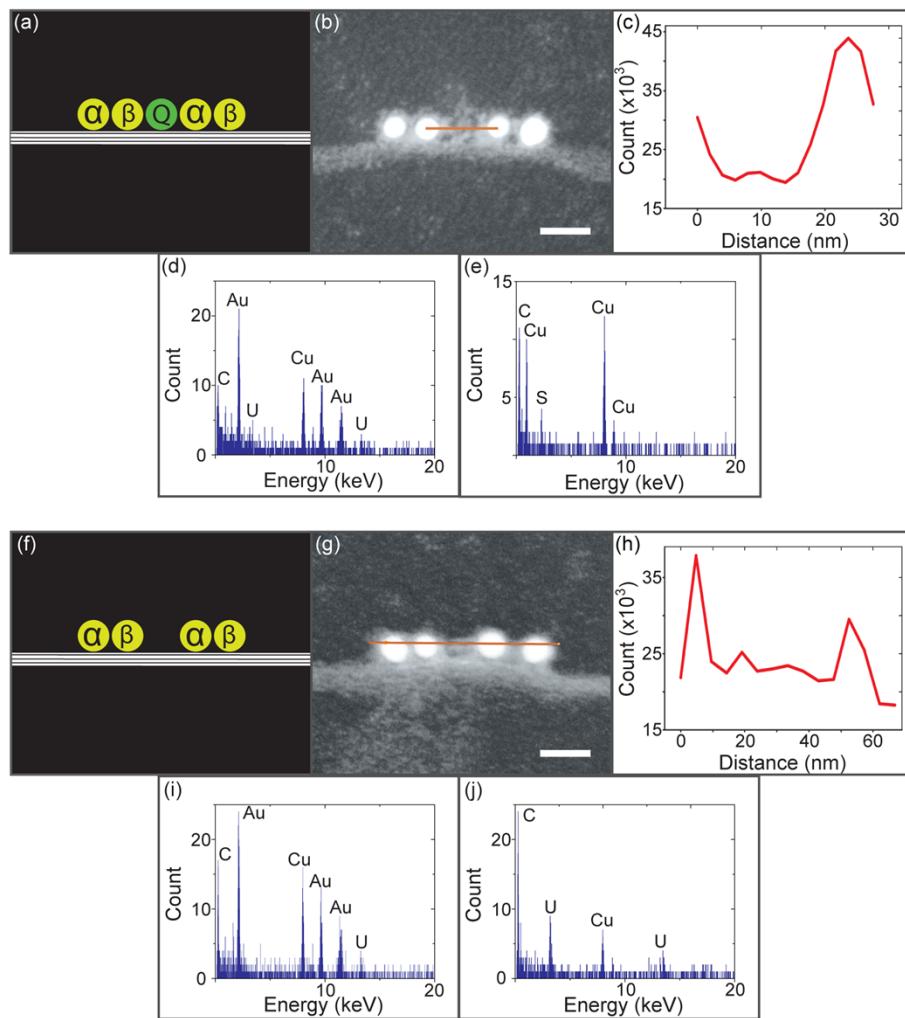


Figure S4 (a) A schematic of H1 with complete nanoparticle attachment, (b) STEM image of the H1 nanostructure with complete nanoparticle attachment, (c) counts vs. distance plot along the EDS line scan shown as orange line in (b), (d) EDS spectrum of the AuNP attached to the nanostructure, (e) EDS spectrum of QD attached to the nanostructure, (f) schematic of H1 nanostructure with a missing QD, (g) STEM image of the H1 nanostructure with a missing QD, (h) counts vs. distance plot along the EDS line scan shown as orange line in (g), (i) EDS spectrum of the AuNP attached to the nanostructure, (j) EDS spectrum at the empty QD binding site. The EDS spectrums of the nanoparticles, (d), (e), and (i), clearly show that they are indeed AuNP and QD. Although the QD consists of CdSe core and ZnS shell, the spectrum only shows the distinct sulfur peak because of the low concentration of the other elements. Also, some of their peaks overlap with the background signal. The EDS spectrum of the empty QD binding site did not show the sulfur peak seen in (e), which confirms that the sulfur peak came from the attached QD. In addition, uranium peaks were observed because of the staining of the sample. The scale bars are 20 nm.

S4 – ADDITIONAL AFM IMAGES OF THE H2 NANOSTRUCTURE

Several AFM images of the H2 heterostructure were acquired using a Bruker MultiMode 8 with a Nanoscope V controller. High-resolution AFM images clearly show QD attachment between the fourth and the fifth AuNP pairs, which agree with the H2 heterostructure design (Figure S5). The QDs for select heterostructures were offset toward either side of the nanostructure axis because of steric hindrance between the QD and neighboring AuNPs. Because there were 10-thymine spacers on the biotin tethers, the QDs were anticipated to have a high degree of freedom after attaching to the nanostructure.

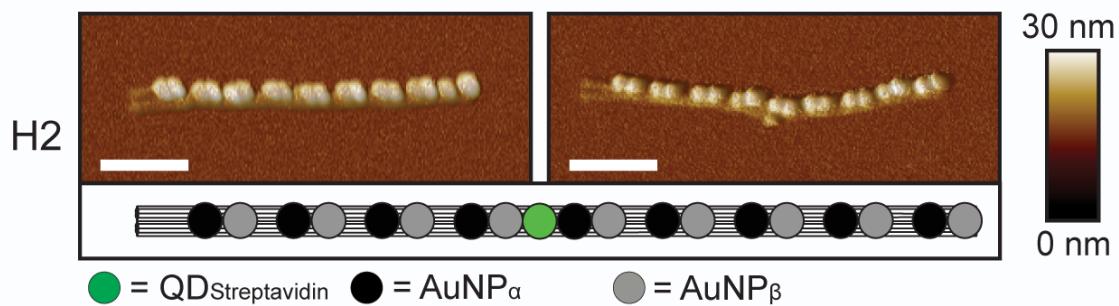


Figure S5: High-resolution AFM images and the schematic of the H2 heterostructure. Both images clearly show QD placement between the fourth and the fifth AuNP pairs, which is consistent with the heterostructure design shown as the schematic. The size of the scale bars is 100 nm.

S5 – DNA NANOSTRUCTURE SYNTHESIS

1x & 2x NANOSTRUCTURES: The 1x and 2x nanostructures were synthesized by mixing 10 nM of DNA from the M13mp18 bacteriophage (New England Biolabs) with 100 μ M of staple strands (IDT) in a 1:10 molar ratio. 10x TAE (40 mM tris, 20 mM acetic acid, 2 mM ethylenediaminetetraacetic acid (EDTA); pH 8.0), and 1 M MgCl₂ were added to the mixture to bring the final buffer concentration to 1x TAE, 14mM MgCl₂. The solution was heated at 90 °C for 20 min and then cooled to 20 °C at 0.6 °C/min via a thermocycler (Eppendorf Mastercycler Nexus Gradient). DNA nanotubes were purified using gel electrophoresis (0.7% agarose gel, 40 V, 35 mA, 180 min). The buffer used for casting and running gels was 0.5x TBE, 12 mM MgCl₂ (89 mM tris-borate, 2 mM EDTA, and 12 mM MgCl₂; pH 8.3). Gels were stained with 1x SYBR Gold (Life Technologies) / 0.5x TBE, 12 mM MgCl₂ solution for 30 min and then placed on a UV light table (Hoefer MACROVUE UVIS-20) to extract bands using a razor blade. Bands were cut into sub millimeter pieces and then loaded into Freeze N' Squeeze microcentrifuge filters (Bio-Rad). Filters were spun at 4800 rcf at 4 °C for 10 min using a centrifuge (Eppendorf 5430R).

4x NANOSTRUCTURES: 4x nanostructures were cross-linked in a parallel orientation by hybridizing two 2x nanostructures. Individual 2x nanostructures were synthesized and purified using the procedure described above and mixed in an equal molar volume. 0.5x TBE 200 mM MgCl₂ buffer was added to the solution to increase the magnesium ion concentration to 40 mM. The mixture was heated to 45 °C for 2 hrs and then cooled to 20 °C at 0.6 °C/min. The yield was approximately 83%, and no purification was required.

S6 – AUNP AND DNA NANOSTRUCTURE FUNCTIONALIZATION

AUNP FUNCTIONALIZATION: A colloidal solution of 10 nm gold (Ted Pella) was mixed with bis(*p*-sulfonatophenyl)phenylphosphine dihydrate dispotassium salt (BSPP) from Sigma Aldrich in 1:748,000 molar ratio. The solution was wrapped with tin foil and shaken for 48 hours to saturate the surface with BSPP. A 5 M NaCl solution was titrated into the colloidal mixture, in 0.2 mL increments, until the gold colloids began to precipitate out of the solution. The mixture was then centrifuged at 3200 rcf for 30 min to acquire a gold colloidal pellet at the bottom of the container. The transparent supernatant was discarded. 400 μ L of 2.5 mM BSPP and 400 μ L of methanol were added to the gold colloidal pellet. The mixture was spun at 1,600 rcf for 30 min, and the supernatant was discarded. Then 200 μ L of 2.5 mM BSPP was added to the remaining solution. The concentration of the gold colloidal solution was calculated using Beer-Lambert law based on the absorbance measured at 490 nm via Eppendorf BioPhotometer Plus.

Tris(2-carboxyethyl) phosphine hydrochloride (TCEP) from Sigma Aldrich was then mixed with thiolated single stranded DNA (T-ssDNA) in a 99:1 ratio in a 0.5x TBE buffer solution. The mixture was agitated for 30 min. Both the concentrated gold colloidal solution and the T-ssDNA solution were mixed in 1:300 molar ratio (later this ratio was cut down to 1:150 molar ratio). The mixture was agitated for 3 days at room temperature. The excess T-ssDNA in the mixture was removed using 100 kDa Amicon Ultra microcentrifuge filter (Millipore). 100 μ L of AuNP solution and 400 μ L of 0.5x TBE buffer were mixed in the filter, and it was spun at 13,000 rcf for 6 min. After the initial spin, 480 μ L of 0.5x TBE buffer were added to the filter, and it was re-spun at 13,000 rcf for 6 min for rinsing. After repeating two additional rinsing processes, the functionalized AuNP solution was transferred into a new filter using a pipette. Four additional rinsing steps were done to assure excess oligos were removed from the solution. Inverting the Amicon filter and then spinning it at 1,000 rcf for 3 min recovered the AuNPs. The absorbance of the AuNP solution was then measured at 521 nm via an Agilent Cary 5000 UV-Vis-NIR to calculate the concentration.

1x & 2x DNA NANOSTRUCTURE FUNCTIONALIZATION: The functionalized AuNPs and the 1x and 2x nanostructures were mixed in a 1:5 binding site to AuNP ratio. The mixture was heated at 45 °C for 41 min then cooled to 20 °C at 0.6 °C/min. The functionalized 1x and 2x nanostructures were then purified by agarose gel electrophoresis (0.7% Agarose Gel, 40 V, 35 mA, and 180 min). The buffer used for casting and running the gels was 0.5x TBE, 12 mM MgCl₂. The band of interest was extracted visually using a razor blade. The extracted bands were chopped into sub millimeter sized pieces. The pieces were then loaded in a Freeze N' Squeeze microcentrifuge filter and spun at 4800 rcf at 4 °C for 10 min for recovery.

4x DNA NANOSTRUCTURE FUNCTIONALIZATION: The MgCl₂ concentration of the 4x nanostructure solution was

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adjusted to 12 mM by adding 0.5x TBE buffer prior to mixing the AuNPs. This prevents the AuNPs from precipitating out of the solution prematurely. The AuNPs and 4x nanostructures were then mixed in a 1:5 binding site to AuNP ratio. The mixture was heated to 45 °C for 41 min then cooled to 20 °C at 0.6 °C/min.

The functionalized 4x nanostructures were purified using agarose gel electrophoresis (1.4% Agarose Gel, 70 V, 66 mA, and 180 min). The buffer used for casting and running the gel was 0.5x TBE, 12 mM MgCl₂. The bands were extracted visually using a razor blade. The extracted bands were then chopped in sub-millimeter pieces that were loaded into Freeze N' Squeeze microcentrifuge filter and spun at 4800 rcf at 4 °C for 10 min for recovery.

S7 – HETEROSTRUCTURE FUNCTIONALIZATION

The magnesium concentration of the heterostructure solution was adjusted to 12 mM by adding 0.5x TBE buffer before mixing it with the AuNPs and QDs. The nanoparticles and heterostructures were mixed in a 1:5 binding site to nanoparticle ratio. The mixture was heated to 45 °C for 41 min then cooled to 20 °C at 0.6 °C/min for AuNP and QD attachment.

The functionalized heterostructures were then purified using agarose gel electrophoresis (0.7% Agarose Gel, 60 V, 65 mA, and 90 min). The buffer used for casting and running the gels was 0.5x TBE 12 mM MgCl₂. The bands were extracted visually using a razor blade. Extracted bands were chopped into sub-millimeter pieces which were loaded into a Freeze N' Squeeze microcentrifuge filter and spun at 4800 rcf at 4 °C for 10 min for recovery.

S8 – AFM AND TEM SPECIMEN PREPARATION

AFM SPECIMEN PREPARATION: Substrates for AFM were prepared by adhering a sheet of mica (V4 grade, SPI Supplies) onto an AFM puck (TED PELLA) using superglue. Five micro-liters of the sample solution was deposited onto the mica substrate and incubated for 2 min. Then 20 μ L of 1x TAE, 12 mM nickel-acetate was dispensed onto the mica surface for 5 min. The mica surface was rinsed with 1 mL of NANOpure water (Thermo Scientific) three times and dried with high purity nitrogen gas.

TEM SPECIMEN PREPARATION: TEM grids with ultrathin carbon films (TED PELLA) were plasma treated using an in-house glow discharge unit for 5 sec to induce hydrophilicity of the carbon surface. Three micro-liters of the sample solution was dispensed onto the grid for 5 min and then the excess solution was extracted using a piece of filter paper. Ten micro-litters of 2% uranyl acetate staining solution was dispensed onto the grids and removed immediately to pre-wet the surface, and another 10 μ L of the staining solution was deposited for 12 sec to stain the samples. The 2% uranyl acetate staining solution was prepared by mixing 15 mg of uranyl acetate dihydrate (bis(aceto-O)dioxouranium) (SPI Supplies) with 1.1 mL of 95 °C NANOpure water. The solution was agitated until all of the uranyl acetate was dissolved, and roughly 1.1 mL of 5 M NaOH solution was added to the uranyl acetate solution.

S9 – NANOPARTICLE ATTACHMENT COUNTING

The nanoparticle attachment counting for the AuNP arrays, except for $2 \times 14_{\alpha}$ were done based on the AFM images, and the nanoparticle counting for $2 \times 14_{\alpha}$ and the three heterostructures were done based on the TEM images. AFM images were acquired using a Bruker MultiMode 8 with a Nanoscope V controller. Nanoscope Analysis software from Bruker was utilized to flatten the images, as well as change the color settings and the height scale to enhance the contrast between the nanostructures and their background. A visual grid was placed on top of the images using Adobe Illustrator to prevent counting the same nanostructure multiple times. Images with the grid were opened in Adobe Reader and the number of AuNPs attached to individual nanostructures was manually counted. For statistical purposes, more than 100 nanostructures were analyzed for each design. The histograms for each design shown in Figure 4 are numerically expressed in Table S3. The analysis was performed objectively by rejecting nanostructures that satisfy any of the following criterion:

- A nanostructure partially overlapping with another nanostructure(s)
- A malformed nanostructure
- A nanostructure with indistinguishable AuNP(s) under AFM analysis
- A nanostructure sharing AuNP(s) with other nanostructure(s)
- A nanostructure extended beyond the image boundary
- A nanostructure with binding sites(s) that have more attached AuNP(s) than available tether(s)

In addition to the criteria above, the criterion below was added to reject inappropriate nanostructures when calculating the average probability of AuNP site-occupation, p_{exp} .

- A nanostructure having more AuNP(s) attached than available binding site(s) on the nanostructure

Below is a representative AFM image for the $4 \times 14_{\alpha}$ design (Figure S6). The nanostructures pointed to with a red arrow were rejected, the nanostructure pointed to with a blue arrow was included in the histogram, shown in Figure 4, but was rejected for calculating the average probability of AuNP site-occupation, p_{exp} , and the nanostructures pointed to with a green arrow were valid for analysis.

Hundreds of TEM micrographs of $2 \times 14_{\alpha}$ and the three heterostructures were acquired using a JEOL JEM-2100 HR analytical transmission electron microscope with the beam voltage of 120 kV. Instead of taking large low-magnification images containing many nanostructures, high-magnification images of individual nanostructures were taken to carefully inspect individual nanostructures for counting. In addition to the nanostructures satisfying the above criterion for the counting using AFM, the structures with insufficient staining were eliminated from counting for maintaining the quality of the analysis.

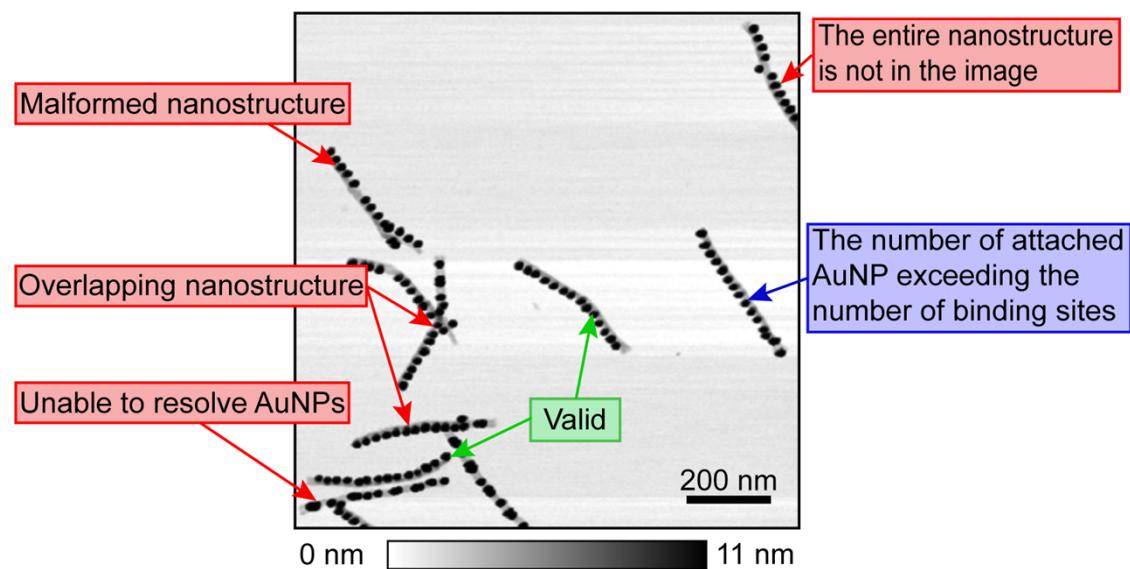


Figure S6: Representative AFM image for the $4 \times 14_{\alpha}$ design. The nanostructures pointed to with a red arrow were rejected, the nanostructure pointed to with a blue arrow was included in the histogram, shown in Figure 4, but was rejected for calculating the average probability of AuNP site-occupation, p_{exp} , and the nanostructures pointed to with a green arrow were valid for both analyses.

Table S3: Numerical Values of the Histogram Shown in Figure 4

Design	1x9_a	1x9_b	2x9_a	2x9_b	4x9_a	4x9_b	1x14_a	2x14_a	4x14_a	2x18_{a/b}	4x18_{a/b}
Number of Structures (N)	160	101	104	116	137	134	108	154	133	306	121
Number of Attached AuNPs	0	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
	1	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00
	2	0.01	0.04	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
	3	0.07	0.07	0.02	0.01	0.00	0.00	0.00	0.00	0.00	0.00
	4	0.17	0.24	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
	5	0.31	0.25	0.06	0.01	0.00	0.00	0.04	0.00	0.00	0.00
	6	0.23	0.28	0.13	0.10	0.01	0.01	0.17	0.01	0.00	0.00
	7	0.13	0.10	0.23	0.21	0.00	0.00	0.18	0.03	0.00	0.00
	8	0.07	0.02	0.32	0.41	0.08	0.04	0.19	0.02	0.00	0.00
	9	0.01	0.01	0.23	0.25	0.70	0.85	0.19	0.12	0.00	0.00
	10				0.00	0.01	0.20	0.10	0.09	0.17	0.00
	11				0.00	0.00	0.01	0.00	0.11	0.19	0.02
	12							0.03	0.26	0.05	0.00
	13							0.01	0.11	0.27	0.02
	14							0.00	0.08	0.54	0.04
	15									0.11	0.14
	16									0.02	0.28
	17										0.31
	18										0.17
	19										0.03
	20										0.01

S10 – NEAREST NEIGHBOR AuNP SEPARATION MEASUREMENT

The nearest neighbor AuNP separation distance was measured via AFM images acquired for the AuNP attachment counting in S4. Images were acquired using a Bruker MultiMode 8 with a Nanoscope V controller. All measurements were done using Windows Scanning X Microscope (WSxM™) from Nanotec.¹ The zoomed-in images of individual nanostructures were acquired with a multiple dynamic zoom tool. Profile lines were carefully drawn along the axis of the nanostructure using the profile tool. The process also generated the height profile window. The profile line was slid on top of the AuNP array to acquire the height profile of the AuNP array (Figure S7a). In the height profile window, the nearest neighbor AuNP separation was measured by moving the sliders to the center of the two adjacent AuNPs (Figure S7b). The center position of the AuNPs were determined based on the AFM image.

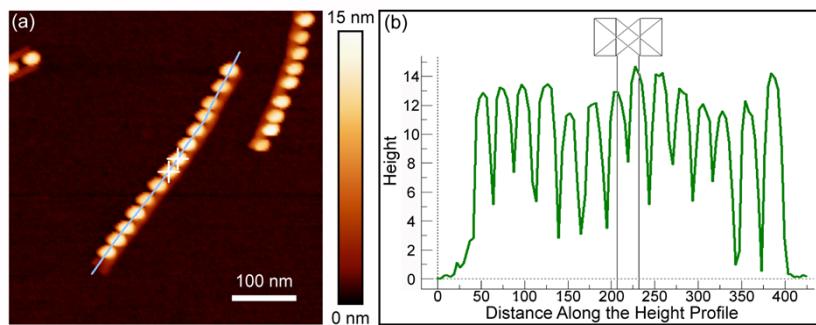


Figure S7: (a) AFM micrograph of $4 \times 14_{\alpha}$ design with a profile line along the axis of the nanostructure. (b) Height profile along the profile line. Both the micrograph and the height profile were generated using WSxM. The peaks of the height profile often did not match with the exact center of the AuNP.

The 4x nanostructures laid on the mica substrate in two orientations: upright and sideways. The upright orientation refers to AuNPs on top of the nanostructure, whereas the sideways orientation refers to AuNPs on the side of the nanostructure (Figure S8). The ratio of the upright oriented nanostructures decreased between 9-site and 14-site designs even though it increased between the 14-site and 18-site designs; 85%, 48%, and 52% for $4 \times 9_{\alpha}$, $4 \times 14_{\alpha}$, and $4 \times 18_{\alpha/\beta}$, respectively. For the $4 \times 9_{\alpha}$ design, there was not a noticeable deviation in the AuNP separation between the two orientations; however, a large deviation was observed for the $4 \times 14_{\alpha}$. When the nanostructures adsorbed onto the mica surface in the upright orientation, the AuNPs formed dimers. Dimer formation is attributed to the capillary force during drying. In comparison, sideways oriented AuNPs on the $4 \times 14_{\alpha}$ design were equally distributed. Unlike the upright oriented samples, the AuNPs on the sideways oriented nanostructures were in contact with the mica. The charge attraction and secondary forces between the AuNPs and the mica surface likely overcame the capillary force between the water and the AuNPs. As a consequence, AuNP separation measurements were taken from sideways oriented

nanostructures for the $4 \times 14_{\alpha}$ design.

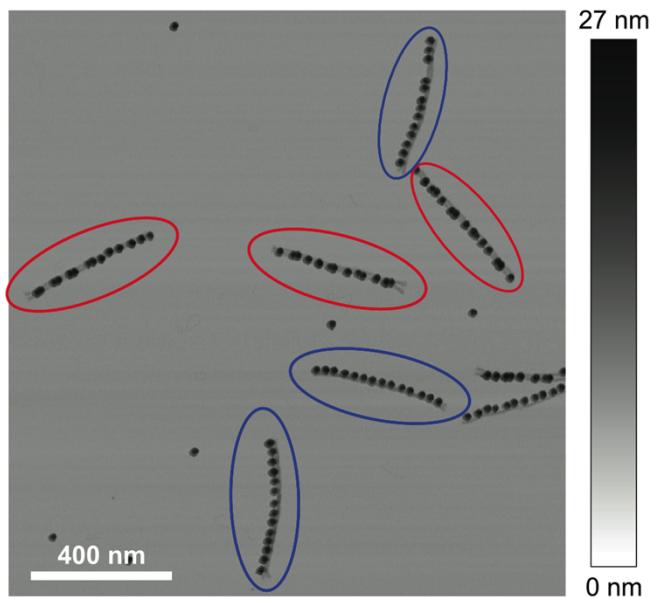


Figure S8: AFM micrograph of a $4 \times 14_{\alpha}$ design showing that the nanostructures lie on a mica surface in two orientations: upright and sideways. The upright oriented nanostructures are highlighted in red ovals and the sideways oriented nanostructures are highlighted in blue ovals. While the AuNPs attached on sideways oriented nanostructures had equal AuNP separation, the AuNPs attached to the upright oriented nanostructures formed dimers. Dimer formation was anticipated to be caused by capillary action when drying the sample.

S11 – TRINOMIAL DISTRIBUTION EQUATION

The distribution of nanoparticles on arrays having events where more than one nanoparticle binds to one site may be analyzed using a trinomial distribution. If the probability of binding two or more nanoparticles per site is low, then a good approximation is acquired by assuming that no more than two nanoparticles bind per site. Analogous to the binomial distribution, a trinomial distribution, $P(m)$, for the probability that m nanoparticles bind to an array with n binding sites can be written in terms of *Prob(0 Bound)*, *Prob(1 Bound)*, and *Prob(2 Bound)* – the site-binding probabilities for zero, one, or two nanoparticles, respectively. All possible ways to attach m particles to n sites are summed over the possible numbers of double-occupancy sites, indexed by j , where for a given j , the variables u_j , v_j , and w_j are the number of zero-, single-, and double-occupancy sites respectively, yielding the trinomial distribution:

$$P_{tri}(m) = \sum_{j=\max(0,m-n)}^{\lfloor m/2 \rfloor} \frac{n!}{u_j!v_j!w_j!} Prob(0\ Bound)^{u_j} Prob(1\ Bound)^{v_j} Prob(2\ Bound)^{w_j} \quad (S5)$$

The trinomial distribution may be reduced to a single unknown parameter by conditioning the probability that one or two particles are bound on the probability that a site is occupied, $Prob(\text{Occupied}) = p_{fit}$, a value which is estimated by fitting to the data. The unknown conditional probability, $Prob(1\ Bound|\text{Occupied}) = c$, is a parameter that can also be estimated by fitting to the data. Combining the law of total probability and the assumption that a maximum of two particles bind per site yields the following:

$$Prob(0\ Bound) = 1 - p_{fit} \quad (S6)$$

$$Prob(1\ Bound) = Prob(1\ Bound | \text{Occupied}) Prob(\text{Occupied}) = cp_{fit} \quad (S7)$$

$$Prob(2\ Bound) = Prob(2\ Bound | \text{Occupied}) Prob(\text{Occupied}) = (1 - c)p_{fit} \quad (S8)$$

Substituting the above equations for the site-binding probabilities and writing u_j , v_j , and w_j in terms of j , n , and t yields the following form of the trinomial distribution:

$$P_{tri}(m) = \sum_{j=\max(0,m-n)}^{\lfloor m/2 \rfloor} \frac{n!}{(n-m+j)!(m-2j)!} (1-p_{fit})^{(n-m+j)} (cp_{fit})^{(m-2j)} ((1-c)p_{fit})^j \quad (S9)$$

The data was fit to this equation to estimate the parameter c , yielding an estimate of how often single occupancy of binding sites can be expected and p_{fit} , the fitted average probability of site-occupation. To validate the fit parameters c and p_{fit} , we compared the expected value of the average site-occupancy based on these parameters, y_{th} :

$$y_{th} = 1 \cdot c \cdot p_{fit} + 2 \cdot (1 - c)p_{fit} \quad (S10)$$

to the measured value of average site-occupancy over all nanostructures, y_{exp} , including those with more nanoparticles than binding sites (Table S4).

Table S4: Comparison of calculated and observed average site-occupancy

Support Information

	2x9_a	2x 9_B	4x 9_a	4x 9_B	2x 14_a	4x 14_a	2x 18_{a/B}	4x 18_{a/B}
p_{fit}	0.83	0.87	0.99	1.00	0.77	0.96	0.91	0.99
c	0.98	1.00	0.97	0.99	0.96	0.99	0.99	0.99
y_{th}	0.85	0.86	1.02	1.01	0.80	0.98	0.91	0.99
y_{exp}	0.83	0.86	1.01	1.01	0.79	0.98	0.91	1.00

S12 – EXPERIMENTAL BINDING SITE PERIODICITY DETERMINATION

Assuming that the difference between the experimental and the theoretical binding site periodicity is equal to the difference between the experimental and the theoretical nanostructure length, the experimental binding site periodicity was calculated by:

$$d_{\text{exp}} = \left(\frac{l_{\text{exp}}}{l_{\text{th}}} \right) \times d_{\text{th}} \quad (\text{S1})$$

where d_{exp} is the experimental binding site periodicity, d_{th} is the theoretical binding site periodicity, l_{exp} is the experimental nanostructure length, and l_{th} is the theoretical nanostructure length. The theoretical binding site periodicity and the nanostructure length were calculated by multiplying the number of base-pairs with the unit length of the base-pair (0.34 nm). The experimental nanostructure length was determined by measuring the end-to-end nanostructure length under AFM using WSxM. The zoomed-in images of individual nanostructures were then acquired via a multiple dynamic zoom tool. The profile line was carefully drawn along the axis of the nanostructure using the profile tool (Figure S9a). In the height profile window, the nanostructure length was then measured by moving the sliders to the ends of the nanostructure (Figure S9b). To account for the width of the probe, the ends of the nanostructure were estimated at the mid points of the slopes at the ends of the nanostructure. Twenty experimental nanostructure lengths were acquired from each design.

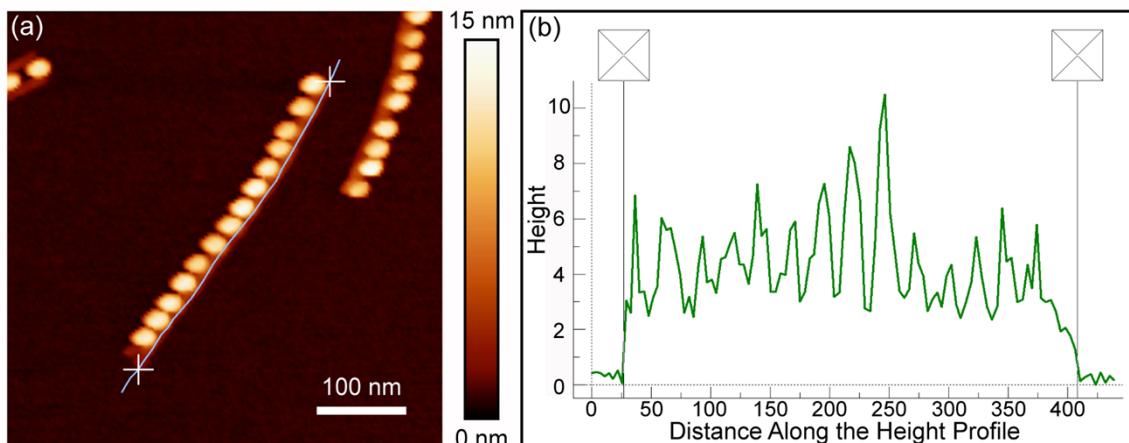


Figure S9: (a) AFM image of 4x14_α with a height profile line. (b) Height profile of the nanostructure. To account for the width of the AFM probe, the ends of the nanostructure were estimated to be at the midpoints of the slopes at the ends of the nanostructure.

S13 – MODIFIED GEOMETRIC DISTRIBUTION EQUATION

In a previous study, steric hinderance and site-bridging were examined by comparing the nearest neighbor QD separation histogram with the geometric distribution plot.² In the absence of steric hinderance or site-bridging, the histogram expected to follow a geometric distribution, $P(k)$, given by:

$$P(k) = p_{exp}(1 - p_{exp})^{(k-1)} \quad (S2)$$

where p_{exp} is the average probability of AuNP site-occupation, and k is the integer number of periods between nearest neighbors. The standard deviation of the AuNP separation was determined by decreasing the bin size of the histogram from 1 to 0.25. When considering a separation between two adjacent AuNPs, the separation, x , should follow the normal distribution because of the random motion of the AuNPs. The normal distribution, $f(x)$, is given by:

$$f(x) = \frac{1}{\sigma\sqrt{2\pi}} \exp\left[-\frac{(x-\mu)^2}{2\sigma^2}\right] \quad (S3)$$

where μ is the mean nearest neighbor separation and σ is the standard deviation of the separation. Therefore, with a smaller bin size, the resulting nearest neighbor separation histogram is expected to follow the distribution that combined both the geometric and normal distributions. In this paper, this is referred to as a “modified geometric distribution”, $P(x)$, and is given by:

$$P_{geo}(x) = \sum_{k=1}^{\infty} \frac{1}{\sigma\sqrt{2\pi}} \exp\left[-\frac{(x-k)^2}{2\sigma^2}\right] p_{exp}(1 - p_{exp})^{(k-1)} \quad (S4)$$

The mean separation, μ , in the Eqn. 3 has become the integer number of periods between nearest neighbors, k , because the x-axis of the histogram was normalized by the experimental binding site periodicity. Likewise, the separation and standard deviations are fractional.

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