Inhibition of microvesiculation sensitizes prostate cancer cells to chemotherapy and reduces docetaxel dose required to limit tumor growth *in vivo*

Samireh Jorfi^a, Ephraim A. Ansa-Addo^{a,b}, Sharad Kholia^{a,}, Dan Stratton^a, Shaunelle Valley^a, Sigrun Lange^{d,*}, Jameel Inal^{a,*}

^aCellular and Molecular Immunology Research Centre, School of Human Sciences, London Metropolitan University, U.K.

^bCurrent Address: Department of Immunobiology and Cancer Immunology, Hollings Cancer Center, Medical University of South Carolina, Charleston, South Carolina 29425, USA.

^dUniversity College London School of Pharmacy, 29-39 Brunswick Square, London WC1N 1AX, U.K.

*co-corresponding authors: Tel: +44 20 7133 2122 or +44 20 7753 5895 *E-mail address*: j.inal@londonmet.ac.uk or sigrun.lange@ucl.ac.uk

Supplementary Figures

Supplementary Fig. 1 Calpain silencing in PC3 reduces CAPNS1 expression. Semiconfluent PC3 cells were transfected (or not) with 50 nM of control (Neg Cont) siRNA or various *CAPNS1* siRNA sequences targeting different regions for 48 h. Flow cytometry analysis of CAPNS1 knockdown in siRNA-treated cells showed that siRNA-#6 had the most significant reduction of CAPNS1 expression. To confirm siRNA-#6 for silencing calpain, PC3 cells were transfected with siRNA-#6 (50 nM) for 48 h. Relative CAPNS1 expression assessed by flow cytometry showed a 42% reduction compared to negative control siRNA. Data presented is the mean \pm S.E.M. of three separate experiments performed in triplicate. ****P* < 0.001; ***P* < 0.01; **P* < 0.05 were considered statistically significant.

Supplementary Fig. 2 Representative HPLC chromatographs of MVs from untreated, control PC3 cells (A), cells treated with 100 μ M MTX and of MTX standards, 2-100 μ M (C).

Supplementary Fig. 3 Control HPLC chromatographs showing solvent peaks (A) and consistency of wavelength scan with MTX spectra from the literature, showing UV detection peak at 303 nm, (B).

Supplementary Fig. 4 Calpeptin reduces MV release from DTX-treated human prostatic cancer cells, PC3. Cells treated with 10 nM DTX, in the presence of 20 μ M CP, released significantly fewer MVs. Released MVs were isolated, and identified as MVs from their typical scatter plot, high phosphotidylserine exposition (95% AnnexinV-positive), size estimation by nanosight (250 nm diameter) and from their characteristic morphology and size (~250 nm diameter) by transmission electron microscopy.

Supplementary Fig. 5 Prostate cancer cell lines LnCaP and Du-145, pretreated with calpeptin are sensitized to DTX treatment. Cells pretreated with CP (45 min; 20 μ M) were treated with a range of DTX concentrations from 0.1 to 100 nM and after 48h were found to show higher levels of apoptosis where microvesiculation was inhibited.

Supplementary Fig. 6 Plasma MV levels at euthanasia and mouse body weights over the study. Plasma MV levels at euthanasia in the subcutaneous PC3 xenograft mouse model are diminished in animals treated with DTX and CP (both at 10 mg/kg), i.p., compared to animals given DTX alone. Mouse body weights throughout the experiment to eliminate toxicity effects due to treatment (B). Additional representative excised xenografted PC3 tumors (C) showing vehicle treated mice versus CP (10 mg/kg) and DTX + CP (both at 10 mg/kg); Bar, 4mm.

Supplementary Figures

Figure S1



siRNA#1, 5' -CAC CTG AAT GAG CAT CTC TAT -3'; siRNA#3, 5' -AAG GTG GCA GGC CAT ATA CAA -3'; siRNA#5, 5' -CAG CGC CAC AGA ACT CAT GAA -3'; siRNA#6, 5' -TCC GAC GCT ACT CAG ATG AAA -3'. Negative control siRNA, 5' -AAT TCT CCG AAC GTG TCA CGT -3'







Figure S4









Figure S6