Table 4A: Bacterial strains		
Name	Genotype	Construction or reference
Caulobacter cres	centus	
NA1000	Synchronizable derivative of the wild-type strain CB15	[1]
CMS2	NA1000 carrying a kan <sup>R</sup> resistance marker 239808–240826 (CC0222–0224)	[2]
CMS9	NA1000 carrying a kan <sup>R</sup> resistance marker 939587–940578 (CC0843–0844)	[2]
CMS21	NA1000 carrying a kan <sup>R</sup> resistance marker 2049425–2050600 (CC1853–1855)	[2]
CMS28	NA1000 carrying a kan <sup>R</sup> resistance marker 2742248–2743137 (CC2537–2538)	[2]
CMS33	NA1000 carrying a kan <sup>R</sup> resistance marker 3256387–3257298 (CC3039–3040)	[2]
JC1149, JC1150, JC1151	NA1000 Δ <i>ccrM</i> ::Ω	[3] This work.
JC1299	Evolved JC1149 clone, population 1.1, 300 generations	This work.
JC1300	Evolved JC1149 clone, population 1.2, 300 generations	This work.
JC1301	Evolved JC1149 clone, population 1.3, 300 generations	This work.
JC1302	Evolved JC1149 clone, population 1.4, 300 generations	This work.
JC1303	Evolved JC1150 clone, population 2.1, 300 generations	This work.
JC1304	Evolved JC1150 clone, population 2.2, 300 generations	This work.
JC1305	Evolved JC1150 clone, population 2.3, 300 generations	This work.
JC1306	Evolved JC1150 clone, population 2.4, 300 generations	This work.
JC1307	Evolved JC1151 clone, population 3.1, 300 generations	This work.
JC1308	Evolved JC1151 clone, population 3.2, 300 generations	This work.
JC1309	Evolved JC1151 clone, population 3.3, 300 generations	This work.

JC1310	Evolved JC1151 clone, population 3.4, 300 generations	This work.
JC1311	Evolved JC1149 clone, population 1.2, 50 generations	This work.
JC1312	Evolved JC1151 clone, population 3.1, 50 generations	This work.
JC1320	NA1000 carrying the kan <sup>R</sup> marker of CMS2	This work.
JC1322	NA1000 carrying the kan <sup>R</sup> marker of CMS28	This work.
JC1329	NA1000 carrying the kan <sup>R</sup> marker of CMS9.	This work.
JC1323	NA1000 carrying the kan <sup>R</sup> marker of CMS2 and the SNP found in <i>hprK</i> in JC1302	This work.
JC1336	NA1000 carrying the kan <sup>R</sup> marker of CMS9 and the SNP found in <i>ptsP</i> in JC1300	This work.
JC1327	NA1000 carrying the kan <sup>R</sup> marker of CMS21 and the SNP found in CCNA_01943 in JC1302	This work.
JC1325	NA1000 carrying the kan <sup>R</sup> marker of CMS28 and the SNP found in the <i>ftsZ</i> promoter region in JC1299	This work.
JC1328	NA1000 carrying the kan <sup>R</sup> marker of CMS28 and the SNP found in the <i>ftsZ</i> promoter region in JC1301	This work.
JC1338	NA1000 carrying the kan <sup>R</sup> marker of CMS28 and the SNP found in the <i>ftsZ</i> promoter region in JC1306	This work.
JC1343	NA1000 carrying the kan <sup>R</sup> marker of CMS33 and the SNP found in <i>ctrA</i> in JC1300	This work.
JC1347	JC1322 Δ <i>ccrM</i> ::Ω	This work.
JC1339	JC1323 Δ <i>ccrM</i> ::Ω	This work.
JC1340	JC1336 Δ <i>ccrM</i> ::Ω	This work.
JC1344	JC1327 Δ <i>ccrM</i> ::Ω	This work.
JC1345	JC1325 Δ <i>ccrM</i> ::Ω	This work.
JC1341	JC1328 Δ <i>ccrM</i> ::Ω	This work.
JC1342	JC1338 Δ <i>ccrM</i> ::Ω	This work.
JC1346	JC1343 Δ <i>ccrM</i> ::Ω	This work.
JC1353	JC1320 placZ290-PftsZ-WT	This work.

JC1354	JC1323 placZ290-PftsZ-WT	This work.
JC1369	JC1336 placZ290-PftsZ-WT	This work.
JC1368	JC1327 placZ290-PftsZ-WT	This work.
JC1370	JC1343 placZ290-PftsZ-WT	This work.
JC1371	JC1353 Δ <i>ccrM</i> ::Ω	This work.
JC1372	JC1354 $\Delta ccrM::\Omega$	This work.
JC1386	JC1369 Δ <i>ccrM</i> ::Ω	This work.
JC1387	JC1368 Δ <i>ccrM</i> ::Ω	This work.
JC1388	JC1370 Δ <i>ccrM</i> ::Ω	This work.
JC1383	NA1000 placZ290-PftsZ-SNP 1.1	This work.
JC1384	NA1000 placZ290-PftsZ-SNP 1.3	This work.
JC1385	NA1000 placZ290-PftsZ-SNP 2.4	This work.
JC1378	JC1320 placZ290-mipZP	This work.
JC1379	JC1323 placZ290-mipZP	This work.
JC1381	JC1336 placZ290-mipZP	This work.
JC1398	JC1329 placZ290-PftsZ-AT	This work.
JC1401	JC1329 placZ290-PftsZ-ctrA	This work.
JC1399	JC1329 placZ290-PftsZ-dnaA	This work.
JC1402	JC1336 placZ290-PftsZ-AT	This work.
JC1405	JC1336 placZ290-PftsZ-ctrA	This work.
JC1403	JC1336 placZ290-PftsZ-dnaA	This work.

Table 4B : Plasmids		
Name	Description	Reference
p <i>lacZ</i> 290	Low copy number vector, usually used to create <i>lacZ</i> transcriptional fusion.	[4]
p <i>lacZ</i> 290- P <i>ftsZ</i> -WT	p <i>lacZ290</i> containing the wild-type <i>ftsZ</i> promoter region.	[3]
placZ290- mipZP	p <i>lacZ290</i> containing the wild-type <i>mipZ</i> promoter region.	[5]
p <i>lacZ</i> 290- P <i>ftsZ</i> -AT	p <i>lacZ290</i> containing the <i>ftsZ</i> promoter region, mutated at a conserved GANTC (A becomes T).	[3].
placZ290- PftsZ-SNP 1.1	p <i>lacZ290</i> containing the <i>ftsZ</i> promoter region carrying the SNP present in strain JC1299.	This work.
placZ290- PftsZ-SNP 1.3	p <i>lacZ290</i> containing the <i>ftsZ</i> promoter region carrying the SNP present in strain	This work.

	JC1301.	
p <i>lacZ</i> 290- P <i>ftsZ</i> -SNP 2.4	p <i>lacZ290</i> containing the <i>ftsZ</i> promoter region carrying the SNP present in strain JC1306.	This work.
placZ290- PftsZ-ctrA	p <i>lacZ290</i> containing the <i>ftsZ</i> promoter region carrying mutations in the CtrA binding sites.	This work.
placZ290- PftsZ-dnaA	p <i>lacZ290</i> containing the <i>ftsZ</i> promoter region carrying mutations in the DnaA binding sites.	This work.

Table 5C : Primers					
Name	Sequence				
HRM_hprK_F	ACG CCG AGA ACC GCA AGG AC				
HRM_hprK_R	CCC GCA GCG AAA GTC ACG AC				
HRM_CCNA_01943_F	CCG CCG GTC GAG GTG GAG				
HRM_CCNA_01943_R	TTT TCG ACG CTG GGC AGG TG				
HRM_PftsZ_F	GCG GCG CGG AAA GAG AAA TAG				
HRM_PftsZ_R	CGA CCG GAC CAC GAC TCA GAT TA				
Seq_ptsP_F	GCA AGC TGG ACG TGG TGG TG				
Seq_ptsP_R	GAG GCC GGC ATT CAT CAA CAG				
Seq_hprK_F	CGA ACG GGG CGG TGG TC				
Seq_hprK_R	CGC GGG CGG TCT TGA TAC				
Seq_ctrA_F	CCT TTT GGG TGG GCG TTT CTG				
Seq_ctrA_R	CGC CTG CGA CCA ATG TGT TCA				
ftsZP_DnaA_mut_R	GTG GTC CGG TCG AGC GGG ACC GTA CGT TAA ATT GTG GTC TAA CAA GTG AAT CCC CCA ATG AAC CAA AGG				
ftsZP_DnaA_mut_F	TCC CGC TCG ACC GGA CCA CGA C				
ftsZP_CtrA_mut_F	CGG TCC CGC TCG ACC GGA CCA CGA CTC AGA TTA TGC AGA ATT CCG CCG ATT AAC GAT GG				
ftsZP_CtrA_mut_R	TGC ATA ATC TGA GTC GTG GTC CGG TCG				

- 1. Evinger, M. and N. Agabian, *Envelope-associated nucleoid from Caulobacter crescentus stalked and swarmer cells.* J Bacteriol, 1977. **132**(1): p. 294-301.
- West, L., D. Yang, and C. Stephens, Use of the Caulobacter crescentus genome sequence to develop a method for systematic genetic mapping. J Bacteriol, 2002. 184(8): p. 2155-66.
- 3. Gonzalez, D. and J. Collier, *DNA methylation by CcrM activates the transcription of two genes required for the division of Caulobacter crescentus.* Mol Microbiol, 2013. **88**(1): p. 203-18.
- 4. Gober, J.W. and L. Shapiro, *A developmentally regulated Caulobacter flagellar promoter is activated by 3' enhancer and IHF binding elements*. Mol Biol Cell, 1992. **3**(8): p. 913-26.
- 5. Fernandez-Fernandez, C., D. Gonzalez, and J. Collier, *Regulation of the Activity of the Dual-Function DnaA Protein in Caulobacter crescentus.* PLoS One, 2011. **6**(10): p. e26028.